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APPENDICES

APPENDIX A

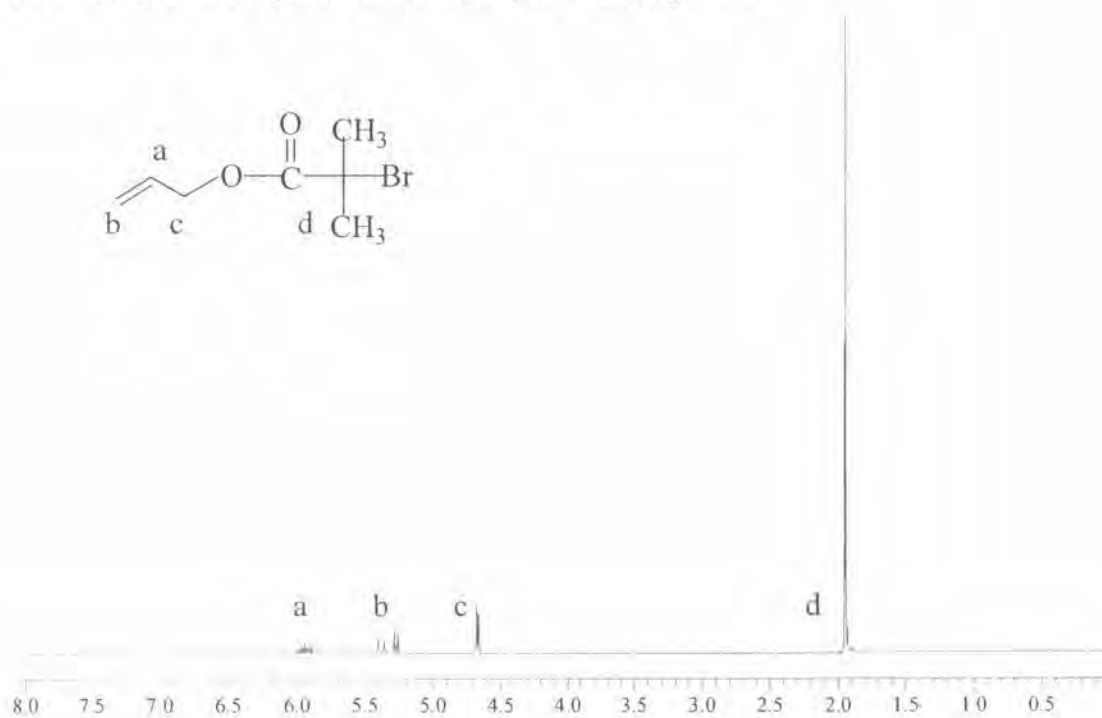
Proton nuclear magnetic resonance (^1H NMR) spectra

Figure A-1 ^1H -NMR spectrum (400 MHz, CDCl_3) of 2-bromo-2-methylpropionic acid allyl ester (1)

