# **REFERANCES**

- Adams NR. 1995. Detection of the effects of phytoestrogens on sheep and cattle. **J. Anim Sci.** 73: 1509-1515.
- Alison M. Duncan, William R. Phipps, Mindy S. Kurzer. 2003. Phytoestrogens. Best practice & research clinical endocrinology and metabolism. 17(2): 253-271.
- Alonso LC and Rosenfield RL. 2002. Oestrogen and puberty. **Best practice & research**. 16(1): 13-30.
- Anusansunthorn. 1931. KWAO KRUA. Cheangmai: Aupapong printing.
- Benassayag C, Perrot-Applant M, Ferre F. 2002. Phytoestrogens as modulators of steroid action in target cells. **Jurnal of Chromatography B.** 777: 233-248.
- Britt KL and Findlay JK. 2002. Estrogen actions in the ovary revisited. **J Endocrinol**. 175: 269-276.
- Burton JL and Wells M. 2002. The effect of phytoestrogens on the female genital tact. J. Clin Pathol. 55: 401-407.
- Chansakaow S, Sekine TIK, Okada M, Higuchi Y, Kudo M and Chaichantipyuth C.
  2000. Identification of Deoxymiroestrol as the Actual Rejuvenating Principle of "
  Kwao Keur ", Pueraria mirifica. The known miroestrol may be an artifact. Journal of Natural products. 63: 173-175.
- Chansakaow S, Sekine TIK, Okada M, Higuchi Y, Kudo M and Chaichantipyuth C. 2000. Isoflavanoids from *pueraria minifica* and their Estroginic Activity. **Planta**Med. 66: 572-575.
- Chawalit N. 1995. Kwao Krua. Bangkok. Pean Company.

- Cherdshewasart W. 2003. Toxicity tests of a phytoestrogen-rich herb; *Pueraria mirifica*. **J. Sci. Res. Chula. Univ.** 28(1): 1-12.
- Chivapat S, chavalittumrong P, Rattanajarasroj S, Chuthaputti A, Panyamang S. 2000.

  Toxicity study of; *Pueraria mirifica* Airy Shaw et Suvatabandhu. **J. medical Sciences**. 42(3): 202-223.
- Clermont Y.1972 Kinetics of spermatogenesis in mammals: seminiferous epithelium cycle and spermatogonial renewal. **Physiological Reviews.** 52 (1): 198-236.
- De Jong FH, Uilenbroek TJ, Van der Moen HJ. 1975. Estradiol-17β, testosterone and gonadotropins in estradiol-17β-treated intact adult male rats. **J Endocrinol**. 65:281-282.
- De Jong FH, Uilenbroek TJ, Van der Molen HJ. 1975. Oestradiol-17β, testosterone and gonadotrophins in oestrodiol-17β-treated intact adult male rats. J Endocrinol. 65: 281-282.
- Duncan AM, Phipps WR, Kurzer MS. 2003. Phyto-oestrogens. Best Practice & Research Clinical Endocrinology and Metabolism. 17(2): 253-271.
- Dyson ALMB, Orgebin-Crist MC. 1973. Effects of hypophysectomy, castration and androgen replacement upon the fertilizing ability of rat epididymal spermatozoa. **Endocrinology.** 93: 391-402.
- Fielden MR, Samy SM, Chou KC, Zacharewski TR. 2003. Effect of human dietary exposure levels of genistein during gestation and lactation on long-term reproductive development and sperm quality in mice. Food Chem Toxicol. 41:447-454.
- Gad SC and Chenglis CP. 1992. Animal models in toxicology. New York: Marcle Dekker.

- Goyal, HO, Braden TD, Mansour M, Wiliams CS, Kamaleldin A and Srivastava KK.

