REFERENCES



Almusallam, A. A., Alzarban, S. S., Fasasi, Y. A., Kroppenstedt, R. M., and Stackebrandt, E. 2003. *Amycolatopsis keratiniphila* sp.nov., a novel keratinolytic soil actinomycete from Kuwait. Int. J. Syst. Evol. Microbiol. 53: 871-874.

Arai, T. 1975. Culture Media for Actinomycetes. In The Society for Actinomycetes. Japan.

- Asolkar, R. N., Maskey, R. P., Helmke, E., and Laatsch, H. 2002. Chalcomysin B, a new macrolide centibiotic from the marine isolate *streptomyces* sp.B7064. <u>J. Antibiot</u>. 55(10): 893-898.
- Bala, S., Khanna, R., Dadhwal, M., Prabaharan, S. R., Shivaji., S., Cullum, J., and Lal, R. 2004 Reclassification of *Amycolatopsis mediterranei* DSM 46095 as *Amycolatopsis rifamycinica* sp.nov. <u>Int. J. Syst. Evol. Microbiol.</u> 54: 1145-1149.
- Ballini, R., and Bosica, G. 1998. Synthesis of (E)-4-oxonon-2-enoic acid, a natural antibiotic produced by *Streptomyces olivaceus*. J. Nat. Prod. 61: 673-674.

Berdy, J. 2005. Bioactive microbial metabolites. J. Antibiot. 58: 1-26.

- Bertasso, M., Holzenkampfer, M., Zeeck, A., Antonia, F., and Fiedler, H. P. 2001. Bagremycin A and B, novel antibiotics from *Streptomyces* sp. TU 4128. J. Antibiot. 54(9): 730-736.
- Bertasso, M., Holzenkampfer, M., Zeeck, A., Stackbrandt, E., Beil, W., and Fiedler, H. P. 2003.
 Ripamycin and other polycyclic macrolactams from *Streptomyces* sp. TU 6239. J. Antibiot. 56(4): 364-371.
- Bordoloi, G. N., Kumari, B., Guha, A. 2001. Isolation and structure elucidation of new antifungal and antibacterial antibiotic produced by *Streptomyces* sp. 201. <u>Biosci. Biotechnol.</u> <u>Biochem.</u> 65(8): 1856-1858.
- Burg, R. W., Miller, B. M., Baker, E.E., and Omura, S. 1979. Avermectins, new family of potent anthelmintic agent producing organism and fermentation J. Antibiot. 15: 361-367.
- Brock, T. D., Madigan, M. T., Martinko, J. M., and Parker, J. 1993. <u>Biology of microorganisms</u>. 7th ed. New Jersy: Prentice Hall.
- Chun, T., Kim, S. B., Oh, Y. K., Seong, C., Lee D., Bae, K. S., Lee, K., Keng S., Hah. Y. C., and Goodfollow, M. 1999. *Amycolatopsis thermoflava* sp. nov., a novel soil actinomycete form Hainan Island, China. <u>Int. J. Syst. Bacteriol</u>. 49: 1369-1373.

- Chung, Y. R., Sung, K.C., Mo, H. K., Son, D.Y., Nam, J.S., Chun, J., and Bae, K. S. 1999. *Kitasatospora cheerisanensis* sp.nov., a new species of the genus *kitasatospora* that produces an antifungal agent. <u>Int. J. Syst. Bacteriol.</u> 49: 753-758.
- Collins, M. D., Pirouz, T., Goodfellow, M., and Minnikin, P. E. 1977. Distribution of menaquinones in actinomycetes and corynebacteria. J. Gen. Microbiol. 100: 221-230.
- Coyne, M. S. 1999. <u>Soil microbiology: An Exploration Approach</u>. Delmar. An International Thomson.
- Cross, T. 1994. Growth and examination of actinomycetes some guidelines. In J. B. Holt, N. R. Krieg, P. H. A. Sneath, J. T. Staley, and S. T. Williams (eds), <u>Bergev's manual of determinative bacteriology</u>. 9th ed., pp. 605-623. Baltimore: The Williams and Wilkins Co.
- Deman, J. M., De man, L., and Gupta, S. 1986. Texture and microstructure of soybean curd (tofu) as affected by different coagulants. <u>Food. Micro. Struct</u>. 5: 83-89.
- Embley, T. M., Smida, J., and Stackebrandt, E. 1988. The phylogeny of mycolate-less wall chemotype IV actinomycetes and the description of *Pseudonocardiaceae* fam. nov. <u>Syst.</u> <u>Appl. Microbiol</u>. 11: 44-52.
- Embley, T. M., O'Donnell, A. G., Rostron, J., and Goodfellow, M. 1988. Chemotaxonomy of wall type IV actinomycetes which lack mycolic acids. <u>J. Gen. Microbiol</u>. 134. 953-960.
- Ezaki, T., Hashimoto, Y., and Yabuuchi, E. 1989. Fluorometric deoxyribonucleic aciddeoxyribonucleic acid hybridization in microdilution well as alternative to membrane filter hybridization in which radioisotopes are used to determine genetic relatedness among bacteria strains. Int J. Syst. Bacteriol. 39: 224-229.
- Felsenstein, J. 1985. Confidence limits on phylogenies: an approach using the bootstrap. Evolution. 39: 783-791.
- Fiedler, H. P., Nega, M., Pfefferle, C., Groth, I., Kempter, C., Stephan, H., and Metzger, J. W.
 1996. Kanchanamycins, now polyol macrolide antibiotics produced by *Streptomyces* olivaceus TU 4018. I. taxonomy, fermentation, isolation and biological activities. J. Antibiot. 49(8): 758-764.
- Flowers, T. H., and Williams, S. T. 1977. Measurements of growth rates of *Streptomyces*: Comparision of turbidimetric and gravimetric techniques. J. Gen. Microbiol. 98: 285-289.
- Franco, C. M., Gandhi, J. H., Chatterjee, S., and Ganguli, B. N. 1987. Swalpamycin, a new macrolide antibiotic. I. taxonomy of the producing organism, fermentation, isolation and biological activity. J. Antibiot. 40(10): 1361-1367.

- Franberge, E., Petersson, C., Lundgren, L. N., and Schnurer, J. 2000. Streptomyces halstedii K122 produces the antifungal compounds bafilomycin B1 and C1. <u>Can. J. Microbiol</u>. 46(8): 753-758.
- Furumai, T., Eto, K., Sasaki, T., Higuchi, H., Onaka, H., Saito, N. Fujita, T., Naoki, H., and Igarashi, Y. 2002. TPU-0037-A, B, C and D, novel lydicamycin congeners with anti-MRSA activity from *Streptomyces platensis* TP-A0598. J. Antibiot. 55(10): 873-880.
- Gebhardt, K., Pukall, R., and Fiedler, H. P. 2001. Streptocidins A-D, novel cyclic decapeptide antibiotics produced by *Streptomyces* sp. Tu 6071. J. Antibiot. 54(5): 428-433.
- Glasby, J. S. 1993. Encyclopedia of Antibiotics 3rded. Chichester: John Wiley and Sons Inc.
- Gonzalez, H. G., Castro, R. J., Canedo, H. M. Diaz, M., Fernandez, J. M. and Santamaría, R. I. 2002. Radamycin, a novel thiopeptide produced by *Streptomyces* sp.RSP9. I. taxonomy, fermentation, isolation and biological activities. <u>J. Antibiot</u>. 55(4): 383-390.
- Goodfellow, M. 1988. Actinomycetes in Biotechnology. pp.1-30. San Diego: Academic Press Inc.
- Goodfellow, M., and Board, R. G. 1980. <u>Microbiology classification and identification</u>. pp. 599-861. London. Academic Press Inc.
- Goodfellow, M., Brown, A. B., Cai, J., Chun, J., and Collins M. D. 1997. *Amycolatopsis japonicum* sp. nov., an actinomycete producing (S,S)-N,N'- ethylenediaminedisuccinic acid. <u>Svst. Appl. Microbiol</u>. 20: 78-84.
- Goodfellow, M., Kim. S. B., Minikin, D. E., Whitehead, D., Zhou, E., and Rose, H. M. 2001. *Amycolatopsis sacchari* sp.nov., a moderately themophilic actinomycete isolated from vegetable matter. <u>Int. J. Syst. Evol. Microbiol</u>. 51:187-193.
- Goodfellow, M., and Williams S. T. 1983. Ecology of actinimycetes. <u>Ann. Rev. Microbiol</u>. 37: 189-216.
- Goodfellow, M., Williams, S. T., and Mordarski, M. 1988. <u>Actinomycetes in Biotechnology</u>, pp. 1-88. London: Academie press.
- Gorajana, A., Venkatensa, M., Vinjamuri, S., Peela, S., Jangam, P., and Zeeck, A. 2006. Resistoflavine, cytotoxic compound from a marine actinomycete, *Streptomyces chibaensis* AUBN₁/7. <u>Microbiol. Res</u>. 26:1-6.
- Gordon, R. E., Barnett, D. A., Handerhan, J. E., and Pang, C. H. 1994. Nocardia coeliaca, Nocardia autotrophica, and the Nocardia strains. Int. J. Syst. Evol. Microbiol. 24: 54-63.