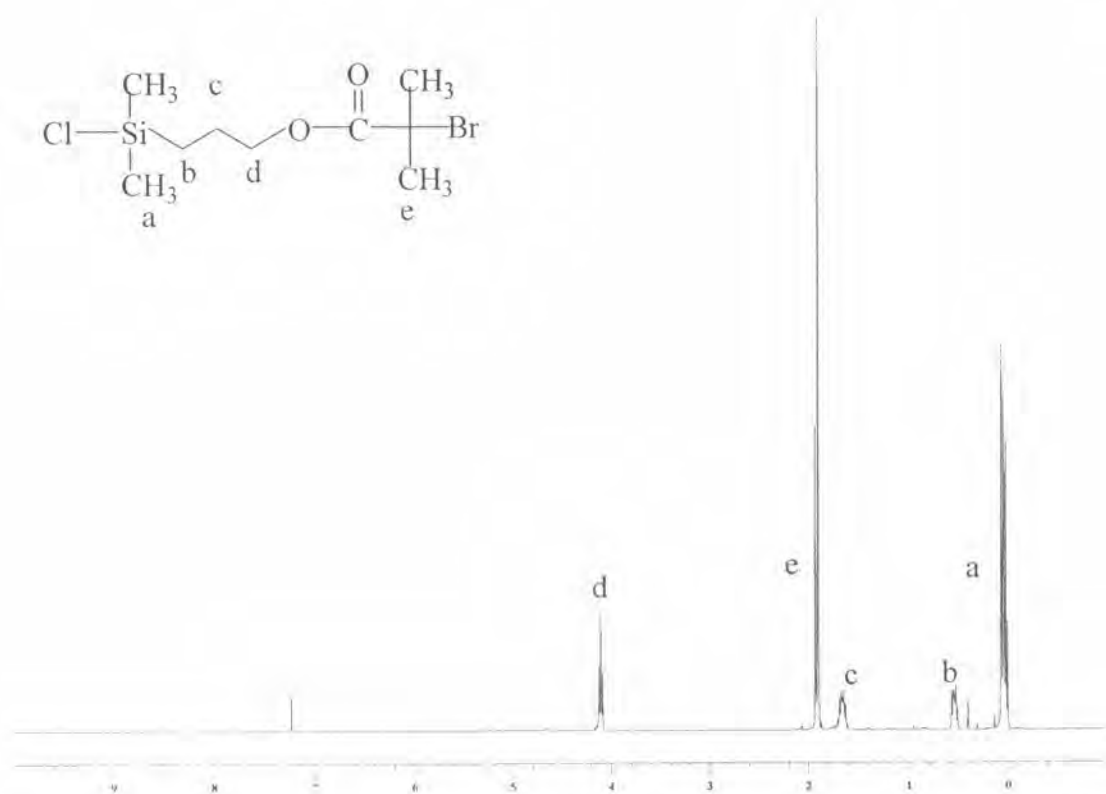


Figure A-2 ¹H-NMR spectrum (400 MHz, CDCl₃) of 2-bromo-2-methylpropionic acid 3-(dimethylchloro silanyl) propyl ester (**2**)

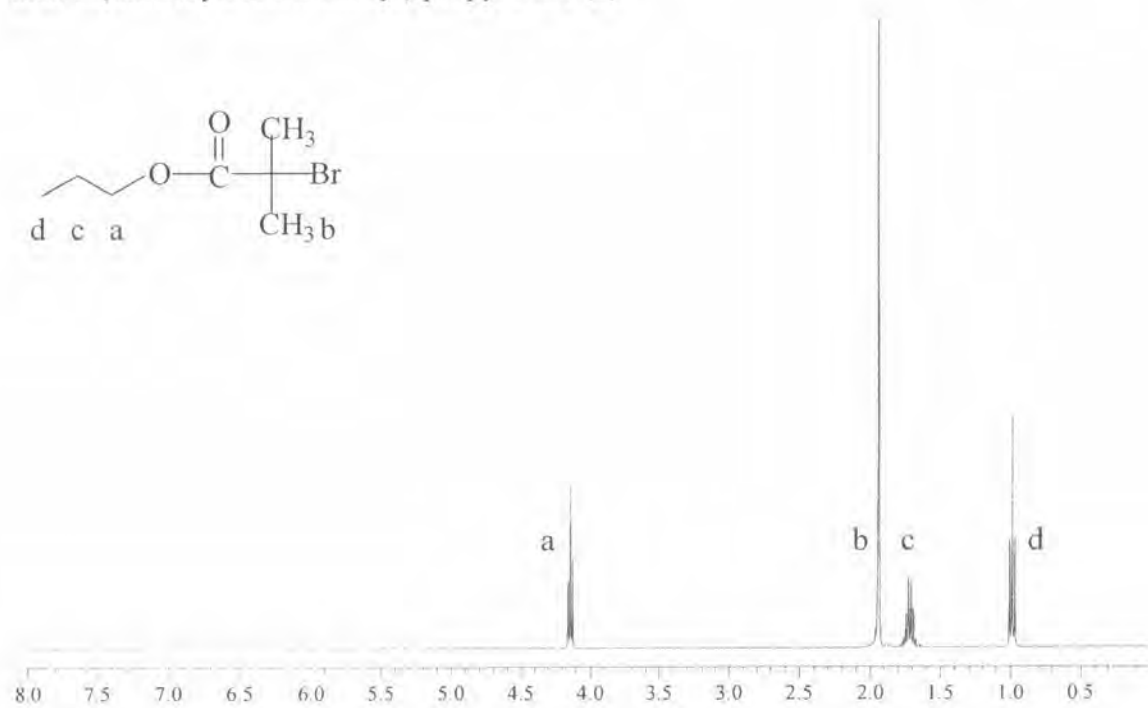


Figure A-3 ¹H-NMR spectrum (400 MHz, CDCl₃) of 2-bromo-2-methylpropionic acid propyl ester (**3**)