  2001. Diethylstilbestrol Treated adult rat with altered Epididymal sperm numbers and sperm motility parameters, but without alterations in sperm production and sperm Morphology. **Biology of Reproduction**. 64: 972-934.
- Green EL. 1975. Biology of the Laboratory Mouse. USA: Dover Publications, Inc.
- Gutendrof B, and Westendorf J. 2001. Comparison of an array of in vitro assays for the assessment of the estrogenic potential of natural and synthetic estrogens, phytoestrogens and xenoestrogens. **Toxicology.** 166: 79-89.
- Hadley ME. 2000. Endocrinology (5th ed.). USA: Prentice-Hall, Inc.
- Hayes FJ, DeCruz S, Seminara SB, Boepple PA, Crowley Jr WF. 2001. Differential regulation of gonadotropin secretion by testosterone in the human male: absence of a negative feedback effect of testosterone on follicle-stimulating hormone secretion. J clin Endocrinol Metab. 86:53-58.
- Hayes FJ, Seminara SB, DeCruz S, Boepple PA, Crowley Jr WF. 2000. Aromatase inhibition in the human male reveals a hypothalamic site of esrogen feedback. J clin Endocrinol Metab. 85:3027-3035
- Hughes CL Jr, kalas RS, weisinger AS. 1991. Acute and subacute effects of naturally occurring estrogens on luteinizing hormone secretion in the ovariectomised rat: part 1. **Reprod Toxicol**. 5: 124-132.
- Humason GL. 1972. Animal tissue techniques (3<sup>rd</sup> ed.). USA: W.H. Freeman and Company.
- Johnson MH and Everitt BJ. 1995. Essential Reproduction (4<sup>th</sup> ed.). New York: Blackwell Science Ltd.

- Jones HEH and Pop GS.1960. A study of the action of miroestrol and other oestrogenens on the reproductive tract of the immature female mouse. J. Endocrine. 20: 229-235.

  Jonson M.H. and Everitt BJ. 1995. Essential Reproductive (4<sup>th</sup> ed.). New York: Blackwell Science Ltd.
- Kaneto M, Kanamoris S, Hishikawa A, Kishi K. 1999. Epididymal sperm motion as a parameter of male reproductive toxicity: sperm motion, fertility, and histopathology in ethiny estradiol-treated rats. Reprod toxicol. 13: 279-289.
- Knight, P. G. 1996. Roles of inhibins, activins, and follistatin in the female reproductive system. Front. **Neuroendocrinology**. 17:476-509.
- Korach KS, Metzler M, McLachlan JA. 1978. Estrogenic activity in vivo and in vitro of some diethylstilbestrol metabolites and analogs. **Proc Natl Acad Sci USA.** 75: 468-471.
- Kuiper GJM, Lemmen JG, Carlsson BO, Corton JC, Safe SH, Van Der Saag PT, Van Der Burg B, Gustafsson JA. 1998. Interaction of estrogenic chemicals and phytoestrogens with estrogen receptor β. Endocrinology. 139(10): 4252-4263.
- Kurzer MS. 2002. Hormonal effects of soy in premenopausal women and men. J Nutr. 132:570s-573s.
- Langkalichan Y and Smitasiri Y. 1984. Effect of White Cwow (*Pueraria mirifica*) on Reproduction in Male Albino Rat. Master's Thesis, Chiang Mai University.
- Macgeregor JI and Jordan VC. 1998. Basic guide to the mechanisms of antiestrogen action. **Phamacological reviews**. 50(2): 151-196.
- Makler A. 1978. A new multiple exposure photography method for objective human spermatozoal motility determination. **Fertil Steril.** 30, 192-199.

- Malaivijitnond S, Kiatthaipipat P, Cherdshewasart W, Watanabe G and Taya K. 2004. Sex differences in response to *Pueraria mirifica* phytoestrogens in rats. **Biolreprod.** Submit 6-2-2004.
- Massaad C, Enterrzami F, Massade L, Benahmed M, Olivennes F, Barouki R, Hamamah S. 2002. How can chemical compounds alter human fertility? **Obstetrics & Gynecology**. 100:127-137.
- Mitchell JH, Cawood E, Kinniburgh D, Provan A, Collins Ar, Irvine DS. 2001. Effect of a phytoestrogen food supplement on reproductive health in normal males. Clinical Science. 100: 613-618.
- Muangdet N and Anuntalabhochai S. 1986. Effects of low doses of white gwow (*Pueraria mirifica*) on female Japanese Quails. J. Sci. Fac. CMU. 12(1): 28-40.
- O'Donnell L, Robertson KM, Jones ME, and Simpson ER. 2001. Estrogen and spermatogenesis. Endocrine Reviews.22: 289-318.
- Panriansaen R. 2000. Characterization of *Pueraria mirifica* populations from various parts of Thailand. Master's Thesis, Chulalongkorn University.
- Panthong K, Chaiananwong S and Jirattikalkit A. 1987. Alternation of female reproductive system in dog by *Pueraria mirifica*. Master's Thesis, Chulalongkorn University.
- Phinilla L, Garnelo P, Gaytan F, Aguilar E. 1992. Hypothalamic-pituitary function in neonatally oestrogen-treated male rats. **J Endocrinol.** 134: 279-286.
  - Pinilla L, Garnelo P, Gaytan F, Aguilar E.1992. Hypothalamic-ptuitary function in neonatally estrogen-treated male rats. **J Endocrinol**. 134:279-286.

