

REFERENCES

- (1) Gupta, K. C. and Ravi Kumar, M. N. V. **Drug release behavior of beads and microgranules of chitosan.** *Biomaterials* 21 (2000): 1115-1119.
- (2) González-Rodríguez, M. L.; Holgado, M. A.; Sánchez-Lafuente, C.; Rabasco, A. M. and Fini, A. et al. **Alginate/chitosan particulate systems for sodium diclofenac release.** *International Journal of Pharmaceutics* 232 (2002): 225-234.
- (3) Remuñán-López, C.; Lorenzo-Lamosa, M. L.; Vila-Jato, J. L. and Alonso, M. J. **Development of new chitosan-cellulose multicore microparticles for controlled drug delivery.** *European Journal of Pharmaceutics and Biopharmaceutics* 45 (1998): 49-56.
- (4) Kim, T. K.; Park, Y. H.; Kim, K. J. and Cho, C. S. **Release of albumin from chitosan-coated pectin beads in vitro.** *International Journal of Pharmaceutics* 250 (2003): 371-383.
- (5) Torre, P. M.; Enobakhare, Y.; Torrado, G. and Torrado, S. **Release of amoxicillin from polyionic complexes of chitosan and poly(acrylic acid). study of polymer/polymer and polymer/drug interactions within the network structure.** *Biomaterials* 24 (2003): 1499-1506.
- (6) Gupta, V. K.; Hariharan, M.; Wheatley, T. A. and Price, J. C. **Controlled-release tablets from carrageenans: effect of formulation, storage and dissolution factors.** *European Journal of Pharmaceutics and Biopharmaceutics* 51 (2001): 241-248.
- (7) Sakiyama, T.; Chu, C. H.; Fujii, T. and Yano T. **Preparation of a polyelectrolyte complex gel from chitosan and κ -carrageenan and its pH-sensitive swelling.** *Journal of Applied Polymer Science* 50 (1993): 2021-2025.

- (8) Tomida, H; Nakamura, C. and Kiryu, S. **A novel method for the preparation of controlled-release theophylline capsules coated with a polyelectrolyte complex of κ -carrageenan and chitosan.** *Chemical & Pharmaceutical Bulletin* 42 (1994): 979-981.
- (9) Tapia, C.; Escobar, Z.; Costa, E.; Sapag-Hager, J.; Valenzuela, F.; Basualto, C.; Gai, M. N. and Yazdani-Pedram, M. **Comparative studies on polyelectrolyte complexes and mixture of chitosan-alginate and chitosan-carrageenan as prolonged diltiazem clorhydrate release system.** *European Journal of Pharmaceutics and Biopharmaceutics* 57 (2004): 65-75
- (10) Gillman, A. G.; Nies, T. W. and Taylor, P. **The pharmacological basis of therapeutics.** vol. 1. 8thed. New York: Pergamon Press, 1991.
- (11) O' Brien, W. M. **Adverse reactions to nonsteroidal anti-inflammatory drugs: diclofenac compared with other nonsteroidal anti-inflammatory drugs.** *The American Journal of Medicine* 80 (1986): 70-80.
- (12) Berger, J.; Reist, M.; Mayer, J. M.; Felt, O.; Peppas, N. A. and Gurny, R. **Structure and interactions in covalently and ionically crosslinked chitosan hydrogels for biomedical applications.** *European Journal of Pharmaceutics and Biopharmaceutics* 57 (2004): 19-34.
- (13) Çetinus, Ş. A. and Öztop H. N. **Immobilization of catalase into chemically crosslinked chitosan beads.** *Enzyme and Microbial Technology* 32 (2003): 889-894.
- (14) Hoffman, A. S. **Hydrogels for biomedical applications.** *Advanced drug delivery reviews* 54 (2002): 3-12.
- (15) Sriamornsak, P. **Effect of calcium concentration, harding agent and drying condition on release characteristics of oral proteins from calcium pectinate gel beads.** *European Journal of Pharmaceutics Sciences* 8 (1999): 221-227.

- (16) Roy, I. and Gupta, M. N. **κ -Carrageenan as a new smart macroaffinity ligand for the purification of pullulanase.** *Journal of Chromatography A* 998 (2003): 103-108
- (17) Kim, C. **Controlled release dosage form design.** Pennsylvania: Technology Publishing Company Book, 2000.
- (18) Baker, R. **Controlled release of biologically active agents.** California: John Wiley & Sons. Inc., 1987.
- (19) Shalaby, S. W.; Ikada, Y.; Langer, R. and Williams, J. **Polymers of biological and biomedical significance.** Washington D. C.: American Chemical Society, 1994.
- (20) Banker, G. S. and Rhodes, C. T. **Drug and the pharmaceutical sciences.** vol. 72. 3rd ed. New York: Marcel Dekker, 1996.
- (21) Higuchi, T. **Rate of release of medicaments from ointment bases containing drugs in suspension.** *Journal of Pharmaceutical Sciences* 50 (1961): 874-875.
- (22) Benita, S. and Donbrow, M. **Release kinetics of sparingly soluble drugs from ethyl cellulose-walled microcapsules: theophylline microcapsules.** *Journal of Pharmacy and Pharmacology* 34 (1981): 77-82.
- (23) Paul, w. and Sharma, C. P. **Chitosan, a drug carrier for the 21st century: a review.** *S.T.P. Pharmasciences* 10 (2000): 5-22.
- (24) Sinha, V. R.; Singla, A. K.; Wadhawan, S.; Kumria, R.; Bonsal, K. and Dhawan, S. **Chitosan microspheres as a potential carrier for drugs.** *International Journal of Pharmaceutics* 274 (2004): 1-33.
- (25) Calvo, P.; Vila-Jato J. L. and Alonso, M. J. **Evaluation of cationic polymer-coated nanocapsules as ocular drug carriers.** *International Journal of Pharmaceutics* 153 (1997): 371-383.
- (26) Azab, A. K.; Orkin, B.; Doviner, V.; Nissan, A.; Klein, M.; Srebnik, m. and Rubinstein, A. **Crosslinked chitosan implants as potential degradable devices for brachytherapy: In vitro and in vivo analysis.** *Journal of Controlled Release* 111 (2006): 281-289.

- (27) Patel, V. R. and Amiji, M. M. **Preparation and characterization of freeze-dried chitosan-poly(ethylene oxide) hydrogels for site-specific antibiotic delivery in the stomach.** *Pharmaceutical Research* 13 (1996): 588-593.
- (28) Ueno, H.; Mori, T. and Fujinaga, T. **Topical formulations and wound healing applications of chitosan** *Advanced drug delivery reviews* 52 (2001): 105-115.
- (29) Munjeri, O.; Collett, J. H. and Fell, J. T. **Hydrogel beads based on amidated pectins for colon-specific drug delivery: the role of chitosan in modifying drug release.** *Journal of Controlled Release* 46 (1997): 273-278.
- (30) Stanley, P. H.; Davis, S. and Illum, L. **In vitro evaluation of the mucoadhesive properties of chitosan microspheres.** *International Journal of Pharmaceutics* 166 (1998): 75-88.
- (31) Muzzarelli, R. A. **Chitosan-based dietary foods.** *Carbohydrate Polymers* 29 (1996): 306-316.
- (32) De Ruiter, G. A. and Rudolph, B. **Carrageenan biotechnology.** *Trends in Food Science & Technology* 8 (1998): 389-394.
- (33) Sankalio, M. G.; Mashru, R. C.; Sankalia, J. M. and Sutariya, V. B. **Stability improvement of alpha-amylase entrapped in kappa-carrageenan beads: physicochemical characterization and optimization using composition index.** *International Journal of Pharmaceutics* 312 (2006): 1-14.
- (34) Picker, K. M. **The use of carrageenan in mixture with microcrystalline cellulose and its functionality for making tablets.** *European Journal of Pharmaceutics and Biopharmaceutics* 48 (1997): 27-36.
- (35) Sipahigil, O. and Doctunç, B. **Preparation and in vitro evaluation of verapamil HCl and ibuprofen containing carrageenan beads.** *International Journal of Pharmaceutics* 228 (2001): 119-128.
- (36) Sallmann, A. R. **The history of diclofenac.** *The American Journal of Medicine* 80 (1986): 29-33.

- (37) Lund, W. **The pharmaceutical codex principles and practice of pharmaceuticals.** 12th ed. London: The pharmaceutical press, 1994.
- (38) Shu, X. Z. and Zhu, K. J. **A novel approach to prepare tripolyphosphate/chitosan complex beads for controlled release drug delivery.** *International Journal of Pharmaceutics* 201 (2000): 51-58.
- (39) Kurkuri, M. D. and Aminabhavi, T. M. **Poly(vinyl alcohol) and poly(acrylic acid) sequential interpenetrating network pH-sensitive micropheres for the delivery of diclofenac sodium to the intestine.** *Journal of Controlled Release* 96 (2004): 9-20.
- (40) Shu, X. Z. and Zhu, K. J. **Controlled drug release properties of ionically cross-linked chitosan beads: influence of anion structure** *International Journal of Pharmaceutics* 233 (2002): 217-225.
- (41) Cassidy, M. B.; Lee, H. and Trevors, J. T. **Survival and activity of lac-lux marked *Pseudomonas aeruginosa* UG2Lr cells encapsulated in κ -carrageenan over four years at 4°C.** *Journal of Microbiological Methods* 30 (1997): 167-170.
- (42) López, A.; Lázaro, N. and Marqués, M. **The interphase technique: a simple method of cell immobilization in gel-beads.** *Journal of Microbiological Methods* 30 (1997): 231-234.
- (43) Durby, M.; Peterson, R. V.; Larsen, J.; Rudolt, B.; Nørgaard, L. and Engelsen, S. B. **Towards on-line monitoring of the composition of commercial carrageenan powders.** *Carbohydrate Polymers* 57 (2004): 337-348.
- (44) Maciel, J. S.; Silva, D. A.; Haroldo, C. B. P. and Paula, R. C. M. **Chitosan/carboxymethyl cashew gum polyelectrolyte complex: synthesis and thermal stability.** *European Polymer Journal* 41 (2005) 2726-2733.

