



REFERENCES

- Agarwal, R., Katiyar, S. K., Zaidi, S. I. A., and Mukhtar, H. 1992. Inhibition of skin tumor promoter-caused induction of epidermal ornithine decarboxylase in SENCAR mice by polyphenolic fraction isolated from green tea and its individual epicatechin derivatives. Cancer Res. 52 (July 1): 3582-3588.
- Agarwal, R. 2000. Cell signaling and regulators of cell cycle as molecular targets for prostate cancer prevention by dietary agents. Biochem pharmacol. 60: 1051-1059.
- Bidlack, W. R., Omaye, S. T., Meskin, M. S. and Jahner, D. 1998. Phytochemicals: a new paradigm. Lancaster : Technomic Publishing Co., Inc., USA.
- Bright, D., Stewart, G. G. and Patino, H. 1999. A novel assay for antioxidant potential of speciality Malts. J. Am. Soc. Brew. Chem. 57(4): 133-137.
- Brown, M. D. 1999. Green tea (*Camellia sinensis*) extract and its possible role in the prevention of cancer. Altern. Med. Rev. 4(5): 360-370.
- Chung, F. L., Wang, M., Rivenson, A. 1998. Inhibition of lung carcinogenesis by black tea in Fischer rats treated with a tobacco-specific carcinogen: caffeine as an important constituent. Cancer Res. 58: 4096-4101.
- Caturla, N., Vera-Samper, E., Villalain, J., Mateo, C.R., Micol, V. 2003, The relationship between the antioxidant and the antibacterial properties of galloylated catechins and the structure of phospholipid model membranes, Free Radical Biology and medicine 34 (6): 648-662.
- Dufresne, C. J. and Farnworth, E. R. 2001. A review of latest research findings on the health promotion properties of tea. J. Nutritional Biochem. 12 (7): 404-421.
- Einspahr, J. G., Stratton, S. P., Bowden, G. T. and Alberts, D. S. 2002. Chemoprevention of human skin cancer. Critical Reviews in Oncology/Haematology. 41: 269-285.

- Fujiki, H., Suganuma, M., Imas, K., Nakachi, K. 2002. Green tea : cancer preventive beverage and/or drug Cancer Letters. 188: 9-13.
- Gilchresst, B. A., Eller, M. S., Gelleer, A. C. and Yaar, M. 1999. The pathogenesis of melanoma induced by ultraviolet radiation. New Eng. J. Med. 340: 1341-8.
- Gerhauser, C., Klimo, K., Heiss E., Neumann, I., Gamal Eldee, Knauft, J., Lice, G., Sitthimonchai, S., Frank, N. 2003. Mechanism-based in vitro screening of potential cancer chemopreventive agents. Mutation Research 523-524: 163-172.
- Huang, K. C. 1999. The pharmacology of chinese herbs. Boca Raton : CRC Press, (2nd ed.) pp: 2209-2213.
- Hundley, H. G., Ko, C. K. U. 1961. List of trees, shrubs, herbs, principal climbers, etc. recorded from Burma. Supdt, Govt. Printing.
- Hatano, T., Edamatsu, R., Hiramatsu, M., Mori, A., Fujita, Y., Yasuhara, T., Yoshida, T. and Okuda, T. 1989. Effects of the interaction of tannins with co-existing substances. VI. ¹⁾ Effects of tannins and related polyphenols on superoxide anion radical, and on 1,1-diphenyl-2-picrylhydrazyl radical. Chem. Pharm. Bull. 37(8): 2016-2021.
- Hong, J., Smith, T. J., Ho, C. T., August, D. A. and Yang, C. S. 2001. Effects of purified green and black tea polyphenols on cyclooxygenase- and lipoxygenase-dependent metabolism of arachidonic acid in human colon mucosa and colon tumor tissue. Bio. Pharmacol. 62: 1175-1183.
- Hu, M. and Skibsted, L. H. 2002. Antioxidative capacity of rhizome extract and rhizome knot extract of edible lotus (*Nelumbo nucifera*). Food chem. 76: 327-333.
- Ichihashi, M., Ahmed, N.U., Budiyo, A., Wu, A., Bito, T., Vedo, M., Osawa, T. 2000. Preventive effect of antioxidant on ultra-violet-induced skin cancer in mice J. Dermatological Sci. 23 Suppl. S 45- S 50.

- Imai, K. D. L., Suga, K. and Nakachi, K. 1997. Cancer-preventive effects of drinking green tea among a Japanese population. Prev. Med. 26: 769-775.
- Jung, Y. D., Ellis, L. M. 2001. Inhibition of tumor invasion and angiogenesis by epigallocatechin gallate (EGCG), a major component of green tea. Int. J. Exp. Pathol. 82 (6) 309 – 316.
- Katiyar, S. K., Afaq, F., Azizuddin, K. and Mukhtar, H. 2001. Inhibition of UVB-induced oxidative stress-mediated phosphorylation of mitogen-activated Protein kinase signaling pathways in cultured human epidermal keratinocytes by green tea polyphenol (-)-Epigallocatechin-3-gallate. Toxicol. Appl. Pharmacol. 176: 110-117.
- Katiyar, S. K., Bergamo, B. M., Vajalill, P. K., Elmets, C. A. 2001. Green Tea polyphenols: DNA photodamage & photoimmunology. J. Photochem. Photobiol. Dec 31; 65 (2 – 3): 109 – 114.
- Lambert, J. D. and Yang, C. S. 2003. Cancer chemopreventive activity and bioavailability of tea and tea polyphenols. Mutation Res. (523-524): 201-203.
- Leighton, F. 1999. Plant polyphenol antioxidants and oxidative stress. Held: July 29-30. Catholic University: Santiago, Chile.
- Leung, L. K., Su Y., Chen, R., Zhang, Z., Huang, Y., Chen, Z. Y. 2001. Theaflavins in black tea and catechins in green tea are equally effective antioxidants. J. Nutr. 131(9); 2248-2251.
- Lin, J. and Liang, Y. 2000. Cancer chemoprevention by tea polyphenols. Proc. Natl. Sci. Counc. ROC (B). 24 (1): 1-13.
- Liu, Z., Ma, L. P., Zhou, B., Yang, L. and Liu, Z. L. 2000. Antioxidant effects of green tea polyphenols on free radical initiated and photosensitized peroxidation of human low density lipoprotein. Chem. phys. Lipids. 106: 53-63.