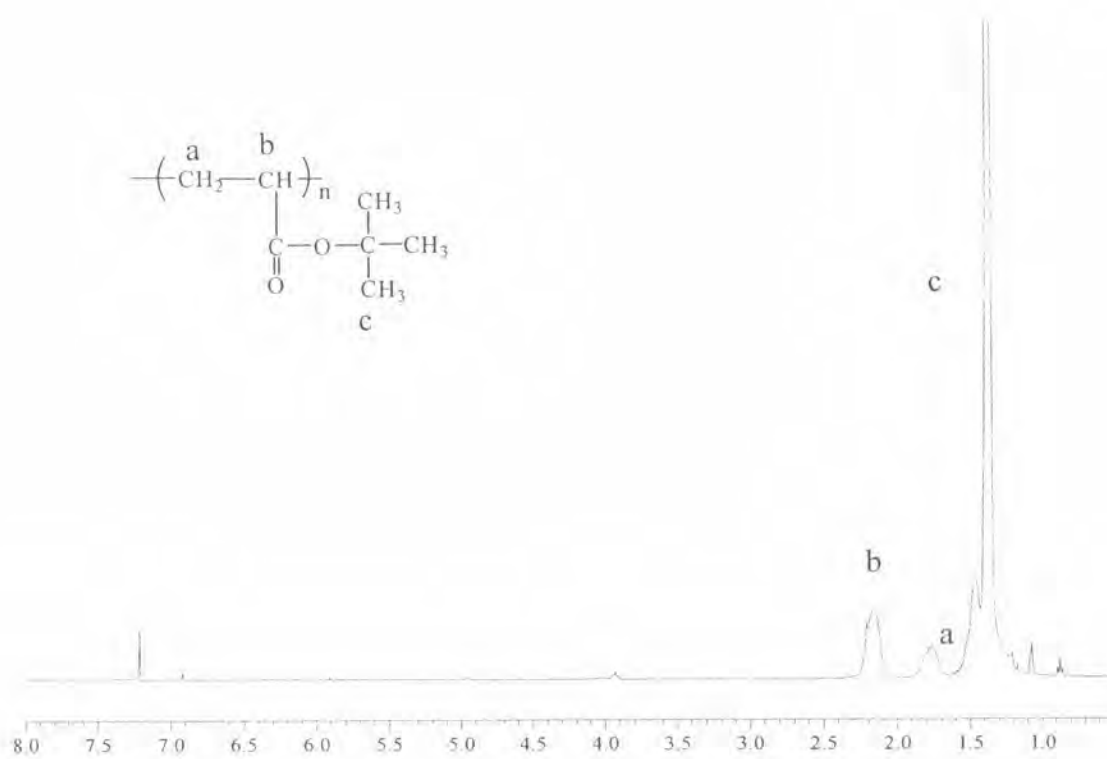


Figure A-4 $^1\text{H-NMR}$ spectrum (400 MHz, CDCl_3) of Pt-BA

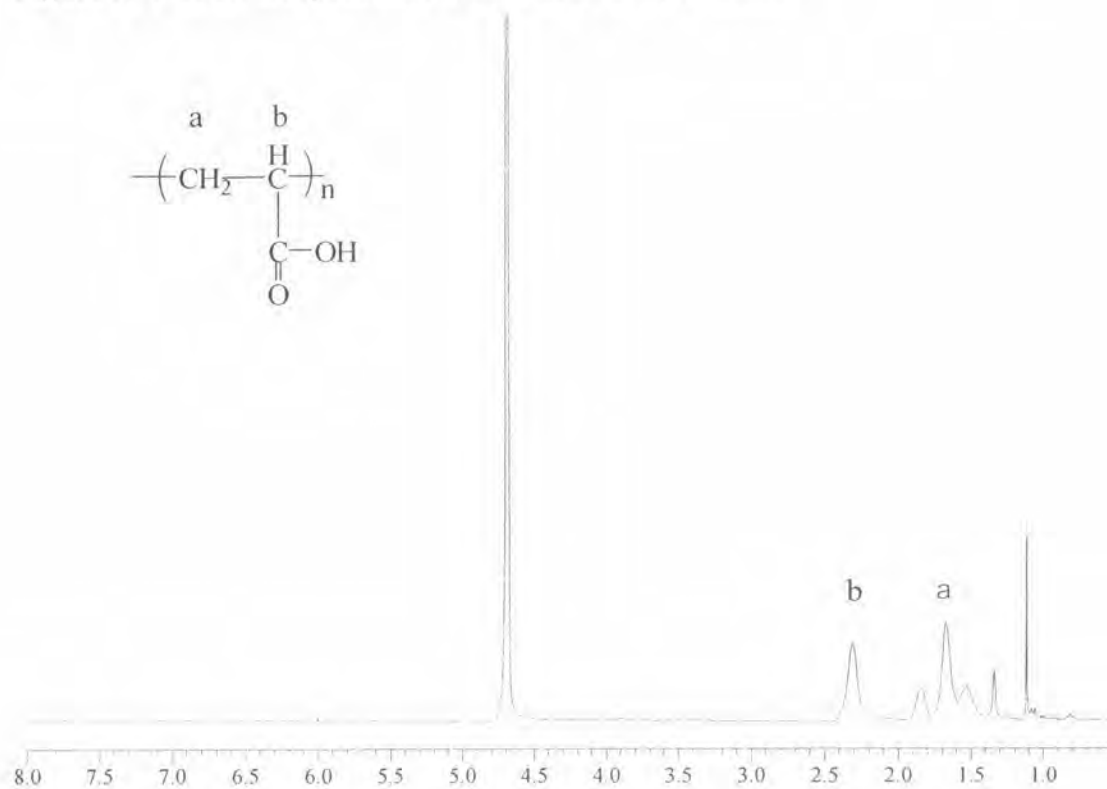


Figure A-5 The $^1\text{H-NMR}$ spectrum (400 MHz, D_2O) of PAA

