

การสังเคราะห์ไฟร์ทรินจากเซลล์เพาชเลี้ยงของไฟร์ทริน
(*Chrysanthemum cinerariaefolium* Bocc.)



นางสาวอภิชา เวชประศิริ

ศูนย์วิทยทรัพยากร

วิทยานิพนธ์นี้ เป็นส่วนหนึ่งของการศึกษาตามหลักสูตรปริญญาวิทยาศาสตร์มหาบัณฑิต
หลักสูตรเทคโนโลยีชีวภาพ

บัณฑิตวิทยาลัย จุฬาลงกรณ์มหาวิทยาลัย

พ.ศ. 2533

ISBN 977-577-786-2

ลิขสิทธิ์ของบัณฑิตวิทยาลัย จุฬาลงกรณ์มหาวิทยาลัย

016515

110312304

SYNTHESIS OF PYRETHRINS IN TISSUE CULTURE OF PYRETHRUM
(*CHRYSANTHEMUM CINERARIAEFOLIUM* BOCC.)



Miss Apitar Vesprasit

A Thesis Submitted in Partial Fulfillment of the Requirements

for the Degree of Master of Science

Programme Biotechnology

Graduate School

Chulalongkorn University

1990

ISBN 974-577-786-2

Thesis Title SYNTHESIS OF PYRETHRINS IN TISSUE CULTURE OF PYRETHRUM
(CHRYSANTHEMUM CINERARIAEFOLIUM Bocc.)

By Miss Apitar Vesprasit

Department Programme Biotechnology

Thesis Advisor Associate Professor Sanha Panichjakul, Ph.D.
 Associate Professor Oradee Sahavacharin, Ph.D.



Accepted by the Graduate School, Chulalongkorn University in
partial fulfillment of the requirements for the Master's degree.

..... *Thavorn Vajrabaya* Dean of Graduate School
(Professor Thavorn Vajrabaya, Ph.D.)

Thesis Committee :

..... *Suthep Taneeyawan* Chairman
(Assistant Professor Suthep Taneeyawan, Ph. D.)

..... *Sanha Panichjakul* Member
(Associate Professor Sanha Panichjakul, Ph. D.)

..... *Oradee Sahavacharin* Member
(Associate Professor Oradee Sahavacharin, Ph. D.)

..... *Soongsri Kulpreeda* Member
(Associate Professor Soongsri Kulpreeda, Ph. D.)



อภิตา เวชประสาท : การสังเคราะห์ไฟร์กินจากเซลล์เพาะเลี้ยงของไฟร์กิน
(CHRYSANTHEMUM CINERARIAEFOLIUM BOCC.) (SYNTHESIS OF PYRETHRINS IN
TISSUE CULTURE OF PYRETHRUM (CHRYSANTHEMUM CINERARIAEFOLIUM BOCC.))
อ.ป.ปริญญา : รศ.ดร.สันติ์ พนิษยกุล และ รศ.ดร.อรดี สหวัชชินทร์, 147 หน้า.
ISBN 974-577-786-2

สารไฟร์กินซึ่งสักดิ้นจากพืชไฟร์กิน (Chrysanthemum cinerariaefolium Bocc.) พบว่ามีคุณสมบัติในการข้ามแมลง ดังนี้เจิงได้มีการศึกษาการสังเคราะห์สารไฟร์กินจากพืชไฟร์กินที่ได้จากการเพาะเลี้ยงเนื้อเยื่อโดยนำเข้าส่วนต่าง ๆ (ก้านใบ ใบ และส่วนลำต้น) ของพืชมาเพาะเลี้ยงภายใต้สภาพแวดล้อมและอาหารสูตรต่าง ๆ ซึ่งมีผลต่อการเจริญเติบโตและการผลิตสารไฟร์กิน นอกจากนี้ได้พัฒนาวิธีสักดิ้นและวิเคราะห์สารไฟร์กินที่เหมาะสม เพื่อให้ได้ผลผลิตสารไฟร์กินในปริมาณมากที่สุด และพนวจเนื้อเยื่อใบให้ผลตอบสนองต่อการเพาะเลี้ยงเนื้อเยื่อสูงสุด และการขึ้นรากนำไปเก็บแครลล์ส ของพืชไฟร์กินสูงสุดเมื่อเพาะเลี้ยงขึ้นส่วนของพืชในอาหารสูตร MS เติมสูตรมาตรฐานเสริมด้วย 2,4-D และ BA (ความเข้มข้น 1.0 และ 3.0 มิลลิกรัม/ลิตร ตามลำดับ) ที่ความเข้มแข็งฟลูออเรสเซนต์ 2,000 ลักษณะ อุณหภูมิ 25 ± 2 °C อย่างไรก็ตามจากการศึกษาผลของสารควบคุมการเจริญเติบโต ต่อการเจริญเซลล์พบว่า IAA และ BA (ความเข้มข้น 1.0 และ 3.0 มิลลิกรัม/ลิตร ตามลำดับ) ให้ผลการผลิตสารไฟร์กินจากเซลล์เพาะเลี้ยงมากกว่าที่เจริญในอาหารที่เสริมด้วย 2,4-D และ BA