- Liu J. D., Chen, S. H., Lin, C. L., Tsai, S. H., Liang, Y. C. 2001. Inhibition of melanoma growth and metastasis by combination with (-) – epigallocatechin – 3 – gallate and decarbazine in mice. J. Cell Biochem. 83(4) : 631-642.
- Malcolm B. 2003. Mae Salong, China of the north. Benjarong magazine. March. vol 6 (3).
- Matsumoto, N., Kohri, T., Okushio, K., Hara, Y. 1996. Inhibitory effects of tea catechins, black tea extract and oolong tea extract on hepatocarcinogenesis in rat. Jpn. J. Cancer Res. Oct 87 (10): 1034 – 1038.
- McCaleb, R. S., Leigh, E. and Morjen, K. 1999. The encyclopedia of popular herbs: your complete guide to the leading medicinal plants. Roseville, California: Prima Health. pp: 238-247.
- Moure, A., Cruz, J. M. , Franco, D., Dominguez, M., Sineiro, J., Dominguez, H., Nunez, M. J. and Parajo, J. C. 2001. Natural antioxidants from residual sources. Food Chemistry 72 (2) 145 – 171.
- Murray, M. T. 1995. The healing power of herbs the enlightened person's guide to the wonders of medicinal plants. Rocklin, CA: Prima Pub., pp 192 – 195.
- Naik, G. H., Priyadarsini, K. I., Satav, J. G., Banavalikar, M. M., Sohoni, D. P., Biyani, M. K. and Mohan, H. 2003. Comparative antioxidant activity of individual herbal components used in Ayurvedic medicine. Phytochem. 63 (1): 97-104.
- Nanjo, F., Goto, K., Seto, R., Suzuki, M., Sakai, M. and Hara, Y. 1996. Scavenging effects of tea catechins and their derivatives on 1,1-diphenyl-2-picrylhydrazyl radical. Free Radic. Biol. Med. 21(6): 895-902. a T.2002.
- Nakagawa, T., Yokozaw, T. 2002. Direct scavenging of nitric oxide and superoxide by green tea. Food and Chemical Toxicology. 40: 1745 – 1750.
- Pawlik, T. M. and Sondak, V. K. 2003. Malignant melanoma: current state of primary and adjuvant treatment. Critical Rev. Oncology/Hematology. 45: 245-264.

- Robards, K., Prenzler, P. D., Tucker, G., Swatsitong, P. and Glover, W. 1999. Phenolic compounds and their role in oxidative process in fruits. Food Chem. 66: 401-436.
- Rice-Evans, C. A., Miller, N. J. and Paganga, G. 1997. Antioxidant properties of phenolic compounds . Trends in Plant science April Vol 2: 152-159.
- Stratton, S. P., Dorr, R. T. and Alberts, D. S. 2000. The state-of-the-art in chemoprevention of skin cancer. Europ. J. Cancer. 36: 1292-1297.
- Sugihara, N., Ohnishi, M., Imamura, M. and Furuno, K. 2001. Differences in antioxidative efficiency of catechins in various metal-induced lipid peroxidations in cultured hepatocytes. J. Health Science. 47(2): 99-106.
- Stach, D. and Schmitz, O. J. 2001. Decrease in concentration of free catechins in tea over time determined by micellar electrokinetic chromatography. J. Chromatography A. 924; 519-522.
- Toit, R., Volstedt, Y., and Apostolides, Z. 2001. Comparison of the antioxidant content of fruits, vegetables and teas measured as vitamin C equivalents. Toxicology. 166 (1-2): 63-69.
- Tsuōono, Y., Nishino, Y., Komatsu, S., Hsieh, C., Kanemuru, S., Tsuji, I., Natkatsuka, H., Fukao, A., Satoh, H. and Hisamachi, S. 2001. Green Tea and the Risk of Gastric Cancer in Japan. New Engl. J. Med. 344: 632-636.
- Valcic, S., Muders, A., Jacobsen, N. E., Liebler D. C. and Timmerman, B. N. 1999. Antioxidant chemistry of green tea catechins. Identification of Products of the reaction of (-)-Epigallocatechin Gallate with Peroxyl radicals. Chem. Res. Toxicol. 12(4) 382 – 386.
- Yang, C. S. and Wang, Z.Y. 1993. Tea and cancer J. Natl. Cancer Ins. 85: 1038 – 1049.
- Yoshida, T., Mori, K., Hatano, T., Okumura, T., Uehara, I., Komagoe, K., Fujita, Y. and Okuda, T. 1989. Studies on inhibition mechanism of autoxidation by tannins and flavonoids. V ¹⁾ radical-scavenging effects of tannins and related

- polyphenols on 1-1-diphenyl-2-picrylhydrazyl radical. Chem. Pharm. Bulletin. 37(7): 1919-1921.
- Zhang, G., Miura, Y. and Yagasaki, K. 1999. Effects of green, oolong and black teas and related components on the proliferation and invasion of hepatoma cells in culture. Cytotech. 31: 37-44.
- Zhu, A., Wang, X. and Guo, Z. 2001. Study of tea polyphenol as a reversal agent for carcinoma cell lines' multidrug resistance (study of TP as a MDR reversal agent). Nucl. Med. Bio. 28: 735-740.
- Zhu, Q. Y., Zhang, A., Tsang, D., Huang, Y. and Chen, Z. Y. 1997. Stability of green Tea catechins. J. Agric. Food Chem. 45: 4624-4628.

APPENDICES

APPENDIX I
UV spectrophotometer results

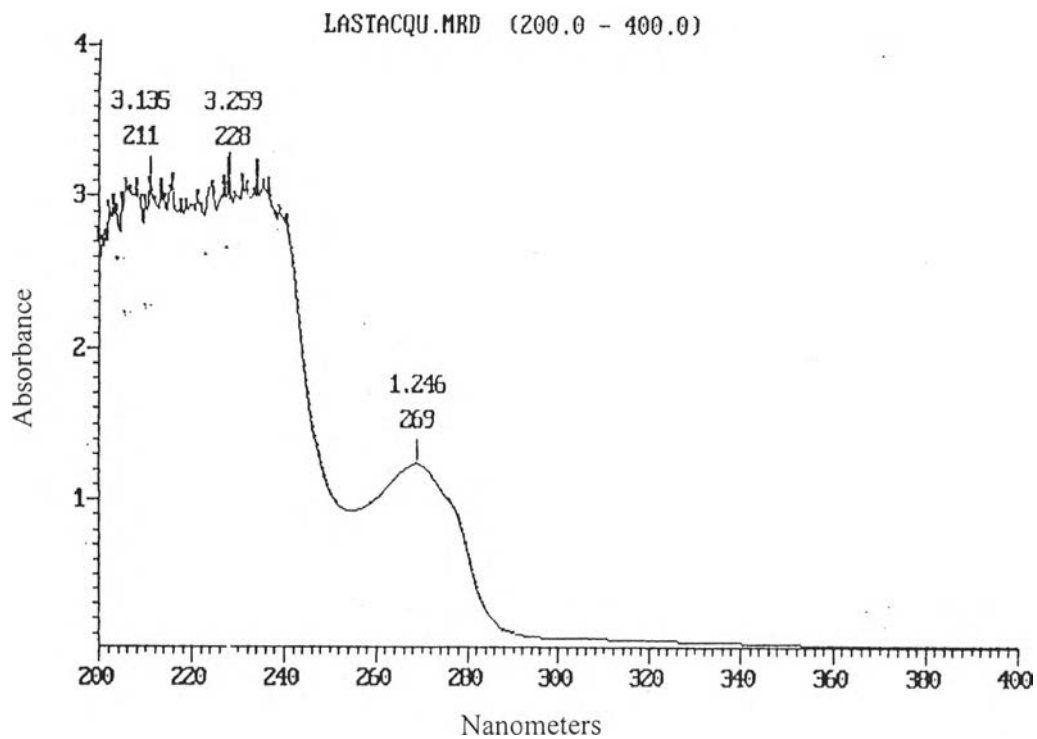


Figure 20. The UV absorption spectrum of Epigallocatechin (EGC)