- Semler DE. The rat in Shayne, CG, and Chengelis, Cp, (ed). 1992. **Animal models** intoxicology. New York: Academic Press. Pp. 21-76.
- Smitasiri Y, Junyatum U, Songjitsawad A, Sripromma P, Trisrilp S and Anuntalahochai S. 1987. Postcoital antifertility effects of *Pueraria mirifica* in rat. **J. Sci. Fac. CMU.** 13(1): 19-28.
- Smitasiri Y, Pangjit S and Anuntalahochai S. 1989. Inhibitation in lactating rats with *Pueraria mirifica* compared with estrogen. **J. Sci. Fac. CMU.** 16: 7-11.
- Sufi SB, Donaldson A and Jeffcoate SL. 1986. Method Manual: World Health
  Organization Collaborating Center for Research and Reference Services in the
  Immunoassay of Hormones in Human Reproduction. London, U.K.
- Sujarit S, Pholphamool C. 1985. Enhancement of sperm transport through the rat epididymis after castration. J Reprod Fertil. 74: 497-502.
- Tena-Sempere M, Navarro J, Pinilla L, Gonzalez LC, Huhtaniemi I, Aguilar E. 2000. Neonatal exposure to estrogen differentially alters estrogen receptor α and β mRNA expression in testis during postnatal development. **J Endocrinol**, 165: 345-357.
- Thrupcharoen P. 1998. Use of white kwao krua in Thai medicine. Seminar of kwao krua. Department of Medical Sciences.
- Turner CD and Bagnara JT. 1971. General endocrinology (4th ed.). Paris. Masson & Cie.
- Whitten PL, Lewis C, Naftolin F. 1993. A phytoestrogen diet induces the premature anovulatory syndrome in lactationally exposed female rats. **Biol Reprod**. 49:1117-1121.

- Whitten PL, Lewis C, Noftolin F. 1993. A phytoestrogen diet induces the premature anovulatory syndrome in lactationally exposed female rats. **Biol Reprod**. 49: 1117-1121.
- Whitten PL, Naftolin F. 1992. Effects of a phytoestrogen diet on estrogen-dependent reproductive processes in immature female rats. **Steroids**. 57:56-61.
- World Health Organization. 1999. WHO laboration manual for the examination of human semen of sperm-cervical mucus interaction (4<sup>th</sup> ed.). Canbridge: Canbridge University Press.

Zhang S. 1999. An Atlas of Histology. Springer-Verlag New York, Inc.

ศูนย์วิทยทรัพยากร จุฬาลงกรณ์มหาวิทยาลัย **Appendix** 

ศูนย์วิทยทรัพยากร จุฬาลงกรณ์มหาวิทยาลัย

#### **APPENDIXES**

### Appendix I: chemical reagents

### 1.1 Radioimmunoassay kits

- 17β-estradiol standard

: WHO RIA Reagent Programme, Switzerland.

(Batch number 79/11)

- antiserum to 17β-estradiol

: Professor Kohen Fortune, Department of

(Cloned 2F9)

Biology Regulation, Weizmann Institute of

Science, Israel.

-  $17\beta$ -estradiol tracer (2,4,6,7- $^{3}$ H)

: Amersham International, PLC, England.

(Batch number 2 Nov 2001)

- testosterone standard

: WHO RIA Reagent Programme, Switzerland.

(Batch number K079810)

- antiserum to testosterone

: WHO RIA Reagent Programme, Switzerland.

(Batch number K888510)

- testosterone tracer  $(1,2,6,7-^3H)$ 

: Amersham International, PLC, England.

(Lot TRK 402)

- FSH standard

: National Institute of Diabetes and Digestive and

(Batch number NIDDK-rFSH-RP-2 Kidney Disease (NIDDK), Japan.

