

REFERENCES

- Ahivenainen, R., and Smolander, M. (2003) The use of freshness indicators in packaging. *Novel Food Packaging Techniques*, 127-143. Cambridge England: Woodhead Publishing Limited.
- Baixas-nogueras S., Bover-cid S., Veciana-nogu  T., and Vidal-carou M. (2002) Chemical and Sensory Changes in Mediterranean Hake (*Merluccius merluccius*) under Refrigeration (6-8  C) and Stored in Ice. *Journal of agricultural and food chemistry*
- Byrne, L., Tong Lau, K., and Diamond, D. (2002) Monitoring of headspace total volatile basic nitrogen from selected fish species using reflectance spectroscopic measurements of pH sensitive films. *Analyst*, 127, 1338–1341.
- Cann, D., Smith, G. L., and Houston, N. G. (1983) Further studies on marine fish stored under modified atmosphere packaging. Aberdeen: Ministry of Agriculture Fisheries and Food, Torry Research Station.
- Chiu, F.C., Lai, S.M., Chen, J.W., and Chu, P.S. (2004) Combined effects of clay modifications and compatibilizers on the formation and physical properties of melt-mixed polypropylene/clay nanocomposites. *Polmer Science*, 42, 4139-4150.
- Choy, J., Kwak, S., Han, Y., and Kim, B. (1997) New organo-montmorillonite complexes with hydrophobic and hydrophilic functions. *Materials Letters*, 33, 143-147.
- Crowley, K., Pacquit, A., Hayes, J., Lau, K.T., and Diamond, D. (2005) A gas-phase colorimetric sensor for the detection of amine spoilage products in packaged fish. *IEEE Sensors*, 754-757.
- Di, Y., Lannac, S., Sanguigno, L., and Nicolais L. (2005) Barrier and mechanical properties of poly(caprolactone)/organoclay nanocomposites. *Macromolecular symposia*, 228, 115-124.
- Ding, C., Jia, D., He, H., Guo, B., and Hong, H. (2004) How organo-montmorillonite truly affects the structure and properties of polypropylene. *Polymer Testing*, 24, 94-100.

- Frounchi, M., Dadbin, S., Salehpour, Z., and Noferesi, M. (2006) Gas barrier properties of PP/EPDM blend nanocomposites. Membrane Science, 282, 146-148.
- Garcia-Lopez, D., Picazo, O., Merino, J.C., and Pastor, J.M. (2003) Polypropylene-clay nanocomposites: effect of compatibilizing agents on clay dispersion. European Polymer, 39, 945-950.
- Gram, L., and Huss, H. (1996) Microbiological spoilage of fish and fish products. Food Microbiology, 33, 121-137.
- He, H., Duchet, J., Galy, J., and Gerard, J-F. (2005) Grafting of swelling clay materials with 3-aminopropyltriethoxysilane. Colloid and Interface Science, 288, 171-176.
- Hurm, E. (2002) Intelligent Systems in Food Packaging –Advances in the Supply Chain. London: Pira International.
- Ismail, H., and Nasir, M. (2002) The effect of various compatibilizers on mechanical properties of polystyrene/polypropylene blend. Polymer Testing, 21, 163-170.
- Jirakittidul, K., Magaraphan, R., Nithithanakul, M., and Manuspiya, H., (2006) Effect of onium ion structure on nanoclays and polypropylene nanocomposites. M.S. Thesis. The Petroleum and Petrochemical College, Chulalongkorn University, Bangkok, Thailand.
- Koutsoumanis, K. (2001) Predictive modeling of the shelf life of fish under non-isothermal conditions. Applied and Environment Microbiology, 67, 1821–1829.
- Kruijf, N.D., Beest, M.V., Rijk, R., Sipilainen-Malm, T., Losada, P.P., and Meulenaer, B.D. (2002) Active and intelligent packaging: applications and regulatory aspects. Food Additives and Contaminants, 19, 144-162.
- Kyranas, V.R., Lougovois, V.P., and Valsamis, D.S. (1997) Assessment of shelf-life of maricultured gilthead sea bream (*Sparus aurata*) stored in ice. Food Science and Technology, 32, 339–347.
- Lai, S., Chen, J., and Chu, P. (2004) Combined effects of clay modifications and compatibilizers on the formation and physical properties of melt-mixed polypropylene/clay nanocomposites. Polymer Science, 42, 4139-4150.

- Lee, Y. J. and Lee, K. H. (2004) Characterization of organobentonite used for polymer nanocomposites. Materials Chemistry and Physics, 85, 410-415.
- Lertwimolnun, W., and Vergnes, B. (2005) Influence of compatibilizer and processing conditions on the dispersion of nanoclay in a polypropylene matrix. Polymer, 46, 3462-3471.
- Ludorff, W. and Meyer, V. (1973) *Fische und Fischerzeugnisse*. Verlag Paul Parey, Hamburg, Berlin, 308 pp.
- Maio, E., Iannace, S., Sorrentino, L., and Nicolais, L. (2004) Isothermal crystallization in PCL/clay nanocomposites investigated with thermal and rheometric methods. Polymer, 45, 8893-8900.
- Milanes, M. (2004) Characterization of sensory indexes of artic charr (*salvelinus*) and determination of its shelf life. Fishries training programme, The United Nations University, Islands.
- Modesti, M., Lorenzetti, A., Bon, D., and Besco, S., (2006) Thermal behaviour of compatibilised polypropylene nanocomposite: Effect of processing conditions. Polymer Degradation and Stability, 91, 672.
- Olafsdottir, G., Martinsdottir, E., Oehlenschlager, J., Dalgaard, P., Jensen, B., and Unde, I. (1997) Methods to evaluate fish freshness in research and industry. Food Science and Technology, 8, 258-265.
- Olafsdottir, G., Nesvadba, P., Natale, C. D., Careche, M., Oehlenschlager, J., and Tryggvadottir, S. V. (2004) Multisensor for fish quality determination. Food Science and Technology, 15, 86-93.
- Pacquit, A., Frisby, J., Diamond, D., Lau, K.T., Farrell, A., Quilty, B., and Diamond, D. (2007) Development of a smart packaging for the monitoring of fish spoilage. Food Chemistry, 102, 466-470. Pacquit, A., Lau, K.T., and Diamond, D. (2004) Development of a colorimetric sensor for monitoring of fish spoilage amines in packaging headspace. IEEE Sensors, 365-367.
- Pacquit, A., Lau, K.T., McLaughlin, H., Frisby, J., Quilty, B., and Diamond, D. (2006) Development of a volatile amine sensor for the monitoring of fish spoilage. Talanta, 69, 515-520.