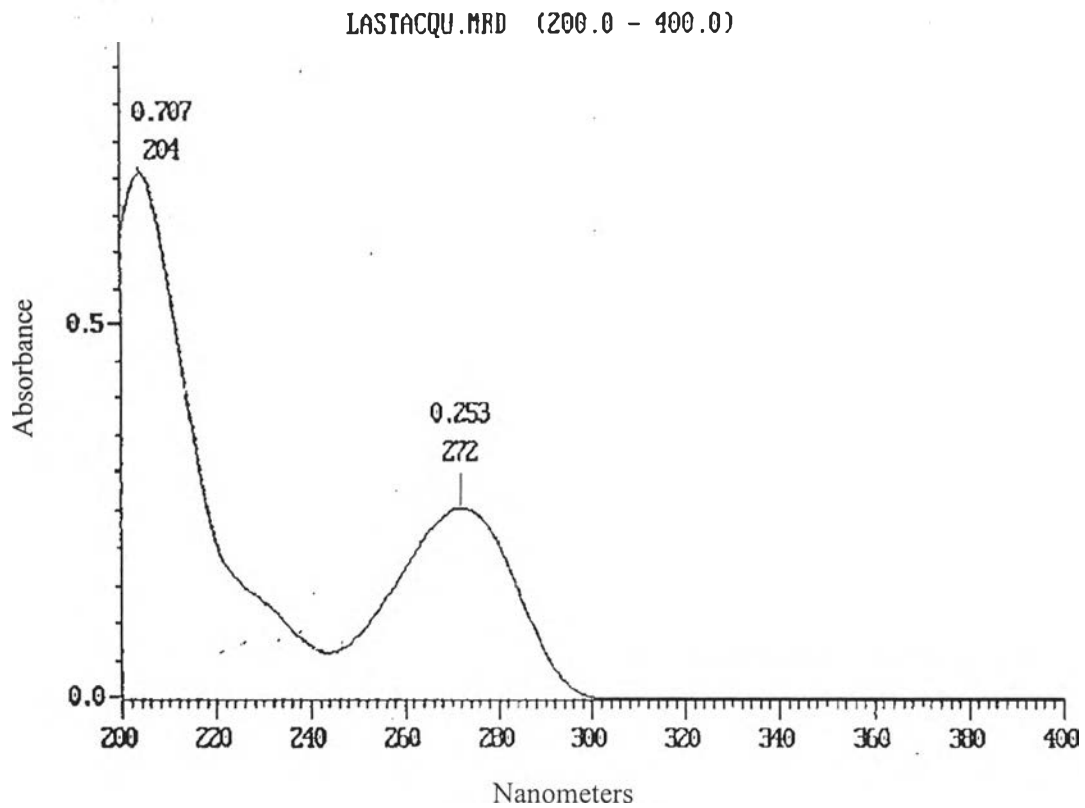


Figure 21. The UV absorption spectrum of caffeine

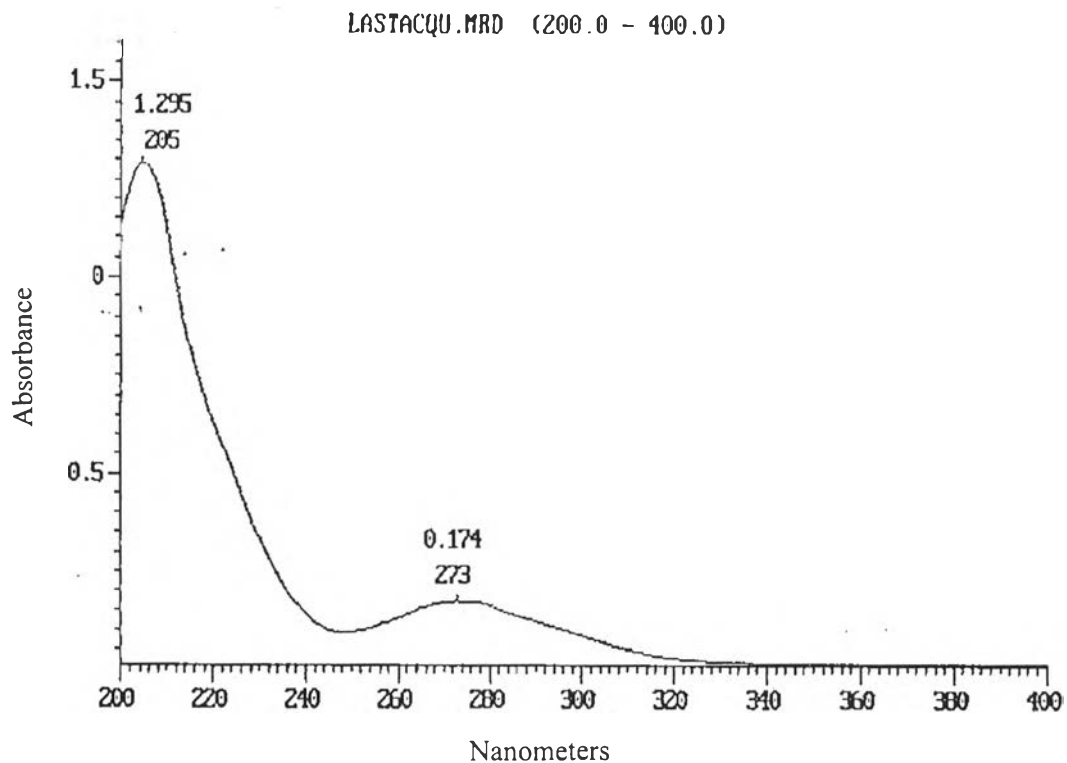


Figure 22. The UV absorption spectrum of Epigallocatechin gallate (EGCG)