การสักดิ้นสารไฟร์กินใช้วิธี Soxhlet ที่อุณหภูมิ 60°C โดยใช้ปิโตรเลียมอีเทอร์เป็นตัวสักดิ้น ซึ่งเป็นวิธีที่สามารถสักดิ้นสารไฟร์กินได้สูงสุด และการศึกษาทางคุณภาพวิเคราะห์ใช้วิธี Thin-layer chromatography โดยใช้ n-hexane:n-heptane:ethylacetate ในอัตราส่วน 40:48:12 เป็นตัวพา สามารถแยกองค์ประกอบทั้ง 6 ชนิดของสารไฟร์กินออกจากกันได้ดีที่สุด การศึกษาปริมาณและองค์ประกอบของสารไฟร์กินโดยละเอียดใช้วิธี gas chromatography และ high performance liquid chromatography โดยใช้ methylstearate เป็นสารมาตรฐาน

จากการศึกษานี้สรุปได้ว่า หากใช้อาหาร และสภาพเพาะเลี้ยงที่เหมาะสมแล้ว ทำให้สามารถเจริญเซลล์และสังเคราะห์สารไฟร์กินจากไฟร์กินที่ได้จากการเพาะเลี้ยงเนื้อเยื่อได้อย่างมีประสิทธิภาพ

ภาควิชา มหาวิทยาลัยเชียงใหม่
สาขาวิชา เทคโนโลยีชีวภาพ
ปีการศึกษา 2532

ลายมือชื่อนักเต็ต
ลายมือชื่ออาจารย์ที่ปรึกษา



เอกสารนี้เป็นของมหาวิทยาลัยมหาสารคาม

APITAR VESPRASIT : SYNTHESIS OF PYRETHRINS IN TISSUE CULTURE OF PYRETHRUM (*CHRYSANTHEMUM CINERARIAEFOLIUM* BOCC.). THESIS ADVISOR : ASSO. PROF. SANHA PANICHAJAKUL, PH.D. AND ASSO. PROF. ORADEE SAHAVACHARIN, PH.D., 147 PP. ISBN 974-577-786-2

Pyrethrins extracted from pyrethrum (*Chrysanthemum cinerariaefolium* Bocc.) has been found to possess an active insecticidal property. Through this property an attempt to synthesize pyrethrins in tissue culture of pyrethrum was carried out. Explants of three organ sources (petiole, leaf and stem piece) were cultured under varying influential factors on the growth and production of pyrethrins such as the medium constituents, and the environmental conditions. In addition, to maximize the pyrethrins extracted from pyrethrum grew under those conditions, the suitable extraction and analysis procedures were established. Through this study it was found that leaf tissue gave the highest response to those treated conditions and the maximum pyrethrum callus initiation could be achieved when the explants were grown on full strength MS medium supplemented with 2,4-D, BA (1.0:3.0 mg/ml) under 2,000 lux fluorescent illumination and maintained at the temperature of $25\pm2^{\circ}\text{C}$. However, it was found that the effect of growth regulators, IAA and BA recombined at a ratio of 1.0 to 3.0 mg/ml was more sustained on the pyrethrins production in culture cells than the 2,4-D and BA recombined at the ratio 1.0 to 3.0 mg/ml respectively.

Maximum yield of pyrethrum extract could be established when petroleum ether was employed as an extracting solvent in the Soxhlet apparatus at performing temperature of 60°C for at least 7 hours. The TLC qualitative analysis developed by n-hexane-n-heptane-ethylacetate (40:48:12) comprised the good separation of all six compounds. GC and HPLC analysis of the culture cells revealed to facilitate for more separation of pyrethrins constituents. It can be concluded that with appropriate media constituents and environmental culture conditions, an efficient synthesis of pyrethrins in tissue culture of pyrethrum can be achieved.