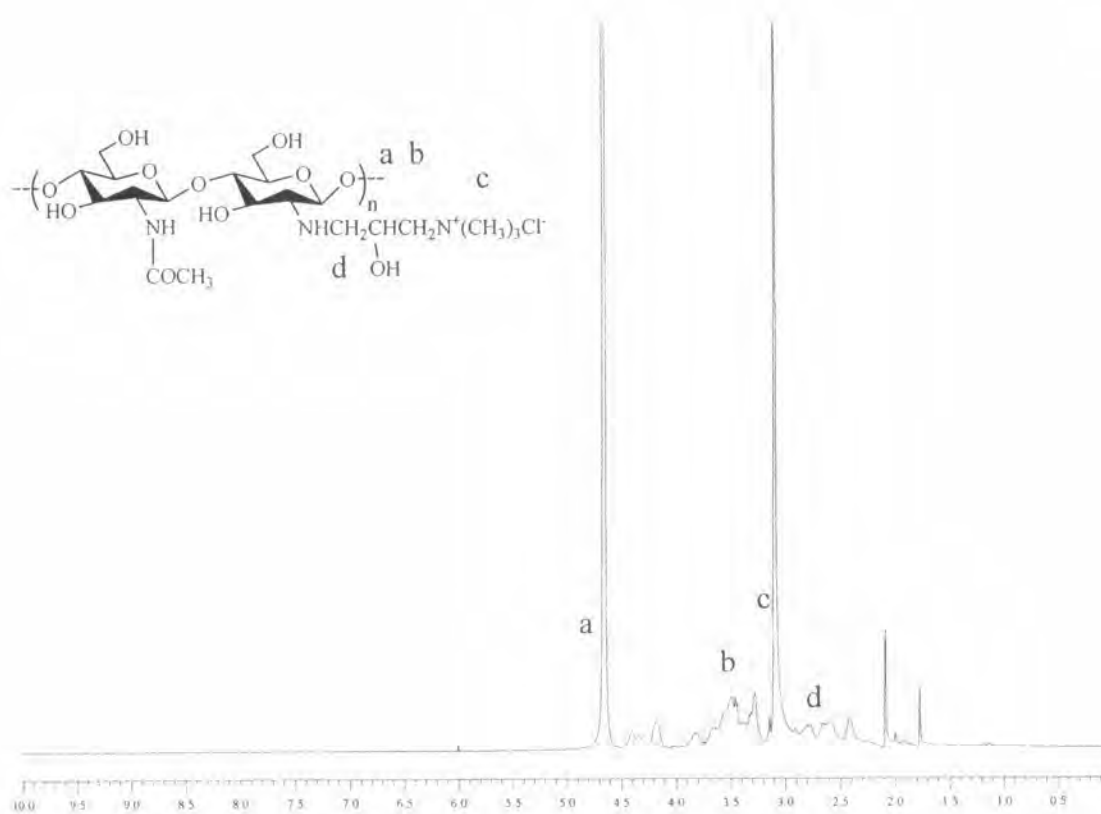


Figure A-6 $^1\text{H-NMR}$ spectrum (400 MHz, D_2O) of HTACC

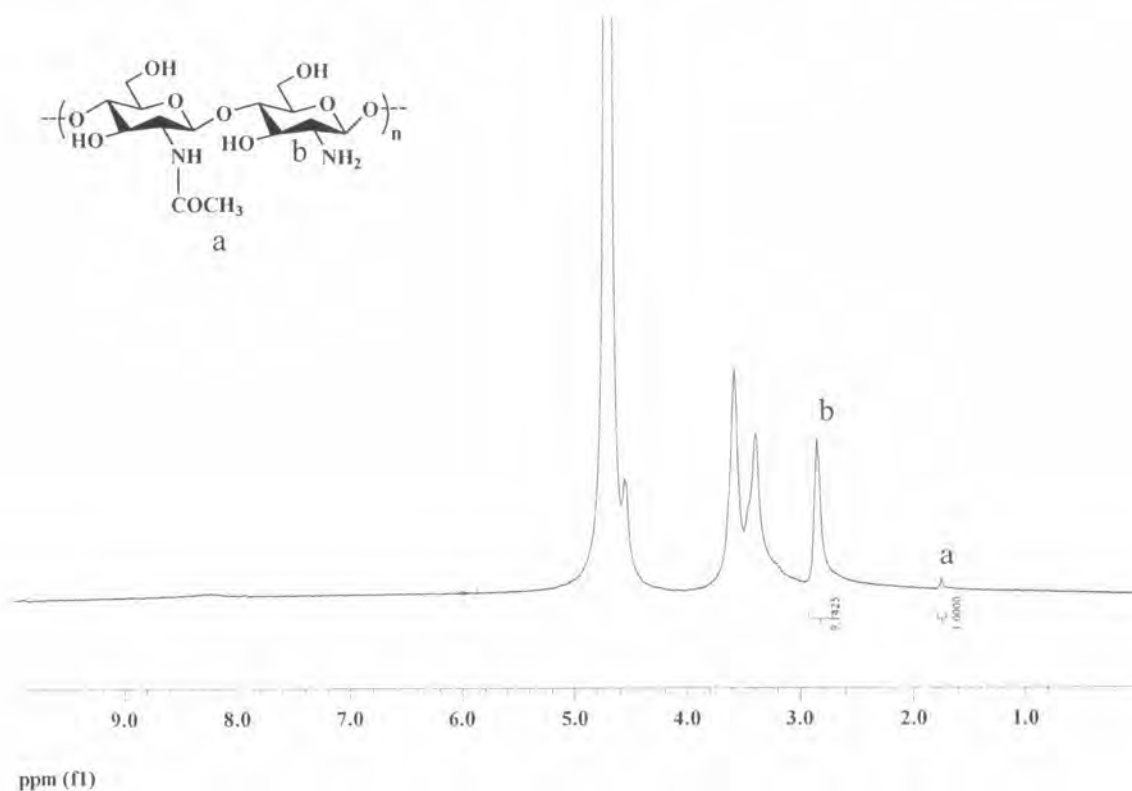


Figure A-7 $^1\text{H-NMR}$ spectrum (400 MHz, D_2O) of Chitosan \overline{M}_w 100,000, 96% DD