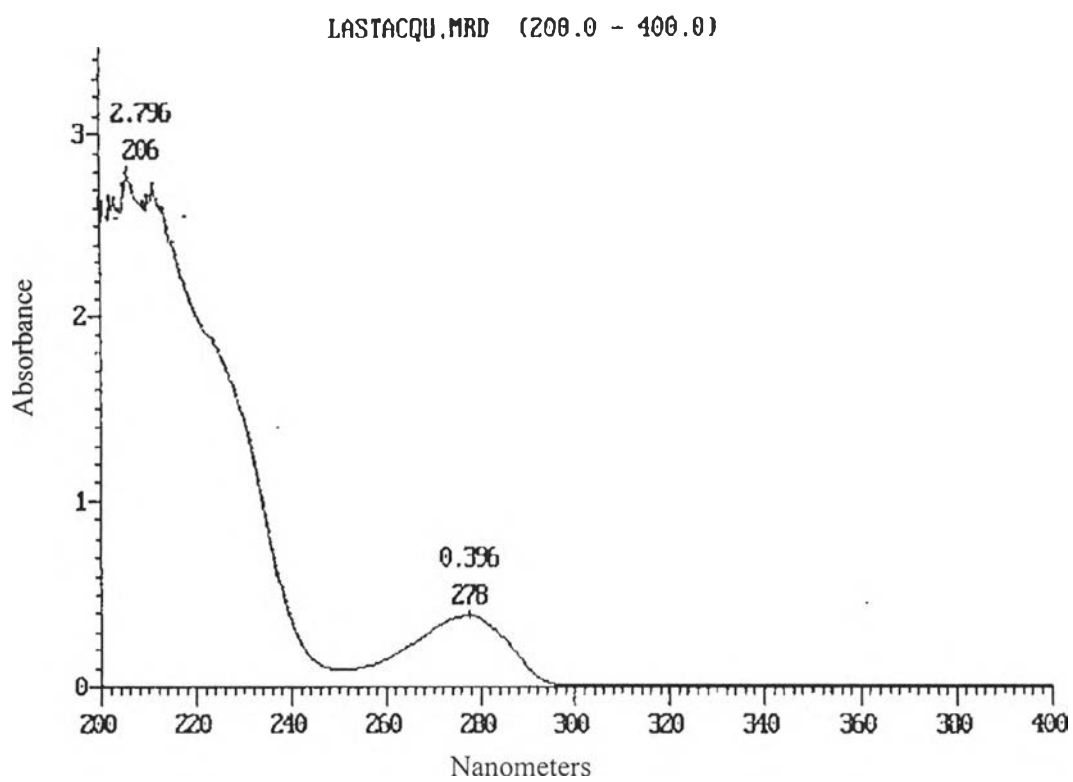


Figure 23. The UV absorption spectrum of Epicatechin (EC)

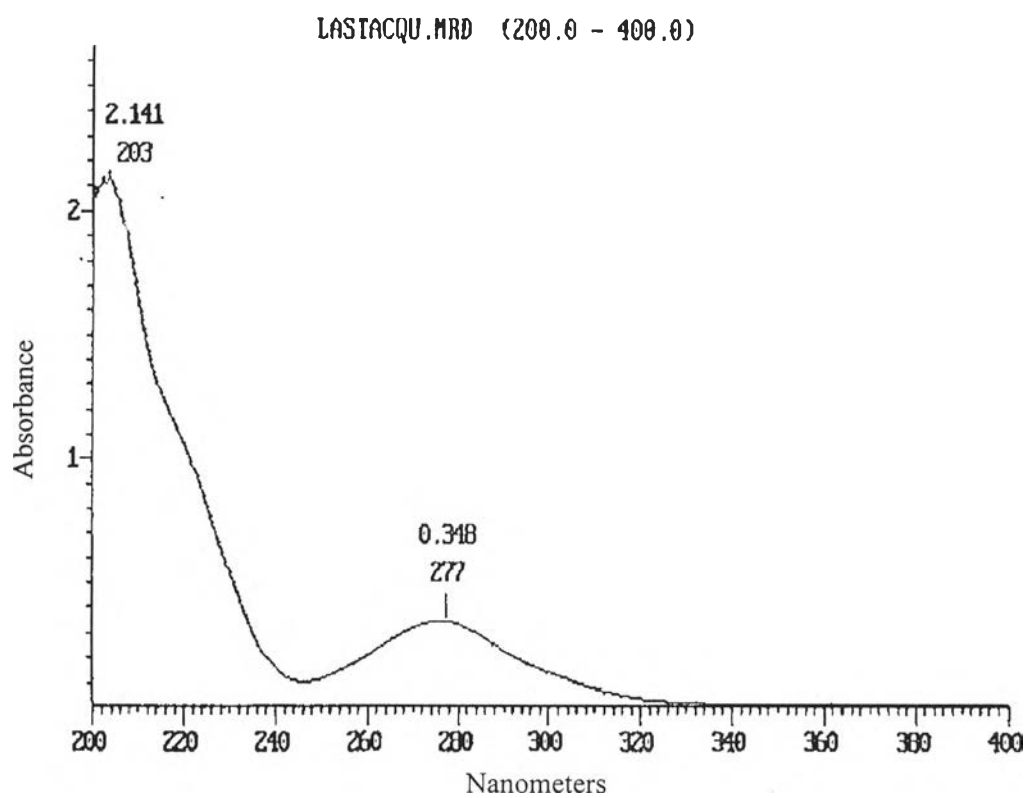


Figure 24. The UV absorption spectrum of Epicatechin gallate (ECG)