- Groth, I., Rodriguez, C., Schutze, B., Schmitz., P., Leistner, E., and Goodfellow, M. 2004. Five novel Kitasatospora species from soil: Kitasatospora arboriphila sp.nov., K. gansuensis sp.nov., K. nipponensis sp.nov., K. paranensis sp.nov. and K. terrestris sp.nov. Int. J. Svst. Evol. Microbiol. 54: 2121-2129.
- Groth, I., Schütze B., Boettcher, T., Pullen, C.B., Rodriguez, C., Liestner, E., and Goodfellow,
 M., 2003. *Kitasatospora putterlickiae* sp.nov., isolated from rhizosphere soil, transfer of *Streptomyces kifunensis* to the genus *Kiitasatospora* as *Kitasatospora kifunensis* comb.
 nov., and emended description of *Streptomyces aureofaciens* Duggar 1948. <u>Int. J. Svst.</u>
 <u>Evol. Microbiol.</u> 53: 2033-2040.
- Harold, C. N. 1983. In vitro activity of Midecamycin, a new macrolide antibiotic.<u>Antimicrobial</u> agent and chromato. 24(3): 443-444.
- Hessen, A., Kothe, H. W., and Kroppenstedt, R. M. 1987. Transfer of *Pseudonocardia azurea* and *Pseudonocardia fastidiosa* to the genus *Amycolatopsis*, with emended species description. <u>Int. J. Svst. Bacteriol</u>. 37: 292-295.
- Holt, J. G. 1989. Filamentous actinomycetes and related bacteria. In S. T. Williams, M. E. Sharpe, and J. G. Holt (eds.), <u>Bergev's Mannual of Systematic Bacteriology</u>, vol. 4, pp. 2333-2450.
 Baltimore: William and Wilkins.
- Hopmann, C., Kurz, M., Bronstrup, M., Wink, J., and Lebeller, D. 2002. Isolation and structure elucidation of vancoresmycin a new antibiotic from *Amycolatopsis* sp. ST 101170. <u>Tetra.</u> <u>Lett.</u> 43(3): 435-438.
- Hosotani, N., Kumagai, K., Nakagawa, H., Schimatani, T., and Saji, I. 2005. Antimycins A 10 approximately A16, seven new antimycin antibiotics produced by *Streptomyces* spp. SPA-10191 and SPA-8893. J. Antibiot. 58(7): 460-467.
- Hotta, K., Yoshida, M., Hamada, M., and Okami, Y. 1980. Studies on new aminoglycoside antibiotics, istamycins, from an actinomycete isolated from a marine environment. III. nutritional effects on istamycin production production and additional chemical and biological properties of istamycins. J. Antibiot. 33(12): 1515-1520.
- Huang, Y., Pasciak, M., Liu, Z., Xie, Q., and Gamian, A. 2004. Amycolatopsis palatopharyngis sp. nov., a potentially pathogenic actinomycete isolated from a human clinical source. Int. J. Svst. Evol. Microbiol. 54: 359 – 363.
- Huang, Y., Qi, W., Lu, Z., Liu, Z., and Goodfellow, M. 2001. Amycolatopsis ribida sp. nov., a new Amycolatopsis species from soil. Int. J. Syst. Evol. Microbiol. 51: 1093 – 1097.

- Igarashi, Y., In, Y., Ishida, T., Fujita, T., Yamakawa, T., Onaka, H., and Furumai, T. 2005. Absolute configuration of TPU-0043, a pentaene macrolide from *Streptomyces* sp. J. <u>Antibiot</u>. 58(8): 523-525.
- Igarashi, M., Tsuchida, T., Kinishita, N., Kamijima, H., Sawa, R., Nagannawa, H., Hamada, M., Takeuchi, T., Yamazaki, K., and Ishizuka, M. 1998. Cremimycin, a novel 19- membered macrocyclic lactam antibiotic, from *Streptomyces* sp. J. Antibiot. 51(2): 123-129.
- Igarashi, Y., Futamata, K., Fujita, T., Sekino, A., Senda, H., Nobki, H., and Furumai, T. 2003. Yatakemycin, a novel antifungal antibiotic produced by *Streptomyces* sp. TP-A0356. <u>J.</u> <u>Antibiot</u>. 56(2): 107-113.
- Igarashi, M., Shida, T., Sasaki, Y., Kinoshita, N., Noganawa, H., Hamada, M., and Takeuchi, T. 1999. Vinylmycin, a new desipeptide antibiotic, from *Streptomyces* sp. J. Antibiot. 52(10): 873-879.
- Ivanova, V. 1997. New macrolactone of the desertomycin family from *Streptomyces spectabilis*. <u>Prep. Biochem. Biotechnol</u>. 27(1): 19-38.
- Jensen, P. R., Dwing, R., and Fenical, W. 1991. Distribution of actinomycetes in near shore tropical marine sediments. <u>Appl. Microbiol</u>. 57(4): 1102-1108.
- Jongrungruanchok, S., Tanasupawat, S., Kitakoop, P., Bavovada, R., Kobayashi, H., and Kudo, T. 2006. Identification of *Streptomyces* and *Kitasatospora* strains from Thai soils with geldanamycin production strain. <u>Actinomycetologica</u>. 20(1): 1-4.
- Kakinuma, K., Hanson, C. A., and Rinehart, K. L. 1976. Spectinabilin, a new nitro-containing metabolite isolated from *Streptomyces spectabilis*. <u>Tetrahedron</u>. 32: 217-222.
- Kim, S. B., and Goodfellow, M. 1999. Reclassification of *Amycolatopsis rugosa* Lechevalier *et al.* 1986 as *Prauserella rugosa* gen. nov., comb. nov. <u>Int. J. Syst. Bacteriol</u>. 49: 507-512.
- Kim, S. B., Lonsdale, J. Seong. C. N., and Goodfellow, M. 2003. *Streptacidiphilus* gen. nov., acidophilic actinomycetes with wall chemotype I and emendation of the family Streptomycetaceae (Waksman and Henrici (1943)^{AL}) emend. Rainey et al. 1997. <u>Antonie van Leeuwenhoek</u>. 83: 107-116.
- Kim, S. B., Sahin, N., Tan G. Y., Zakrzewska, J., and Goodfellow, M. 2002. Amycolatopsis eurytherma sp.nov., a thermophilic actinomycete isolated from soil. <u>Int. J. Syst. Evol.</u> <u>Microbiol</u>. 52: 889 – 894.
- Komagata, K. and Suzuki, K. 1987. Lipid and cell-wall analysis in bacterial systematics. <u>Methods In Microbiology</u> 19: 161-207. London: Academic Press.

- Kroppenstedt, R. M. 1985. Fatty acid and menaquinone analysis of actinomycetes and related organism. In M. Goodfellow, and D. E. Minnikin (eds.), <u>Chemical Methods in Bacterial</u> <u>Systematics</u>, pp. 173-199. London: Academic Press.
- Kumar, S., Tamura, K., Jakobsen, I. B., and Nei, M. 2001. MEGA2 : molecular evolutionary genetics analysis software. <u>Bioinformatics</u>. 17: 1244-1245.
- Kusakabe, H. and Isono, K. 1988. Taxonomic studies on *Kitasatosporia cystarginea* sp. nov., which produces a new antifungul antibiotic cystargin. J. Antibiot. 41: 1758-1762.
- Kutzner, H. J. 1981. <u>A handbook on habitats. isolation and identification of bacteria</u>. 2: 2028-2029. Berlin: springer-Verlag.
- Labeda, D. P. 1988. Kitasatosporia mediocidica sp. nov. Int. J. Syst. Bacteriol. 38: 287-290.
- Lackner, H., Bahner, I., Shigematsu, N., Pannell, L. K., and Mauger, A. B. 2000. Structures of five compounds of the actinomycin Z complex from *Streptomyces fradiae*, two of which contain 4- chlorothreonine. J. Nat. Prod. 63(3): 352-356.
- Labeda, D. P., Donahue, J. M., Wiiliams N. M., Sells, S. F., and Henton, H. M. 2003. *Amycolatopsis kentuckyensis* sp.nov., *Amycolatopsis lexingtonensis* sp.nov., and *Amycolatopsis pretoriensis* sp.nov., isolated from equine placentas. <u>Int. J. Svst. Evol.</u> <u>Microbiol.</u> 53: 1601-1605.
- Labeda, D. P., and Kroppenstedt, R. M. 2000. Phylogenetic analysis of *Saccharothix* and related taxa : proposal for *Actinosynnemataceae* fam. nov. <u>Int J. Syst. Evol. Microbiol</u>. 50: 331-336.
- Lechevalier, M. P., De Bievre, C., and Lechevalier, H. A. 1977. Chemotaxonomy of aerobic actinomycetes : phospholipid composition. <u>Biochem. Syst. Ecol</u>. 5: 249-260.
- Lechevalier, M. P., Prauser, H., Labeda, D. P., and Ruan, J. S. 1986. Two new genera of nocardioform actinomycetes : *Amycolata* gen. nov. and *Amycolatopsis* gen. nov. <u>Int. J.</u> <u>Svst. Bacteriol.</u> 36: 29-37.
- Lee, S. D. 2006. *Amycolatopsis jejuensis* sp. nov. and *Amycolatopsis halotolerans* sp. nov., novel actinomycetes isolated from a natural cave. Int J. Syst. Evol. Microbiol. 56: 549-553.
- Lee, J. Y., Moon, S. S., and Hwang, B. K. 2005. Isolation and antifungal activity of 4-phenyl-3buteonoic acid from *Streptomyces koyangensis* strain VK-A60. J. Agric Food. Chem. 53(20): 7696-7700.
- Lee, S. D., and Hah, Y. C. 2001. Amycolatopsis albidoflavus sp. nov. Int J. Svst. Evol. Microbiol. 51: 645-650.