(AFP-4621B))

- antiserum to FSH (Batch number : NIDDK, Japan.

NIDDK-anti-rFSH-S-11

(AFPO 972881))

- FSH tracer (Batch number : NIDDK, Japan.

NIDDK-rFSH-I-5 (AFP-11454B))

- LH standard (Batch number : NIDDK, Japan.

NIDDK-rLH-RP-3

(AFP-7187B))

- antiserum to LH (Batch number : NIDDK, Japan.

NIDDK-anti-rLH-S-11)

- luteinizing hormone tracer : NIDDK, Japan.

(NIDDK-rLH-I-5 (AFP-11536B))

1.2 Hormones

- diestrilbestrol : Sigma Chemical Company, Merck, USA.

1.3 Others

- charcoal reagent : WHO RIA Reagent Programme, Switzerland.

- dextran reagent : WHO RIA Reagent Programme, Switzerland.

- gelatin : Difco laboratory, USA.

- diethyl ether  $(C_2H_5)_2O$  : E. Merck, Damstadt, Germany.

- natrium dihydrogen phosphate-

: E. Merck, Damstadt, Germany.

monohydrated (NaH<sub>2</sub>PO<sub>4</sub>. H<sub>2</sub>O)

- toluene P.a. (C<sub>7</sub>H<sub>8</sub>)

: E. Merck, Damstadt, Germany.

- ethanol (95%)

: E. Merck, Damstadt, Germany.

- ethanol (Absolute)

: E. Merck, Damstadt, Germany.

- methanol (CH<sub>3</sub>OH)

: E. Merck, Damstadt, Germany.

- art.3115-1-4-Dioxane  $(C_7H_8O_2)$ 

: E. Merck, Damstadt, Germany.

- art.2946 [(2,5-diphenyloxazol)-

: E. Merck, Damstadt, Germany.

phenyl-oxazolyl phenyl anhydrous)]

 $(C_{15}H_{11}NO)$ 

- formalin (40%)

: E. Merck, Damstadt, Germany.

- xylene  $(C_6H_4(CH_3)_2)$ 

: E. Merck, Damstadt, Germany.

- n-butyl alcohol (absolute)

: E. Merck, Damstadt, Germany.

- hematoxylin

: E. Merck, Damstadt, Germany.

- eosin

: E. Merck, Damstadt, Germany.

- glacial acetic acid

: E. Merck, Damstadt, Germany.

- glycerine

: E. Merck, Damstadt, Germany.

- POPOP

: Sigma Chemical Company, USA.

[1,4-bis(2-(5-phenyloxazol))]

Benzene, phenyl-oxazolyl-phenyl-

oxazolyl phenyl anhydrous

- thiomersal (merthiolate)

: Sigma Chemical Company, USA.

- sesame oil

: Sigma Chemical Company, USA.

- sodium hydroxide (NaOH)

: BDH Chemical Ltd. England.

- disodium hydrogen phosphate-

: BDH Chemical Ltd. England.

anhydrous (Na<sub>2</sub>HPO<sub>4</sub>)

- sodium chloride (NaCl)

: BDH Chemical Ltd. England.

- paraffin

- egg albumin



ศูนย์วิทยทรัพยากร จุฬาลงกรณ์มหาวิทยาลัย

#### Appendix II: equipments

- beta counter : Model 1218 Rack Beta LKB Wallac, Findland.

- dunoff incubator shaker : Model 3575-1, Lab-Line Instrument Inc., USA.

- dynac centrifuge : Clay Adams, Becton Dickinson and Company,

USA.

- ultrasonic cleanser : Right A Weight, WM, Benver, USA.

- magnetic stirrer bars S-18520 : Thermolyne Corporation Iowa, USA.

- micropipette size 100 ul : Nichiyo Model 5000 Japan.

200 ul : Nichiyo Model 8100 Japan.

1,000 ul : Eppendorff 3130 Germany

- vortex mixer: M-16715 : Thermolyne Corporation Iowa, USA.

- pH meter : Corning pH meter 240 Cat No. 476530,

Corning-EEI Scientific Instrument, England.

- refrigerated centrifuge : Model PR-J, International Equipment

Company, USA.

- machinery weight : Right A Weight, WM, Benver, USA.