ภาควิชา เทคโนโลยีชีวภาพ
สาขาวิชา เทคโนโลยีชีวภาพ
ปีการศึกษา 2532

ลายมือชื่อนักศึกษา *Darin Deekul*
ลายมือชื่ออาจารย์ที่ปรึกษา *Phasit*



ACKNOWLEDGEMENTS

The author wishes to express her sincere gratitude to the following people :-

Associate Professor Dr. Sanha Panichajakul of the department of Biochemistry, Faculty of Science, Chulalongkorn University, for his valuable suggestion, useful guidance and keen interest throughout the course of practical work and presentation of the thesis.

Associate Professor Oradee Sahavacharin of the Department of Horticulture, Faculty of Agriculture, Kasetsart, University, for her kindness to advise in tissue culture practical work and her supervision of this research ideas.

Associate Professor Preeda Chaisiri of Department of Biochemistry, Faculty of Science, Chulalongkorn University, for her kindness and valuable advise in determining the Gas Chromatography during the present work.

Mr. Pin Thawanvichajakij of East Asiatic Company Limited for his kind providing of the pyrethrum dry power.

Mr. Wichai Supanimitrakul of Wellcome Thailand Limited for his kind providing of the pyrethrum extract.

Mrs. Yupa Mongkolsuk of Central Laboratory Instrument Center, Kasetsart University, for her helpful guidance and providing shoot culture.

All staff members of the Department of Biotechnology and Biochemistry, Faculty of Science Chulalongkorn University, for their kindness and helpful.

Finally, the author's grateful thanks are due to National Center for Genetic Engineering and Biotechnology for granting her partial financial support through conduct some parts of this investigation.



CONTENTS

	Page
ABSTRACT (Thai)	iv
ABSTRACT (English)	v
ACKNOWLEDGEMENTS	vi
CONTENTS	viii
LIST OF TABLES	xiii
LIST OF FIGURES	xv
ABBREVIATIONS	xx
CHEMICAL FORMULA	xxii
CHAPTER	
I INTRODUCTION	1
Botany	1
Historical	1
Pyrethrum as the economical plant	3
Dominant characteristics	4
The situation in Thailand	4
Chemistry of pyrethrins	5
1. General information of pyrethrins	5
2. Basic structure	9
Biogenesis of pyrethrins	12
1. Acid moiety	12
2. Alcohol moiety	14
Plant tissue culture as a unique and valuable tool.....	15

I	1. Vegetative propagation of pyrethrum <i>in vitro</i>	17
	2. Establishing callus culture	19
	3. Optimum volume of medium per culture vessel	24
	4. Sterilization of medium	24
	The use of cell culture technique for the production of economically valuable products	24
	1. Techniques of Cell Suspension Culture ..	25
	The suitability of tissue culture for secondary metabolites biosynthesis	26
	Objectives	27
II	EXPERIMENTALS	29
	MATERIAL	29
	Source of plant material	29
	Chemical agents	29
	1. Chemical agents for medium preoparation	29
	2. Surface sterilitant	30
	3. Growth regulators	30
	4. Staining reagent	30
	5. Chemical agents for extraction	30
	Glasswares	31
	1. Glasswares for tissue culture	31
	2. Glasswares for cell selection	31
	3. Glasswares for medium preparation	31
	Miscelleneous	31

II	METHODS	33
	Callus initiation	33
	1. Tissue sterilization	33
	2. Callus Culture establishment	36
	Establishment and development of suspension culture	41
	1. Free cell and aggregates of cell formation	41
	2. Batch culture of plateform shaker	45
	Extraction of pyrethrins	45
	1. Selection of the organic solvent for pyrethrins	47
	2. Screening for extracting procedure	47
	Pyrethrins analysis	48
	1. Preparation of TLC spreying reagent	48
	2. Preparation of the GC standard solution	49
	3. Standard calibrating curve	50
	4. Quailitative analysis of pyrethrins by TLC	50
	5. Quantitative analysis of pyrethrins by GC	51
	6. Quantitative determination of pyrethrins by HPLC	52
III	RESULTS	54
	Callus initiation	54
	1. Tissue sterilization	54
	2. Establishing callus culture	56