- Liu, Z., Rodriguez, C., Wang, L., Cui, Q., Huang, Y., Quintana, E. T. and Goodfellow, M. 2005.
 Kitasatospora viridis sp. nov., a novel actinomycete from soil. <u>Int J. Syst. Evol. Microbiol.</u> 55: 707-711.
- Lorain, V. 1991. <u>Antibiotics in Laboratory Medicine</u>. pp. 1-51. Baltimore: The Williams and Wilkins.
- Majumdar, S., Prabhangaran, S. R., Shivaji, S., and Lal, R. 2006. Reclassification of Amycolatopsis orientalis DSM 43387 as Amycolatopsis benzoatilytica sp. nov. Int J. Syst. Evol. Microbiol. 56: 199-204.
- Matsumoto, N., Sasaki, Y., Kinoshita, N., Fujita, T., and Sekino, K. 1999. Lactonamycin, a new antimicrobial antibiotic produced by *Streptomyces rishiriensis* MJ773-88K4. <u>J. Antibiot</u>. 52(3): 269-275.
- Mayilraj, K., Jakobsen, I. B., and Nei, M. 2006. Kitasatospora sampliensis. sp nov. Int J. Syst. Evol. Microbiol. 50: 245-250.
- Meja, A., Barria, J., and Gonzalez, G. 1997. Overproduction of rifamycin B by Amycolatopsis mediterranei and its relationship with the toxic effect of barbital on growth. J. Antibiot. 51(1): 58-63.
- Mertz, F. P., Yao, R. C. 1993. Amycolatopsis alba sp. nov. isolated from soil. Int J. Svst. Bacteriol. 43: 715-720.
- Naggar, E., Assar, S. A., and Gawad, S. M. 2006. Merosparamycin. Production by newly isolated *Streptomyces* sp. Strain MAR01: taxonomy, fermentation, purification and structural elucidation. J. Microbiol. 44(4): 432-438.
- Okami, K. and Hotta, Y. 1988. Search and discovery of new antibiotics. In M. Goodfellow,
 S.T. Williams, and M. Mordaski (eds), <u>Actinomycetes in Biotechnology</u>. pp. 33-67.
 London: Academic Press.
- Oki, T. 1994. Recent progress of antibiotics research. S4-8. In Komagata, *et.al*.(eds.)
 <u>Application control of microorganisms in Asia</u>. pp 258-294 Science and Technology
 Agency, The Institute of Physical and Chemical Research, Japan International Science and Technology Exchange Center.
- Omura, S., Takahashi, Y., Iwai, Y and Tanaka, H.1982. *Kitasatospora*, a new genus of the order Actinomycetales. J. Antibiot. 35: 1013-1019.
- Pittenger, R. C., and Brigham, R. B. 1956. *Streptomyces orientalis*. sp., the source of vancomycin. <u>Antibiot. Chemother</u>. 6: 642-647.

- Puder, C., Zeeck, A., and Beil, W. 2000. New biologically active rubiginones from *Streptomyces* sp. J. Antibiot. 53(4): 329-336.
- Ryu, G., Choi, W. C., Hwang, S., Yeo, W. H., Lee, C. S., and Kim, S. K. 1999. Tetrin C, a new glycosylated polyene macrolide antibiotic produced by *Streptomyces* sp. GK 9244. J. Nat. <u>Prod.</u> 62(6): 917-919.
- Saintpierre, D., Amir, H., Pineau, R., Tan, G. Y., and Goodfellow, M 2005. Amycolatopsis plumensis, referring to the plum region of the main island of new Caledonian brown hypermagnesian ultramafic soil. Int. J. Syst. Evol. Microbiol. 55: 2057 – 2061.
- Saitou, N. and Nei, M. 1987. The neighbor-joining Method: a new method for reconstructing phylogenetic trees. <u>Mol. Biol. Evol</u>. 4: 406-429.
- Sasaki, T., Iragashi, Y., Saito, N., and Furumai, T. 2001. Cedamycins A and B, new antimicrobial antibiotics from *Streptomyces* sp. TP-A0456. J. Antibiot. 54(7): 567-572.
- Sasaki, T., Iragashi, Y., Saito, N., and Furumai, T. 2001. TPU-0031-A and B, new antibiotics of the novobiocin group produced by *Streptomyces* sp. TP-A0556. <u>J. Antibiot</u>. 54(5): 441-447.
- Sasaki, T., Iragashi, Y., Saito, N., and Furumai, T. 2002. Watsemycins A and B, new antibiotics produced by *Streptomyces* sp. TP-A0597. J. Antibiot. 55(3): 249-255.
- Sasaki, T., Otani, T., Matsumoto, H., Unemi, N. 1998. MJ347-81F4 A and B, novel antibiotics from *Amycolatopsis* sp.: taxonomic characteristics, fermentation, and antimicrobial activity. J. Antibiot. 51(8): 715-721.
- Schimana, J., Gebnardt, K., Holtzel, A., Schmid, D. G., Sussmuth, R., Muller, J., Pukall, R., and Fiedler, H. P. 2002. Arylomycins A and B, new biaryl-bridged lipopeptide antibiotics produced by *Streptomyces* sp. TU6075. I. taxonomy, fermentation, isolation and biological activities. J. Antibiot. 55(6): 565-570.
- Shimanaka, K., Kinoshita, N., Iinuma, H., Hamada, M., and Iakeuchi, T. 1994. Novel antibiotics, amytiamicins.I, taxonomy, fermentation, isolation, physicochemical properties, and antimicrobial activity. J. Antibiot. 47(6): 668-674.
- Shirato, S. and H. Motoyama, H. 1966 Fermentation studies with *Streptomyces griseus* II. Synthetic media for the production of streptomycin. <u>Appl. Microbiol</u>. 14: 706-710.
- Shirling, E. B. and Gottlieb, D. 1966. Methods for characterizaton of *Streptomyces* species. Int. J. Syst. Bacteriol. 16: 313-340.

- Sitachitta, N., Gadepalli, M., and Davidson, B. S. 1996. New pyrone containing metabolites from a marine-derived actinomycetes. <u>Tetra. Lett</u>. 52(24): 8073-8080.
- Spasova, D., Vesselinova, N., and Gesheva, R. 1997. Comparative investigation of a streptovaricin- producing strain of *Streptomyces spectabilis* and its selectant. <u>Folia.</u> <u>Microbia</u>. 42(1): 35-38.
- Speitling, M., Nattewan, P., Yazawa, K., Mikami, Y., Ritzau, M., Laatsch, H., and Grafe, U. 1998. Demethyl mutactimycins, new anthracycline antibiotics from *Nocardia* and *Streptomyces* strains. J. Antibiot. 51(8): 693-698.
- Staley, A. L., and Rinehart, K. L. 1994. Spectomycins, new antibacterial compounds produced by Streptomyces spectabilis: isolation, structures, and biosynthesis. J. Antibiot. 47(12): 1425-1433.
- Sujatha, P., Ruju, K. V., and Ramana, T. 2005. Studies on a new marine Streptomycete. BT-408 producing polyketide antibiotic SBR-22 effective against methicillin resistant Staphylococcus aureus. <u>Res in Microbiol</u>. 160(2): 119-126.
- Tan, G. A., Robison, S., Lacey, E., and Goodfellow, M. 2006. Amycolatopsis australiensis, pertaining to Australia, the source of the soil from which the first strains were isolated. Int. J. Syst. Evol. Microbiol. 56: 2297-2301.
- Tang, Y.O., Sattler, I., Thiericke, R., Grabley, S., and Eeng, X. Z. 2000. Feigrisolides A, B, C and D, new lactones with antibacterial activities from *Streptomyces griseus*. <u>J. Antibiot.</u> 53(9): 934-943.
- Tajima, K., Takahashi, Y., Seino, A., Iwai, Y., and Omura, S. 2001 Description of two novel species of the genus *Kitasatospora* omura et al. 1982, *Kitasatospora cineracea* sp.nov. and *Kitasatospora niigatensis* sp.nov. Int. J. Syst. Evol. Microbiol. 51: 1765-1771.
- Takahashi, Y., Iwai, Y., and Omura, S. 1984. Two new species of the genus Kitasatosporia, Kitasatosporia phosalacinea sp. nov. and Kitasatosporia griseola sp. nov. J. Gen. Appl. Microbial. 30:377-387.
- Takahashi, Y. and Omura, S. 2003. Isolation of new actinomycete strains for the screening of new bioactive compounds. <u>J. Gen. Appl. Microbiol</u>. 49: 141-154.
- Tamaoka, J. 1994. Determination of DNA base composition. In M. Goodfellow, and A.G. O'Donnell (eds.). <u>Chemical methods in prokaryotic systematics</u>. pp. 463-470. Chichester. John Wiley and Sons.