Table A-1 Information from ^1H NMR spectrum of chitosan used in this study

	δ (ppm)	Integration	Relative Amount of units
$-\text{CHNH}_2$ of GlcN	2.80	9.1	9.1/1
$-\text{CH}_3$ of GlcNAc	1.75	1.0	1.0/3

Considering the total repeating units in chitosan as 100%, %DD of 96 % was calculated based on the data in Table 4.2 using the following equation.

$$\begin{aligned} \%DD &= \frac{\text{Amount of GlcN unit in chitosan}}{\text{The total amount of GlcN and GlcNAc units in chitosan}} \times 100 \\ &= \frac{(9.1/1)}{(1.0/3) + (9.1/1)} \times 100\% = 96\% \end{aligned}$$

APPENDIX B

B. Data corresponding to the plots in Chapter IV

Table B-1 Average molecular weight and molecular weight distribution of *Pt*-BA brushes analyzed by GPC as a function of time (DP =100, 200)

Time (h)	GPC data					
	DP 100			DP 200		
	\overline{M}_n	\overline{M}_w	$\overline{M}_w/\overline{M}_n$	\overline{M}_n	\overline{M}_w	$\overline{M}_w/\overline{M}_n$
3	1990	2157	1.10	3058	3302	1.08
6	3869	4063	1.05	5499	5992	1.09
12	6834	7309	1.07	11765	12467	1.06
24	11965	12563	1.05	22101	24976	1.13

Table B-2 The thickness of linear *Pt*-BA brushes calculated from ellipsometric data as a function of average molecular weight of *Pt*-BA (DP = 100)

Time (h)	Thickness (nm)	GPC data (DP 100)		
		\overline{M}_n	\overline{M}_w	$\overline{M}_w/\overline{M}_n$
3	NA	2047	2251	1.10
6	3.11	4592	4816	1.05
12	4.47	6612	7273	1.10
24	6.86	12237	13338	1.09

Table B-3 The thickness of linear *Pt*-BA brushes calculated from ellipsometric data as a function of average molecular weight of *Pt*-BA (DP = 200)

Time (h)	Thickness (nm)	GPC data (DP 200)		
		\overline{M}_n	\overline{M}_w	$\overline{M}_w/\overline{M}_n$
3	2.70	3058	3302	1.08
6	4.21	5769	6345	1.10
12	7.27	12465	13586	1.09
24	10.20	20101	21307	1.06

Table B-4 Advancing (θ_A) and receding (θ_R) water contact angles of *Pt*-BA (DP = 100, 200) as a function of time

Time (h)	DP = 100		DP = 200	
	θ_A (°)	θ_R (°)	θ_A (°)	θ_R (°)
0 ^a	68 ± 2.4	56 ± 2.2	67 ± 2.2	58 ± 2.0
3	75 ± 2.1	66 ± 2.5	75 ± 2.1	67 ± 1.7
6	77 ± 1.7	71 ± 2.2	83 ± 1.7	69 ± 2.4
12	85 ± 1.5	78 ± 1.9	86 ± 2.1	75 ± 2.1
24	89 ± 2.0	80 ± 1.8	92 ± 1.8	80 ± 2.2

^a is advancing and receding water contact angle of α -bromoisobutyl ester initiator on silicon surface.

Table B-5 UV-Vis absorbance at 640 nm of CHI/ PPCDA vesicles multilayer (\overline{M}_n of CHI =100,000 g/mole) on glass-tethered PAA brushes having $\overline{M}_n = 9125$ and 12661 as a function of number of layer

Number of layers	UV-Vis absorbance at 640 nm	
	$\overline{M}_n = 9125$	$\overline{M}_n = 12661$
6	0.045	0.086
8	0.084	0.161
10	0.091	0.171
12	0.110	0.154
14	0.120	0.169
16	0.169	0.219

Table B-6 UV-Vis absorbance at 640 nm of a bilayer CHI/PPCDA vesicles (\overline{M}_n of CHI = 743,000 g/mole) on glass-tethered PAA brushes having $\overline{M}_n = 8343$ and 11760 as a function of deposition time

Deposition Time (min)	UV-Vis absorbance at 640 nm	
	$\overline{M}_n = 8343$	$\overline{M}_n = 11760$
5	0.25	0.34
10	0.29	-
15	-	0.5
20	0.36	0.62
30	-	0.91

Table B-7 Ellipsometric thickness of CHI/PAA multilayer on silicon-tethered PAA brushes having $\bar{M}_n = 24,797$ and 13,835 as a function of number of layer