- Pacquit, A., Frisby, J., Diamond, D., Lau, K.T., Farrell, A., Quilty, B., and Diamond, D. (2007) Development of a smart packaging for the monitoring of fish spoilage. Food Chemistry, 102, 466–470.
- Papadopoulos, V., Chouliara, I., Badeka, A., Savvaidis, I.N., and Kontominas, M.G. (2003) Effect of gutting on microbiological, chemical, and sensory properties, of aqua-cultured sea bass (*Dicentrarchus labrax*) stored in ice. Food Microbiol, 20, 414–420.
- Phandee, A., Magaraphan, R., Nithithanakul, M., and Manuspiya, H., (2006) Effect of surfactant structure on nanoclays and PP reactive nanocomposites. M.S. Thesis, The Petroleum and Petrochemical Collage, Chulalongkorn University, Bangkok, Thailand.
- Ramos, F.G., Melo, T.A., Rabello, M.S., and Silva, S.M. (2005) Thermal stability of nanocomposites based on polypropylene and bentonite. Polymer Degradation and Stability, 89, 383-392.
- Simeonidou, S., Govaris, A., and Vareltzis, K. (1998) Quality assessment of seven Mediterranean fish species during storage on ice. Food Research International, 30, 479–484.
- Shah, R.K., Hunter, D.L., and Paul, D.R. (2005) Nanocomposites from poly(ethylene-co-methacrylic acid) ionomers: effect of surfactant structure on morphology and properties. Polymer, 46, 2646-2662.
- Sorrentino A., Gorrasi G., and Vittoria, V. (2007) Potential perspectives of bio-nanocomposites for food packaging applications. Trends in Food Science and Technology, 18, 84-95.
- Suprakas, R., and Masami, O. (2003) Polymer/layered silicate nanocomposites: a review from preparation to processing. Progress in Polymer Science, 28, 1539-1641.
- Százdi, L., Pukánszky, B., Jr, Vancso, G. J., and Pukánszky, B. (2006) Quantitative estimation of the reinforcing effect of layered silicates in PP nanocomposites. Polymer, 47, 4638-4648.
- Tang Y., Hu Y., Song L., Zong R., Gui Z., Chen Z., and Fan W. (2003) Preparation and thermal stability of polypropylene/montmorillonite nanocomposites. Polymer Degradation and Stability, 82, 127-131.

- Tassanawat, S., Manuspiya, H., Magaraphan, R., and Nithithanakul, M. (2006) Polypropylene/Organoclay Nanocomposite for pH-sensitive Packaging. M.S. Thesis, The Petroleum and Petrochemical Collage, Chulalongkorn University, Bangkok, Thailand.
- Tejada, M. and Huidobro, A. (2002) Quality of farmed gilthead seabream (*Sparus aurata*) during ice storage related to the slaughter method and gutting. European Food Research Technology, 215, 1–7
- Wang, Y., Easteal, A.J., and Chen, X.D. (1998) Ethylene and oxygen permeability through polyethylene packaging films. Packaging Technology and Science, 11, 169-178.
- Zeng Q., Yu A., Lu G., and Paul, D. (2005) Clay-Based Polymer Nanocomposites: Research and Commercial Development. Nanoscience and Nanotechnology, 5, 1574–1592.

APPENDICES

Appendix A Determination of Total Volatile Basic Nitrogen (TVB-N)

TVB-N can be calculated by following equation:

$$\text{TVB-N (mg/100g)} = \frac{(V_S - V_B) \times (N_{\text{HCl}} \times A_N) \times [W_S \times (M/100) + V_E] \times 100}{W_S}$$

- Where,
- V_S = Titration volume of 0.01 N HCl for sample extract (ml)
 - V_B = Titration volume of 0.01 N HCl for blank (ml)
 - N_{HCl} = Normality of HCl (= 0.01 N × factor of HCl)
 - A_N = Atomic weight of nitrogen (14.00)
 - W_S = Weight of tissue sample (g)
 - M = Percentage moisture of tissue sample (Assume 80%)
 - V_E = Volume of 4% TCA used in extraction

Table A1 Change in TVB-N values of fresh milk during storage at ambient temperature

Hours	0.01 HCl (ml)			TVB-N (mg/100g)
	1	2	Av	
3	0.12	0.12	0.12	8.064
6	0.125	0.12	0.1225	8.232
9	0.2	0.2	0.2	13.44
12	0.37	0.39	0.38	25.536
15	0.45	0.62	0.535	35.952
18	0.54	0.56	0.55	36.96
21	0.65	0.8	0.725	48.72
24	1.75	1.8	1.775	119.28
27	2	2.1	2.05	137.76

Table A2 Change in APC values of Barramundi fish during storage at ambient temperature

Hours	APC (cfu/g)	Log Count(cfu/g)
3	1.40×10^5	5.146128
6	3.80×10^5	5.579784
9	6.20×10^6	6.792392
12	2.10×10^7	7.322219
15	2.50×10^7	7.39794
18	3.60×10^7	7.556303
21	5.80×10^7	7.763428
24	2.80×10^7	7.447158
27	1.60×10^7	7.20412
30	3.00×10^7	7.477121
33	8.20×10^7	7.913814
36	5.80×10^7	7.763428
39	8.40×10^7	7.924279
42	8.40×10^7	7.924279

Appendix B Change in Hunter and TCD Values to standard ammonia using Chroma Meter

Table B1 Change in Hunter color values of PP/1000 rpm/1%wt BCG film during storage at ambient temperature using 20g of fish tissue.

Conc. (ppm)	b	a	L	delta b	delta a	delta L	delta E
10	56.30	-15.41	54.74	36.95	-10.79	-32.15	50.15
25	10.46	-35.44	37.10	-8.89	-30.82	-49.79	59.23
50	-3.29	-34.60	33.03	-22.64	-29.98	-53.86	65.67
75	-13.56	-31.92	29.82	-32.91	-27.30	-57.07	71.31
100	-19.35	-28.15	28.07	-38.70	-23.53	-58.82	74.24
250	-30.24	-20.16	27.37	-49.59	-15.54	-59.52	79.02
500	-32.61	-16.61	26.05	-51.96	-11.99	-60.84	80.90
750	-39.73	-9.42	25.83	-59.08	-4.80	-61.06	85.10
1000	-41.03	-8.18	25.76	-60.38	-3.56	-61.13	85.99

Table B2 Change in Hunter color values of PP/2000 rpm/1%wt BCG film during storage at ambient temperature using 20g of fish tissue.