- Thompsons, J. D., Gibson, T. J., Plewniak, F., Jeanmougin, F., and Higgins, D. G. 1997. The CLUSTAL_X windows interface: flexible strategies for multiple sequence alignment aided by quality analysis tools. <u>Nucleic Acids Res</u>. 25: 4876-4882.
- Tortora, G. I., Funke, B. R. and Case, C. L. 1995. <u>Microbiology an introduction</u>. (5thed.). pp. 290-291; 491-512. Bridge Parkway: The Benjamin Cumming Publishing Company.
- Tseng, M., Yang, S. F., Li, W. J., and Jiang, C. L. 2006. Amycolatopsis taiwanensis sp. nov., from soil. Int. J. Syst. Evol. Microbiol. 56: 1811 – 1815.
- Tsuchida, T., Inuma, H., Kinoshita, N., Ikeda, T., Sawa, T., Hamada, M., and Takeuchi, T. 1995. Azicemicines A and B, a new antimicrobial agent produced by *Amycolatopsis*.I, taxonomy, fermentation, isolation, characterization and biological activities. <u>J.Anitbiot</u>. 62(11): 1562-1564.
- Ubukata, H., Osada, T., Kudo, K., and Isno, K. 1993. Respinomycins A1, A2, B, C and D, a novel group of anthracycline antibiotics: taxonomy, fermentation, isolation and biological activities. J.Anitbiot. 46: 936-941.
- Ubukata, M., Shiraishi, N., Kobinata, K., Kudo, T., Yamaguchi, I., Osada, H., Shen, Y. G. and Isono, K. 1995. RS-22A, B and C : new macrolide antibiotics from *Streptomyces violaceusniger*. I. taxonomy, fermentation, isolation and biological activities. J. Anitbiot. 48(4): 289-292.
- Vijaya, E. K., Kenia, J., Mukhopadhyay, T., and Nadkarni, S. R. 1999. Methylsulfomycin I, a new cyclic peptide antibiotic from a *Streptomyces* sp. HILY-9420704. <u>J. Nat. Prod.</u> 63(3): 352-356.
- Wang, E. L., Hamada, M., Okami. Y., and Umezawa, H. 1966. A new antibiotic spinamycin. J.Anitbiot. 19(4): 216-221.
- Warwick, S., Bowen, T., Mcveigh, H., and Embley, T. M. 1994. A phylogenetic analysis of the family Pseudonocardiaceae and the genera *Actinokineospora* and *Saccharothix* with 16S rRNA sequences and a proposal to combine the genera *Amycolata* and *Pseudonocardia* in an emended genus *Pseudonocardia*. Int. J. Syst. Bacteriol. 44: 293-299.
- Wayne, L. G., Brenner, D. J., Colwell, R. R., Grimont, A. D., Kandler, O., Hrichevsky, M. I., Moore, L. H., Murray, R. E., Stackebrandt, E., Starr, M. P., and Truper, H. G. 1987. Report of the Ad Hoc Committee on Reconciliation of Approaches to bacteria Systematics. <u>Int. J.</u> <u>Svst. Bacteriol.</u> 37: 463-464.

Walksman, S. A. 1953. Letter to editor. American Scientist. 41: 8-12.

- Williams, S. T., and Cross, T. 1971. Actinomycetes: slide and coverslip methods. In Method in microbiology (4th ed.), pp. 320. London: Academic Press.
- Wellington, E. H., Stackebrandt, E., Sanders, D., Wolstrup, J., and Jorgensen, N. O. 1992. Taxonomic status of *Kitsatosporia*, and proposed unification with *Streptomyces*. On the basis of phenotypic and 16S rRNA analysis and emendation of *Streptomyces* Waksman and Henrici 1943, 339AL. Int. J. Syst. Bacteriol. 42: 156-160.
- Wink, J., Gandhi, J., Kroppenstedt, R. M., Seibert, G., Straubler, B., Schumann, P., and Stackebranolt, E. 2004. *Amycolatopsis decaplanina* sp. nov., a novel member of the genus with unusual morphology. Int. J. Syst. Bacteriol. 54: 235-239.
- Wink, J. M., Kroppenstedt, R. M., Ganguli, B., Nadkarni, S. R., Schumann, P., Seibert, G., and Stackebrandt, E. 2003. Three new antibiotic producing species of the genus *Amycolatopsis*, *Amycolatopsis balhimycina* sp. nov., *A. tolypomycina* sp. nov., *A. vancoresmycina* sp. nov., and description of *Amycolatopsis keratiniphila* subsp. *Keratiniphila* subsp. Nov. and *A. keratiniphila* subsp. *Nogabecina* subsp. nov. <u>Syst. Appl. Microbiol</u>. 26: 38-46.
- Yamada, K., and Komagata, K. 1970. Taxonomic studies on coryneform bacteria. III. DNA base composition of coryneform bacteria. J. Gen. Appl. Microbiol. 16: 215-224.
- Yang, S. W., Chan, T. M., Terracciano, J., Patel, R., Loebenberg, D., Chen, G., Patel, M., Gullo, V., Pramonik, B., and Chu, M. 2005 New antibiotic Sch 725424 and its dehydration product Sch 725428 from *Kitasatospora* sp. J. Antibiot. 58(3): 192-195.
- Yao, Y., Zhang, W., Jiao, R., Zhao, G., and Jiang, W. 2002. Efficient isolation of total RNA from antibiotic-producing bacterium *Amycolatopsis mediterranei*. J. Microbiol. 51(2): 191-195.
- Yeo, W. H., Kim, S. K., Kim S. S., Yu, S. H., and Park, E. K. 1994. Taxonomy and fermentation of *Kitasatosporia kimorexae* producing new thiopeptide antibiotics, kimorexins. J. <u>microbial. Biotechnol</u>. 4: 354-359.
- Yu, Q., and Fan, C. 1994. Taxonomy of 1043 strain producing actinospectin. <u>Wei. Sheng. Wu.</u>
 <u>Xue. Bao.</u> 43(2): 160-163.
- Zhang, H., Tomoda, H., Tabata, N., Oohori, M., Shinose, M., Takahashi, Y., and Omura, S. 1999. Zelkovamycin, a new cyclic peptide antibiotic from *Streptomyces* sp. K96-0670.I, production, isolation and biological properties. <u>J. Antibiot</u>. 52(1): 29-33.
- Zhang, Z., Wang, Y. Ruan, J. 1997. A proposal to revive the genus *Kitasatospora* (Omura, Takahashi, Iwai, and Tanaka 1982). Int. J. Syst. Bacteriol. 47: 1048-1054.

APPENDICES

APPENDIX A

Culture media

All media were dispensed and sterilized in autoclave for 15 minutes at 15 pound pressure (121 $^{\circ}$ C) except for the carbon utilization test medium which was sterilized at 10 pound for 10 minutes

1.	Boullion	Gelatin	broth

Peptone	1.0 g
Meat extract	0.5 g
NaCl	0.5 g
Gelatin	15.0 g
Distilled water	100 ml
pH 7.0-7.2	

2. Basal Inorganic Nitrogen medium

$(NH_4)_2HPO_4$	0.1 g
KC1	0.02 g
MgSO ₄ .7H ₂ 0	0.02 g
Agar	7.5 g
рН 7.0	

3. Carbon utilization medium (ISP-9)

Basal mineral salt agar	
Carbohydrate	1.0 g
$(NH_4)_2SO_4$	0.264 g
K ₂ HPO ₄ (anhydrous)	0.238 g
K ₂ HPO ₄ .3H ₂ O	0.565 g
MgSO ₄ .7H ₂ 0	0.1 g
Pridham and Gottlieb trace salt (B)	0.1 ml
Agar	1.5 g
Distilled water	100 ml
рН 6.8-7.0	

Pridham and Gottlieb trace salt (B)

CuSO ₄ .5H ₂ O	0.64 g
FeSO ₄ .7H ₂ O	0.11 g
MnCl ₂ .4H ₂ O	0.79 g
ZnSO ₄ .7H ₂ O	0.15 g
Distilled water	100 ml

4. Esculin broth

Esculin	0.1 g
Ferric citrate	0.05 g
Peptone	1.5 g
NaCl	0.5 g
Distilled water	100 ml

5. Glyceral-Asparagine agar

L-asparagine (anhydrous basis)	0.1 g
Glycerol	1.0 g
K_2 HPO ₄ (anthdrous basis)	0.1 g
Pridham and Gottlieb trace salt (A)	0.1 ml
Agar	1.5 g
Distilled water	100 ml

pH 7.0-7.4

Pridham and Gottlieb trace salt (A)	
FeSO ₄ .7H ₂ O	0.1 g
MnCl ₂ .4H ₂ O	0.1 g
ZnSO ₄ .7H ₂ O	0.1 g
Distilled water	100 ml

6. Inorganic salt- starch agar (ISP-4)

Soluble starch (Difco)	1 g
K_2 HPO ₄ (anhydrous)	0.1 g

MgSO ₄ .7H ₂ 0	0.1 g
NaCl	0.1 g
(NH ₄) ₂ SO ₄	0.2 g
CaCO ₃	0.2 g
Pridham and Gottlieb trace salt (A)	0.1 ml
Agar	1.5 g
Distilled water	100 ml
рН 7.0-7.4	

