

References



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## Appendix I

### 1. Preparation of Media

#### 1.1 Antibiotic Medium 1 (Difco Laboratories)

##### Ingredients per liter

Bacto-Beef Extract	1.5 gm
Bacto-Yeast Extract	3 gm
Bacto-Casitone	4 gm
Bacto-Peptone	6 gm
Bacto-Dextrose	1 gm
Bacto-Agar	15 gm

To rehydrate the medium, suspend 30.5 gm in 1,000 ml cold distilled water and heat to boiling to dissolve the medium completely. Sterilize in the autoclave for 15 minutes at 15 pounds pressure (121°C). The final pH would be obtained to  $6.6 \pm 0.1$ .

#### 1.2 Antibiotic Medium 2 (Difco Laboratories)

##### Ingredients per liter

Bacto-Beef Extract	1.5 gm
Bacto-Yeast Extract	3 gm
Bacto-Peptone	6 gm
Bacto-Agar	15 gm

To rehydrate the medium, suspend 25.5 gm in 1,000 ml cold distilled water and heat to boiling to dissolve the medium completely.



Adjust the pH to  $6.0 \pm 0.1$ . Sterilize in the autoclave for 15 minutes at 15 pounds pressure ( $121^{\circ}\text{C}$ ).

### 1.3 Mueller Hinton Medium (Difco Laboratories)

#### Ingredients per liter

Beef, Infusion from	300 gm
Bacto-Casamino Acids, Technical	17.5 gm
Starch	1.5 gm
Bacto-Agar	17 gm

To rehydrate the medium, suspend 38 gm in 1 liter distilled or deionized water and heat to boiling to dissolve completely. Adjust the pH to  $7.3 \pm 0.1$ . Sterilize in the autoclave for 15 minutes at 15 pounds pressure ( $121^{\circ}\text{C}$ ).

### 1.4 Nutrient broth <sup>(58)</sup>

#### Ingredients per liter

Beef extract	10 gm
Peptone	10 gm
Sodium chloride	5 gm
Distilled Water qs.	1,000 ml

Mix the ingredients in the distilled water and heat to boiling to dissolve the ingredients completely. Adjust the pH to  $7.3 \pm 0.1$ . Sterilize in the autoclave for 15 minutes at 15 pounds pressure ( $121^{\circ}\text{C}$ ).

1.5 Nutrient agar<sup>(58)</sup>

## Ingredients per liter

Peptone	5 gm
Beef extract	3 gm
Sodium chloride	5 gm
Agar	20 gm

Mix all ingredients in 1 liter distilled or deionized water and heat to boiling to dissolve completely. Adjust the pH to  $7.3 \pm 1$ . Sterilize in the autoclave for 15 minutes at 15 pounds pressure ( $121^{\circ}\text{C}$ ).

1.6 Blood agar<sup>(58)</sup>

Mix the 190 ml sterile nutrient agar (1.5) with 10 ml sheep blood.

1.7 Plate Count agar<sup>(59)</sup>

## Ingredient per liter

Tryptone	5 gm
Glucose	1 gm
Yeast Extract	3 gm
Agar	15 gm

Mix all ingredients in 1 liter distilled or deionized water and heat to boiling to dissolve completely. Adjust the pH to  $7.0 \pm 0.1$ . Sterilize in the autoclave for 15 minutes at 15 pounds pressure ( $121^{\circ}\text{C}$ ).



## 2. Preparation of solutions

### 2.1 Normal saline solution

Dissolve 9 gm sodium chloride with 1000 ml distilled water. Sterilize in the autoclave for 15 minutes at 15 pounds pressure (121°C).

### 2.2 0.1 M phosphate buffer pH 4.5

Dissolve 13.6 gm monobasic potassium phosphate in distilled water and dilute to 1 liter with distilled water. Sterilize in the autoclave for 15 minutes at 15 pounds pressure (121°C).