Conc. (ppm)	b	a	L	delta b	delta a	delta L	delta E
10	46.63	-18.36	62.29	33.74	-14.82	-24.84	44.45
25	7.87	-30.92	49.56	-5.02	-27.38	-37.57	46.76
50	-17.26	-28.98	43.08	-30.15	-25.44	-44.05	59.13
75	-26.32	-24.62	40.26	-39.21	-21.08	-46.87	64.64
100	-29.82	-21.93	39.88	-42.71	-18.39	-47.25	66.29
250	-40.94	-14.46	38.30	-53.83	-10.92	-48.83	73.49
500	-45.28	-11.80	37.93	-58.17	-8.26	-49.20	76.63
750	-48.78	-8.13	35.95	-61.67	-4.59	-51.18	80.27
1000	-49.38	-7.68	34.84	-62.27	-4.14	-52.29	81.42

Table B3 Change in Hunter color values of PP/3000 rpm/1%wt BCG film during storage at ambient temperature using 20g of fish tissue.

Conc.(ppm)	b	a	L	delta b	delta a	delta L	delta E
10	40.78	-12.72	73.23	26.86	-8.98	-14.39	31.77
25	5.48	-25.08	51.95	-8.44	-21.34	-35.67	42.41
50	-8.43	-23.89	49.32	-22.35	-20.15	-38.30	48.71
75	-17.73	-23.04	47.58	-31.65	-19.30	-40.04	54.56
100	-24.55	-22.85	46.33	-38.47	-19.11	-41.29	59.58
250	-32.45	-18.85	45.88	-46.37	-15.11	-41.74	64.19
500	-39.94	-15.93	42.49	-53.86	-12.19	-45.13	71.32
750	-41.79	-12.98	39.61	-55.71	-9.24	-48.01	74.12
1000	-49.66	-8.61	38.46	-63.58	-4.87	-49.16	80.52

Table B4 Change in Hunter color values of PP/1000 rpm/3%wt BCG film during storage at ambient temperature using 20g of fish tissue.

Conc.(ppm)	b	a	L	delta b	delta a	delta L	delta E
10	75.36	15.99	70.63	52.99	20.54	-15.93	59.02
25	10.34	-21.07	21.68	-12.03	-16.52	-64.88	68.02
50	-0.31	-18.38	9.28	-22.68	-13.83	-77.28	81.72
75	-9.74	-15.28	7.53	-32.11	-10.73	-79.03	85.98
100	-13.34	-14.54	7.41	-35.71	-9.99	-79.15	87.41
250	-18.58	-7.74	7.15	-40.95	-3.19	-79.41	89.41
500	-21.56	-4.57	6.92	-43.93	-0.02	-79.64	90.95
750	-22.96	-3.24	6.89	-45.33	1.31	-79.67	91.67
1000	-26.30	-1.95	6.65	-48.67	2.60	-79.91	93.61

Table B5 Change in Hunter color values of PP/1000 rpm/5%wt BCG film during storage at ambient temperature using 20g of fish tissue.

Conc.(ppm)	b	a	L	delta b	delta a	delta L	delta E
10	85.37	14.68	73.80	41.34	19.62	-9.79	46.80
25	5.02	-4.55	15.81	-39.01	0.39	-67.78	78.21
50	0.44	-1.24	14.00	-43.59	3.70	-69.59	82.20
75	-2.24	-0.66	7.52	-46.27	4.28	-76.07	89.14
100	-3.30	-0.45	4.75	-47.33	4.49	-78.84	92.07
250	-5.16	-0.11	2.57	-49.19	4.83	-81.02	94.91
500	-6.80	1.63	1.81	-50.83	6.57	-81.78	96.52
750	-7.12	1.38	1.76	-51.15	6.32	-81.83	96.72
1000	-8.69	1.99	1.11	-52.72	6.93	-82.48	98.14

Appendix C Change in Hunter and TCD Values to TVB-N using Chroma Meter

Table C1 Change in Hunter color values of PP/1000 rpm/1%wt BCG film during storage at ambient temperature using 20g of fish tissue.

Time(hours)	b	a	L	delta b	delta a	delta L	delta E
3	50.41	-3.32	82.56	17.47	1.00	-3.87	17.92
6	45.47	-1.97	80.36	12.53	2.35	-6.07	14.12
9	43.22	-0.21	78.50	10.28	4.11	-7.93	13.62
12	43.43	-0.59	77.30	10.49	3.73	-9.13	14.40
15	31.30	-6.73	69.90	-1.64	-2.41	-16.53	16.79
18	36.83	-13.39	62.01	3.89	-9.07	-24.42	26.34
21	43.15	-15.16	58.07	10.21	-10.84	-28.36	32.03
24	37.82	-20.55	53.39	4.88	-16.23	-33.04	37.13
27	12.01	-51.07	30.97	-20.93	-46.75	-55.46	75.50
30	10.94	-51.44	30.55	-22.01	-47.12	-55.88	76.34
33	4.90	-52.13	26.54	-28.04	-47.81	-59.90	81.61
36	6.04	-52.07	26.64	-26.90	-47.75	-59.79	81.11
39	5.65	-52.04	26.24	-27.29	-47.72	-60.20	81.52
42	5.71	-52.34	26.78	-27.24	-48.02	-59.66	81.28

Table C2 Change in Hunter color values of PP/2000 rpm/1%wt BCG film during storage at ambient temperature using 20g of fish tissue.

Time(hours)	b	a	L	delta b	delta a	delta L	delta E
3	48.59	-3.52	83.26	21.08	0.88	-3.85	21.45
6	40.25	-3.46	81.92	12.74	0.93	-5.19	13.79
9	39.76	-2.17	79.93	12.25	2.23	-7.18	14.38
12	37.97	-0.25	78.90	10.46	4.15	-8.21	13.93
15	30.62	-10.27	70.40	3.11	-5.87	-16.71	17.98
18	28.17	-20.43	67.19	0.66	-16.03	-19.92	25.58
21	33.07	-21.76	58.24	5.56	-17.36	-28.87	34.15
24	18.95	-26.25	56.39	-8.56	-21.85	-30.72	38.66
27	-21.27	-26.95	45.05	-48.78	-22.55	-42.06	68.24
30	-12.90	-20.28	45.59	-40.41	-15.88	-41.52	60.07
33	-28.27	-21.66	39.62	-55.78	-17.26	-47.49	75.26
36	-28.08	-22.47	39.29	-55.59	-18.07	-47.823	75.52
39	-28.08	-22.47	39.30	-55.59	-18.07	-47.806	75.51
42	-28.08	-22.47	39.36	-55.59	-18.07	-47.746	75.47

Table C3 Change in Hunter color values of PP/3000 rpm/1%wt BCG film during storage at ambient temperature using 20g of fish tissue.