7. Muller-Hinton medium (MHM)

Muller-Hinton (Difco)	3.4 g
Distilled water	100 ml
рН 7.3	

8. Nutrient agar

Nutrient agar (Difco)	2.3 g
Distilled water	100 ml

9. Nitrate broth

Peptone	1.0 g
KNO3	0.1 g
NaCl	0.5 g
Distilled water	100 ml
pH 7.0	

10. Oatmeal agar (ISP-3)

Oatmeal	20 g
Agar	18 g
Distilled water	100 ml
pH 7.0-7.4	

11. Potato starch agar

Potato Starch	1.0 g

Glycerol	1.0 g
K ₂ HPO ₄	0.2 g

- $(NH_4)_2SO_4$ 0.2 g
- MgSO₄·7H₂O 0.1 g
- NaCl 0.1 g
- CaCO₃ 0.2 g Agar 1.2 g
- Distilled water 100 ml

pH 7.0-7.3

12. Production medium

Yeast extract	0.4 g
Glucose	0.4 g
Malt extract	1.0 g
CaCO ₃	0.1 g
рН 7.3	

13. Sabouraud dextrose agar (SDA)

Sabouraud dextrose agar	3.0 g
Distilled water	100 ml

14. Skim milk

Skim milk (Difco)	10.0 g
Distilled water	100 ml

15. Starch-casein nitrate agar

Starch	1.0 g
Sodium caseinate	0.03 g
KNO3	0.2 g
Agar	1.5 g

Distilled water	100 ml
-----------------	--------

pH 7.0-7.4

16. Seed medium

Yeast extract	0.4 g
Glucose	0.4 g
Malt extract	1.0 g
Distilled water	100 ml
рН 7.3	

17. Tyrosine agar

Glycerol	1.5 g
L-Tyrosine	0.05 g
L-Asparagine	0.1 g
K_2 HPO ₄ (anhydrous)	0.05 g
MgSO ₄ .7H ₂ O	0.05 g
NaCl	0.05 g
FeSO ₄ .7H ₂ O	0.01 g
Pridham and Gottlieb trace salt (A)	0.1 ml
Agar	1.5 g
Distilled water	100 ml
pH 7.2-7.4	

18. Yeast Extract- Malt Extract agar (ISP-2)

Yeast extract	0.4 g
Malt extract	1.0 g
Glucose	0.4 g
Agar	1.5 g
Distilled water	100 ml
рН 7.3	

128

APPENDIX B

130

Reagent and Buffers

1. 6N HCl

2.

Conc. HCl	60 ml
Distilled water	60 ml
Add. Conc. HCl into the distilled water.	
Ninhydrin solution	
Ninhydrin	0.3 g
l-Butanol	100 ml
Glacial acetic acid	3 ml

3. Nitrate reduction test reagent

Sulphanilic acid solution

Sulphanilic acid	0.8 g
5 N Acetic acid	100 ml

Dissolve by gentle heating in a fume hood.

N,N- dimethyl-1-naphthylamine solution

N,N- dimethyl-1-naphthylamine solution	0.5 g
5 N Acetic acid	100 ml

Dissolve by gentle heating in a fume hood.

Two drops of sulphanilic acid solution and three drops of N,N- dimethyl-1naphthylamine into peptone nitrate broth inoculing with the test microorganisms.

4. Phenol : Chloroform (1:1 v/v)

Crystalline phenol was liquidified in water bath at 65 $^{\circ}$ C and mixed with chloroform in the ratio of 1:1 (v/v). The solution was stored in a light tight bottle.

5. 10% Sodium dodecyl sulphate (SDS)

The stock solution of 10% SDS was prepared by dissolved 10 g of sodium dodecyl sulphate in 100 ml steried distilled water. Sterilization is not required for the preparation of this stock solution.

6. 20XSSC

3M NaCl

0.1 M Tri-sodiumcitrate

The 20XSSC was adjusted the pH to 7.0 with 1 N NaOH. The solution was sterilized by autoclaving for 15 minutes at 15 lb/in^2 .

7. RNase A solution

Rnase A	20 mg
0.15 M NaCl	10 ml

Dissolved 20 mg Rnase A in 10 ml 0.15 m NaCl and heat at 95 $^{\circ}$ C for 5-10 minutes. Keep Rnase A solution in -20 $^{\circ}$ C.

8. RNase T₁ solution

Rnase T ₁	80 µl
0.1 M Tris-HCl (pH7.5)	10 ml

9. Nuclease P₁ solution

Nuclease P ₁	0.1 mg

40 mM CH₃COONa+12 mM ZnSO₄ (pH 5.3)

10. Alkaline phosphatase solution

Alkaline phosphatase	2.4 units
0.1 M Tris-HCl (pH8.1)	l ml

11. 0.1 M Tris-HCl buffer, pH 9

Tris	1.21 mg
Distilled water	100 ml

Adjust the pH to 9 with HCl

12. Saline-EDTA

0.1 M NaCl 50 mM EDTA (pH 8)

13. Ethidium bromide solution (10 mg/ml)

The ethidium bromide solution was prepared by dissolved 1 g of ethidium bromide in 100 ml of distilled water. The solution was stored in light-tight container at room temperature.

14. Agarose gel

Agarose	1.6 g
1XTBE buffer	200 ml

15. Reagent and buffer for DNA-DNA hybridization

15.1 Prehybridization solution	
100x Denhardt solution	5 ml
10 mg/ml Salmon sperm DNA	1 ml
20x SSC	10 ml
Formamide	50 ml
Distilled water	34 ml
15.2 Hybridization solution	
Prehybridization solution	100 ml
Dextran-sulfate	5 g
15.3 Solution I	
Bovine serum albumin	0.25 g
Titron X-100	50 µl
PBS	50 ml
15.4 Solution II	
Streptavidin-POD	1 µl
Solution I	4 ml
15.5 Solution III	
3,3',5,5'-Tetramethylbenzidine (TMB)	100 µl
(10 mg/ml in DMSO)	
0.3% H ₂ O ₂	100 µl
0.4 M Citric acid-0.2 M Na ₂ HPO ₄ buffer	100 µl
pH 6.2 in 10% DMSO	

$15.6\ 2M\ H_2SO_4$

H_2SO_4	22 ml

Distilled water 178 ml

The solution was sterilized by autoclaving

APPENDIX C

Primers and Nucleotide sequences of the PCR amplified 16S rDNA

1. List of primer for 16S rDNA PCR amplification and Sequencing

8-27f 5'-AGAGTTTGATC(A/C)TGGCTCAG-3'

530f 5'-GTGCCAGC(A/C)GCCGCGG-3'

1114f 5'-GCAACGAGCGCAACCC-3'

1392r 5'-ACGGGCGGTGTGT(A/G)C-3'

2. Nucleotide sequences of the PCR amplified 16S rDNA

GGCGTGCTTACACATGCAAGTCGAACGATGAACCGGTTTCGGCCGGGGATTAGTGGCGAACGGGTGAGTAACA CGTGGGCAATCTGCCCTGCACTCTGGGACAAGCCCTGGAAACGGGGTCTAATACCGGATATGACTGCCGACCG CATGGTCTGGTGGTGGAAAGCTCCGGCGGTGCAGGATGAGCCCGCGGCCTATCAGCTTGTTGGTGGGGTGATG GCCTACCAAGGCGACGACGGGTAGCCGGCCTGAGAGGGCGACCGGCCACACTGGGACTGAGACACGGCCCAGA CTCCTACGGGAGGCAGCAGTGGGGGAATATTGCACAATGGGCGCAAGCCTGATGCAGCGACGCCGCGTGAGGGA TGACGGCCTTCGGGTTGTAAACCTCTTTCAGCAGGGAAGAAGCGCAAGTGACGGTACCTGCAGAAGAAGCGCC GGCTAACTACGTGCCAGCAGCCGCGGTAATACGTAGGGCGCAAGCGTTGTCCGGAATTATTGGGCGTAAAGAG CTCGTAGGCGGCTTGTCGCGTCGGATGTGAAAGCCCGGGGCTTAACTCCGGGTCTGCATTCGATACGGGCAGG CTAGAGTTCGGTAGGGGAGATCGGAATTCCTGGTGTAGCGGTGAAATGCGCAGATATCAGGAGGAACACCGGT GGCGAAGGCGGATCTCTGGGCCGATACTGACGCTGAGGAGCGAAAGCGTGGGGAGCGAACAGGATTAGATACC CTGGTAGTCCACGCCGTAAACGTTGGGAACTAGGTGTGGGCGACATTCCACGTTGTCCGTGCCGCAGCTAACG CATTAAGTTCCCCGCCTGGGGAGTACGGCCGCAAGGCTAAAACTCAAAGGAATTGACGGGGGCCCGCACAAGC GAGACAGGGCCCCCTTGTGGTCGGTGTACAGGTGGTGCATGGCTGTCGTCAGCTCGTCGTGAGATGTTGGG TTAAGTCCCGCAACGAGCGCAACCCTTGTCCTGTGTTGCCAGCATGCCCTTTGGGGTGATGGGGAACTCACAG GAGCACTGCCGGGGATCAACTCGGAGGAAGGTGGGGGCACGACGTCAAGTCATGCCCCTTATGTCTTGGGC TGCACACGTGCTACAATGGCCGGTACAATGAGCTGCGAAGCCGTGAGGTGGAGCGAATCTCAAAAAGCCGGTC TCAGTTCGGATTGGGGTCTGCAACTCGACCCCATGAAGTCGGAGTCGCTAGTAATCGCAGATCAGCATTGCTG CGGTGAATACGTTCCCGGGCCTTGTACACACCGCCCGTCACGTCACGAAAGTCGGTAACACCCGAAGCCGGTG GCCCAACCCTTGTGGGGGGGGGGGCCGTCAAGGTGGGACTGGCGATTGGGACAAGTCTAACAAGGTACCGTAAACT