### 2.3 Solution for intraperitoneal and intramuscular administration (1 mg/ml)

Dissolve 100 mg tetracycline HCl with normal saline solution and adjust to 100 ml with the solvent. Filter the solution through millipore filter. Discard the first 10 ml filtrate.

### 2.4 Solution for oral administration (20 mg/ml)

Dissolve 2.0 gm tetracycline HCl with normal saline solution and adjust to 100 ml with the solvent.

## Appendix II

### 1. Determination of the biological half-life ( $t_{1/2}$ ) (60,61)

The biological half life was calculated from the following equation. (60,61)

$$t_{1/2} = \frac{\ln 2}{\beta} = \frac{0.693}{\beta}$$

In the present study, it could be observed that after 24 h, the drug levels in serum and muscle after the three routes of administration were decreased slowly. The linear equation<sup>(46)</sup> was established from 24 h to the terminal observed time.  $\beta$  is the slope of the linear line.

The tetracycline levels in muscle after the intraperitoneal administration was selected for the example of the determination of the biological half-life (see Table 6 and Figure 4). The detail was shown in Figure 16.

### 2. Determination of under the concentration-time curve ( $AUC_0^{\alpha}$ ) (60,61)

The area under the concentration-time curve was calculated by using the trapezoidal rule and the estimation of the rest area from the terminal sampling time to infinity was calculated by using the following equation.

$$AUC_{x \rightarrow \alpha}^t = \frac{C_x}{\beta}$$



where,  $C_x$  is the drug level at the terminal sampling time.

The tetracycline levels in muscle after the intraperitoneal administration was selected for the example of the determination of the  $AUC_0^\alpha$ . The detail was shown in Figure 17 and Table 13.



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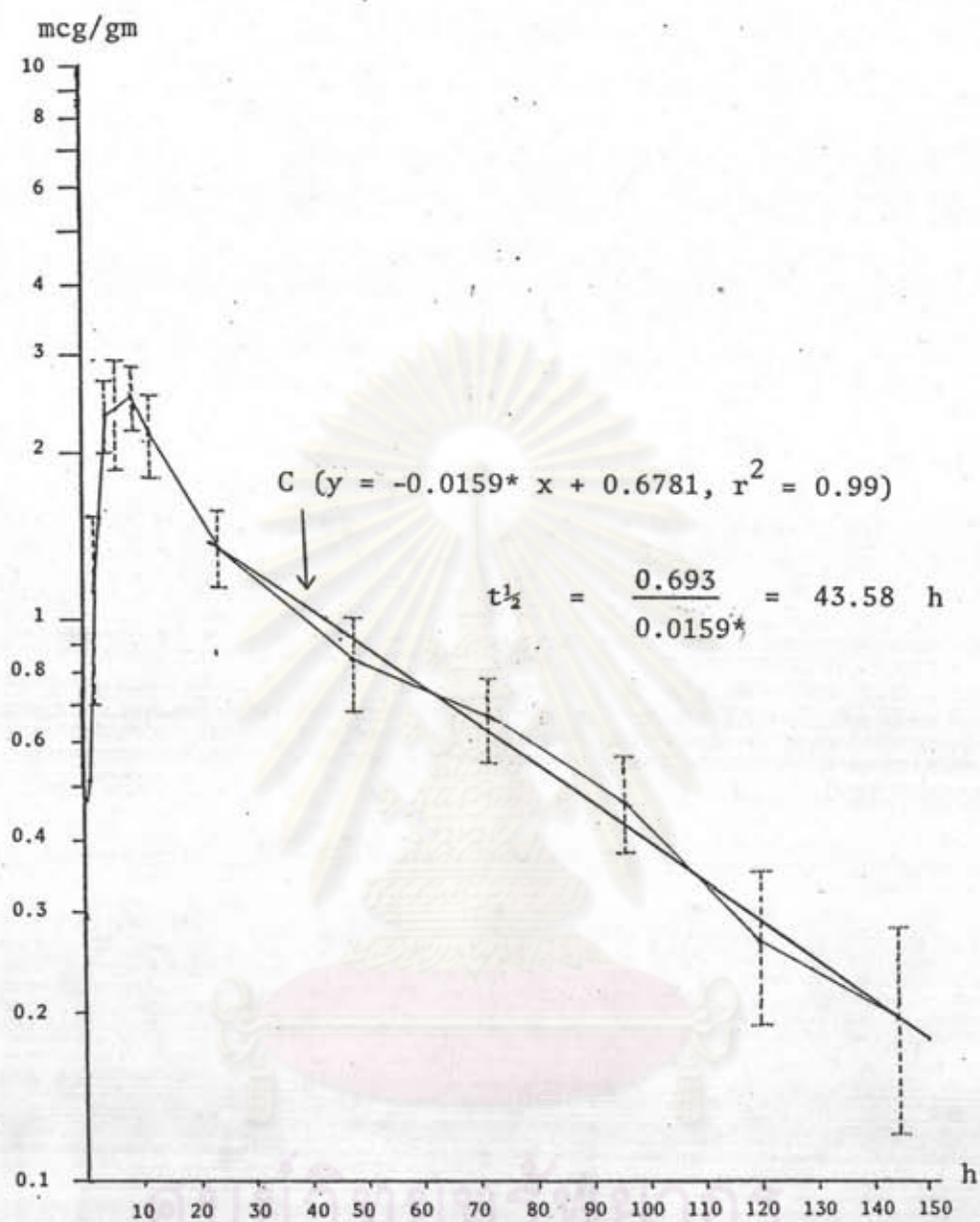