Time(hours)	b	a	L	delta b	delta a	delta L	delta E
3	36.16	-4.83	85.33	6.36	-1.02	-2.21	6.81
6	41.50	-3.99	83.59	11.70	-0.18	-3.95	12.35
9	40.79	-2.71	80.58	10.99	1.10	-6.96	13.06
12	37.76	-7.37	77.81	7.96	-3.56	-9.73	13.07
15	35.15	-11.31	73.18	5.35	-7.50	-14.36	17.06
18	29.66	-16.29	66.63	-0.14	-12.48	-20.91	24.36
21	20.08	-22.38	62.40	-9.72	-18.57	-25.14	32.74
24	22.56	-19.46	62.04	-7.24	-15.65	-25.50	30.78
27	-29.46	-23.01	48.36	-59.26	-19.20	-39.18	73.59
30	-29.46	-20.67	48.36	-59.26	-16.86	-39.18	73.02
33	-30.22	-20.61	48.00	-60.03	-16.80	-39.54	73.82
36	-30.18	-20.44	48.67	-59.98	-16.63	-38.88	73.39
39	-30.22	-20.61	47.87	-60.03	-16.80	-39.67	73.89
42	-30.28	-20.43	48.00	-60.08	-16.62	-39.54	73.82

Table C4 Change in Hunter color values of PP/1000 rpm/1%wt BCG film during storage at ambient temperature using 20g of fish tissue.

Time(hours)	b	a	L	delta b	delta a	delta L	delta E
3	40.80	-3.73	81.33	7.86	0.90	-5.16	9.44
6	50.80	-1.97	80.36	17.86	2.65	-6.13	19.07
9	68.21	-0.21	78.50	35.27	4.41	-7.99	36.43
12	66.10	-0.59	77.30	33.16	4.03	-9.19	34.64
15	62.30	-6.73	69.90	29.36	-2.11	-16.59	33.79
18	59.03	-13.39	64.01	26.09	-8.77	-22.48	35.54
21	43.15	-15.16	58.07	10.21	-10.54	-28.42	31.99
24	37.82	-20.55	53.39	4.88	-15.93	-33.10	37.06
27	12.01	-51.08	30.97	-20.93	-46.45	-55.52	75.35
30	10.94	-51.44	30.55	-22.01	-46.82	-55.94	76.19
33	4.90	-52.13	28.54	-28.04	-47.51	-57.96	80.02
36	6.04	-52.07	28.97	-26.90	-47.45	-57.52	79.27
39	5.65	-52.05	29.24	-27.29	-47.42	-57.26	79.20
42	5.71	-52.35	29.78	-27.24	-47.72	-56.72	78.97

Table C5 Change in Hunter color values of PP/1000 rpm/3%wt BCG film during storage at ambient temperature using 20g of fish tissue.

Time(hours)	b	a	L	delta b	delta a	delta L	delta E
3	77.14	1.31	79.68	27.74	4.63	-3.79	28.38
6	76.69	3.66	74.34	27.29	6.98	-9.13	29.61
9	82.16	5.41	73.77	32.76	8.73	-9.70	35.26
12	87.82	8.51	72.51	38.42	11.83	-10.96	41.67
15	89.56	14.52	67.19	40.16	17.84	-16.28	46.86
18	86.64	12.08	58.55	37.24	15.40	-24.92	47.38
21	49.80	-14.52	41.64	0.40	-11.20	-41.83	43.30
24	27.36	-15.77	32.51	-22.04	-12.45	-50.96	56.90
27	2.14	-21.54	21.68	-47.26	-18.22	-61.79	79.89
30	-15.44	-11.19	21.04	-64.84	-7.87	-62.43	90.35
33	-3.34	-22.87	20.63	-52.74	-19.55	-62.84	84.33
36	-2.85	-23.51	20.78	-52.25	-20.19	-62.69	84.07
39	-2.46	-23.03	20.6	-51.86	-19.71	-62.87	83.85
42	-1.51	-24.75	21.11	-50.91	-21.43	-62.36	83.30

Table C6 Change in Hunter color values of PP/1000 rpm/5%wt BCG film during storage at ambient temperature using 20g of fish tissue.

Time(hours)	b	a	L	delta b	delta a	delta L	delta E
3	88.96	3.94	78.71	34.34	7.13	-5.08	35.44
6	88.46	6.10	73.83	33.84	9.29	-9.96	36.48
9	97.32	17.51	71.37	42.70	20.70	-12.42	49.06
12	97.35	16.89	70.59	42.73	20.08	-13.20	49.03
15	89.32	24.46	66.59	34.70	27.65	-17.20	47.59
18	87.89	23.44	51.64	33.27	26.63	-32.15	53.39
21	33.80	-4.85	25.08	-20.82	-1.66	-58.71	62.32
24	25.11	-8.80	20.29	-29.51	-5.61	-63.50	70.25
27	10.91	-12.04	11.49	-43.71	-8.85	-72.30	84.95
30	0.51	-4.16	11.72	-54.11	-0.97	-72.07	90.13
33	0.89	-4.68	10.40	-53.73	-1.49	-73.39	90.97
36	0.37	-4.39	11.22	-54.25	-1.20	-72.57	90.62
39	0.47	-4.69	10.59	-54.15	-1.50	-73.20	91.06
42	0.90	-3.86	10.72	-53.72	-0.67	-73.07	90.69

Table C7 Change in Hunter color values of PP/1000 rpm/3%wt BCG film during storage at ambient temperature using 20g of fish tissue.