- (45) Puttipipatkachorn, S.; Pongjanyakul, T. and Priprem, A. **Molecular interaction in alginate beads reinforced with sodium starch glycolate or magnesium aluminum silicate, and their physical characteristic.** *International Journal of Pharmaceutics* 293 (2005): 51-62.
- (46) Kincl, M.; Vrečer, F. and Veber, M. **Characterization of factors affecting the release of low-solubility drug from prolonged release tablets.** *Analytica Chimica Acta* 502 (2004): 107-113.
- (47) Sheu, M.; Chou, H.; Kao, C.; Liu, C. and Sokoloski, T. D. **Dissolution of diclofenac sodium from matrix tablets.** *International Journal of Pharmaceutics* 85 (1992): 57-63.

APPENDICES

APPENDIX A

UV-Vis Spectrum and Calibration Curve of Sodium Diclofenac

UV-Vis Spectrum of Sodium Diclofenac

UV-Vis spectrophotometer was used to determine the amount of sodium diclofenac. The λ_{max} of drug absorbance in 0.1 N HCl (pH 1.2), phosphate buffer saline pH 6.6 and pH 7.4, NaOH solutions (2.5-7.5% (w/v)) and 5.0% (w/v) NaOH/KCl (0.1-0.5M) was identical (276 nm). The absorbance spectra in these dissolution media were illustrated in Figures A1-A9.

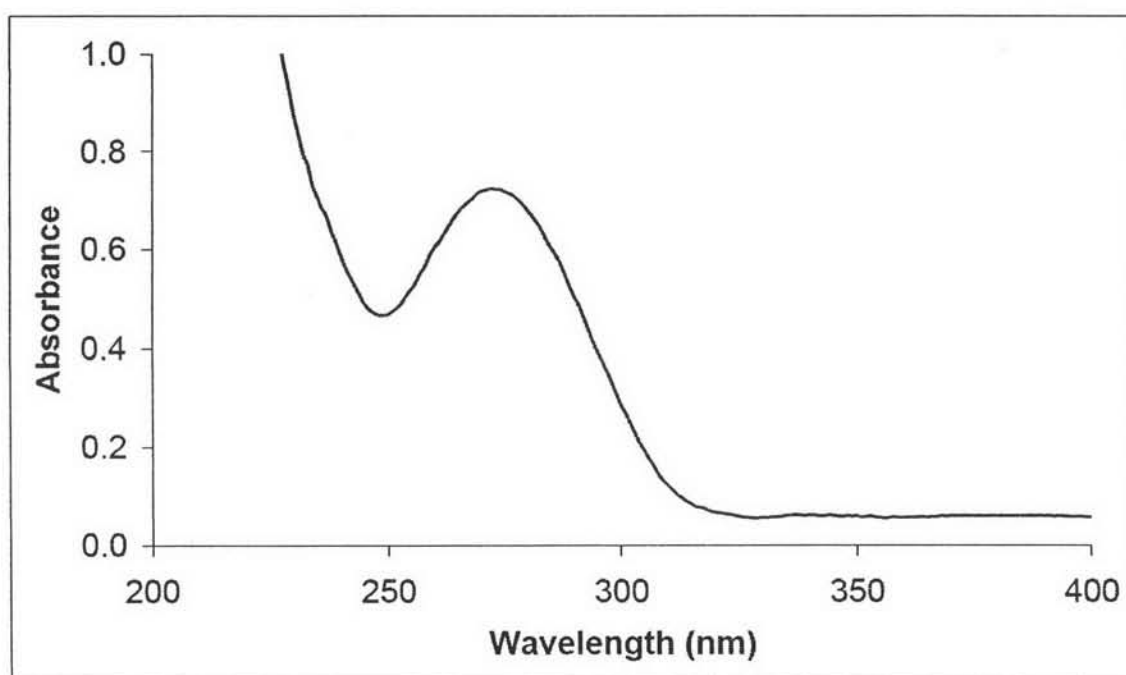


Figure A1 UV-Vis spectrum of sodium diclofenac in 0.1N HCl.

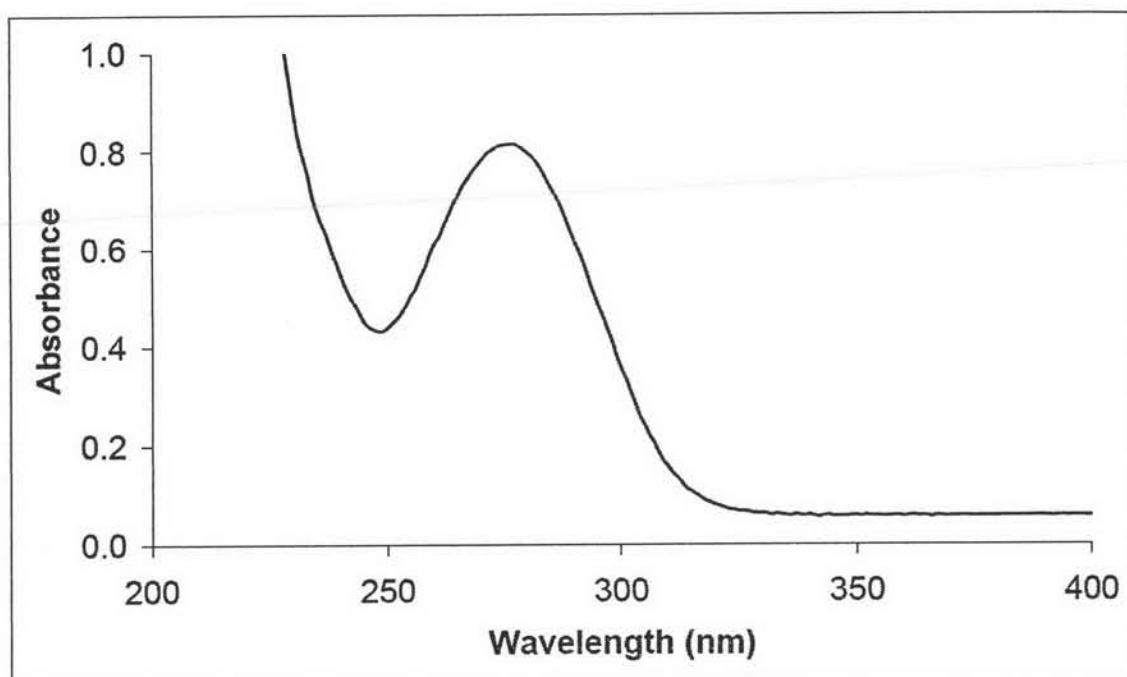


Figure A2 UV-Vis spectrum of sodium diclofenac in phosphate buffer saline pH 6.6.

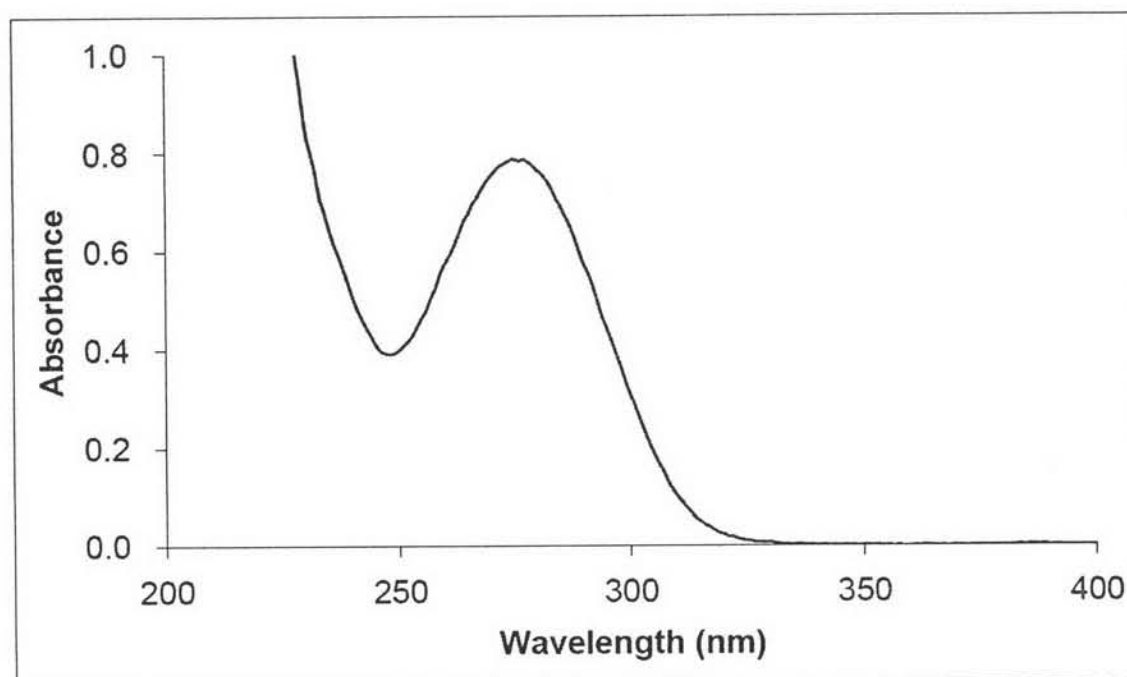


Figure A3 UV-Vis spectrum of sodium diclofenac in phosphate buffer saline pH 7.4.