Figure 16 The tetracycline levels in muscle after the intraperitoneal administration and the established linear regression line (C) (from 24 h to 144 h)

$y = \ln(\text{tetracycline levels in muscle})$

$x = \text{the time (h) after the intraperitoneal administration}$

$r^2 = \text{the coefficient of determination}$

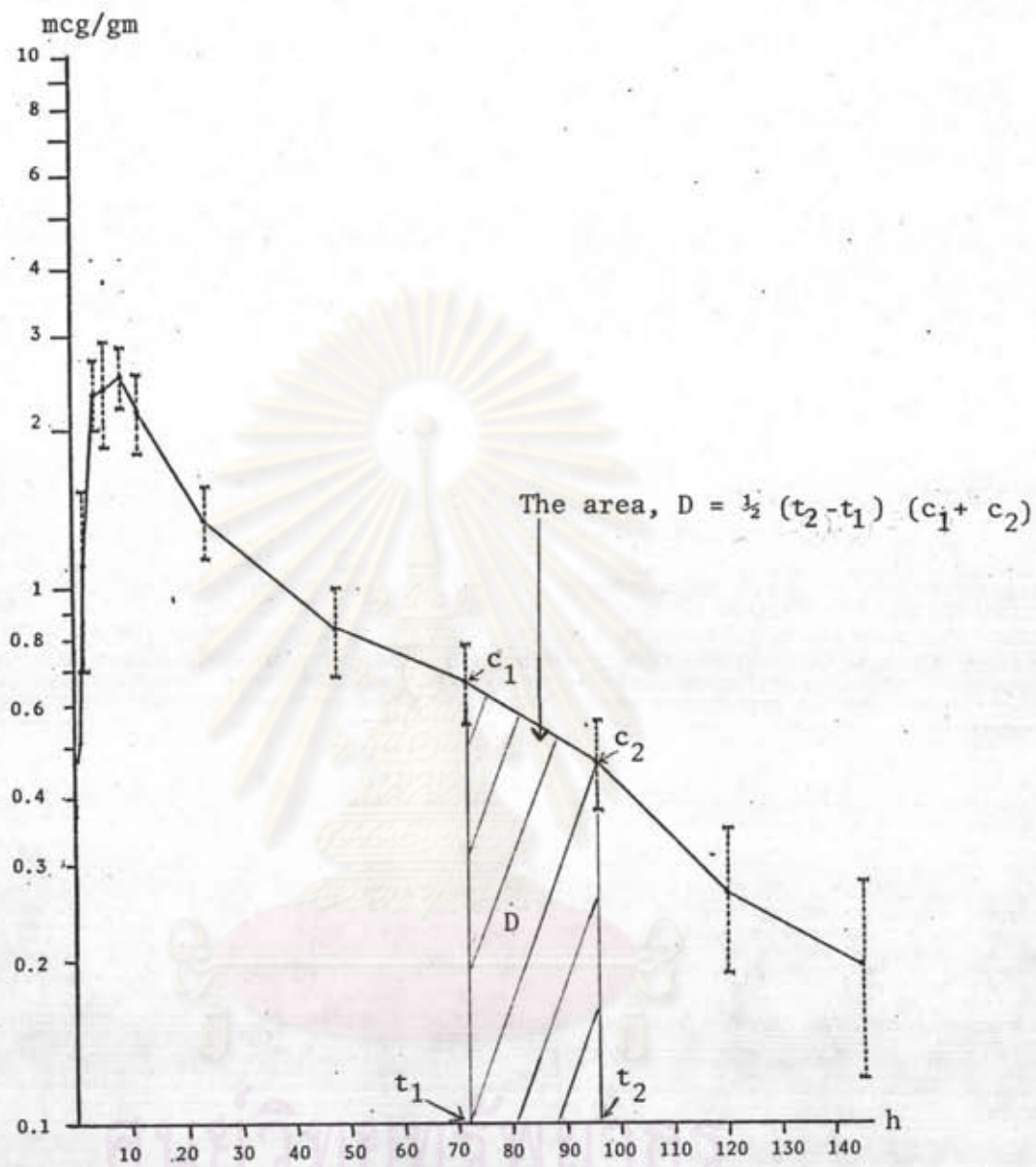


Figure 17 The determination of the area,  $D$  of tetracycline levels in muscle after the intraperitoneal administration ( $c_1$  and  $c_2$  are the tetracycline levels in muscle at the sampling time,  $t_1$  and  $t_2$ , respectively.)



Table 13 The determination of the  $AUC_0^\alpha$  of tetracycline in muscle (mcg.h/gm) after the intraperitoneal administration (see Table 7, Figure 16 and Figure 17)

time (n)	$AUC_0^\alpha$ of tetracycline in muscle (mcg.h/gm.)