Time(hours)	b	a	L	delta b	delta a	delta L	delta E
3	78.93	-0.64	79.45	29.80	4.68	-4.49	30.50
6	98.87	7.88	75.54	49.74	13.20	-8.40	52.15
9	106.33	13.57	73.68	57.20	18.89	-10.26	61.11
12	94.73	19.37	69.03	45.60	24.69	-14.91	53.96
15	45.69	-17.84	31.74	-3.44	-12.52	-52.20	53.79
18	47.45	-11.26	31.21	-1.68	-5.94	-52.73	53.09
21	40.70	-15.75	27.87	-8.43	-10.43	-56.07	57.66
24	11.40	-22.88	9.69	-37.73	-17.56	-74.25	85.12
27	7.22	-21.74	8.32	-41.91	-16.42	-75.62	88.01
30	5.89	-21.45	8.28	-43.24	-16.13	-75.66	88.63
33	3.26	-28.23	11.42	-45.87	-22.91	-72.52	88.82
36	-6.65	-22.11	9.75	-55.78	-16.79	-74.19	94.33
39	-3.00	-11.36	15.56	-52.13	-6.04	-68.38	86.20
42	-15.99	-10.83	8.38	-65.12	-5.51	-75.56	99.90

Table C8 Change in Hunter color values of PP/Surllyn/1000 rpm/3%wt BCG film during storage at ambient temperature using 20g of fish tissue.

Time(hours)	b	a	L	delta b	delta a	delta L	delta E
3	84.68	2.13	78.07	36.35	7.19	-5.69	37.49
6	88.64	4.27	77.15	40.31	9.33	-6.61	41.90
9	95.14	3.24	76.07	46.81	8.30	-7.69	48.16
12	100.59	8.82	75.10	52.26	13.88	-8.66	54.76
15	71.40	-0.63	47.48	23.07	4.43	-36.28	43.23
18	70.39	-2.13	46.35	22.06	2.93	-37.41	43.53
21	13.71	-7.99	15.21	-34.62	-2.93	-68.55	76.85
24	-1.10	-20.63	16.78	-49.43	-15.57	-66.98	84.69
27	23.09	-25.65	24.23	-25.24	-20.59	-59.53	67.86
30	18.23	-24.75	24.85	-30.10	-19.69	-58.91	69.02
33	14.50	-30.74	15.53	-33.83	-25.68	-68.23	80.37
36	1.87	-28.54	16.15	-46.46	-23.48	-67.61	85.33
39	0.13	-14.14	24.80	-48.20	-9.08	-58.96	76.70
42	1.92	-25.07	17.55	-46.41	-20.01	-66.21	83.30

Table C9 Change in Hunter color values of 1%wt Clay/1000 rpm/3%wt BCG film during storage at ambient temperature using 20g of fish tissue.

Time(hours)	b	a	L	delta b	delta a	delta L	delta E
3	82.12	1.19	79.24	32.38	6.33	-4.38	33.28
6	94.09	5.34	77.12	44.35	10.48	-6.50	46.03
9	97.87	6.47	77.47	48.13	11.61	-6.15	49.89
12	100.73	10.91	74.93	50.99	16.05	-8.69	54.15
15	90.50	23.10	55.80	40.76	28.24	-27.82	56.85
18	55.86	-13.51	39.31	6.12	-8.37	-44.31	45.51
21	33.92	-18.48	35.39	-15.82	-13.34	-48.23	52.48
24	35.42	-16.61	38.34	-14.32	-11.47	-45.28	48.86
27	32.64	-19.57	23.73	-17.10	-14.43	-59.89	63.93
30	15.90	-20.22	24.57	-33.84	-15.08	-59.05	69.71
33	9.05	-32.91	17.63	-40.69	-27.77	-65.99	82.35
36	8.93	-22.44	20.80	-40.81	-17.30	-62.82	76.89
39	4.96	-21.04	20.90	-44.78	-15.90	-62.72	78.69
42	-0.58	-24.48	12.22	-50.32	-19.34	-71.40	89.47

Table C10 Change in Hunter color values of 3%wt Clay/1000 rpm/3%wt BCG film during storage at ambient temperature using 20g of fish tissue.

Time(hours)	b	a	L	delta b	delta a	delta L	delta E
3	83.55	2.21	78.54	31.59	7.26	-4.51	32.72
6	92.04	4.92	76.83	40.08	9.97	-6.22	41.77
9	94.44	6.91	75.99	42.48	11.96	-7.06	44.69
12	102.64	11.70	75.18	50.68	16.75	-7.87	53.95
15	81.49	2.82	52.94	29.53	7.87	-30.11	42.90
18	63.04	-7.50	46.02	11.08	-2.45	-37.03	38.73
21	38.84	-14.94	40.04	-13.12	-9.89	-43.01	46.04
24	23.25	-22.18	31.54	-28.71	-17.13	-51.51	61.41
27	24.40	-24.05	29.07	-27.56	-19.00	-53.98	63.52
30	16.17	-24.52	25.33	-35.79	-19.47	-57.72	70.65
33	6.36	-32.97	15.39	-45.60	-27.92	-67.66	86.24
36	-1.66	-17.38	18.92	-53.62	-12.33	-64.13	84.50
39	-12.39	-10.10	17.86	-64.35	-5.05	-65.19	91.74
42	-11.41	-19.97	10.74	-63.37	-14.92	-72.31	97.30

Table C11 Change in Hunter color values of 5%wt Clay/1000 rpm/3%wt BCG film during storage at ambient temperature using 20g of fish tissue.

Time(hours)	b	a	L	delta b	delta a	delta L	delta E
3	96.37	6.97	77.58	41.54	11.37	-5.59	43.43
6	95.72	5.62	78.30	40.89	10.02	-4.87	42.38
9	101.21	11.43	75.41	46.38	15.83	-7.76	49.62
12	102.95	9.72	76.19	48.12	14.12	-6.98	50.63
15	83.71	9.12	57.04	28.88	13.52	-26.13	41.23
18	53.87	-13.31	48.61	-0.96	-8.91	-34.56	35.71
21	47.77	-11.71	42.85	-7.06	-7.31	-40.32	41.59
24	34.21	-20.33	35.38	-20.62	-15.93	-47.79	54.44
27	28.56	-21.97	22.10	-26.27	-17.57	-61.07	68.77
30	25.95	-20.97	20.6	-28.88	-16.57	-62.57	70.89
33	13.26	-16.74	16.57	-41.57	-12.34	-66.60	79.48
36	4.49	-19.34	17.15	-50.34	-14.94	-66.02	84.36
39	-6.98	-11.05	18.04	-61.81	-6.65	-65.13	90.04
42	1.48	-24.37	10.03	-53.35	-19.97	-73.14	92.71

Table C12 Change in Hunter color values of 5%wt Clay/1000 rpm/3%wt BCG film during storage at ambient temperature using 20g of fish tissue.