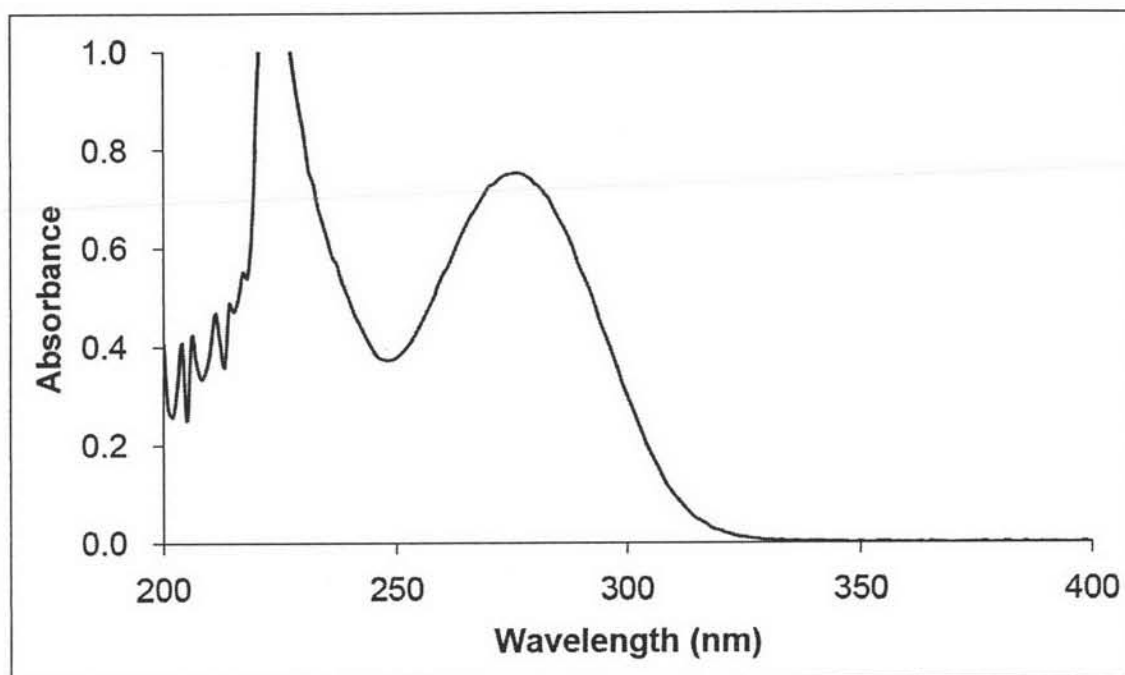


Figure A4 UV-Vis spectrum of sodium diclofenac in 2.5% (w/v) NaOH solution.

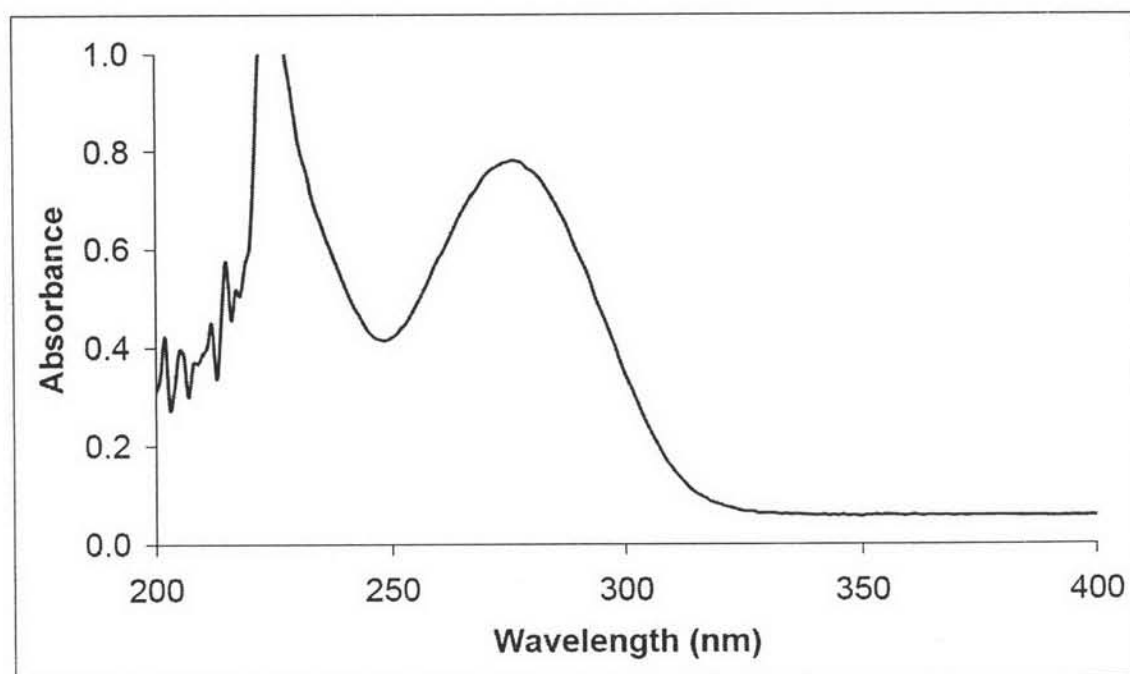


Figure A5 UV-Vis spectrum of sodium diclofenac in 5.0% (w/v) NaOH solution.

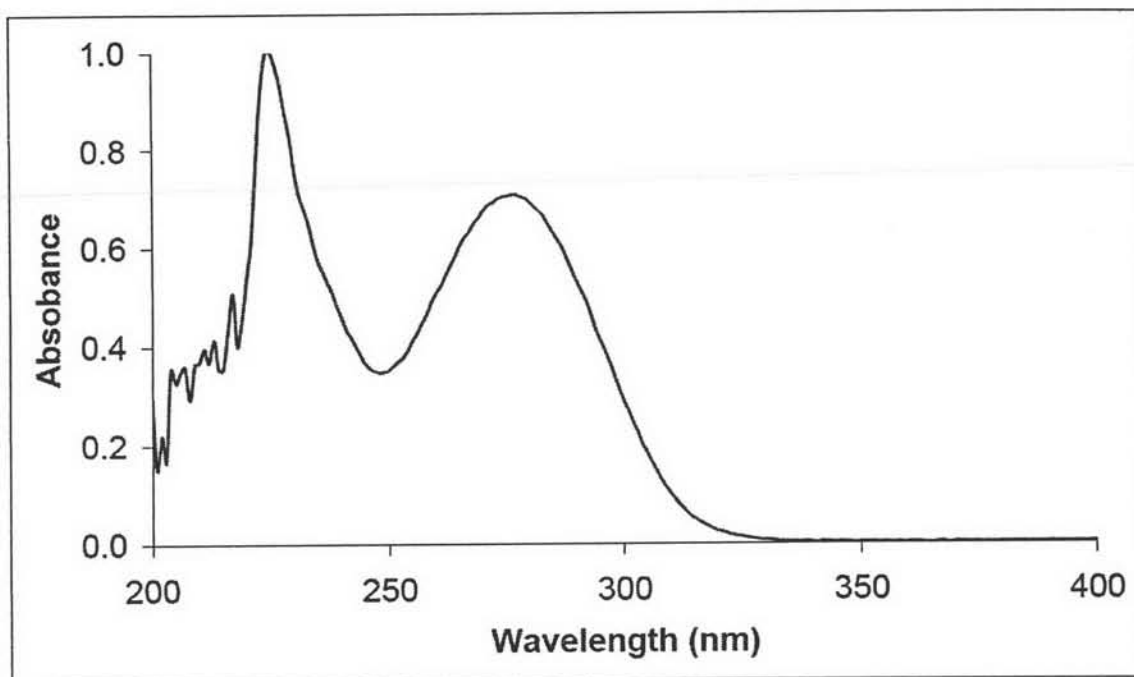


Figure A6 UV-Vis spectrum of sodium diclofenac in 7.5% (w/v) NaOH solution.