III	Callus maintenance	67
	Establishment of development of suspension	
	culture	69
	1. Free cell and aggregates of cells	
	formation	71
	2. Batch cultures on plateform shaker	76
	Suspension maintenance	80
	Extraction of pyrethrins	81
	1. Selection of extracting solvent	81
	2. Selection of extracting procedure	81
	Qualitative analysis of pyrethrins by TLC	84
	Quantitative of pyrethrins by GC	89
IV	DISCUSSION	103
	Callus initiation	103
	1. Tissue sterilization	103
	2. Establishing callus culture	104
	Callus maintenance	112
	Establishment and development of suspension	
	culture	112
	1. Free cells and aggregate cells	
	formation	113
	2. Batch culture on plateform shaker	115
	Suspension maintenance	116
	Extraction of pyrethrins	116
	1. Selection of extracting solvent	116
	2. Selection of extracting procedure	117
	Quanlitative analysis by TLC	118

IV	Quantitative analysis of pyrethrins by gas chromatography	119
	Quantitative analysis of pyrethrins by HPLC ...	120
	CONCLUSION	122
	Study on establishment of callus and cell suspension culture	122
	Study on establishment of pyrethrins extraction and analysis	122
	Study on the active ingredient of pyrethrum extract and their constituents in culture cells	123
	Study on effect of growth regulator on pyrethrins production in culture cells	123
REFERENCES		124
APPENDIX		136
VITA		147



**ศูนย์วิทยทรัพยากร
จุฬาลงกรณ์มหาวิทยาลัย**

LIST OF TABLES

Table		Page
1	Insecticides from plants and tissue culture	1
2	Steps of surface sterilization of pyrethrum tissues.....	34
3	Tissue preparation prior to callus establishment.....	34
4	Interaction of 2,4-D and BA effecting on pyrethrum callus initiation.....	38
5	Interaction of 2,4-D and BA effecting on pyrethrum cell culture	38
6	Interaction of IAA and BA effecting on pyrethrum cell culture	38
7	Apparent scores of sterilized pyrethrum leaf tissues after 7 days inoculation under selection treatments.....	55
8	Influence of growth regulator (2,4-D + BA) on initiation and establishment of leaf-derived callus of pyrethrum.....	57
9	Influence of salt concentration on establishment of leaf-derived callus of pyrethrum	63
10	Influence of light and dark period on the establishment of leaf-derived callus of pyrethrum	66
11	Effect of temperature on pyrethrum leaf-derived callus initiation.....	66
12	Influence of growth regulators (2,4-D + BA) on growth and establishment of pyrethrum cell suspension culture.....	73
13	Influence of growth regulators (IAA + BA) on growth and establishment of pyrethrum cell suspension culture.....	74
14	Influence of light and dark on the establishment of pyrethrum suspension cells that had been maintained under different conditions	77

LIST OF TABLES (cont.)

Table		Page
15	Influence of temperature on the establishment of pyrethrum suspension cells that had been maintained under different conditions	78
16	Apparent scores of pyrethrin I extracted by selection treatment	83
17	Apparent scores of pyrethrin I extracted by selection methods	83
18	Retention time of pyrethrin I and pyrethrin II in relation to column temperature	90
19	Appearance scores of retention time of methylstearate in relation to flow rate of GC operation	91
20	HPLC analysis of pyrethrins content in suspension cells in relation to GC analysis.....	92



**ศูนย์วิทยทรัพยากร
จุฬาลงกรณ์มหาวิทยาลัย**

LIST OF FIGURES

Figure	Page
1 Active ingredients of pyrethrum	10
2 Basic structure of pyrethrins, insecticidal esters	11
3 Synthesis of chrysanthemumdicarboxylic acid	13
4 Synthesis of chrysanthemummonocarboxylic acid	13
5 Synthesis of chrysanthemumdicarboxylic acid from chrysanthemummonocarboxylic acid	16
6 Synthesis of pyrethrolone	16
7 Degree signifies size of leaf-derived callus	39
8 Signifies intensity of cell formation	43
9 The process for preparing refined pyrethrum extract	46
10 Effect of growth regulator (2,4-D and BA) at various levels on callus initiation from leaf which had been cultured on MS basal medium under 2,000 lux of fluorescent illumination about an 8-hour dark period and maintained at temperature range of $25\pm2^{\circ}\text{C}$	58
11 Effect of growth regulator (2,4-D and BA) at various levels on callus initiation from petiole which had been cultured on MS basal medium under 2,000 lux of fluorescent illumination about an 8-hour dark period and maintained at temperature range of $25\pm2^{\circ}\text{C}$	59
12 Effect of growth regulator (BA) at various levels recombined with 2,4-D (1.0 mg/l) on callus initiation of leaf blade which had been cultured on MS basal medium under 2,000 lux of fluorescent illumination about an 8-hour dark period and maintained at temperature range of $25\pm2^{\circ}\text{C}$	60

LIST OF FIGURES (cont.)