Time(hours)	b	a	L	delta b	delta a	delta L	delta E
3	96.37	6.97	77.58	47.24	12.29	-6.36	49.23
9	103.21	11.43	75.41	54.08	16.75	-8.53	57.26
15	83.71	9.12	57.04	34.58	14.44	-26.90	46.13
21	47.77	-11.71	42.85	-1.36	-6.39	-41.09	41.61
27	28.56	-21.97	22.10	-20.57	-16.65	-61.84	67.27
33	13.26	-16.74	16.57	-35.87	-11.42	-67.37	77.18
39	-6.98	-11.05	18.04	-56.11	-5.73	-65.90	86.74

Table C13 Change in Hunter color values of 5%wt Clay/1000 rpm/3%wt BCG film during storage at ambient temperature using 40g of fish tissue.

Time(hours)	b	a	L	delta b	delta a	delta L	delta E
3	105.47	12.22	74.03	58.20	17.42	-10.17	61.60
9	99.76	23.03	69.25	52.49	28.23	-14.95	61.45
15	17.95	-17.97	22.49	-29.32	-12.77	-61.71	69.50
21	33.57	-23.08	27.51	-13.70	-17.88	-56.69	61.00
27	14.08	-24.90	11.54	-33.19	-19.70	-72.66	82.27
33	7.97	-23.74	18.98	-39.30	-18.54	-65.22	78.37
39	-0.34	-12.76	7.78	-47.61	-7.56	-76.42	90.35

Table C14 Change in Hunter color values of 5%wt Clay/1000 rpm/3%wt BCG film during storage at ambient temperature using 60g of fish tissue.

Time(hours)	b	a	L	delta b	delta a	delta L	delta E
3	112.3	7.37	75.70	60.33	12.29	-7.86	62.07
9	94.93	27.57	53.58	42.96	32.49	-29.98	61.64
15	18.41	-11.03	21.57	-33.56	-6.11	-61.99	70.75
21	16.56	-26.26	12.21	-35.41	-21.34	-71.35	82.46
27	9.33	-18.33	15.85	-42.64	-13.41	-67.71	81.13
33	8.63	-19.78	11.32	-43.34	-14.86	-72.24	85.54
39	0.13	-27.07	11.83	-51.84	-22.15	-71.73	91.23

Appendix D Mechanical Measurement of PP/organoclay nanocomposites

Table D1 Mechanical properties of PP/organoclay nanocomposites

Sample	Young's Modulus (MPa)	Tensile Strength (MPa)	Elongation At break (%)
PP	513.23 ± 29.82	30.55 ± 1.70	15.96 ± 3.38
PP/6%Surlyn	576.25 ± 49.19	25.80 ± 1.11	10.46 ± 1.46
PP/6%Surlyn/1%Clay	596.29 ± 10.94	25.84 ± 0.81	12.53 ± 1.54
PP/6%Surlyn/3%Clay	549.18 ± 29.82	23.00 ± 1.74	11.72 ± 3.42
PP/6%Surlyn/5%Clay	532.95 ± 27.89	23.06 ± 2.53	11.28 ± 2.61

Table D2 Oxygen transmission rate, OTR (cc/m²/day) of PP/organoclay nanocomposites

Sample	Oxygen Gas Permeability (cc/m ² .day)
PP	3580 ± 84
PP/6%Surlyn	1649 ± 139
PP/6%Surlyn/1%Clay	634 ± 19
PP/6%Surlyn/3%Clay	627 ± 16
PP/6%Surlyn/5%Clay	615 ± 40

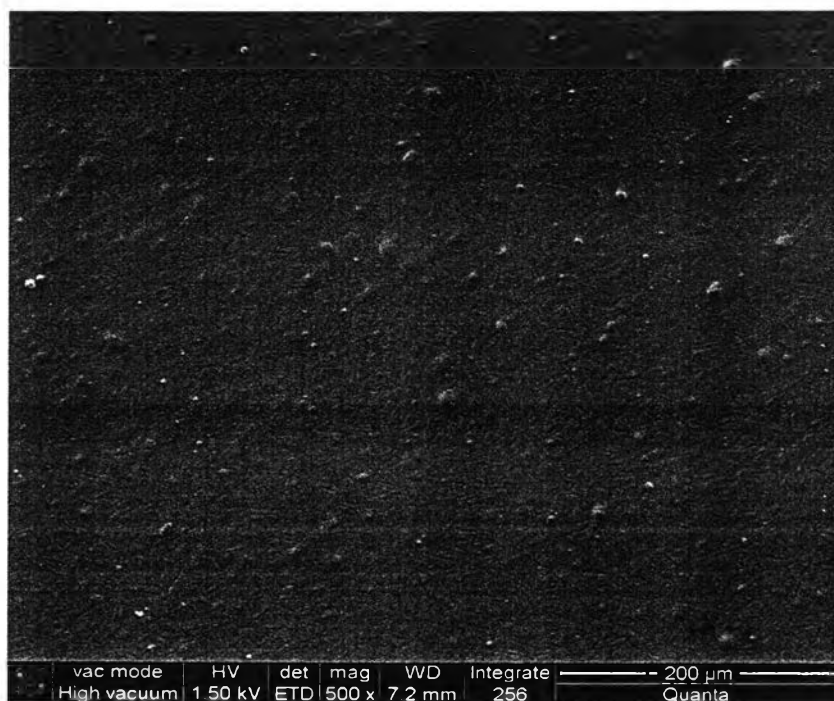
Appendix E Bentonite Clay, Max-Gel[®] GRADE SAC

Table E1 Typical chemical analysis of bentonite on dry basis at 105°C

Element	Percentage
SiO ₂	65-70
Al ₂ O ₃	13-17
Fe ₂ O ₃	1.0-2.0
Na ₂ O	1.5-2.5
LOI	10-12
MgO	2.0-3.0
CaO	1.5-2.5
K ₂ O	0.4-0.8
TiO ₂	0.2-0.3