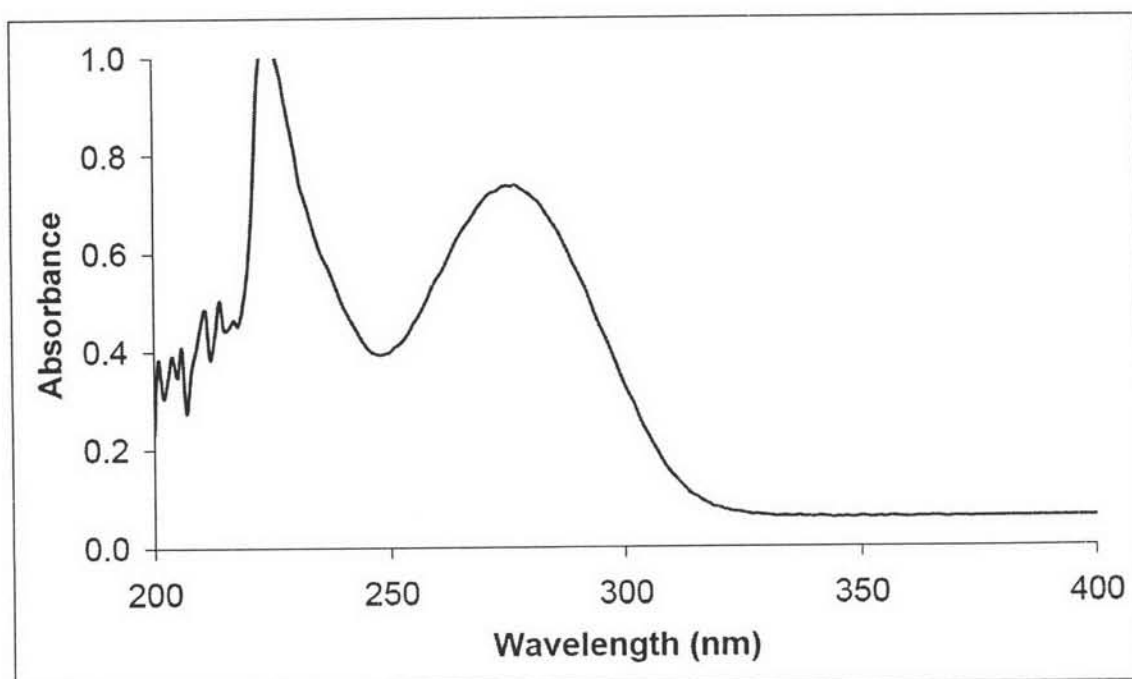


Figure A7 UV-Vis spectrum of sodium diclofenac in 5.0% (w/v) NaOH/0.1M KCl solution.

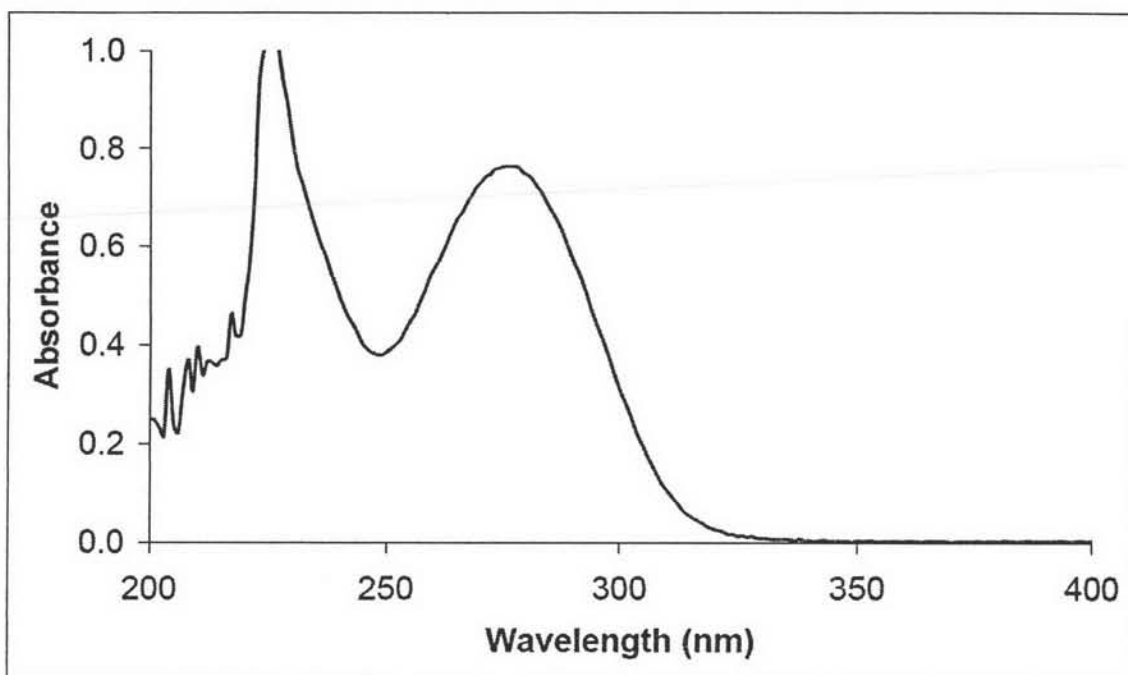


Figure A8 UV-Vis spectrum of sodium diclofenac in 5.0% (w/v) NaOH/0.3M KCl solution.

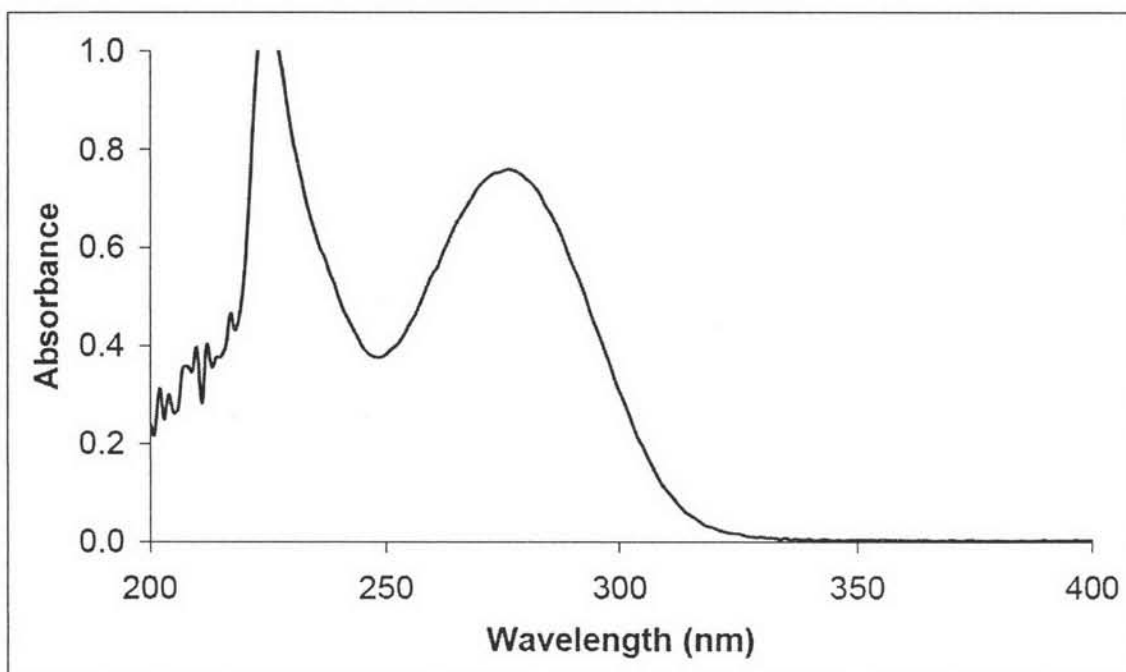


Figure A9 UV-Vis spectrum of sodium diclofenac in 5.0% (w/v) NaOH/0.5M KCl solution.

Calibration Curve of Sodium Diclofenac

The concentration versus absorbance of sodium diclofenac in 0.1 N HCl (pH 1.2), phosphate buffer saline pH 6.6 and pH 7.4, NaOH solutions (2.5-7.5% (w/v)) and 5.0% (w/v) NaOH/KCl (0.1-0.5M) at 276 nm are presented in Tables A1-A9. The standard curves of sodium diclofenac in these dissolution media are illustrated in Figures A10-A18.