- foam decanting rack : DPC, USA.

- machinery weight : Right A Weight, WM.

- microtome : Model 820 serial 66305: American optical,

Scientific Instrument Division, Buffalo,

New York, USA.

- microtome blade : S 35, USA.

- hot air oven

: Griffin Grundy.

- refrigerator

: J-elegance Mitsubishi MR-F51GY.

- light microscope

: Olympus B071, Japan.

- hot plate

: Model PS-D, Sakura Finetechnical Co.Ltd.,

Tokyo, Japan.

- paraffin dispenser

: Ashcroft, USA.

- laminar flow

- syringe terumo with needle

: Terumo, Inc., Japan.

size 1, 2.5 and 5 ml

- gamma counter

### Appendix III: reagent preparations

## 3.1 Preparation of reagents for determination of sex steroid hormone by RIA technique

The reagent preparations were followed WHO (1986) procedure.

## 3.1.1 Steroid assay buffer (buffer S)

Natrium dihydrogen phosphate-monohydrated: NaH <sub>2</sub> PO <sub>4</sub> . H <sub>2</sub> O	3.05	g
Disodium hydrogen phosphate anhydrous: Na <sub>2</sub> HPO <sub>4</sub>	11.6	g
Sodium chloride: Nacl	8.8	g
Thiomersal	0.1	g
Gelatin	1.0	g

The gelatin must be dissolved in 300 ml of warm di-distilled water. After the solution was cool, the rest of the certain reagents were added. The volume was made up to 1 liter and the pH of this buffer is adjusted to 7.2 to 7.4 by dropwise addition of sodium hydroxide (NaOH) or hydrochloric acid (HCl).

The buffer was stored at 4 °C. It should be stable for at least 1 month. This buffer was used as the diluent for all reagents in sex steroid assays of hormone.

### 3.1.2 Charcoal suspension

Charcoal	0.625	g
Dextran	0.0625	g
Assay buffer	100	ml

Dextran was dissolved in 100 ml of assay buffer in a stoppered container, and then charcoal was added and shaked vigorously for 30 seconds. The charcoal reagent should be stable at 4 °C for at least 1 month. The settle down suspension should be stirred vigorously during use at 4 °C.

### 3.1.3 Scintillation fluid

2,5-diphenyloxazole (PPO)	5.0	g
1,4-bis(2-(5-phenyloxazol))-Benzene (POPOP)	0.3	g
Toluene	1.0	g
Dioxane	200	ml

These constituents were homogenously mixed and stored in the dark bottle. The solution could be stable at the room temperature. Scintillation fluid should be prepared before use at least 7 days.

### 3. 2 Preparation of estradiol tracer, antiserum and standards

#### 3.2.1 Estradiol tracer

The stock solution (concentration 10  $\mu$ Ci/ml) was prepared from estradiol tracer [(2,4,6,7- $^3$ H) estradiol] in amounts of 250  $\mu$ Ci by mixing with toluene:ethanol (9:1). 100  $\mu$ l of stock solution was pipetted and evaporated, then redissolved in 10 ml of assay buffer. The final concentration contained 100 nCi/ml or 10,000 cpm. The estradiol tracer was stored at 4  $^{\circ}$ C.

#### 3.2.2 Estradiol antiserum

Lyophilized form of estradiol antiserum obtained from Prof. Kohen Fortune, Israel was added with 0.5 ml of di-distilled water two times, then transferred 100 µl of the solution in each microfuge tube and stored at 0 °C. Each microfuge tube was added with 900 µl of di-distilled water, this concentration was 1:10. One-hundred microlitters of this solution was transferred in each microfuge tube and stored at 0 °C. The solution was dissolved again as mentioned above, but this time the assay buffer was used instead of di-distilled water. The concentration was 1:1,000. This concentration served as stock solution of estradiol antiserum and stored at 4 °C. When working solution was required, stock solution was added with assay buffer and mixed until the final concentration was 1:20,000.