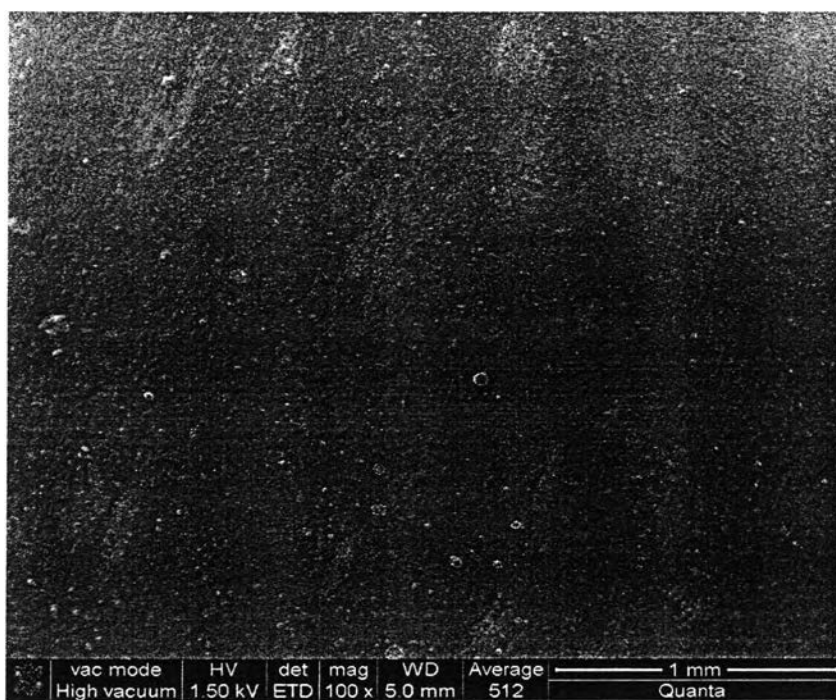
Table E2 Physical properties of bentonite

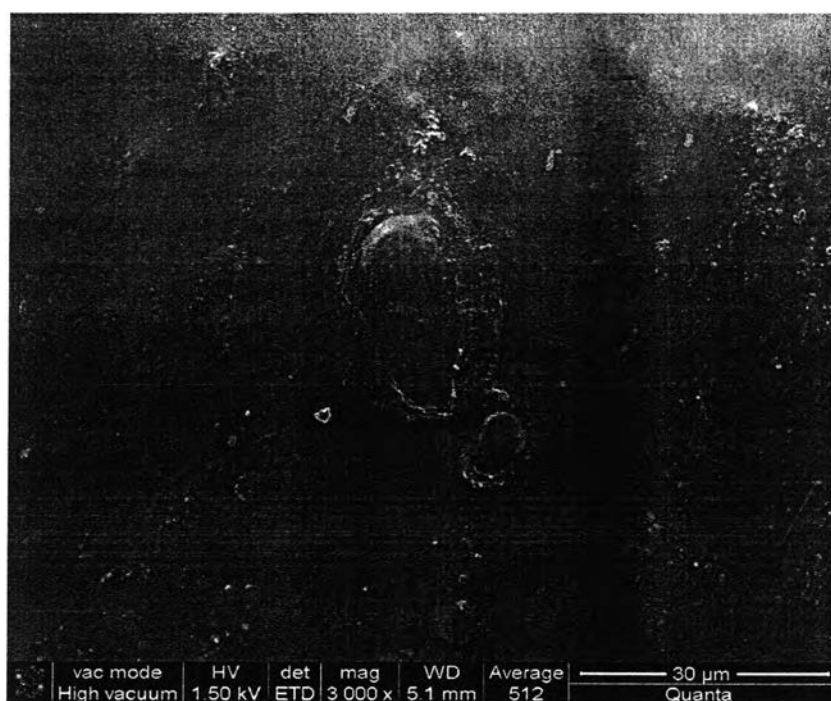
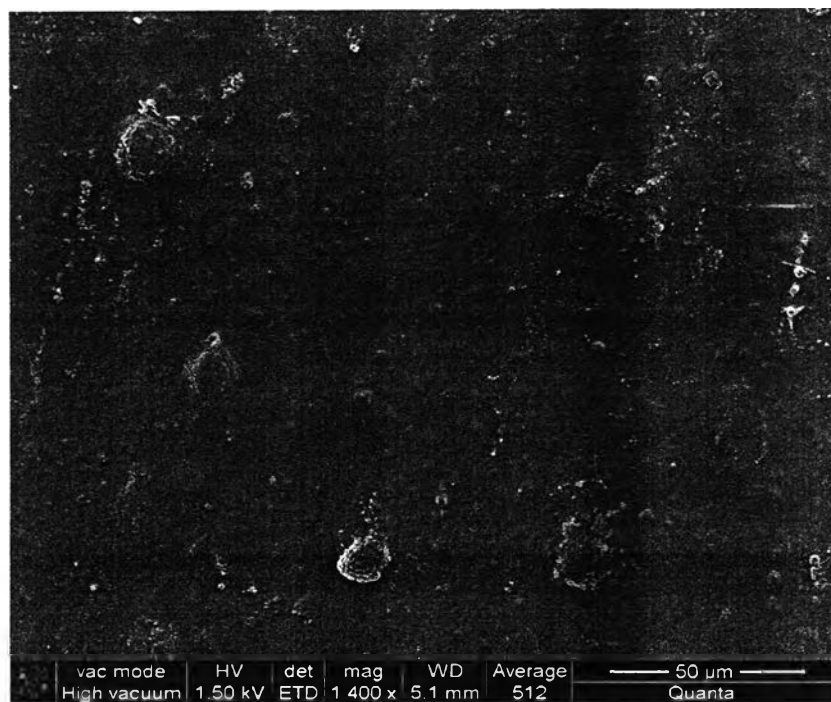
Physical properties	
Moisture content, %	8-12
5% suspension, pH	9.5-11.0
Swelling index, ml per 2 g of clay	15
Viscosity dial reading at 600 rpm	12-20
Dry particle size (pass 200 meshes), %	80 min
Wet particle size (pass 325 meshes),	98 min
Specific gravity	2.3-2.4
CEC, meq/100g of clay	50