The PCR amplified 16S rDNA nucleotide sequences of S1-2

CGTGCTTACCATGCAAGTCGAACGATGAAGCCCTTCGGGGTGGATTAGTGGCGAACGGGTGAGTAACACGTGG GCAATCTGCCCTGCACTCTGGGACAAGCCCTGGAAACGGGGTCTAATACCGGATATGACACGGGATCGCATGA TCTTGTGTGGAAAGCTCCGGCGGTGCAGGATGAGCCCGCGGCCTATCAGCTAGTTGGTGAGGTAATGGCTCAC CAAGGCGACGACGGGTAGCCGGCCTGAGAGGGGCGACCGGCCACACTGGGACTGAGACACGGCCCAGACTCCTA CGGGAGGCAGCAGTGGGGAATATTGCACAATGGGCGAAAGCCTGATGCAGCGACGCCGCGTGAGGGATGACGG CCTTCGGGTTGTAAACCTCTTTCAGCAGGGAAGAAGCGAAAGTGACGGTACCTGCAGAAGAAGCGCCGGCTAA CTACGTGCCAGCAGCCGCGGTAATACGTAGGGCGCGAGCGTTGTCCGGAATTATTGGGCGTAAAGAGCTCGTA GGCGGCTTGTCACGTCGGTTGTGAAAGCCCGGGGCTTAACCCCGGGTCTGCAGTCGATACGGGCAGGCTAGAG TTCGGTAGGGGAGATCCGGAATTCCTGGTGTAGCGGTGAAATGCGCAGATATCAGGAGGAACACCGGTGGCGAA GGCGGATCTCTGGGCCGATACTGACGCTGAGGAGCGAAAGCGTGGGGAGCGAACAGGATTAGATACCCTGGTA GTCCACGCCGTAAACGGTGGGCCACTAGGTGTGGGCGACATTCCACGTCGTCCGTGCCGCAGCTAACGCATTAA GTGCCCCGCCTGGGGAGTACGGCCGCAAGGCTAAAACTCAAAGGAATTGACGGGGGCCCGCACAAGCGGCGGA GCATGTGGCTTAATTCGACGCAACGCGAAGAACCTTACCAAGGCTTGACATACACCGGAAACGGCCAGAGATG GTCGCCCCCTTGTGGTCGGTGTACAGGTGGTGCATGGCTGTCGTCAGCTCGTGTCGTGAGATGTTGGGTTAAG TCCCGCAACGAGCGCAACCCTTGTCCCGTGTTGCCAGCAGGCCCTTGTGGTGCTGGGGACTCACGGGCAGACC GCCGGGGTTCAACTCGGAGGAAGGTGGGGCACGACGTCAAGTCATCATGCCCCTTATGTCTTGGGCTGCACAC GTGCTACAATGGCCGGTACAATGAGCTGCGATACCGCGAGGTGGAGCGAATCTCAAAAAGCCGGTCTCAGTTC GGATTGGGGTCTGCAACTCGACCCCATGAAGTCGGAGTCGCTAGTAATCGCAGATCAGCATTGCTGCGGTGAA CCCTTGTGGGAGGGAGCTGTCGAAGGTGGGACTGGCGATTGGGACGAAGTCGTAACAAGGTAACCGTAAT

The PCR amplified 16S rDNA nucleotide sequences of S3-1

I 27168578

GCGTGCTTACACATGCAAGTCGAACGATGAAGCCCTTCGGGGTGGATTAGTGGCGAACGGGTGAGTAACACGTGGG CAATCTGCCCTGCACTCTGGGACAAGCCCTGGAAACGGGGTCTAATACCGGATATGACACGGGATCGCATGATCTT CGTGTGGAAAGCTCCGGCGGTGCAGGATGAGCCCGCGGCCTATCAGCTAGTTGGTGAGGTAACGGCTCACCAAGGC GACGACGGGTAGCCGGCCTGAGAGGGCGACCGGCCACACTGGGACTGAGACACGGCCCAGACTCCTACGGGAGGCA GCAGTGGGGAATATTGCACAATGGGCGAAAGCCTGATGCAGCGACGCCGCGTGAGGGATGACGGCCTTCGGGTTGT AAACCTCTTTCAGCAGGGAAGAAGCGAAAGTGACGGTACCTGCAGAAGAAGCGCCGGCTAACTACGTGCCAGCAGC CGCGGTAATACGTAGGGCGCGAGCGTTGTCCGGAATTATTGGGCGTAAAGAGCTCGTAGGCGGCTTGTCACGTCGG TTGTGAAAGCCCGGGGGCTTAACCCCGGGTCTGCAGTCCATACGGGCAGGCTAGAGTTCGGTAGGGGAGATCGGAA TTCCTGGTGTAGCGGTGAAATGCGCAGATATCAGGAGGAACACCGGTGGCGAAGGCGGATCTCTGGGCCGATACTG ACGCTGAGGAGCGAAAGCGTGGGGAGCGAACAGGATTAGAACCCTGGTAGTCCACGCCGTAAACGGTGGGCACTAG GTGTGGGCGACATTCCACGTCCGTGCCGCAGCTAACGCATTAAGTGCCCCGCCTGGGAGTACGGCCGCAAGGC TAAAACTCAAAGGAATTGACGGGGGCCCGCACAAGCGGCGGGGGCATGTGGCTTAATTCGACGCAACGCGAAGAACC TTACCAAGGCTTGACATACACCGGAAACGGCCAGAGATGGTCGCCCCCTTGGTCGGTGTACAGGTGGTGCATGGCT GTCATCAGCTCGTGCTGAGATGTTGGGTTAAGTCCCGCAACGAGCGCAACCCTTGTCCCTGTTGCCACAAGGCCCC TGCCCCTTATGTCTTGGGCTGCACACGTGCTACAATGGCCGGTACAATGAGCTGCGATACCGCGAGGTGGAGCGAA TCTCAAAAAGCCGGTCTCAGTTCGGATTGGGGTCTGCAACTCGACCCCATGAAGTCGGAGTCGCTAGTAATCGCAG ATCAGCATTGCTGCGGTGAATACGTTCCCGGGCCTTGTACACCGCCCGTCACGTCACGAAAGTCGGTAACACCC GAAGCCGGTGGCCCAACCCCTTGTGGGAGGGAGCTGTCGAAGGTGGGACTGGCGATTGGGACGAAGTCGTAACAGG GTAACCGTA

The PCR amplified 16S rDNA nucleotide sequences of SB12-1

TGGCGGCGTGCTTACACATGCAAGTCGAACGGTGAAGCCCTTCGGGGTGGATCAGTGGCGAACGGGTGAGTAACAC GTGGGCAATCTGCCCTGCACTCTGGGACAAGCCCTGGAAACGGGGTCTAATACCGGATATGACCTTCCTCCGCATG GGGGTTGGTGTAAAGCTCCGGCGGTGCAGGATGAGCCCGCGGCCTATCAGCTTGTTGGTGGGGTAATGGCCTACCA AGGCGACGACGGGTAGCCGGCCTGAGAGGGGCGACCGGCCACACTGGGACTGAGACACGGCCCAGACTCCTACGGGA GGCAGCAGTGGGGAATATTGCACAATGGGCGAAAGCCTGATGCAGCGACGCCGCGTGAGGGATGACGGCCTTCGGG TTGTAAACCTCTTTCAGCAGGAAGAAGCGCAAGTGACGGTACCTGCAGAAGAAGCACCGGCTAACTACGTGCCAG CAGCCGCGGTAATACGTAGGGTGCGAGCGTTGTCCGGAATTATTGGGCGTAAAGAGCTCGTAGGCGGCCTGTCACG TCGGATGTGAAAGCCCGGGGCTTAACCCCGGGTCTGCATTCGATACGGGCAGGCTAGAGTGTGGTAGGGGAGATCG GAATTCCTGGTGTAGCGGTGAAATGCGCAGATATCAGGAGGAACACCGGTGGCGAAGGCGGATCTCTGGGCCATTA CTGACGCTGAGGAGCGAAAGCGTGGGGGAGCGAACAGAATTAGATACCCTGGTAGTCCACGCCGTAAACGTTGGGAA CTAGGTGTTGGCGACATTCCACGTCGTCGGTGCCGCAGCTAACGCATTAAGTTCCCCGGCCTGGGGAGTACGGCCGC AAGGCTAAAACTCAAAGGAATTGACGGGGGGCCCGCACAAGCAGCGGGGGCATGTGGCTTAATTCGACGCAACGCGAA GAACCTTACCAAGGCTTGACATATGCCGGAAACATCCAGAGATGGGTGCCCCCTTGTGGTCGGTATACAGGTGGTG CATGGTTGTCGTCAGCTCGTGTCGTGAGATGTTGGGTTAAGTCCCGCAACGAGCGCAACCCTTGTTCTGTGTTGCC AGCATGCCTTTCGGGGTGATGGGGACTCACAGGAGACTGCCGGGGTCAACTCGGAGGAAGGTGGGGGACGACGTCAA ATCATCATGCCCCTTATGTCTTGGGCTGCACACGTGCTACAATGGTCGGTACAAAGGGCTGCGATGCCGCGAGGCG GACGAATCCCAAAAAAGCCGGCCTCAGTTCGGATTGGGGTCTGCAACTCGACCCCATGAAGTTGGAGTTGCTAGTA ATCGCAATCACATGCTGCGGTGAATACGTTCCCGGGCCTTGTACACACCGCCCGTCACGTC