0 - 0.5	$\frac{1}{2} (0.5 - 0) (0 + 0.47) = 0.118$
0.5 - 1	$\frac{1}{2} (1 - 0.5) (0.47 + 0.51) = 0.245$
1 - 2	$\frac{1}{2} (2 - 1) (0.51 + 1.12) = 0.815$
2 - 4	$\frac{1}{2} (4 - 2) (1.12 + 2.31) = 3.430$
4 - 6	$\frac{1}{2} (6 - 4) (2.31 + 2.39) = 4.700$
6 - 9	$\frac{1}{2} (9 - 6) (2.39 + 2.52) = 7.365$
9 - 12	$\frac{1}{2} (12 - 9) (2.52 + 2.17) = 7.035$
12 - 24	$\frac{1}{2} (24 - 12) (2.17 + 1.35) = 21.120$
24 - 48	$\frac{1}{2} (48 - 24) (1.35 + 0.85) = 26.400$
48 - 72	$\frac{1}{2} (72 - 48) (0.85 + 0.67) = 18.240$
72 - 96	$\frac{1}{2} (96 - 72) (0.67 + 0.47) = 13.680$
96 - 120	$\frac{1}{2} (120 - 96) (0.47 + 0.27) = 8.880$
120 - 144	$\frac{1}{2} (144 - 120) (0.27 + 0.20) = 5.640$
144 - $\alpha$	$\frac{0.20}{0.0159} = 12.579$
0 - $\alpha$	=130.247

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