Number of layers	Thickness (nm)	
	$\bar{M}_n = 13,835$	$\bar{M}_n = 24,797$
0	6.70 ± 1.81	10.37 ± 0.92
1	9.95 ± 0.55	13.80 ± 1.55
2	11.90 ± 0.47	18.00 ± 0.87
3	20.98 ± 1.57	36.00 ± 1.57
4	28.65 ± 1.06	47.00 ± 1.06
5	36.78 ± 2.80	52.00 ± 2.80
6	42.34 ± 2.37	66.00 ± 2.37
7	51.00 ± 1.88	76.00 ± 1.88
8	55.30 ± 2.43	88.00 ± 2.22
9	54.97 ± 2.08	91.90 ± 1.90

Table B-8 Water contact angle of CHI/PAA multilayer on surface-tethered PAA brushes

Number of layer	Water contact angle (degree)
0	39 ± 1.1
1	57 ± 1.0
2	44 ± 3.1
3	54 ± 4.4
4	42 ± 1.0
5	54 ± 1.8
6	42 ± 1.5
7	62 ± 4.4
8	45 ± 0.9
9	58 ± 1.6
10	46 ± 1.4
11	55 ± 4.5

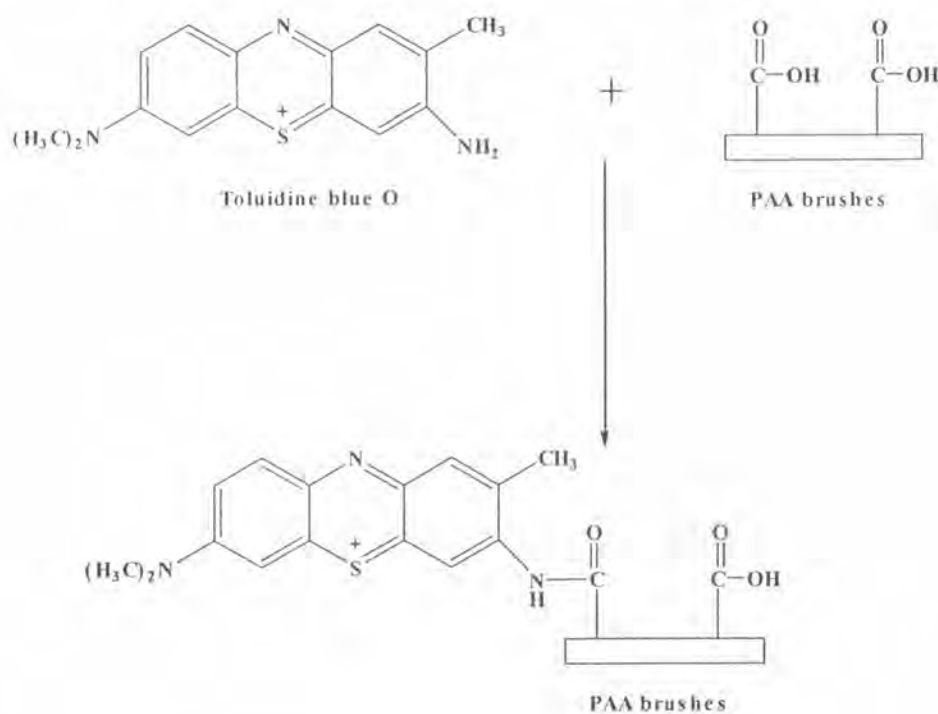
Table B-9 Amount of adsorbed albumin on CHI/PAA multilayer on PAA brushes having $\bar{M}_n = 11,092$ and 18,202

Samples	Adsorped protein ($\mu\text{g}/\text{cm}^2$) of Mutilayer film on PAA brushes substrate	
	$\bar{M}_n = 11,092$	$\bar{M}_n = 18,202$
PAA brushes	0.38 ± 0.15	0.45 ± 0.15
CHI	0.71 ± 0.14	0.88 ± 0.12
CHI/PAA	0.64 ± 0.31	0.76 ± 0.49
(CHI/PAA)CHI	0.80 ± 0.19	0.86 ± 0.34
(CHI/PAA)2	1.23 ± 0.31	1.82 ± 0.40
(CHI/PAA)2CHI	1.46 ± 0.27	1.98 ± 0.48