Figure	Page
13 Effect of 2,4-D and BA (1.0:3.0 mg/l) on callus initiation of various organ sources which had been cultured on MS basal medium under 2,000 lux of fluorescent illumination about an 8-hour dark period and maintained at temperature range of $25\pm 2^{\circ}\text{C}$	62
14 Effect of medium concentration, half strength (left) and full strength (right) on callus initiation of leaf (A) and petiole (B) which had been cultured on MS basal medium under 2,000 lux of fluorescent illumination about an 8-hour dark period and maintained at temperature range of $25\pm 2^{\circ}\text{C}$	64
15 Effect of light and dark period on callus initiation from leaf (A) and petiole (B) which cultured on MS basal medium and maintained at temperature range of $25\pm 2^{\circ}\text{C}$	65
16 Effect of temperature on callus initiation from leaf which had been cultured on MS basal medium under 2,000 lux of fluorescent illumination about an 8-hour dark period and maintained at temperature range of $25\pm 2^{\circ}\text{C}$ (A) and 30°C (B)	68
17 Growth and developmental stages of leaf-derived callus which had been cultured on MS basal medium containing 2,4-D (1.0 mg/l) and BA (3.0 mg/l) under 2,000 lux of fluorescent illumination about an 8-hour dark period and maintained at temperature range of $25\pm 2^{\circ}\text{C}$	70

LIST OF FIGURES (cont.)

Figure		Page
18	Growth pattern of leaf-derived callus cultured on MS basal medium containing 2,4-D (1.0 mg/l) and BA (3.0 mg/l) under 2,000 lux of fluorescent illumination about an 8-hour dark period and maintained at temperature range of $25\pm2^{\circ}\text{C}$	70
19	Growth pattern of 2,4-D:BA-grown culture (0.25:3.0 mg/l) and IAA:BA-grown culture (2.0:3.0 mg/l) which had been culture in liquid MS basal medium under 2,000 lux of fluorescent illumination about an 8-hour dark period and maintained at temperature range of $25\pm2^{\circ}\text{C}$	75
20	Growth pattern of suspension cells cultured on MS basal medium containing 2,4-D (0.25 mg/l) and BA (3.0 mg/l) under 2,000 lux of fluorescent illumination about an 8-hour dark period and maintained at temperature range of $25\pm2^{\circ}\text{C}$	79
21	Growth and development of 4 weeks-old pyrethrum suspension culture	82
22	Appearance of TLC chromatography of PBK standard solution world standard solution and crude Shirayuki I callus extract which was developed within these condition (solvent system benzene in petroleum ether ($60\text{--}80^{\circ}\text{C}$)	85
23	Appearance of TLC chromatography of PBK standard solution world standard solution and crude Shirayuki I callus extract which was developed within these conditions (solvent system petroleum ether ($30\text{--}60^{\circ}\text{C}$)-ethylacetate	86

LIST OF FIGURES (cont.)

Figure		Page
24	Appearance of TLC chromatography of PBK standard solution world standard solution and crude Shirayuki I callus extract which was developed within these conditions (solvent system n-hexane-ethylacetate)	87
25	Appearance of TLC chromatography of PBK standard solution world standard solution and crude Shirayuki I callus extract which was developed within these conditions (solvent system 85/15 n-hexane-ethylacetate and 48:40:12 n-hexane-n-heptane-ethylacetate)	88
26	An isothermal plot of HETP in relation to flow rate of carrier gas (N_2)	93
27	The GC separation of mixture of the six pyrethrins components; equipped with flame ionization detector and 2,000 cm x 3.175 mm i.d. stainless steel column packed with 3% OV-17 on 80-100 mesh Chromosorb W(HP).	94
28	GC standard calibrating curve of word standard pyrethrum extract	95
29	Growth and pyrethrins quantity of leaf-derived callus cultured on MS basal containing 2,4-D (1.0 mg/l) and BA (3.0 mg/l) under 2,000 lux of fluorescent illumination about an 8-hour dark period and maintained at temperature range of $25\pm2^{\circ}C$ in relation of age of culture	96

LIST OF FIGURES (cont.)