#### 3.2.3 Estradiol standard

Estradiol standard batch number 79/11 at the concentration of 10 ng/ml obtained from WHO RIA Reagent Programme, Switzerland was served as stock solution of estradiol standard. The concentration of estradiol standard serial dilution was 9.8, 19.6, 39, 78, 156.5, 312.5, 625, 1250, 2500 and 5,000 pg/500 µl/tube

### 3.3 Preparation of testosterone tracer, antiserum and standards

### 3.3.1 Testosterone tracer

The stock solution (concentration 10  $\mu$ Ci/ml) was prepared from testosterone tracer [(1,2,6,7- $^3$ H) testosterone] in amounts of 250  $\mu$ Ci by mixing with toluene:ethanol (9:1). 100  $\mu$ l of stock solution was pipetted and evaporated, then redissolved in 10 ml of assay buffer. The final concentration contained 100 nCi/ml or 10,000 cpm. The testosterone tracer was stored at 4°C.

## 3.3.2 Testosterone antiserum

Lyophilized form of testosterone antibody batch number K888510 obtained from WHO RIA Reagent Programme, Switzerland was stable for several years if stored at 4°C. One bottle of testosterone antibody was added with 10 ml of assay buffer when required with the final dilution in 1:210,000.

#### 3.3.3 Testosterone standard

Testosterone standard batch number K079810 obtained from WHO RIA Reagent Programme, Switzerland at the concentration of 220 nmol/l or 2,200 fmol/l was aliquoted to the vials provided, each vial contained 100 μl of testosterone standard. These aliquotes were stored at 4°C until need. When required, 10 ml of assay buffer was added to 100 μl of testosterone standard and heated at 40°C in water bath for 30 minutes. After the solution was mixed vigorously, it allowed cooling at the room temperature before use. The concentration of testosterone standard serial dilution was 17.2, 34.4, 68.8, 137.5, 275, 550 and 1,100 fmol/500 μl/tube.

### 3.4 Preparation for determination of gonadotropins by RIA technique.

The reagent preparations were followed Watanabe et al. (1990) procedure.

#### 3.4.1 Assay buffer for 0.5 M PBS-0.1% NaN<sub>3</sub> pH 7.6

- 1. Preparation of solution A and B was as followed;
  - 900 ml of 0.05 M of solution A was prepared from 35.814 grams of  $Na_2HPO_4.12H_2O\ (MW: 358.14), \ which \ it \ was \ dissolved \ in \ 2 \ liters \ of \ distilled \ water$

- 250 ml of 0.05 M of solution B was prepared from 7.801 grams of  $NaH_2PO_4.2H_2O$  (MW: 156.10), which it was dissolved in 1 liters of didistilled water.
- 2. Solution B was poured into solution A, then the pH of this mixed solution should be checked to be 7.6
- 3. NaN<sub>3</sub> 2 grams / 2 liters was added.
- 4. 0.14 M of NaCl 16.364 grams / 2 liters was added.
- 2. The solution was stored at 4°C. It should be stable for 1 month.

### 3.4.2 Assay buffer for 0.5 M PBS-0.1% NaN<sub>3</sub>-0.05 M EDTA-1% NRS pH 7.6

- 1. The assay buffer above was added with EDTA, then the pH of this mixed solution should be checked to be 7.6 by dropwise addition of 5 M NaOH.
- 2. NRS was added and mixed.

#### 3.5 Preparation of FSH tracer, antiserum and standards

#### 3.5.1 FSH tracer

Lyophilized form of FSH tracer (Batch number NIDDK-anti-rFSH-S-11 (AFPO 972881)), which it obtained from NIDDK, Japan and contained its substance 100 µg/ampule, was dissolved in 1 ml of assay buffer, then the solution was aliquoted into vials. 20-25 µg of FSH tracer per vial. The solution was stored at 4°C, it should be stable for 4-6 months. FSH

tracer was dissolved in 0.05 M PBS-0.1% NaN<sub>3</sub>-0.1% BSA. The value of cpm used in this assay was 4,000-5,000 cpm/50  $\mu$ l.

#### 3.5.2 FSH antiserum

FSH antisera (NIDDK-anti-rFSH-S-11 (AFPO 972881)) obtained from NIDDK, Japan was dissolved in 1 ml of 1:12.5 (2% normal rabbit serum (NRS):assay buffer), then the solution was lyophilized. The lyophilized form of FSH antisera was mixed with 1 ml of didistilled water served as stock solution. This solution was stored at 0°C. When working solution was required, stock solution of FSH antisera was added with 0.05 M PBS-0.1% NaN<sub>3</sub>-0.05 M EDTA-1% NRS (pH 7.6) and mixed until the final concentration was 1:125,000.