APPENDIX C

C-1 Toluidine blue O assay

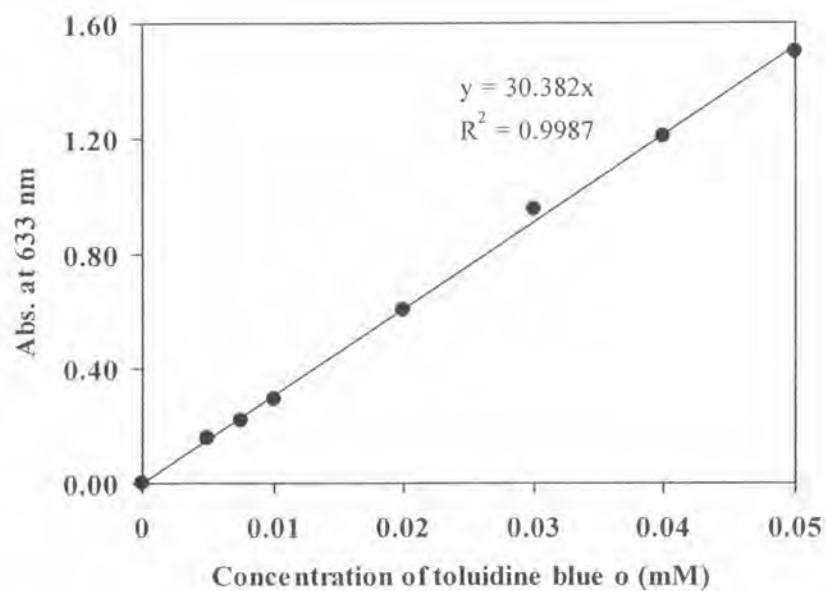
Toluidine blue O assay is a method used for determination of the amount of carboxyl groups. The carboxyl groups of PAA brushes can form a complex with toluidine blue o. The absorbance of the solution containing the desorbed complex was measured at 633 nm. The COOH content was obtained from a calibration plot of the optical density versus dye concentration which is displayed in Figure C-2.



Scheme C-1 Formation of toluidine blue O complex with carboxyl group

Table C-1 Standard Toluidine blue O solution for the calibration curve.

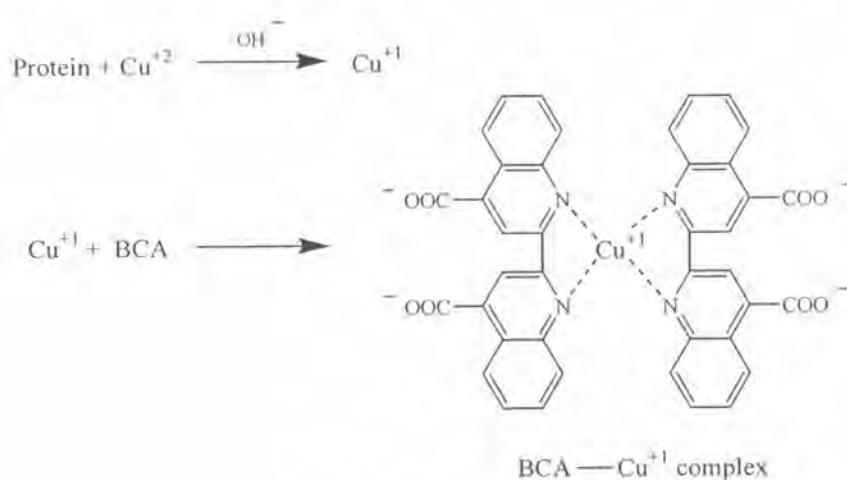
Standard	Solution pH 10 (mL)	NaOH pH 10 (mL)	Toluidine blue O conc (mM)
S ₁	1.00 toluidine blue O (0.5 mM)	9.00	0.0500
S ₂	8.00 of S ₁	2.00	0.0400
S ₃	7.50 of S ₂	2.50	0.0300
S ₄	6.70 of S ₃	3.20	0.0200
S ₅	5.00 of S ₄	5.00	0.0100
S ₆	7.50 of S ₅	2.50	0.0075
S ₇	1.00 of S ₆	9.00	0.0050
S ₈	0.00 of S ₇	10.00	0.0000

**Figure C-1** Calibration curve of UV-Vis absorbance as a function of toluidine blue O concentration

C-2 Bicinchoninic acid assay

Bicinchoninic acid assay is a method used for determination of the amount of proteins. The standard reagents used in this method are reagent A, reagent B and reagent C. Reagent A consists of an aqueous solution of $\text{Na}_2\text{tartrate}$, Na_2CO_3 , NaHCO_3 in 0.2 M NaOH , pH 11.25. Reagent B is 4% (W/V) bicinchoninic acid solution, pH 8.5. Reagent C is 4% $\text{CuSO}_4 \cdot 5\text{H}_2\text{O}$ in deionized water.

The principle of the bicinchoninic assay (BCA) relies on the formation of a Cu^{2+} -protein complex under alkaline conditions, followed by reduction of the Cu^{2+} to Cu^{1+} . The amount of reduction is proportional to protein present. It has been shown that the peptide bond is able to reduce Cu^{2+} to Cu^{1+} . BCA forms a purple-blue complex with Cu^{1+} in alkaline environments, thus providing a basis to monitor the reduction of alkaline Cu^{2+} by proteins.³⁰ Figure B-1 shows complexation between bicinchoninic acid and Cu^{1+} .



Scheme C-2. Formation of purple complex between BCA and cuprous ion generated from the biuret reaction.

Calculation of Protein Adsorption

Table C-2 Standard BSA solution for the calibration curve.