APPENDIX II
High performance liquid chromatograms of
green tea extracts

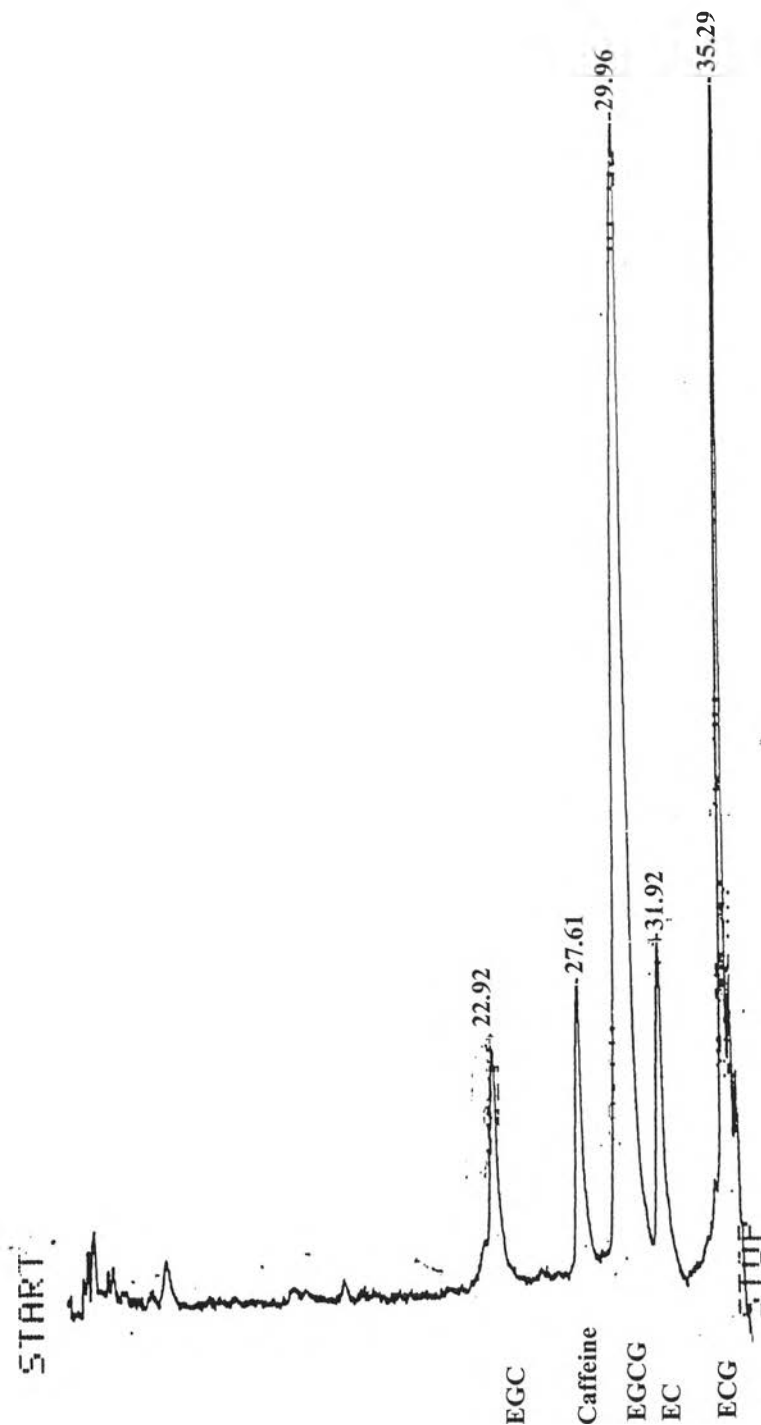


Figure 25. High-performance liquid chromatogram of Japanese green tea extract. EGC = (-)-epigallocatechin, Caffeine, EGCG = (-)-epigallocatechin 3-*O*-gallate, EC = (-)-epicatechin, ECG = (-)-epicatechin 3-*O*-gallate.

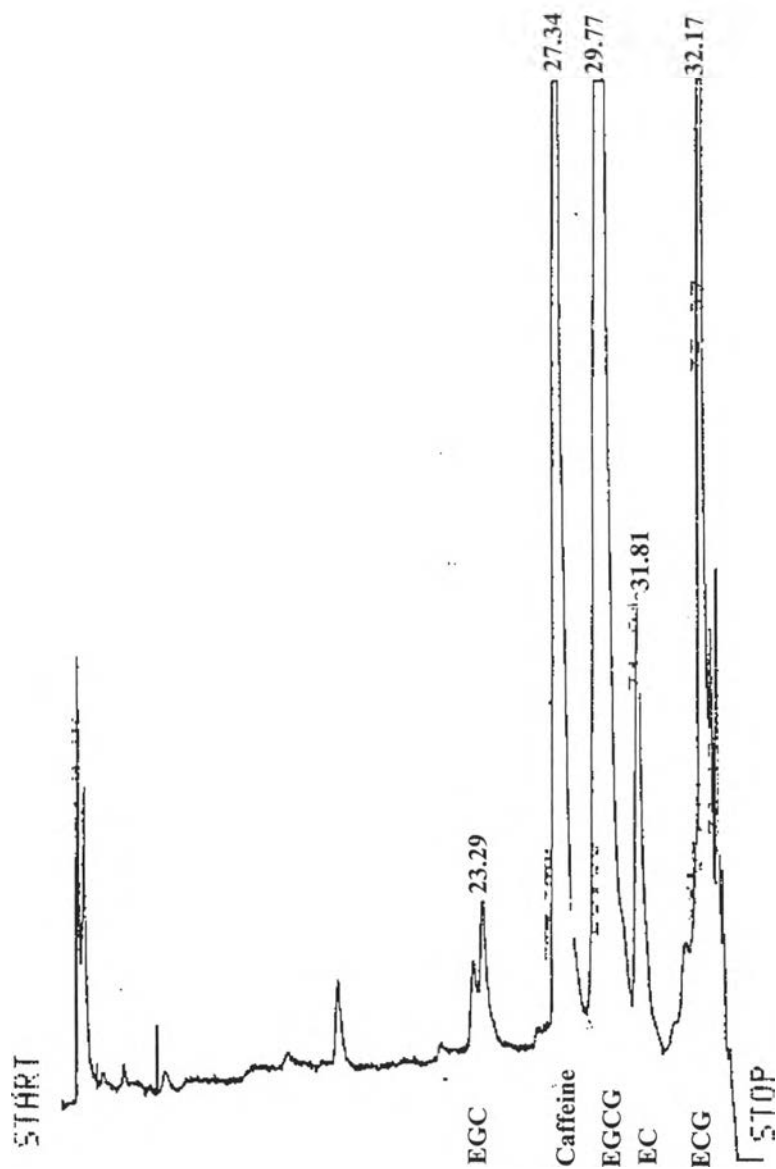


Figure 26. High-performance liquid chromatogram of Myanmar green tea extract.

EGC = (-)-epigallocatechin, Caffeine, EGCG = (-)-epigallocatechin 3-*O*-gallate,

EC = (-)-epicatechin, ECG = (-)-epicatechin 3-*O*-gallate.

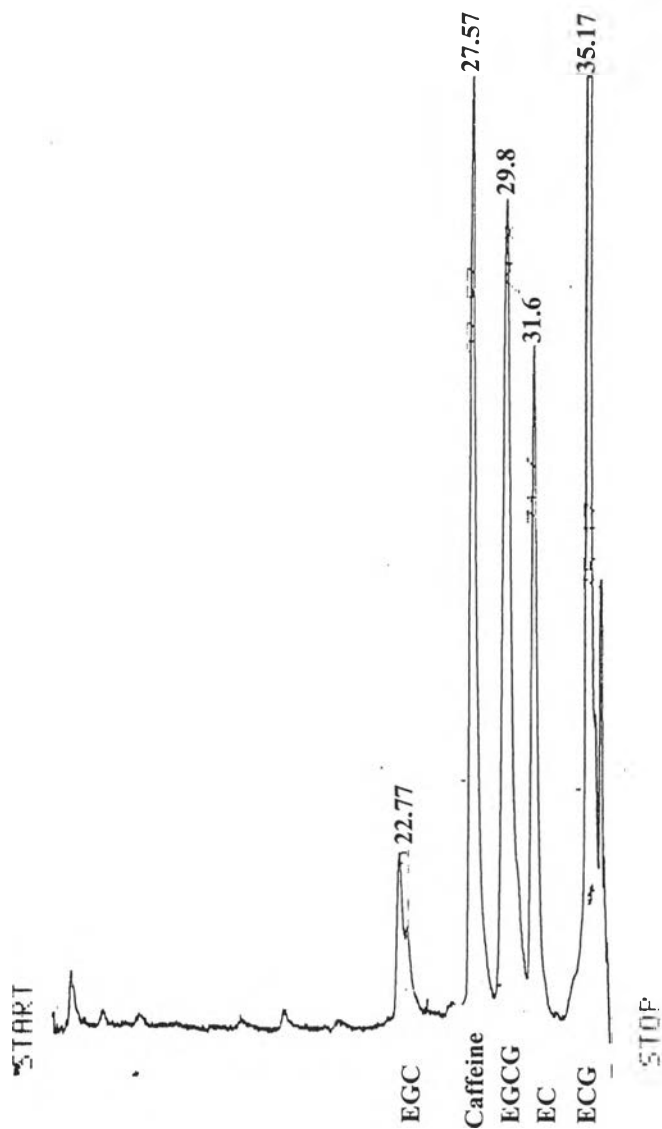


Figure 27. High-performance liquid chromatogram of Thai green tea extract. EGC = (-)-epigallocatechin, Caffeine, EGCG = (-)-epigallocatechin 3-*O*-gallate, EC = (-)-epicatechin, ECG = (-)-epicatechin 3-*O*-gallate.

