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APPENDIX

BUFFERS AND REAGENTS

BUFFERS:

Phosphate buffer stock solution (PB), pH 7.4

0.2 M $\text{NaH}_2\text{PO}_4 \cdot 2\text{H}_2\text{O}$, pH 4.5 31.202 gm/l

0.2 M $\text{NaHPO}_4 \cdot 7\text{H}_2\text{O}$, pH 9.1 53.614 gm/l

Add NaH_2PO_4 to Na_2HPO_4 dropwise to adjust the pH to 7.2 - 8.0 .

Phosphate buffer saline (PBS), pH 7.4

0.2 M PB, pH 7.2 - 7.4 stock solution 50 ml

NaCl 8.76 gm

Distilled water to make 1000 ml

0.1 M Citrate buffer , pH 3.0 - 7.0 (for an affinity chromatography)

0.1 M Citric acid ($\text{C}_6\text{H}_8\text{O}_7 \cdot 1\text{H}_2\text{O}$) 21.014 g/l

0.1 M $\text{Na}_2\text{HPO}_4 \cdot 7\text{H}_2\text{O}$ 26.807 g/l

For pH 5.0 is approximately a 50:50 mixture of citric acid to phosphate ,
below pH5.0 : titrate pH of citric acid with phosphate and above pH 5.0 : titrate
pH of phosphate with citric acid . Autoclave before add thimerosal to 0.01 % as
preservative .

1.0 M Tris - hydrochloric acid buffer , pH 8.5 and 9.0

1 M C ₄ H ₁₁ NO ₃	121.1 g/l
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1 M HCl	
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For pH 8.5 prepare by mix 25 ml of 1 M Tris with 7.2 ml 1 M HCl.

For pH 9.0 prepare by mix 25 ml of 1 M Tris with 2.5 ml 1 M HCl .

Autoclave before add thimerosal (0.01 %).

0.2 M Cacodylate buffer , pH 7.4

Sodium cacodylate	42.8 gm
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Distilled water to make	1000 ml
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Dissolve Sodium cacodylate in distilled water and mix well . Adjust pH to 7.4 with 0.1 N HCl

0.1 M Cacodylate buffer , pH 7.4

0.2 M cacodylate buffer, pH 7.4	1 part
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Distilled water	1 part
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Veronal acetate buffer

stocking solution:

NaCl	3.4 %
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Sodium acetate	1.94 %
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Sodium barbital	2.94 %
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working solution:

0.22 M Veronal acetate buffer , pH 7.4

veronal acetate stocking solution	5 ml
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Distilled water	13 ml
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1 M CaCl ₂	0.25 ml
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Add 0.1 N HCL to adjust pH.

0.11 M Veronal acetate buffer, pH 7.4

0.22 M veronal acetate buffer	1 part
Distilled water	1 part

Gold Buffer (for dilute Protein-A gold conjugate)

Tris-HCL	6.61 gm
Tris base	0.97 gm
NaCl	8.77 gm
Distilled water	800 ml

Adjust pH to 7.4 with 0.1 N HCL and bright volume to 1000 ml with distilled water.

REAGENTS:

1.5 % Agar

Agar	0.15 gm
Dissolve water to make	10 ml

Dissolve agar with distilled water. Heat until boiling and cool at room temperature until the temperature become 37° C before use.

100% Ammonium sulfate (NH₃)₂SO₄

Ammonium Sulfate	100 gm
Distilled water	100 ml

Dissolve this amount of ammonium sulfate at 50°C until the solvent is cleared, stand overnight at room temperature, adjust the pH to 7.2 with dilute ammonia solution.

0.5 % BSA

Bovine serum albumin	0.5 gm
0.01 M PBS with 0.01 % thimerosal	100 ml

Stirring the two components until clearly. Filter with filter paper before store at 4°C.

1 % Glutaraldehyde

50 % Glutaraldehyde	0.2 ml
0.2 M Phosphate buffer, pH 7.4	5.0 ml
Distilled water to make 10 ml	

OPD Substrate (for ELISA Assay)

OPD	0.008 gm
Phosphate citrate buffer, pH 5.0	20 ml
H ₂ O ₂ 30 %	8 µl

Dissolve OPD in phosphate citrate buffer pH 5.0 and mix well.

Immediately use after add 30 % H₂O₂ (reducing agent).

Protease inhibitor (stock solution)

Benzamidine	10 mM
Epsilon amino carproic acid	10 mM
Phenyl methyl sulfonyl Fluoride	10 mM
50 % Ethanol	

Dissolve all the described above in 50 % Ethanol. The stock solution was diluted to 1:10 or 1:100. Store at 4°C and protect from light .