The PCR amplified 16S rDNA nucleotide sequences of S33-3

GCGGCGTGCTTAACACATGCAAGTCGAACGGTGAAGCCCTTCGGGGTGGATCAGTGGCGAACGGGTGAGTAACACG ATGGGCAATCTGCCCTGCACTCTGGGACAAGCCCTGGAAACGGGGTCTAATACCGGATATGACCTTCCTCCGCATG GGGGTTGGTGGAAAGCTCCGGCGGTGCAGGATGAGCCCGCGGCCTATCAGCTTGTTGGTGGGGTAATGGCCTACCA AGGCGACACGGGTAGCCGGCCTGAGAGGGCGACCGGCCACACTGGGACTGAGACACGGCCCAGACTCCTACGGAGG CAGAGTGGGGAATATTGCACAATGGGCGCAAGCCTGATGCACGACGCCGCGTGAGGGATGACGCCTTCGGTTGTAA ACCTCTTTCAGCAGGGAAGAAGCGCAAGTGACGGTACCTGCAAAGAACACCGGCTAACTACGTGCCATCAGCCGCG TAATACTAGGGTGCAGCGTTGTCCGGAATTATTGGACGTAAAGAGCTCGTAGGCGGCCTGTCGCGTCGGATGTGAA AGCCCCGGGGCTTAACCACGGGTCTGCATTCGATACGGGCAGGCTAGAGTGTGGTAGGGGAGATCGGAATTCCTGG TGTAGCGGTGAAATGCGCAGATATCAGGAGGAACACCGGTGGCGAAGGCGGATCTCTGGGCCATTACTGACGCTGA GGAGCGAAAGCGTAGGGAGCGAACAGGATTAGATACCCTGGTAGTCCACGCCGTAAACGTTGGGAACTAGGTGTTG GCGACATTCCACGTCGTCGGTGCCGCAGCTAACGCATTAAGTTCCCCGCCTGGGGAGTACGGCCGCAAGGCTAAAA CTCAAAGGAATTGACGGGGGCCCGCACAAGCAGCGGAGCATGTGGCTTAATTCGACGCAACGCGAAGAACCTTACC AAGGCTTGACATATGCCGGAAACACCTGGAGACAGGTGCCCCCTTGTGGTCGTATACAGTGGTGCATGGTTGTCTC AGCTCGTGTCTGAATGTTGGGTTAAGTCCCGCAACGAGCGCAACCCTTGTTCTGTGTTGACAGCAGAGTAATGTCG GGGACTCACAGGAGACTGCCGGGGGGTCAACTCGGAGGAGGGGGGGACAACGTCAAATCATCAAGCCCCTTATGTA CTTGGGCTGCACACGTGCTACAATGGTCGGTACAAAGGGCTGCGATGCCGTGAGGCGGAGCGAATCCCAAAAAGCC GGCCTCAGTTCGGATTGGGGTCTGCAACTCGACCCCATGAAGTTGGAGTTGCTAGTAATCGCAGATCAGCATGCTG CGGTGAATACGTTCCCGGGCCTTGTACACACCGCCCGTCACGTCACGAAAGTCGGTAACACCCCGAAGCCGGTGGCC TAACCCTCTGGGATGGAGCCGTCGAAGGTGGGACCAGCGATTGGGACGAAGTCGTAACAAGGTAAC

The PCR amplified 16S rDNA nucleotide sequences of S38-2

GCTTACACATGCAAGTCGAACGATGAACCGGCTTCGGCCGGGGGATTAGTGGCGAACGGGTGAGTAACACGTGGGCA ATCTGCCCTTCACTCTGGGACAAGCCCTGGAAACGGGGTCTAATACCGGATACGACGCGCGACCGCATGGTCTGTG CGTGGAAAGCTCCGGCGGTGAAGGATGAGCCCGCGGCCTATCAGCTTGTTGGTGGGGTAATGGCCTACCAAGGCGA CGACGGGTAGCCGGCCTGAGAGGGCGACCGGCCACACTGGGACTGAGACACGGCCCAGACTCCTACGGGAGGCAGC AGTGGGGAATATTGCACAATGGGCGAAAGCCTGATGCAGCGACGCCGCGTGAGGGATGACGGCCTTCGGGTTGTAA ACCTCTTTCAGCAGGGAAGAAGCGAGAGTGACGGTACCTGCAGAAGAAGCGCCGGCTAACTACGTGCCAGCAGCCG CGGTAATACGTAGGGCGCAAGCGTTGTCCGGAATTATTGGGCGTAAAGAGCTCGTAGGCGGCTTGTCACGTCGGAT GTGAAAGCCCGGGGCTTAACCCCGGGTCTGCATTCGATACGGGCAGGCTAGAGTTCGGTAGGGGGAGATCGGAATTC CTGGTGTAGCGGTGAAATGCGCAGATATCAGGAGGAACACCGGTGGCGAAGGCGGATCTCTGGGCCGATACTGACG CTGAGGAGCGAAAGCGTGGGGGGGGGGCGAACAGGATTAGATACCCTGGTAGTCCACGCCGTAAACGTTGGGAACTAGGT GTGGGCGACATTCCACGTCGTCCGTGCCGCAGCTAACGCATTAAGTTCCCCGCCTGGGGAGTACGGCCGCAAGGCT AAAACTCAAAGGAATTGACGGGGGCCCGCACAAGCAGCGGAGCATGTGGCTTAATTCGACGCAACGCGAAGAACCT TACCAAGGCTTGACATACACCGGAAAACCCTGGAGACAGGTCCCCCTTGTGGTCGGTGTACAGGTGGTGCATGGCT GTCGTCAGCTCGTGTCGTGAGATGTTGGGTTAAGTCCCGCAACGAGCGCAACCCTTGTTCTGTGTTGCCAGCATGC CCTTCGGGGTGATGGGGACTCACAGGAGACTGCCGGGGTCAACTCGGAGGAAGGTGGGGGACGACGTCAAGTCATCA TGCCCCTTATGTCTTGGGCTGCACACGTGCTACAATGGCCGGTACAATGAGCTGCGATACCGCGAGGTGGAGCGAA TCTCAAAAAGCCGGTCTCAGTTCGGATTGGGGTCTGCAACTCGACCCCATGAAGTCGGAGTTGCTAGTAATCGCAG ATCAGCATTGCTGCGGTGAATACGTTCCCGGGCCTTGTACACCGCCCGTCACGTCACGAAAGTCGGTAACACCC GAAGCCGGTGGCCCAACCCCTTGTGGGAGGGAATCGTCGAAGGTGGGACTGGCGATTGGGACGAAGTCGTAACAAG GTAACCGT

The PCR amplified 16S rDNA nucleotide sequences of S55-4

TGGCGGCGTGCTTAACACATGCAAGTCGAACGGATGAAGCCCTTCGGGGTGGATTAGTGGCGAACGGGTGAGTAAC ACGTGGGTCAATCTGCCCTGCACTCTGGGACAAGCCCTGGCAAACGGGGTCTAATACCGGATACGACCTGCCGAGG CATCTCGGCGGGTGGAAAGCTCCGGCGGTGCAGATGAGCCCGCGGCCTATCAGCTTGTTGGTGGGGTAATGGCCTA CCAAGGCGACGACGGGTAGCCGGCCTGAGAGGGCGACCGGCCACACTGGGACTGAGACACGGCCCAGACTCCTACG GGAGGCAGCAGTGGGGAATATTGCACAATGGGCGAAAGCCTGATGCACGACGCCGCGTGAGGGATGACGGCCTTCG GGTTGTAAACCTCTTTCAGCAGGGAAGAAGCGAAAGTGACGGTACCTGCAAAGAAGCGCCCGGCTAACTACGTGCAG CAGCCGCGGTAATACGTAGGGCGCAAGCGTTGTCCAGGAATTATTGGGCGTAAAGAGCTCGTAGGCGGCTTGCTCG CGTCAGATGTGAAAGCCCGGGGCTTAACCCCGGGATCTGCATTCGATACGGGCAGGCTAGAGTTCGGTAGGGGAGA TCGGAATTCCTGGGTAGCGGTGAAATGCGCAGATATCAGGAGGAACACCGGTGGCGAAGGCGGATCTCTGGGCCGA TACTGACGCTGAGGAGCGAAAGCGTGGGGAGCGAACAGGATAAGATACCTGTAGTCCACGCCGTAAAGTTGGAACT AGGTGTGGGCGACATTCCACGTCGTCCGTGCCGCAGCTAACGCATTAAGTTCCCCGGCCTGGGGAGTACGGCCGCAA GGCTAAAACTCAAAGGAATTGACGGGGGCCCGCACAAGCAGCGGAGCATGTGGCTTAATTCGACGCAAAGCGCAAAA CCTTACCAAGGCTTGACATACACCGGAAACACCTAGAGATAGGTGCCCCCTTGTGGTCGGTGTACAGGTGGTGCAT GGCTGTCGTCAGCTCGTGTCGTGAGATGTTGGGTTAAGTCCCCCAACGAGCGCAACCCTTGTCCTGTGTTGCCACAT ATGCCCCTTATGTCTTGGGCTGCACACGTGCTACAATGGCCGGTACAATGAGCTGCCATACCGCAAGGTTGGAGCG AATCTCAAAAAGCCGGTCTCAGTTCGGATTGGGGTCTGCAACTCGACCCCATGAAGTTGGAGTTGCTAGTAATCGC AGATCAGCATTGCTGCGGTGAATACGTTCCCGGGCCTTGTACACACCGCCCGTCACGACAGTCGGTAACAC ${\tt CCGAAGCCGGTGGCCCAACCCTTGTGGAGGGAGCCGTCGAAGGTGGGACTGGCGATTGGGACAAGTCGTAACAAGG}$ TAACCGTAA