Figure	Page
30 HPLC chromatograms of A, World standard pyrethrins extract, B and C represent pyrethrins extracts of suspension cells which had been cultured in MS medium supplemented with 2,4-D and BA (0.25:3.0 mg/ml), IAA and BA (1.0:3.0 mg/ml)	98
31 Growth and pyrethrins quantity in relation to the cultivation time of 2,4-D:BA-Grown culture (0.25:3.0 mg/ml) and IAA:BA-Grown culture (2.0:3.0 mg/ml)	99
32 Ratio of pyrethrins contents extracted from natural dry flower.	101
33 Ratio of pyrethrins contents extracted from three types of culture cells derived from leaf blade.	101
34 Stability of pyrethrins synthesized in one-year leaf-derived callus, suspension cell and shoot.	102

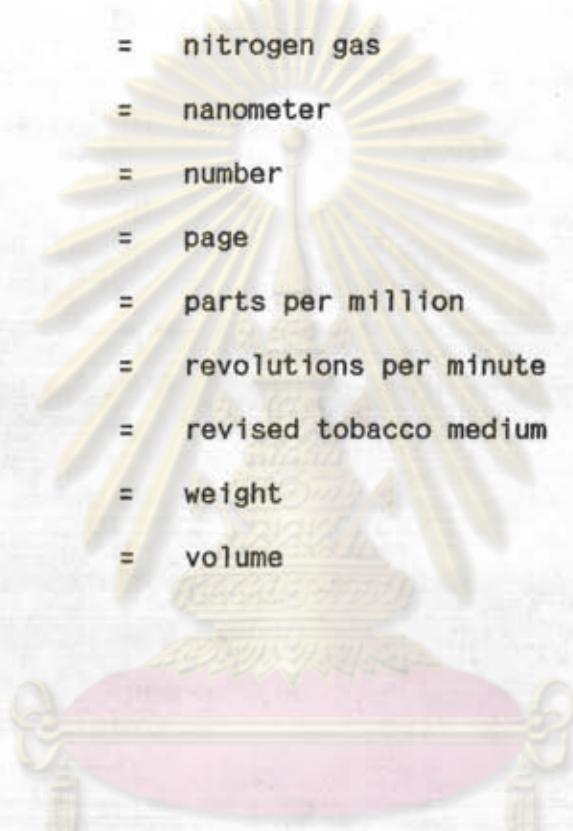
ศูนย์วิทยทรัพยากร
จุฬาลงกรณ์มหาวิทยาลัย



ABBREVIATION

AcOH	= acetic acid
BA	= Benzyladenine
p-BAP	= p-benzylaminopurine
°C	= degree celcius
ca.	= circa (approximately)
cm	= centimeter
2,4-D	= 2,4-dichlorophenoxyacetic acid
EtOAc	= ethylacetate
EtOH	= ethanol
et al.	= et alli (and others)
g	= gram
Glu	= glucose
GC	= gas chromatography
GI	= growth index
hr	= hour
hg	= hughnur (mercury)
HPLC	= high performance liquid chromatography
H ₂	= hydrogen gas
IAA	= indoleacetic acid
kg	= kilogram
l	= liter
M	= molarity
MS medium	= Murashige and Skoog's medium
MeOH	= methanol
μl	= microliter
mg	= milligram

min.	= minute
ml	= milliliter
mm	= millimeter
m.p.	= melting point
NAA	= α -naphthaleneacetic acid
N ₂	= nitrogen gas
nm	= nanometer
no.	= number
P	= page
ppm	= parts per million
rpm	= revolutions per minute
RT medium	= revised tobacco medium
wt	= weight
v	= volume



ศูนย์วิทยทรัพยากร
จุฬาลงกรณ์มหาวิทยาลัย

CHEMICAL FORMULA



CH_3COCH_3	= acetone
	= benzene
CHCl_3	= chloroform
CrO_3	= chromic acid
H_2O	= water
H_2SO_4	= sulfuric acid

ศูนย์วิทยทรัพยากร
จุฬาลงกรณ์มหาวิทยาลัย