Standard	Solution (mL)	SDS (mL)	BSA conc
S ₁	0.25 of BSA (1000 (µg/mL) ^a)	4.75	50
S ₂	2.0 of S ₁	3.00	20
S ₃	2.5 of S ₂	2.5	10
S ₄	2.5 of S ₃	2.5	5
S ₅	0.5 of S ₄	4.5	0.5
S ₆	-	5.0	0.0

a : standard BSA was pipette from 1 mg/mL ampule

After reading the UV absorbance of standard BSA solution at $\lambda = 562$ nm, a calibration curve between UV absorbance versus standard BSA solution can be plotted (Figure B-2).

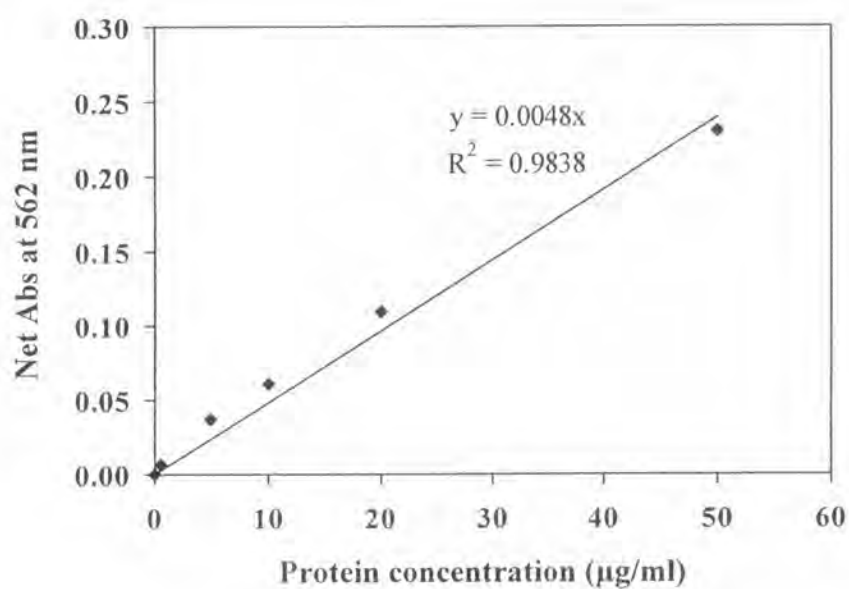


Figure C-2 Calibration curve of the amount of albumin and the absorbance obtained from BCA microassay.

The total amount of protein (P) in the original solution (2 mL) was calculated from the sampling sample (100 μL) + BCA working solution (100 μL). The protein concentration (C; $\mu\text{g}/\text{mL}$) in each well was determined from the calibration curve.

Table C-3 UV absorbance at λ_{562} , measured from sampling solution of various outermost layer

Samples	$N_{\text{net}A_{562}}$
PAA brushes \bar{M}_n 22391	0.006
(CHI/PAA) ₂	0.007
(CHI/PAA) ₂ CHI	0.013
(CHI/PAA) ₂ HTACC	0.008
(CHI/PAA) ₂ CHI/heparin	0.005

$$\text{Net Absorbance at 562 nm } (N_{\text{net}A_{562}}) = A_{562} \text{ sample} - A_{562} \text{ blank} \quad \text{C-1}$$

$$\text{Total amount of protein (P)} = \frac{C (\mu\text{g}/\text{mL}) \times 100 (\mu\text{L})}{1000 (\mu\text{L}/\text{mL})} \times \frac{1000 (\mu\text{L})}{100 (\mu\text{L})} \quad \text{C-2}$$

$$\text{Adsorbed protein/surface area } P_{\text{ads}} = P/\text{surface area (2 sides)} (\mu\text{g}/\text{cm}^2) \quad \text{C-3}$$

Example: (CHI/PAA)₂ CHI which have $N_{\text{net}A_{562}}$ 0.013 Abs. so equal 2.66 $\mu\text{g}/\text{mL}$

$$\begin{aligned} \text{From C-2 Eq. } P &= \frac{C (\mu\text{g}/\text{mL}) \times 100 (\mu\text{L}) \times 1000 (\mu\text{L})}{1000 (\mu\text{L}/\text{mL}) \times 100 (\mu\text{L})} \\ &= \frac{2.66 (\mu\text{g}/\text{mL}) \times 100 (\mu\text{L}) \times 1000 (\mu\text{L})}{1000 (\mu\text{L}/\text{mL}) \times 100 (\mu\text{L})} \\ &= 2.66 \mu\text{g} \end{aligned}$$

$$\begin{aligned} \text{From C-3 Eq. } P_{\text{ads}} &= P/\text{surface area (2 sides)} (\mu\text{g}/\text{cm}^2) \\ &= 2.66 \mu\text{g}/\{(1.1 \times 1.1)/2 \text{ cm}^2\} \\ &= 1.10 \mu\text{g}/\text{cm}^2 \end{aligned}$$

VITAE

Miss Sudarat Pookboonmee was born on August 29, 1981 in Bangkok, Thailand. She received a bachelor degree of science from Department of Chemistry, Faculty of Science, Kasetsart University, Bangkean, Bangkok, Thailand in 2004. In the same year she was admitted to a Master's Degree in Program of Petrochemistry and Polymer Science, Faculty of Science, Chulalongkorn University and completed program in 2007. Her address is Moo 5 Seekun, Donmuang, Bangkok 10210.