Appendix III
HPLC validation

1. HPLC assay for EGC analysis

The analytical method for EGC was performed and validated for its accuracy, precision, specificity and linearity.

1.1 Accuracy

Initial amount (μg)	Analytical amount	% Recovery
1	0.99	99
1	0.99	99
1	1	100
		Mean = 99.33
		SD = 0.58

1.2 Precision

1.2.1 Within run precision data

Amount (μg)	Peak area			Mean	SD	%CV
	N ₁	N ₂	N ₁			
1	3025	3094	2981	3033	56.96	1.88
3	19392	18976	19561	19310	301.06	1.56
5	30611	30942	31074	30876	238.52	0.77
7	49375	48699	49944	49339	623.26	1.26

1.2.2 Between run precision data

Amount (μg)	Peak area			Mean	SD	%CV
	D ₁	D ₂	D ₃			
1	3025	2976	3084	3028	54.08	1.78
3	19392	18772	19413	19192	364.17	1.90
5	30611	31119	30438	30722	353.97	1.15
7	49375	48654	49981	49336	664.33	1.35

1.3 Specificity

Figure 6. shows the chromatogram of the standard catechins and caffeine mixture.

1.4 Linearity

Table 1 shows the peak area of the standard EGC solution and Figure 7. shows the calibration curve of the standard EGC.

2. HPLC assay for caffeine analysis

The analytical method for caffeine was performed and validated for its accuracy, precision, specificity and linearity.

2.1 Accuracy

Initial amount (μg)	Analytical amount	% Recovery
1.77	1.78	100.56
1.77	1.78	100.56
1.77	1.77	100
		Mean = 100.38
		SD = 0.33

2.2 Precision

2.2.1 Within run precision data

Amount (μg)	Peak area			Mean	SD	%CV
	N ₁	N ₂	N ₁			
0.59	51329	51287	51122	51216	84.87	0.16
1.47	166711	166650	166271	166544	238.38	0.14
1.77	165032	165279	164798	165036	240.53	0.14
2.36	216126	217159	216235	216507	567.56	0.26

2.2.2 Between run precision data

Amount (μg)	Peak area			Mean	SD	%CV
	D ₁	D ₂	D ₃			
0.59	51329	51419	51278	51342	71.39	0.14
1.47	166711	163200	165431	165114	1776.84	1.08
1.77	165032	163496	166201	164909	1356.64	0.82
2.36	216126	219299	217584	217669	1588.23	0.73

2.3 Specificity

Figure 6. shows the chromatogram of the standard catechins and caffeine mixture.

2.4 Linearity

Table 2 shows the peak area of the standard caffeine solution and Figure 8. shows the calibration curve of the standard caffeine.

3. HPLC assay for EC analysis

The analytical method for EC was performed and validated for its accuracy, precision, specificity and linearity.

3.1 Accuracy

Initial amount μg	Analytical amount	% Recovery
1.5	1.54	102.67
1.5	1.49	99.33
1.5	1.54	102.67
		Mean = 101.56
		SD = 1.92

3.2 Precision

3.2.1. Within run precision data

Amount (μg)	Peak area			Mean	SD	%CV
	N ₁	N ₂	N ₁			
0.5	12280	12158	12397	12278	119.51	0.97
1.5	56565	54442	56147	55718	1124.64	2.02
2.5	94795	94012	95118	94642	568.72	0.600
3.5	138334	135774	133690	135933	2326.06	1.71

3.2.2. Between run precision data

Amount (μg)	Peak area			Mean	SD	%CV
	D ₁	D ₂	D ₃			
0.5	12280	12086	12475	12280	194.50	1.58
1.5	56565	54391	55876	55611	1111.02	1.99
2.5	94795	94551	95468	94938	474.93	0.50
3.5	138334	137864	139426	138541	801.37	0.58

3.3. Specificity

Figure 6. shows the chromatogram of the standard catechins and caffeine mixture.

3.4. Linearity

Table 4 shows the peak area of the standard EC solution and Figure 10. shows the calibration curve of the standard EC.

4. HPLC assay for EGCG analysis

The analytical method for EGCG was performed and validated for its accuracy, precision, specificity and linearity.

4.1 Accuracy

Initial amount μg	Analytical amount	% Recovery
6.15	6.08	98.86
6.15	6.12	99.51
6.15	6.09	99.02
		Mean = 99.13
		SD = 0.34

4.2 Precision

4.2.1. Within run precision data

Amount (μg)	Peak area			Mean	SD	%CV
	N ₁	N ₂	N ₁			
4.1	67675	68461	67132	67756	668.19	0.99
6.15	131020	132180	131318	131506	602.42	0.46
10.25	275383	273495	275884	274921	1259.82	0.46
16.4	546619	557631	549996	551415	5641.54	1.02

4.2.2. Between run precision data

Amount (μg)	Peak area			Mean	SD	%CV
	D ₁	D ₂	D ₃			
4.1	67675	68543	65966	67395	1311.17	1.95
6.15	131020	130487	131159	130889	354.73	0.27
10.25	275383	285467	275113	278654	5901.49	2.12
16.4	546619	528764	539466	538283	8986.09	1.67

4.3. Specificity

Figure 6. shows the chromatogram of the standard catechins and caffeine mixture.

4.4. Linearity

Table 3 shows the peak area of the standard EGCG solution and Figure 9. shows the calibration curve of the standard EGCG.

5. HPLC assay for ECG analysis

The analytical method for ECG was performed and validated for its accuracy, precision, specificity and linearity.

5.1. Accuracy

Initial amount (μg)	Analytical amount	% Recovery
4.35	4.35	100
4.35	4.46	102.52
4.35	4.33	99.54
		Mean = 100.69
		SD = 1.61

5.2 Precision

5.2.1 Within run precision data

Amount (μg)	Peak area			Mean	SD	%CV
	N ₁	N ₂	N ₁			
1.45	69608	68756	69777	69380	547.25	0.79
2.9	153059	164321	153779	157053	6304.56	4.01
4.35	276858	284366	274844	278689	5018.21	1.80
5.8	389712	386651	375963	384109	7218.47	1.88

5.2.2 Between run precision data

Amount (μg)	Peak area			Mean	SD	%CV
	D ₁	D ₂	D ₃			
1.45	69608	69445	68653	69235	510.86	0.74
2.9	153059	151746	154819	153208	1541.91	1.00
4.35	276858	276654	284132	279215	4259.76	1.53
5.8	389712	381136	387354	386067	4430.42	1.15

5.3. Specificity

Figure 6. shows the chromatogram of the standard catechins and caffeine mixture.

5.4. Linearity

Table 5 shows the peak area of the standard ECG solution and Figure 11. shows the calibration curve of the standard ECG.