Table A1 Absorbance of sodium diclofenac in 0.1N HCl determined at 276 nm

Concentration (ppm)	Absorbance
2	0.060
4	0.112
6	0.163
8	0.220
10	0.273

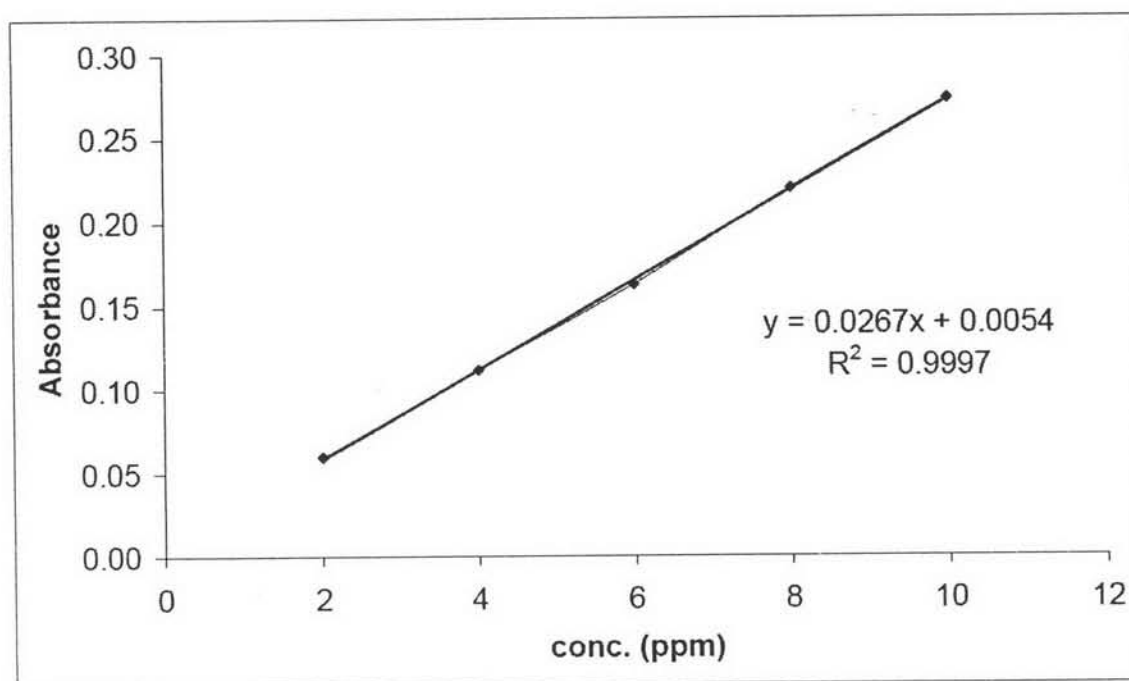


Figure A10 Calibration curve of sodium diclofenac in 0.1N HCl at 276 nm.

Table A2 Absorbance of sodium diclofenac in phosphate buffer saline pH 6.6 determined at 276 nm

Concentration (ppm)	Absorbance
2	0.066
4	0.131
6	0.193
8	0.257
10	0.318

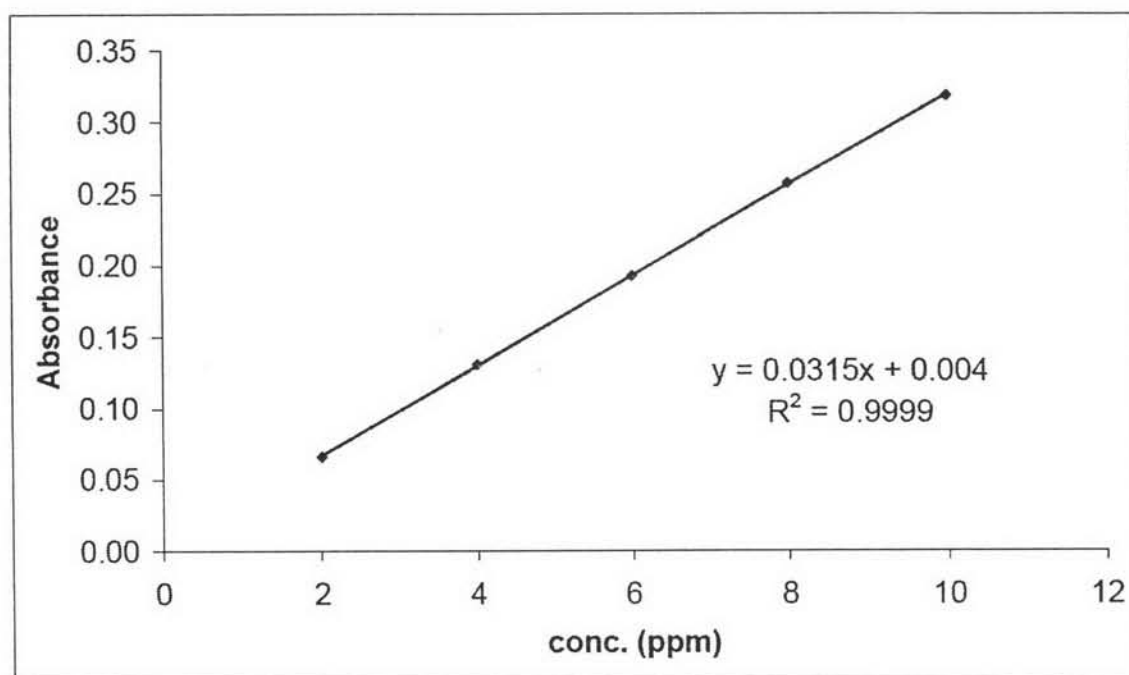


Figure A11 Calibration curve of sodium diclofenac in phosphate buffer saline pH 6.6 at 276 nm.

Table A3 Absorbance of sodium diclofenac in phosphate buffer saline pH 7.4 determined at 276 nm

Concentration (ppm)	Absorbance
5	0.169
10	0.335
15	0.496
20	0.656
25	0.816
30	0.990
35	1.149

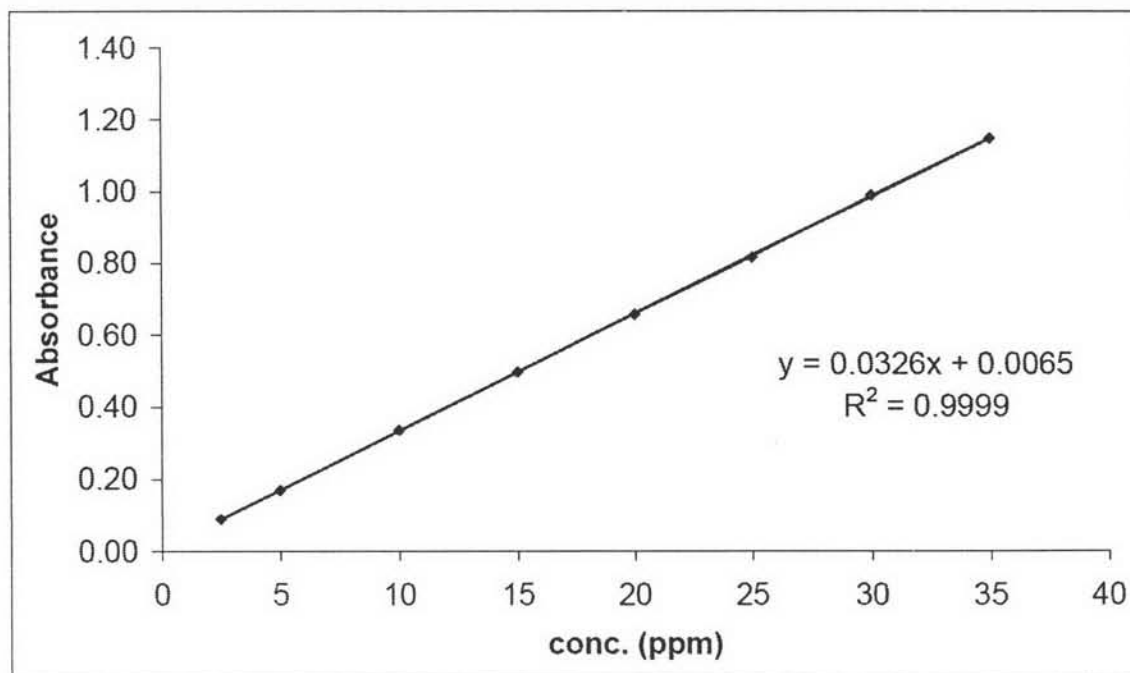


Figure A12 Calibration curve of sodium diclofenac in phosphate buffer saline pH 7.4 at 276 nm.

Table A4 Absorbance of sodium diclofenac in 2.5% (w/v) NaOH solution determined at 276 nm

Concentration (ppm)	Absorbance
5	0.165
10	0.326
20	0.642
30	0.944
40	1.257

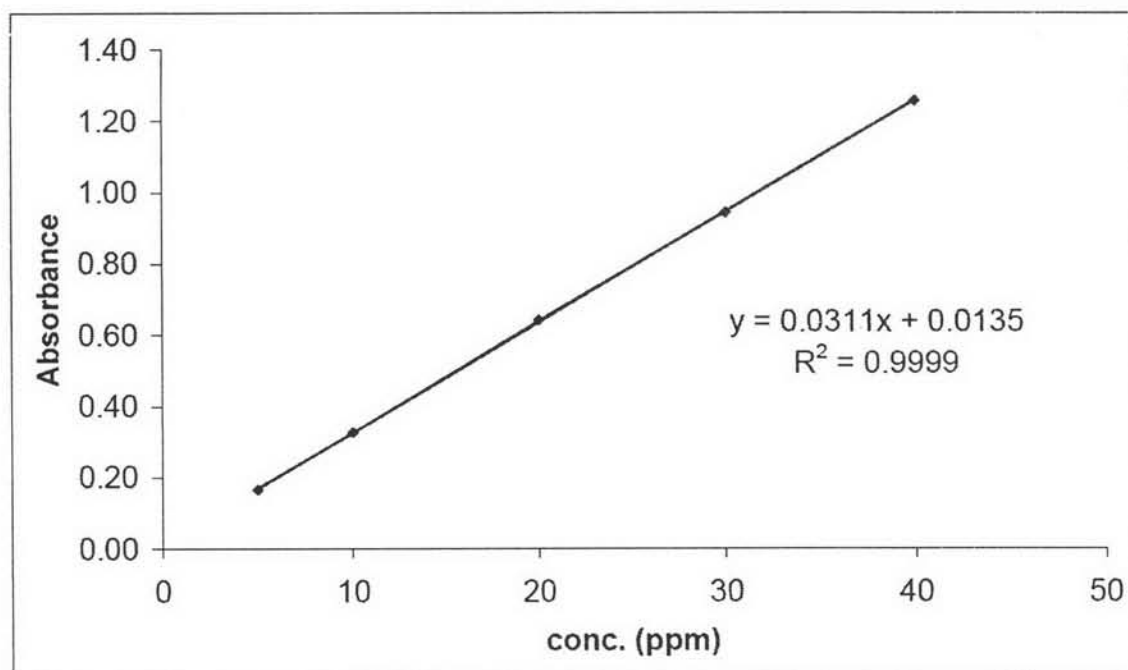


Figure A13 Calibration curve of sodium diclofenac in 2.5% (w/v) NaOH solution at 276 nm.