Reduced Osmium

Stocking solution:

1) 4 % Osmium Tetroxide

OsO ₄	1 gm
Distilled water	25 ml

2) 2 % K₄Fe (CN)₆

K ₄ Fe (CN) ₆	0.1 gm
0.3 M cacodylate buffer, pH 7.4	5 ml
CaCl ₂ (2.5 mM)	0.027 gm

Working solution:

Mix equal amount of 4 % osmium tetroxide and 2 % K₄Fe(CN)₆
(preparation before use)

Spurr resin (Embedding media)

VCD (4 -vinycyclohexene dioxide)	10 gm
DER resin	6 gm
NSA (Nonenyl Succinic Anhydride)	26 gm
DMAE (Dimethylaminoethanol)	0.4 gm

Mix all together immediately before use and after add the DMAE.

0.4 % Trypan blue

Trypan blue (vital stain)	400 mg
Distilled water	90 mg

NaCl	810 mg
K ₂ HPO ₄	60 mg
Methyl p- hydroxy benzoate	50 mg

Dissolve all of the components in distilled water. Mixture was heated to boiling. After the mixture cooled and the pH was adjust to 7.2-7.3 with 1 N NaOH (approximate 8 drops). Finally, adjust to final volume of 100 ml with distilled water .

0.05% Trypsin

Stosking solution:

1) 0.02 % EDTA (Versene)

NaCl	8 gm
KCl	0.2 gm
Na ₂ HPO ₄	1.15 gm
KH ₂ PO ₄	0.2 gm
EDTA	0.2 gm
Distilled water	1 L
Phenol red	0.01 gm

Dissolve all chemicals in distilled water.

2) PBS (CMF) , pH 7.4 (CA,Mg free)

NaCl	8 gm
KCl	0,2 gm
Na ₂ HPO ₄	1.15 gm
KH ₂ PO ₄	0.2 gm
Distilled water	1 L
Phenol red	0.01 gm

Dissolve all chemical in distilled water and adjust pH to 7.4 with 1 N NaOH .

3) 2.5 % Trypsin

PBS (CMF)	100 ml
Trypsin	2.5 gm

Dissolve trypsin in PBS(CMF) and mix well. Sterile with 0.22 μm microfilter before store at -20°c . Thawing before use.

working solution:

2.5 % Trypsin	2 ml
EDTA	49 ml
PBS(CMF)	49 ml

Mix all solution into complete solution. For sterilization by filter with 0.22 μm millipore.

Staining solution (for EM thin section)

1) Lead citrate

Lead nitrate	0.3325 gm
Sodium citrate	0.44 gm
Distilled water	7.5 ml
NaOH 1 N	2 ml
Distilled water	2.5 ml

Dissolve lead nitrate and sodium citrate in distilled water 7.5 ml , mix well and stand at room temperature 30 minutes . Shaking until it become milky then add 1 N NaOH 2 ml and mix will . When it is clear , add 2.5 ml of distilled water . It must be filter before use .

2) 0.5 % Uranyl acetate

Uranyl acetate	0.5 gm
30 % EtOH to make	100 ml

Dissolve and mix gentle, filter through 0.22 μ m milipore. This solution is light sensitive. It is possible to be stored frozen in small aliquotes.

1 % Toluidine blue stain (for EM thick section)

Toluidine blue	1 gm
Sodium borate , pH 12	1 gm
Distilled water to make	100 ml

Dissolve sodium borate in distilled water to make 100 ml . Then, add toluidine blue and mix well.

Culture media:

RPMI media

RPMI 1640	10.41 gm/L
Sterile water	1000 ml
NaHCO ₃	2 gm
Pen-Strep stock solution	10 ml

Dissolve and shake until all component are completely dissolved. For sterilization, filter through 0.22 μ m milipore.

RPMI hybrid media

RPMI 1640	10.41 gm/L
Sterile water	1000 ml
NaHCO ₃	2 gm

Pen-Strep stock solution	10 ml
L-glutamine	0.1 gm
D-glucose	2 gm
Pyruvic acid	0.11 gm

Dissolve and shake until all component are completely dissolved. For sterilization, filter through 0.22 μ m milipore.

Penicillin-Streptomycin (stock solution)

Penicillin G sodium	1,000,000 units
Streptomycin	1 gm
Distilled water	100 ml

Dissolve all of the components in distilled water and mix well.



AUTHOR BIOBLOGRAPHY

Miss Siripen Thongpassano was born on Febuary 1st, 1969 in Nakhonsithammarat, Thailand. She received her Bachelor of Science in Nursing and Midwifery (first class honors) in 1991 from the Faculty of Nursing, Prince of Songkla University, Songkla, Thailand. She has enrolled Chulalongkorn University in the Master of Science Program with a specialization in academic year 1992. During her study, she was supported by grant from the National Science and Technology Development Agency (NSTDA).