Appendix IV
Antioxidant activity study

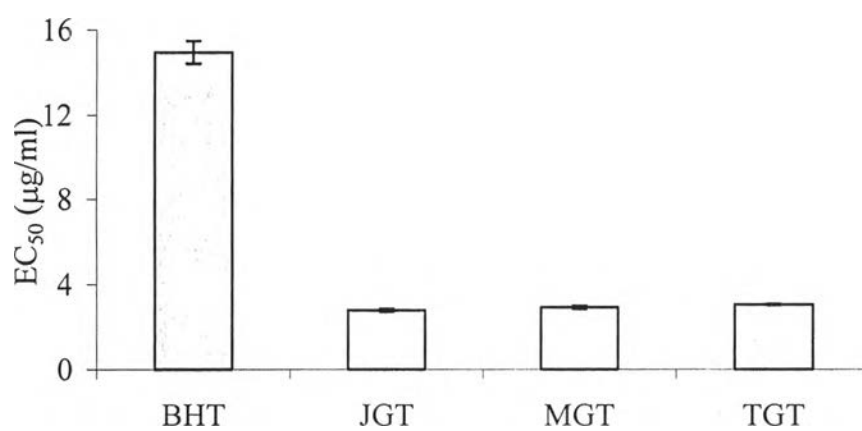


Figure 28. Comparison of the radical scavenging activity of green tea extracts at EC₅₀ studied by DPPH assay

The significance difference was observed in comparison between BHT with green tea extracts and also in comparison among three green tea extracts ($p < 0.05$)

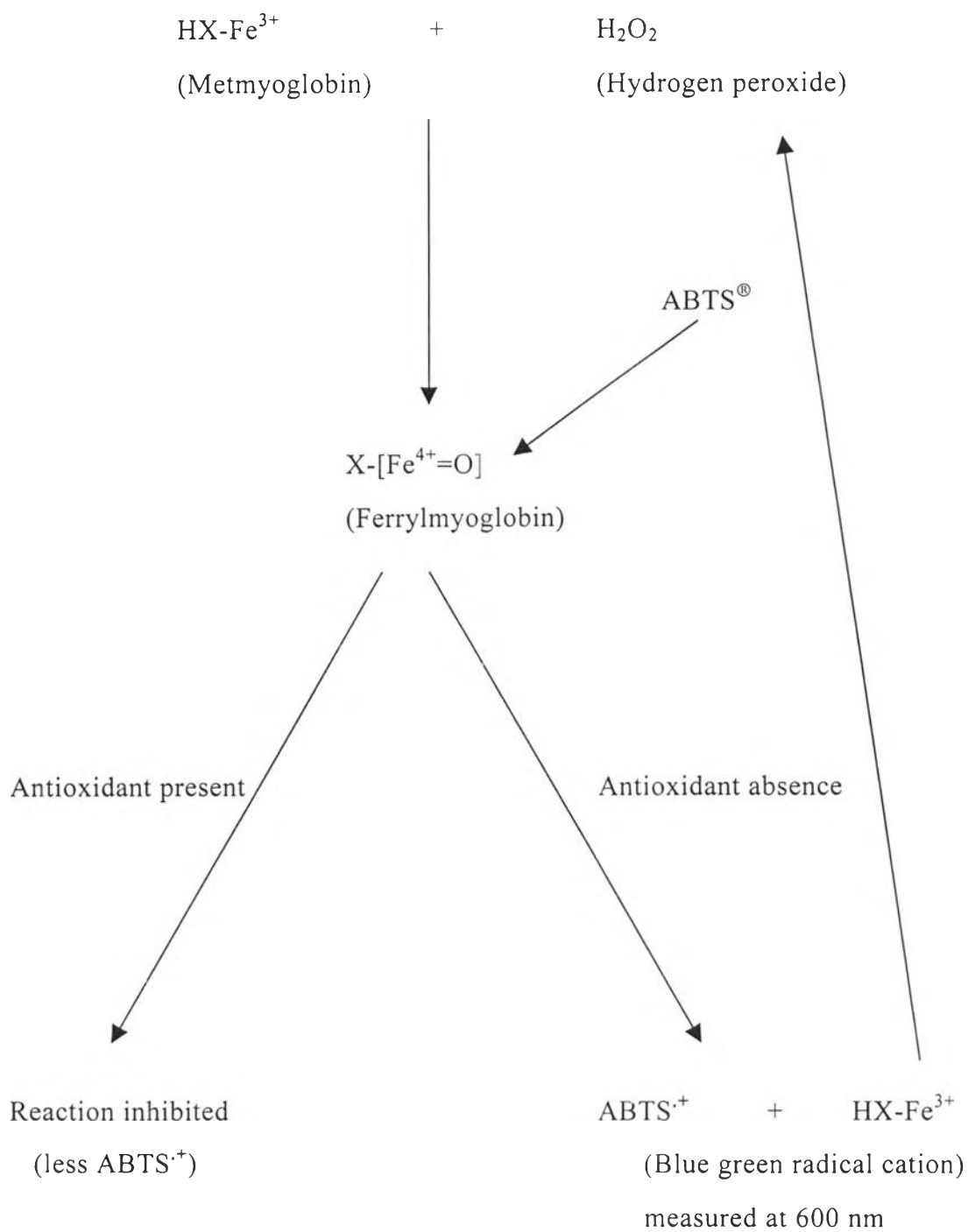


Figure 29. Diagram showing the reaction of total antioxidant status assay (Randox[®] kit)

Appendix V
Cell culture preparation

1. Melanoma Cell line (A375)

Organism	:	<i>Homo Sapiens</i> (human)
Morphology	:	Epithelial
Tissue type	:	skin, malignant melanoma
Growth properties	:	adherent type

2. Cell culture medium

RPMI medium 1640	16.2 g / packet
Antibiotic (Penicillin + Streptomycin) solution	20 ml
Sodium bicarbonate	2 g
Distilled water up to	1 L

16.2 g of RPMI mediumn 1640 powder was dissolved in 750 ml of deionized water by stirring gently at room temperature. Then 2 g of NaHCO₃ and 20 ml of the antibiotic mixture, which contain penicillin and streptomycin, were added and were stirred by magnetic stirrer until dissolved. After that it was made up to 1 L volume by adding more distilled water. The final pH of the medium (7.2) was adjusted by adding slowly with 1 molar NaOH or 1 molar HCl.

Culture medium was sterilized immediately through filtration at 0.22 micro size membrane in laminar airflow hood, sealed and kept at 4°C in the refrigerator. After inactivation of fetal bovine serum at 56°C for 30 min., it was added into the culture medium just before use.

3. MTT solubilization medium

Triton	20 g
HCl	1.72 ml
Isopropanol to	200 ml

20 g of triton was weighted and mixed with one third of isopropanol. Then 1.72 ml of HCl was added drop by drop and the mixture was shaken thoroughly. Then it was made up to 200 ml volume by adding more isopropanol into the volumetric flask.