Table A5 Absorbance of sodium diclofenac in 5.0% (w/v) NaOH solution determined at 276 nm

Concentration (ppm)	Absorbance
5	0.165
10	0.343
20	0.677
30	1.010
40	1.347

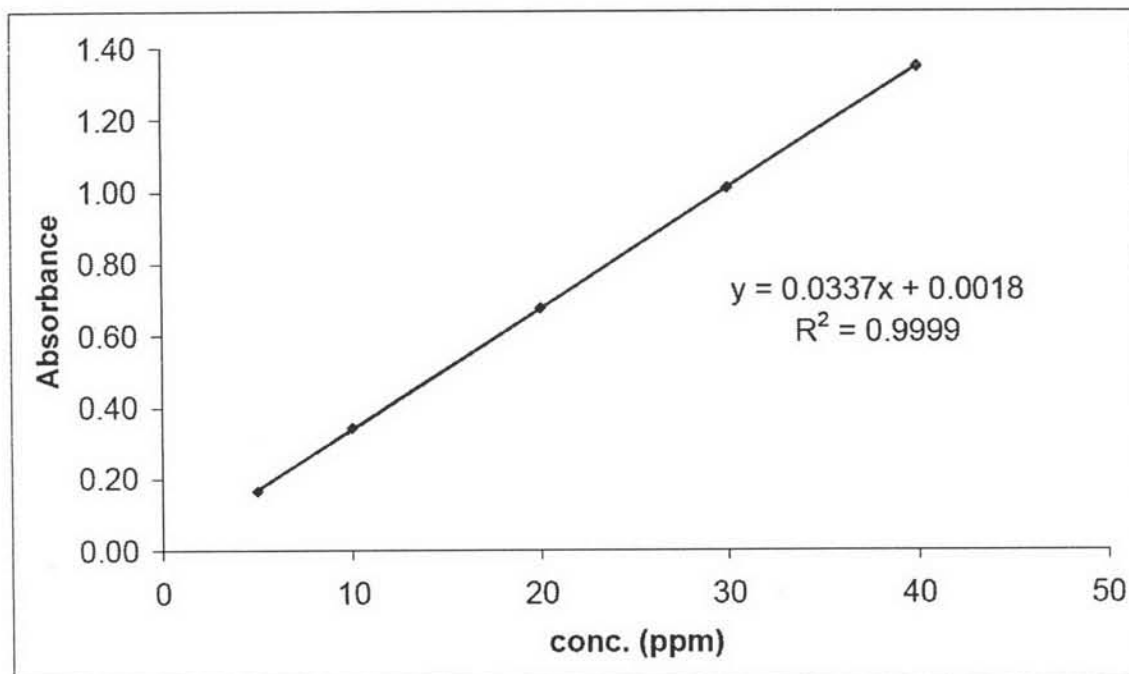


Figure A14 Calibration curve of sodium diclofenac in 5.0% (w/v) NaOH solution at 276 nm.

Table A6 Absorbance of sodium diclofenac in 7.5% (w/v) NaOH solution determined at 276 nm

Concentration (ppm)	Absorbance
5	0.178
10	0.344
20	0.667
30	0.990
40	1.301

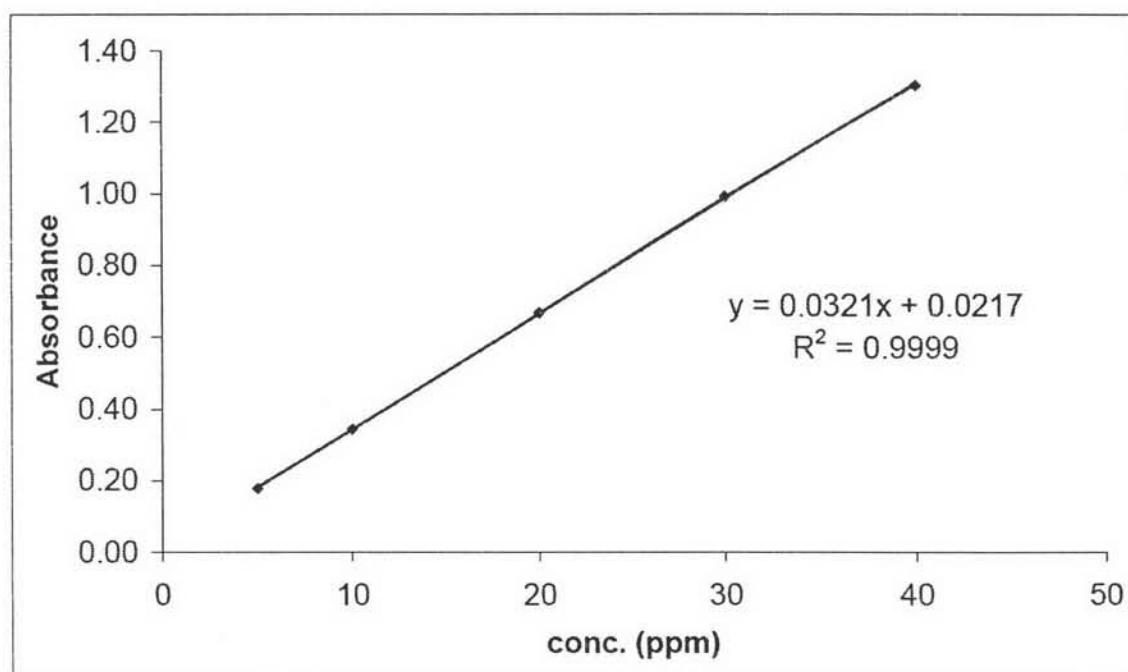


Figure A15 Calibration curve of sodium diclofenac in 7.5% (w/v) NaOH solution at 276 nm.

Table A7 Absorbance of sodium diclofenac in 5.0% (w/v) NaOH/0.1M KCl solution determined at 276 nm

Concentration (ppm)	Absorbance
5	0.174
10	0.334
20	0.676
30	1.001
40	1.369

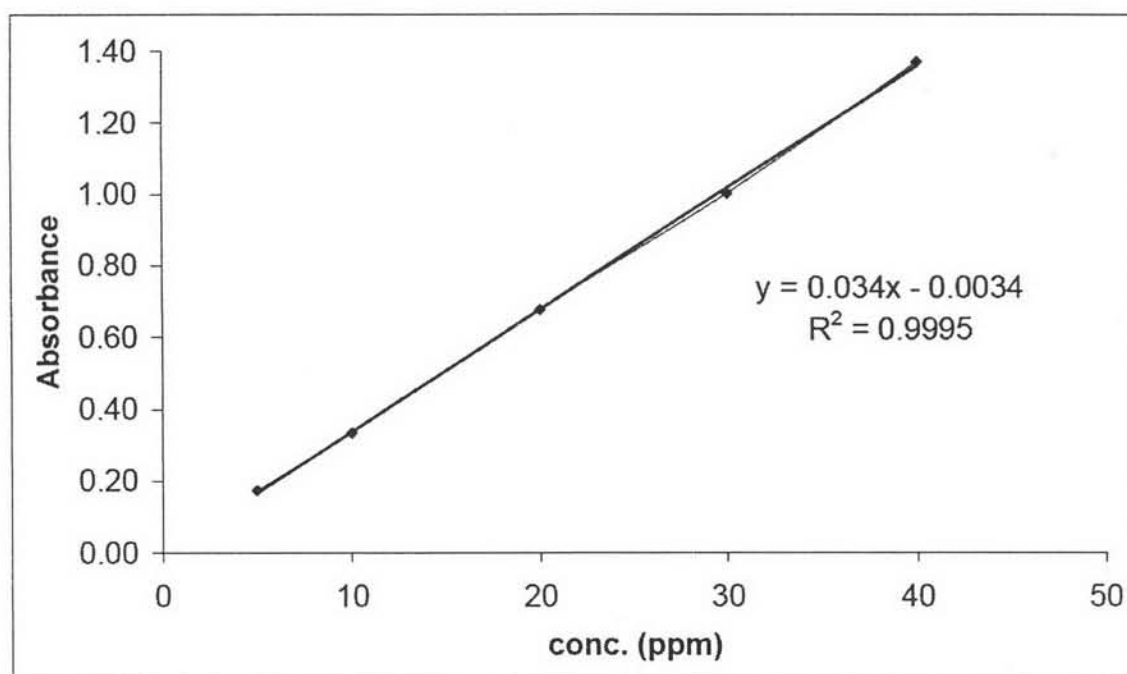


Figure A16 Calibration curve of sodium diclofenac in 5.0% (w/v) NaOH/0.1M KCl solution at 276 nm.