Appendix F SEM Images of PP/Organomodified-BTN Nanocomposite Films

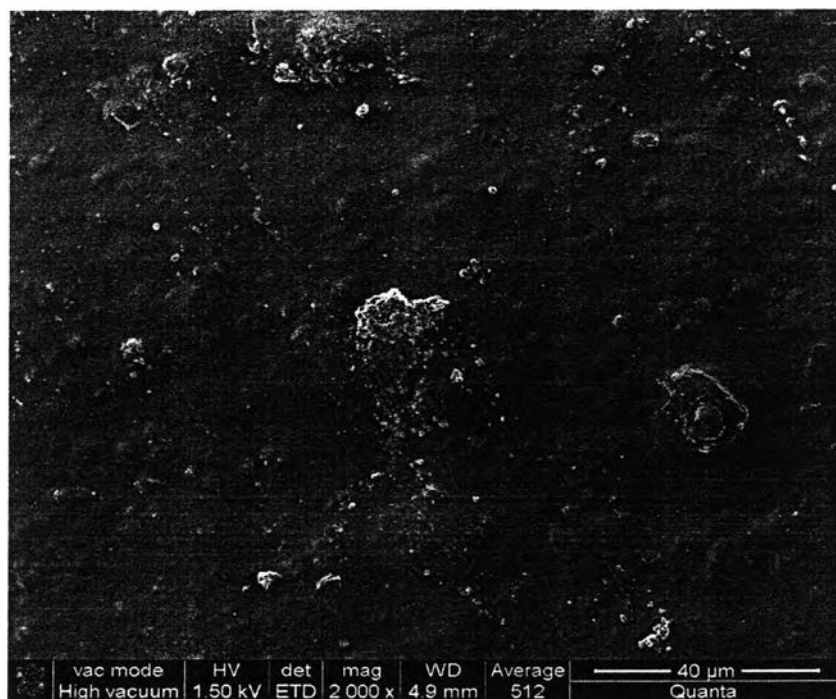
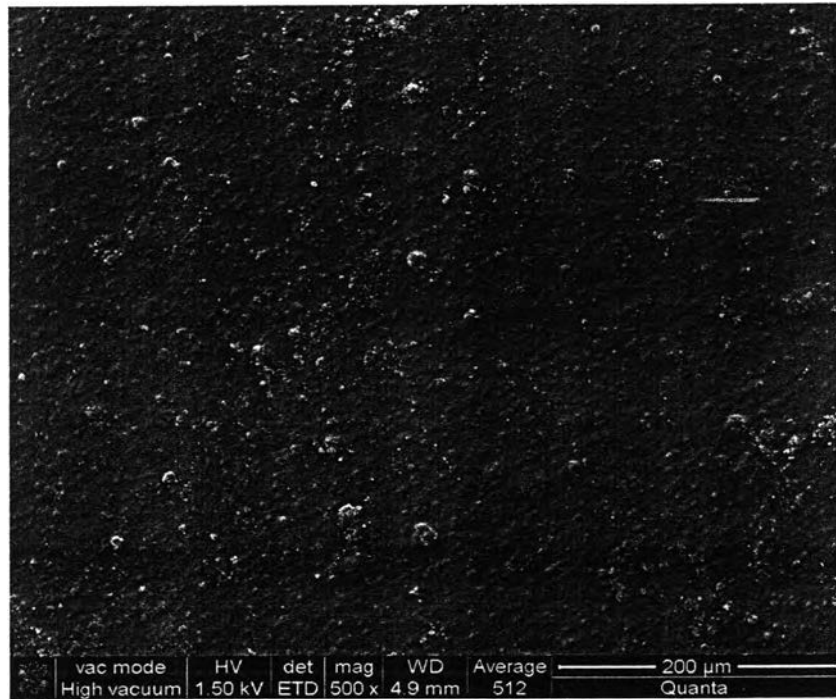


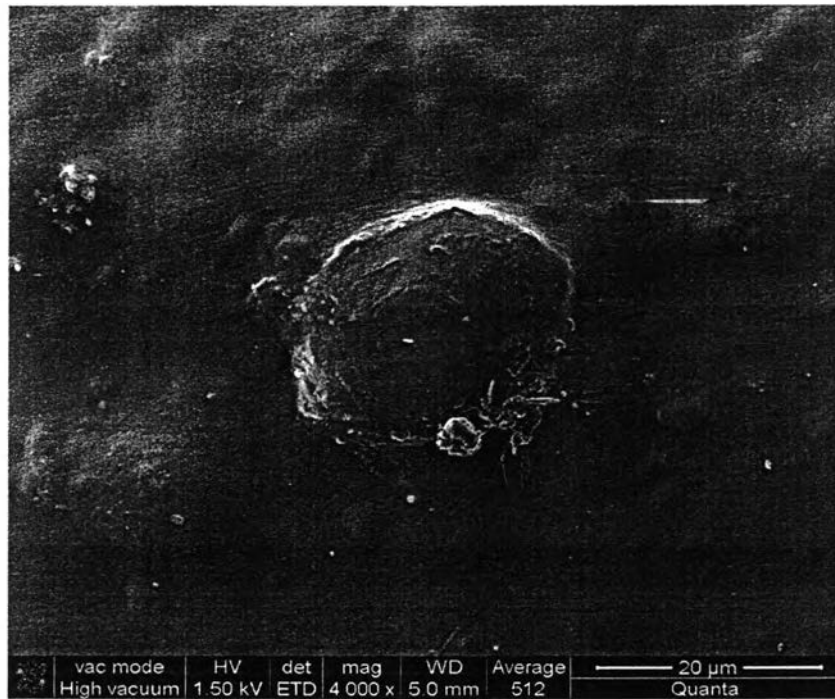
(A)





(B)





(C)

Figure F1 SEM images of nanocomposite films at 500, 2000, and 4000 magnification; (A) 1%wt organoclay, (B) 3%wt organoclay, (C) 5%wt organoclay

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Proceedings:

1. Seephueng, A., Magaraphan, R., Nithitanakul, M., and Manuspiya, H. (2008, April 6-10) Smart Packaging for Fish Spoilage Indicator. Proceedings of the 235th American Chemical Society Conference, New Orleans, Louisiana, USA.
2. Seephueng, A., Magaraphan, R., Nithitanakul, M., and Manuspiya, H. (2008, April 23) Smart Packaging for Fish Spoilage Indicator. Proceedings of the 14th PPC Symposium on Petroleum, Petrochems, and Polymers, Bangkok, Thailand.

Presentation:

1. Seephueng, A., Magaraphan, R., Nithitanakul, M., and Manuspiya, H. (2008, April 6-10) Smart Packaging for Fish Spoilage Indicator. Poster presented at the 235th American Chemical Society Conference, New Orleans, Louisiana, USA.
2. Seephueng, A., Magaraphan, R., Nithitanakul, M., and Manuspiya, H. (2008, April 23) Smart Packaging for Fish Spoilage Indicator. Poster presented at the 14th PPC Symposium on Petroleum, Petrochems, and Polymers, Chulalongkorn University, Bangkok, Thailand.