Appendix VI

Statistical data for comparison among green tea extracts

1. Comparison on the total polyphenol content

Table 17. The statistical data for the comparison of total polyphenol content

Extract	Total polyphenol content		
	N ₁	N ₂	N ₃
JGT	55.95	57.34	59.18
MGT	56.28	55.90	52.35
TGT	51.14	53.75	55.60

Analysis of Variance Procedure

1. Dependent Variable: CONTENT

Source	DF	Sum of Squares	Mean Square	F value	Pr>F
Model	2	24.76506667	12.382553333	3.01	0.1244
Error	6	24.68953333	4.11492222		
Corrected Total	8	49.45460000			

R-Square	C.V.	Root MSE	CONTENT Mean
0.500764	3.669771	2.02852711	55.27666667

Source	DF	Anova SS	Mean Square	F Value	Pr>F
EXTRACT	2	24.76506667	12.38253333	3.01	0.1244

2. T tests (LSD) for variable : CONTENT

NOTE :This test controls the type I comparisonwise error rate not the experimentwise error rate.

$$\text{Alpha} = 0.05 \quad \text{df} = 6 \quad \text{MSE} = 4.114922$$

$$\text{Critical Value of T} = 2.45$$

$$\text{Least Significant Difference} = 4.0528$$

Means with the same letter are not significantly different.

T Grouping	Mean	N	EXTRACT
A	57.490	3	J
A	54.843	3	M
A	53.497	3	T

2. Comparison on the radical scavenging activity of green tea extracts on DPPH radical

Table 18. The statistical data for the comparison of EC₅₀ on DPPH radical

Extract	EC ₅₀ (µg/ml)		
	N ₁	N ₂	N ₃
JGT	2.72	2.83	2.85
MGT	2.88	2.92	3.02
TGT	3.04	3.09	3.08

Analysis of Variance Procedure

1. Dependent Variable: EC₅₀

Source	DF	Sum of Squares	Mean Square	F value	Pr>F
Model	2	0.10940000	0.05470000	15.19	0.0045
Error	6	0.02160000	0.00360000		
Corrected Total	8	0.13100000			

R-Square	C.V.	Root MSE	EC ₅₀ Mean
0.835115	2.043133	0.06000000	2.93666667

Source	DF	Anova SS	Mean Square	F Value	Pr>F
EXTRACT	2	0.10940000	0.05470000	15.19	0.0045

2. T tests (LSD) for variable : EC₅₀

NOTE : This test controls the type I comparisonwise error rate not the experimentwise error rate.

$$\text{Alpha} = 0.05 \quad \text{df} = 6 \quad \text{MSE} = 0.0036$$

$$\text{Critical Value of T} = 2.45$$

$$\text{Least Significant Difference} = 0.1199$$

Means with the same letter are not significantly different.

T Grouping	Mean	N	EXTRACT
A	3.07000	3	T
B	2.94000	3	M
C	2.80000	3	J

3. Comparison on the radical scavenging activity of BHT and green tea extracts on DPPH radical

Table 19. The statistical data for the comparison of EC₅₀ on DPPH radical

Extract	EC ₅₀ (µg/ml)		
	N ₁	N ₂	N ₃
BHT	15.49	14.92	14.44
JGT	2.72	2.83	2.85
MGT	2.88	2.92	3.02
TGT	3.04	3.09	3.08

Analysis of Variance Procedure

1. Dependent Variable: EC₅₀

Source	DF	Sum of Squares	Mean Square	F value	Pr>F
Model	3	324.82980000	108.27660000	1508.56	0.0001
Error	8	0.57420000	0.07177500		
Corrected Total	11	325.40400000			

R-Square	C.V.	Root MSE	EC ₅₀ Mean
0.998235	4.510245	0.26790857	5.94000000

Source	DF	Anova SS	Mean Square	F Value	Pr>F
EXTRACT	3	324.82980000	108.27990000	1508.56	0.0001

2. T tests (LSD) for variable : EC₅₀

NOTE : This test controls the type I comparisonwise error rate not the experimentwise error rate.

$$\text{Alpha} = 0.05 \quad \text{df} = 8 \quad \text{MSE} = 0.071775$$

$$\text{Critical Value of T} = 2.31$$

$$\text{Least Significant Difference} = 0.5044$$

Means with the same letter are not significantly different.

T Grouping	Mean	N	EXTRACT
A	14.9500	3	B
B	3.0700	3	T
B	2.9400	3	M
B	2.8000	3	J

4. Comparison of the total antioxidant activity

Table 20. The statistical data for the comparison of the total antioxidant activity

Extract	Total antioxidant activity (mmol/l)		
	N ₁	N ₂	N ₃
JGT	1.02	0.87	0.95
MGT	0.99	0.76	0.84
TGT	0.72	0.87	0.91

Analysis of Variance Procedure

1. Dependent Variable: TOTAL

Source	DF	Sum of Squares	Mean Square	F value	Pr>F
Model	2	0.02068889	0.01034444	1.06	0.4037
Error	6	0.05860000	0.00976667		
Corrected Total	8	0.07928889			

R-Square	C.V.	Root MSE	TOTAL Mean
0.259313	11.21612	0.09882645	0.88111111

Source	DF	Anova SS	Mean Square	F Value	Pr>F
EXTRACT	2	0.02068889	0.01034444	1.06	0.4037

2. T tests (LSD) for variable : TOTAL

NOTE : This test controls the type I comparisonwise error rate not the experimentwise error rate.

$$\text{Alpha} = 0.05 \quad \text{df} = 6 \quad \text{MSE} = 0.009767$$

$$\text{Critical Value of T} = 2.45$$

$$\text{Least Significant Difference} = 0.1974$$

Means with the same letter are not significantly different.

T Grouping	Mean	N	EXTRACT
A	0.94667	3	J
A	0.86333	3	M
A	0.83333	3	T

5. Comparison on the cytotoxicity assay

Table 21. The statistical data for the comparison of EC₅₀ on melanoma cell line by MTT assay

Extract	EC ₅₀ (µg/ml)		
	N ₁	N ₂	N ₃
JGT	171	173	175
MGT	199	195	200
TGT	203	206	209

Analysis of Variance Procedure

1. Dependent Variable: EC₅₀

Source	DF	Sum of Squares	Mean Square	F value	Pr>F
Model	2	1778.00000000	889.00000000	133.35	0.0001
Error	6	40.00000000	6.66666667		
Corrected Total	8	1818.00000000			

	R-Square	C.V.	Root MSE	EC ₅₀
Mean	0.977998	1.342455	2.58198890	192.33333333

Source	DF	Anova SS	Mean Square	F Value	Pr>F
EXTRACT	2	1778.00000000	889.00000000	133.35	0.0001

2. T tests (LSD) for variable : EC₅₀

NOTE : This test controls the type I comparisonwise error rate not the experimentwise error rate.

$$\text{Alpha} = 0.05 \quad \text{df} = 6 \quad \text{MSE} = 6.666667$$

$$\text{Critical Value of T} = 2.45$$

$$\text{Least Significant Difference} = 5.1585$$

Means with the same letter are not significantly different.

T Grouping	Mean	N	EXTRACT
A	206.000	3	T
B	198.000	3	M
C	173.000	3	J

BIOGRAPHY

Miss Cho Sanda Aung was born on 3rd November, 1975 in Yangon, Myanmar. She received her Bachelor of Pharmacy (B.Pharm.) from the Institute of Pharmacy, Ministry of Health, Yangon, Myanmar in 2000.