Table A8 Absorbance of sodium diclofenac in 5.0% (w/v) NaOH/0.3M KCl solution determined at 276 nm

Concentration (ppm)	Absorbance
5	0.181
10	0.345
20	0.658
30	0.998
40	1.342

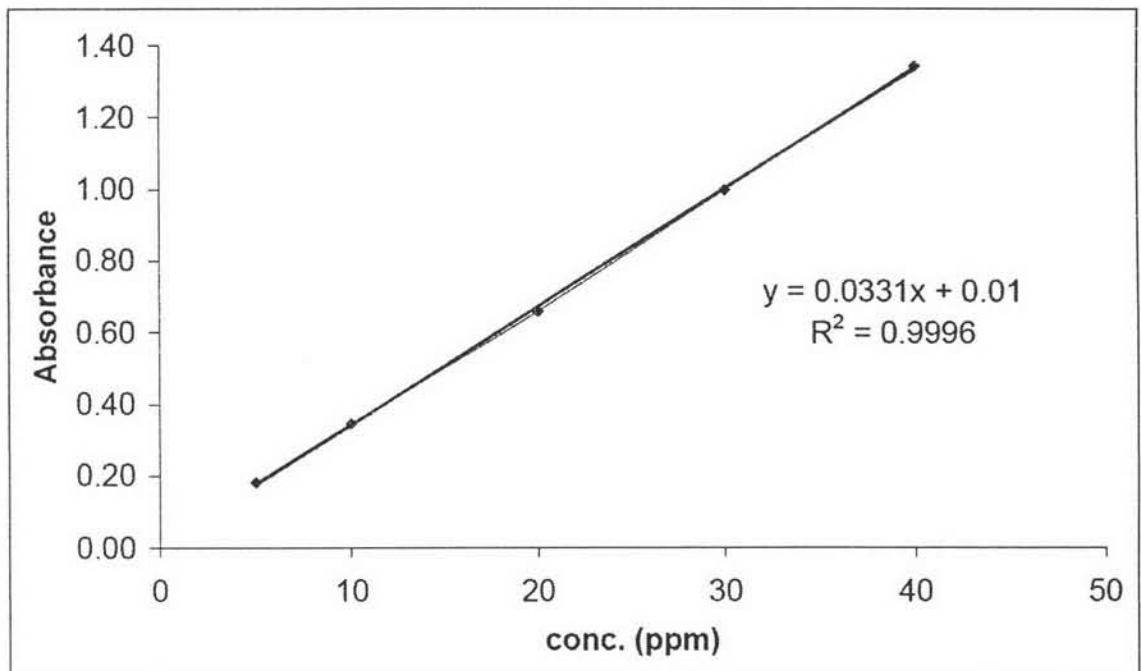


Figure A17 Calibration curve of sodium diclofenac in 5.0% (w/v) NaOH/0.3M KCl solution at 276 nm.

Table A9 Absorbance of sodium diclofenac in 5.0% (w/v) NaOH/0.5M KCl solution determined at 276 nm

Concentration (ppm)	Absorbance
5	0.182
10	0.337
20	0.652
30	1.002
40	1.297

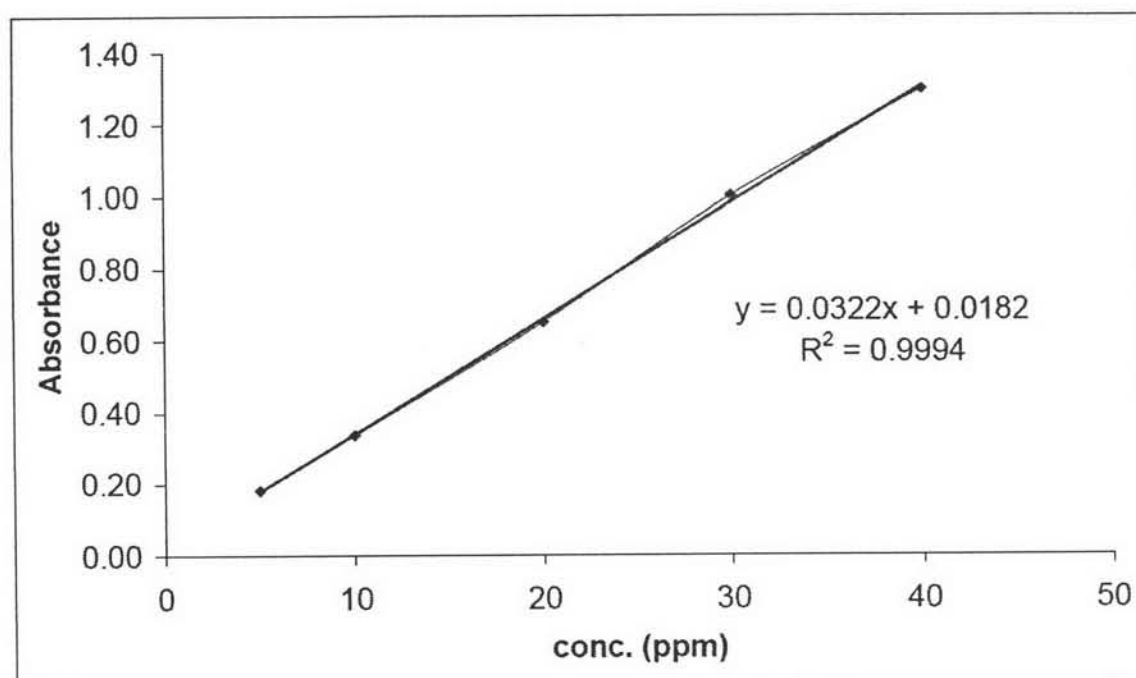


Figure A18 Calibration curve of sodium diclofenac in 5.0% (w/v) NaOH/0.5M KCl solution at 276 nm.

APPENDIX B

Swelling Ratio

Table B1 The swelling ratio of the chitosan/carrageenan (CS/CR : 2/1) bead with 5% (w/v) DFNa content in three dissolution systems

Time (hr:min)	pH 1.2 system		pH 7.4 system		pH-alternating system	
	Swelling ratio	S.D.	Swelling ratio	S.D.	Swelling ratio	S.D.
0:00	1.00	0.00	1.00	0.00	1.00	0.00
0:05	0.97	0.12	1.03	0.10	1.01	0.02
0:10	1.01	0.10	1.06	0.06	1.01	0.02
0:15	1.00	0.08	1.19	0.02	1.05	0.00
0:20	1.01	0.10	1.19	0.02	1.05	0.00
0:30	0.95	0.10	1.22	0.07	1.06	0.01
0:45	0.94	0.08	1.21	0.05	1.10	0.04
1:00	0.95	0.07	1.18	0.03	1.10	0.01
1:15	0.96	0.06	1.19	0.05	1.08	0.02
1:30	0.97	0.07	1.16	0.02	1.06	0.01
2:00	0.95	0.10	1.14	0.01	1.05	0.00
2:30	0.94	0.08	1.16	0.02	0.93	0.06
3:00	0.94	0.08	1.16	0.02	0.93	0.06
4:00	0.94	0.08	1.16	0.02	0.96	0.01
5:00	0.95	0.07	1.16	0.02	0.96	0.01

APPENDIX C

Percentage of Drug Release and Release Rate

Table C1 Percentage of DFNa release from commercial products and the beads from formulation A-S in pH-alternating method

Time (hr:min)	Voltaren® SR tablet		Subsyde® CR capsule		Formulation A		Formulation B		Formulation C	
	%drug release	S.D.	%drug release	S.D.	%drug release	S.D.	%drug release	S.D.	%drug release	S.D.
0:00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00
0:40	0.49	0.19	1.29	0.34	14.33	1.19	14.36	1.41	13.98	0.30
1:20	0.42	0.12	1.54	0.30	13.14	0.82	13.17	1.01	14.16	0.39
2:00	0.36	0.12	1.61	0.06	13.08	0.57	13.10	0.77	13.81	0.33
2:15	0.42	0.06	2.03	0.25	15.01	0.92	15.04	1.14	16.36	0.49
2:30	0.42	0.06	6.08	0.52	15.45	1.04	15.48	1.26	16.71	0.20
2:45	0.42	0.06	11.90	0.56	15.95	1.06	15.98	1.29	17.05	0.16
3:00	0.42	0.06	16.54	0.75	16.32	0.95	16.35	1.18	17.52	0.25
3:15	0.42	0.06	22.28	0.84	33.99	0.20	33.93	0.79	35.88	0.82
3:30	43.20	6.60	30.17	0.61	43.22	0.49	43.14	0.27	43.74	1.01
3:45	110.85	6.74	39.25	0.72	45.27	0.66	45.19	0.14	49.01	1.11
4:00	120.61	6.31	48.40	0.91	46.89	1.11	46.80	0.28	52.13	1.88
4:30	110.03	0.96	67.19	1.68	50.19	2.41	50.09	1.53	57.18	2.92
5:00	113.37	2.58	83.51	1.44	51.42	2.50	51.32	1.59	60.50	1.42
5:30	112.59	0.45	90.13	0.88	51.67	2.57	51.56	1.66	62.87	1.71
6:00	114.71	0.61	91.41	1.35	52.44	2.71	52.33	1.79	64.67	1.82
6:30	110.69	2.02	92.24	1.19	52.29	2.78	52.19	1.86	66.01	1.86
7:00	110.88	1.27	93.36	1.71	52.34	2.71	52.23	1.79	66.53	2.36
7:30	114.65	1.46	93.88	1.46	52.48	2.78	52.37	1.86	67.14	2.41
8:00	114.65	1.46	94.34	1.11	52.48	2.52	52.37	1.60	67.48	2.47
24:00	114.65	1.46	98.13	1.03	52.98	3.35	52.86	2.42	68.07	1.97