#### 3.5.3 FSH standard

Lyophilized form of FSH standard (NIDDK-rFSH-RP-2 (AFP-4621B)) obtained from NIDDK, Japan was dissolved in 1 ml of 1% BSA phosphosaline buffer. Its concentration was  $10 \mu g/ml$ , then  $25 \mu l$  of the solution was aliquoted into each vial and stored at 0°C. It should be stable for 3-5 months. The concentration of FSH standard serial dilution was 3.9, 7.8, 15.6, 31.2, 62.5, 125, 250, 500 and  $1,000 pg/100 \mu l/tube$ .

### 3.6 Preparation of LH tracer, antiserum and standards

#### 3.6.1 LH tracer

Lyophilized form of LH tracer (NIDDK-rLH-I-5 (AFP-11536B)), which it obtained from NIDDK, Japan and contained its substance 100 μg/ampule, was dissolved in 1 ml of assay buffer, then the solution was aliquoted into vials. 20-25 μg of LH tracer per vial. The solution was stored at 0°C, it should be stable for 2-3 months. LH tracer was dissolved in 0.05 M PBS-0.1% NaN<sub>3</sub>-0.1% BSA. The value of cpm used in this assay was 4,000-5,000 cpm/50 μl.

#### 3.6.2 Preparation of LH antiserum

LH antisera (NIDDK-anti-rLH-S-11 (AFPO 972881)) obtained from NIDDK, Japan was dissolved in 1 ml of 1:18.75 (2% NRS:assay buffer), then the solution was lyophilized. The lyophilized form of LH antisera was mixed with 1 ml of di-distilled water served as stock solution. This solution was stored at 0°C. When working solution was required, stock solution of LH antisera was added with 0.05 M PBS-0.1% NaN<sub>3</sub>-0.05 M EDTA-1% NRS (pH 7.6) and mixed until the final concentration was 1:180,000.

### 3.6.3 Preparation of LH standard

Lyophilized form of LH standard (NIDDK-rLH-RP-3 (AFP-7187B)) obtained from NIDDK, Japan was dissolved in 1 ml of 1% BSA phosphosaline buffer. Its concentration was 5  $\mu$ g/ml, the solution was stored at 0°C. It should be stable for 3-5 months. The concentration of LH standard serial dilution was 3.9, 7.8, 15.6, 31.2, 62.5, 125, 250, 500 and 1,000 pg/100 $\mu$ l/tube.

### 3.7 Preparation of histopathological reagents

#### 3.7.1 10% buffer formalin

Formalin (40%)	100	ml
Di-distilled water	900	ml
Natrium dihydrogen phosphate-monohydrated: NaH <sub>2</sub> PO <sub>4</sub> . H <sub>2</sub> O	4	g
Disodium hydrogen phosphate anhydrous: Na <sub>2</sub> HPO <sub>4</sub>	6.5	g

These chemical substances were mixed together in the dark bottle, the solution was shaked until it was completely dissolved. This solution was stored at the room temperature.

### 3.7.2 Ehrlich's acid haematoxylin and eosin

Haematoxylin	8	g
Ethanol (absolute)	400	ml
Ammonium alum	8	g
Di-distilled water	400	ml
Glycerine	400	ml
Glacial acetic acid	40	ml

Haematoxylin was dissolved in absolute ethanol in water bath at 40-50□C. When the solution was cool, it was filtered with filtered paper, then ammonium alum was dissolved in warm di-distilled water. These two solutions were mixed together, then glycerine and glacial acetic acid were added and stirred until these substances were completely dissolved. The solution needs to expose to daylight to ripen for at least 6 weeks.

#### 3.7.3 **Eosin**

Eosin Y	0.5	5 g
Ethanol (95%)	100	) ml

Eosin was dissolved in absolute ethanol until the solution was completely dissolved and stored at the room temperature.

# **BIOGRAPHY**

Miss. Sukanya Jaroenporn was born on April 11, 1972 in Chachoengsao province, Thailand. She received the Bachelor degree of Nursing Science in 1995 from Mahidol University, Bangkok, Thailand. She has entrolled at Chulalongkorn University in graduate program for the Degree of Master of Science in Physiology and graduated in 2004.