Table C1 (continued) Percentage of DFNa release from commercial products and the beads from formulation A-S in pH-alternating method

Time (hr:min)	Formulation D		Formulation I		Formulation J		Formulation K		Formulation L	
	%drug release	S.D.	%drug release	S.D.	%drug release	S.D.	%drug release	S.D.	%drug release	S.D.
0:00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00
0:40	15.18	1.64	7.24	0.37	5.89	0.29	5.69	0.34	5.29	0.42
1:20	12.99	0.96	7.08	0.33	5.95	0.48	5.71	0.20	5.07	0.48
2:00	13.24	1.37	6.75	0.15	5.98	0.59	5.79	0.25	4.78	0.46
2:15	15.52	1.30	7.56	0.18	6.95	0.54	6.62	0.26	5.94	0.67
2:30	16.25	1.32	8.08	0.21	7.44	0.65	7.08	0.41	6.19	0.62
2:45	16.83	1.40	8.44	0.32	7.57	0.56	7.31	0.35	6.37	0.58
3:00	17.27	1.38	8.61	0.22	7.69	0.64	7.43	0.33	6.39	0.58
3:15	29.95	2.74	16.18	0.96	23.07	2.73	19.60	2.88	16.30	0.69
3:30	38.26	2.58	22.07	1.57	29.13	3.45	25.26	2.74	22.74	1.61
3:45	44.95	1.97	26.44	2.60	33.01	3.41	34.64	2.41	26.95	2.03
4:00	48.10	1.69	30.16	2.98	35.99	3.30	38.45	1.31	31.09	2.06
4:30	52.02	1.64	36.49	3.22	40.21	3.15	41.23	3.12	43.17	2.87
5:00	56.18	2.37	42.28	3.80	43.63	3.18	45.14	2.81	51.69	1.05
5:30	56.78	2.17	46.23	3.94	46.10	3.22	49.28	2.68	57.18	0.64
6:00	58.04	2.48	50.79	3.73	48.65	3.23	49.92	1.32	61.45	0.01
6:30	57.25	1.09	54.43	4.20	50.31	3.14	52.10	2.70	64.87	0.92
7:00	57.74	0.77	57.61	3.39	51.81	3.27	58.49	3.42	66.93	1.46
7:30	58.96	0.02	65.42	3.67	53.50	3.50	63.78	3.37	68.67	1.73
8:00	58.66	0.33	67.65	2.45	54.90	3.63	64.30	1.89	69.74	1.57
24:00	58.16	0.10	87.39	0.17	77.33	0.28	75.27	0.85	89.78	4.42

Table C1 (continued) Percentage of DFNa release from commercial products and the beads from formulation A-S in pH-alternating method

Time (hr:min)	Formulation O		Formulation P		Formulation Q		Formulation R		Formulation S	
	%drug release	S.D.	%drug release	S.D.	%drug release	S.D.	%drug release	S.D.	%drug release	S.D.
0:00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00
0:40	4.31	0.30	4.90	0.25	3.78	0.18	4.02	0.13	0.95	0.07
1:20	4.33	0.12	4.84	0.12	3.76	0.10	3.71	0.08	1.40	0.10
2:00	4.47	0.38	5.13	0.33	3.82	0.14	3.66	0.03	1.87	0.07
2:15	5.78	0.70	7.28	0.47	4.76	0.14	9.51	1.51	1.34	0.08
2:30	6.22	0.61	7.93	0.85	5.19	0.09	11.83	1.69	1.42	0.08
2:45	6.40	0.66	8.46	0.88	5.33	0.16	12.97	2.18	1.47	0.07
3:00	6.47	0.66	8.61	0.97	5.70	0.27	13.94	2.72	1.53	0.06
3:15	20.25	1.64	21.68	0.43	17.26	2.43	25.54	1.61	2.59	0.20
3:30	27.48	1.80	26.85	0.25	22.29	2.74	30.52	1.69	5.30	0.20
3:45	31.66	1.30	30.57	0.73	27.00	1.15	33.73	1.67	8.43	0.31
4:00	34.00	1.32	32.85	0.56	29.90	1.03	36.43	1.44	11.59	0.27
4:30	37.06	1.26	36.09	0.63	34.13	1.38	39.59	1.31	17.14	0.17
5:00	39.50	1.31	38.45	0.52	38.66	1.44	42.59	1.22	21.86	0.50
5:30	41.33	1.34	40.20	0.33	40.52	1.49	44.64	1.25	26.54	0.55
6:00	42.40	1.39	41.88	0.21	42.56	1.61	46.42	1.21	30.58	0.69
6:30	44.21	1.46	42.96	0.22	44.57	1.80	47.92	1.02	34.26	0.52
7:00	45.08	1.58	44.64	0.28	46.06	1.92	49.53	1.39	38.28	0.62
7:30	45.90	1.63	45.62	0.39	47.54	2.16	50.54	1.18	41.71	0.73
8:00	46.74	1.77	46.44	0.62	49.17	2.26	51.60	1.14	44.89	0.50
24:00	50.11	3.01	52.45	1.65	57.53	3.30	56.43	2.02	69.93	2.82

Table C2 The release rate of commercial products and the beads from formulation A-S in pH-alternating method

Mean Time (hr:min)	Release Rate (%/hour)				
	Voltaren® SR	Subsyde® CR	Formulation A	Formulation B	Formulation C
0:00	0.00	0.00	0.00	0.00	0.00
0:20	0.74	1.94	21.50	21.54	20.67
1:00	-0.11	0.37	-1.78	-1.79	0.00
1:40	-0.10	0.11	-0.09	-0.09	0.54
2:30	0.07	14.92	3.24	3.25	3.10
3:15	85.89	27.28	53.80	53.59	50.89
3:45	154.82	36.45	7.34	7.32	11.44
4:15	-21.17	37.59	6.60	6.57	8.49
4:45	6.67	32.63	2.47	2.46	10.58
5:15	-1.56	13.24	0.49	0.49	3.12
5:45	4.25	2.03	1.54	1.53	4.33
6:15	-8.04	2.19	-0.28	-0.28	3.45
6:45	0.38	2.23	0.09	0.09	2.60
7:15	7.53	1.04	0.28	0.28	0.35
7:45	0.00	0.92	0.00	0.00	1.13
16:00	0.00	0.24	0.03	0.03	0.02

Table C2 (continued) The release rate of commercial products and the beads from formulation A-S in pH-alternating method

Mean Time (hr:min)	Release Rate (%/hour)				
	Formulation D	Formulation I	Formulation J	Formulation K	Formulation L
0:00	0.00	0.00	0.00	0.00	0.00
0:20	22.77	10.86	8.84	8.54	7.93
1:00	-3.27	-0.24	0.09	0.03	-0.33
1:40	0.36	-0.50	0.05	0.11	-0.43
2:30	4.04	1.86	1.71	1.64	1.62
3:15	41.98	33.48	42.87	39.96	32.69
3:45	19.67	17.20	13.72	17.38	16.70
4:15	7.84	12.79	8.45	10.26	24.17
4:45	8.34	11.37	6.83	7.81	17.04
5:15	1.18	7.43	4.95	8.28	10.98
5:45	2.54	8.83	5.10	6.23	8.54
6:15	-1.58	6.63	3.31	3.82	6.83
6:45	0.97	6.20	3.00	3.10	4.12
7:15	2.44	4.27	3.38	8.17	3.49
7:45	-0.60	5.55	2.82	0.90	2.14
16:00	-0.03	1.24	0.88	0.54	0.89

Table C2 (continued) The release rate of commercial products and the beads from formulation A-S in pH-alternating method

Mean Time (hr:min)	Release Rate (%/hour)				
	Formulation O	Formulation P	Formulation Q	Formulation R	Formulation S
0:00	0.00	0.00	0.00	0.00	0.00
0:20	6.47	7.35	5.68	6.03	1.43
1:00	0.02	-0.08	-0.03	-0.47	0.67
1:40	0.21	0.43	0.08	-0.08	0.70
2:30	2.01	3.48	1.88	10.28	-0.34
3:15	42.00	36.48	33.19	33.15	7.54
3:45	13.05	12.00	15.22	11.82	12.59
4:15	6.12	6.49	8.45	6.33	11.09
4:45	4.89	4.72	9.06	6.00	9.44
5:15	3.65	3.49	3.73	4.10	9.36
5:45	2.15	3.37	4.09	3.55	8.08
6:15	3.61	2.16	4.02	3.01	7.37
6:45	1.75	3.34	2.97	3.22	8.02
7:15	1.62	1.98	2.97	2.02	6.87
7:45	1.69	1.63	3.24	2.11	6.36
16:00	0.21	0.38	0.52	0.30	1.57

VITA

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