The PCR amplified 16S rDNA nucleotide sequences of S49-1

 ${\tt GCAATCTGCCCTTCACTCTGGGACAAGCCCTGGAAACGGGGTCTAATACCGGATATGACACGGGGTCGCATGATCT}$ CCGTGTGGAAAGCTCCGGCGGTGAAGGATGAGCCCGCGGCCTATCAGCTTGTTGGTGAGGTATGGCTCACCAAGGC GACGACGGGTAGCCGGCCTGAGAGGGCGACCGGCCACACTGGGACTGAGACACGGCCCAGACTCCTACGGGAGGCA GCAGTGGGGAATATTGCACAATGGGCGAAAGCCTGATGCAGCGACGCCGCGTGAGGGATGACGGCCTTCGGGTTGT AAACCTCTTTCAGCAGGGAAGAAGCGAGAGTGACGGTACCTGCAGAAGAAGCGCCGGCTAACTACGTGCCAGCAGC ${\tt CGCGGTAATACGTAGGGCGCGAGCGTTGTCCGGAATTATTGGGCGTAAAGAGCTCGTAGGCGGCTTGTCGCGTCGG$ ATGTGAAAGCCCGGGGCTTAACCCCGGGTCTGCATTCGATACGGGCAGGCTAGAGTTCGGTAGGGGAGATCGGAAT TCCTGGTGTAGCGGTGAAATGCGCAGATATCAGGAGGAACACCGGTGGCGAAGGCGGATCTCTGGGCCGATACTGA CGCTGAGGAGCGAAAGCGTGGGGGGGGGACCAGGATTAGATACCCTGGTAGTCCACGCCGTAAACGTTGGGAACTAG GTGTGGGCGACATTCCACGTCGTCCGTGCCGCAGCTAACGCATTAAGTTCCCCGCCTGGGGAGTACGGCCGCAAGG CTAAAACTCAAAGGAATTGACGGGGGCCCGCACAAGCAGCGGAGCATGTGGCTTAATTCGACGCAACGCGAAGAAC CTTACCAAGGCTTGACATACACCGGGAAAACCGTGGAGACACGGTCCCCCTTGTGGTCGGTGTACAGGTGGTGCATG GCTGTCGTCAGCTCGTGTCGTGAGATGTTGGGTTAAGTCCCGCAACGAGCGCAACCCTTGTTCTGTGTTGCCAGCA TGCCTTTCGGGGTGATGGGGGACTCACAGGAGACTGTCCGGGGGTCAACTCGGAGGAAGGTGGGGACGACGTCAAGT CATCATGCCCCTTATGTCTTGGGCTGCACACGTGCTACAATGGCCGGTACAATGAGCTGCGATACCGCGAGGTGGA GCGAATCTCAAAAAGCCGGTCTCAGTTCGGATTAGGGTCTGCAACTCGACCCCATGAAGTCGGAGTTGCTAGTAAT CGCAGATCAGCATTGCTGCGGTGAATACGTTCCCGGGCCTTGTACACACCGCCCGTCACGTCACGAAAGTCGGTAA CACCCGAAGCCGGTGGCCCAACCCCTTGTGGAAGGGAATCGTCAAGGTGGGACTGGCGATTGGGACAAGTCTAACA AGGAACCGTAATAACCAA

The PCR amplified 16S rDNA nucleotide sequences of S71-1

GCGTGCTTACCATGCAAGTCGAACGATGAAGCCCTTCGGGGGTGGATTAGTGGCGAACGGGTGAGTAACACGTGGGC AATCTGCCCTTCACTCTGGGACAAGCCCTGGAAACGGGGTCTAATACCGGATACGACCCGCCGAGGCATCTCGGTG GGTGGAAAGCTCCGGCGGTGAAGGATGAGCCCGCGGCCTATCAGCTTGTTGGTGGGGGTAACGGCCCACCAAGGCGA CGACGGGTAGCCGGCCTGAGAGGGCGACCGGCCACACTGGGACTGAGACACGGCCCAGACTCCTACGGGAGGCAGC AGTGGGGAATATTGCACAATGGGCGAAAGCCTGATGCAGCGACGCCGCGTGAGGGATGACGGCCTTCGGGTTGTAA ACCTCTTTCAGCAGGGAAGAAGCGCCAAGTGACGGTACCTGCAGAAGAAGCGCCGGCTAACTACGTGCCAGCAGCCG CGGTAATACGTAGGGCGCAAGCGTTGTCCGGAATTATTGGGCGTAAAGAGCTCGTAGGCGGCTTGTCACGTTCGGG TGTGAAAGCCCGGGGCTTAACCCCGGGTCTGCATCCGATACGGGCAGGCTAGAGTGTGGTAGGGGGGGAGATCGGAATT CCTGGTGTAGCGGTGAAATGCGCAGATATCAGGAGGAACACCGGTGGCGAAGGCGGATCTCTGGGCCATTACTGAC GCTGAGGAGCGAAAGCGTGGGGAGCGAACAGGATTAGATACCCTGGTAGTCCACGCCGTAAACGTTGGGAACTAGG TGTTGGCGACATTCCACGTCGTCGGTGCCGCAGCTAACGCATTAAGTTCCCCGCCTGGGGAGTACGGCCGCAAGGC TAAAACTCAAAGGAATTGACGGGGGCCCGCACAAGCAGCGGGGCATGTGGCTTAATTCGACGCAACGCGAAGAACC TTACCAAGGCTTGACATATACCGGAAACATCCAGAGATGGGTGCCCCCTTGTGGTCGGTATACAGGTGGTGCATGG CTGTCGTCAGCTCGTGTCGTGAGATGTTGGGTTAAGTCCCGCAACGAGCGCAACCCTTGTCCTGTGTTGCCAGCAT GCCCTTCGGGGTGATGGGGACTCACAGGAGACCGCCGGGGTCAACTGGAGGAAGGTGGGGACGACGTCAAGTCATC ATGCCCCTTATGTCTTGGGCTGCACACGTGCTACAATGGCCGGTACAAAGAGCTGCGATGCCGTGAGGCGGAGCGA ATCTCAAAAAGCCGGTCTCAGTTCGGATTGGGGTCTGCAACTCGACCCCATGAAGTCGGAGTTGCTAGTAATCGCA GATCAGCATTGCTGCGGTGAATACGTTCCCCGGGCCTTGTACACGCCCCGTCACGTCACGAAAGTCGGTAACACC CGAAGCCGGTGGCCCAACCCCTTGTGGGAGGGAGCTGTCGAAGGTGGGACCAGCGATTGGGACGAAGTCGTAACAA GGTAACCGTAA

The PCR amplified 16S rDNA nucleotide sequences of S72-10

CGTGCTTACACATGCAAGTCGAACGATGAAACTTCCTTCGGGAGGGGTATTAGTGGCGAACGGGTGAGTAACACGT GGGCAATCTGCCCTTCACTCTGGGACAAGCCCTGGAAACGGGGTCTAATACCGGATATGACACGGGGTCGCATGAT CTTCGTGTGGAAAGCTCCGGCGGTGAAGGATGAGCCCGCGGCCTATCAGCTTGTTGGTGGGGTGATGGCCTACCAA GGCGACGACGGGTAGCCGGCCTGAGAGGGCGACCGGCCACACTGGGACTGAGACACGGCCCAGACTCCTACGGGAG GCAGCAGTGGGGAATATTGCACAATGGGCGAAAGCCTGATGCAGCGACGCCGCGTGAGGGATGACGGCCTTCGGGT TGTAAACCTCTTTCAGCAGGGAAGAAGCGAGAGAGCGGCGGTACCTGCAGAAGAAGCGCCGGCTAACTACGTGCCAGC AGCCGCGGTAATACGTAGGGCGCAAGCGTTGTCCGGAATTATTGGGCGTAAAGAGCTCGTAGGCGGCTTGTCGCGT CGGATGTGAAAGCCCGGGGCTTAACCCCGGGTCTGCATTCGATACGGGCAGGCTAGAGTTCGGTAGGGGAGATCGG AATTCCTGGTGTAGCGGTGAAATGCGCAGATATCAGGAGGAACACCGGTGGCGAAGGCGGATCTCTGGGCCGATAC TGACGCTGAGGAGCGAAAGCGTGGGGAGCGAACAGGATTAGATACCCTGGTAGTCCACGCCGTAAACGTTGGGAAC TAGGTGTGGGCGACATTCCACGTCGTCCGTGCCGCAGCTAACGCATTAAGTTCCCCGCCTGGGGAGTACGGCCGCA AGGCTAAAACTCAAAGGAATTGACGGGGGCCCCGCACAAGCAGCGGAGCATGTGGCTTAATTCGACGCAACGCGAAG AACCTTACCAAGGCTTGACATACACCGGAAAAACCGTGGAGACACTCCCCCTTGTGGTCGGTGTACAGGTGGTGCAT GGCTGTCGTCAGCTCGTGTCGTGAGATGTTGGGTTAAGTCCCGCAACGAGCGCAACCCTTGTTCTGTGTTGCCAGC ACGTCCTTTCGGGGATGGTGGGGACTCACAGGAGACTGCCGGGGTCAACTCGGAGGAAGGTGGGGACGACGTCAAG TCATCATGCCCCTTATGTCTTGGGCTGCACACGTGCTACAATGGCCGGTACAATGAGCTGCGATACCGTGAGGTGG AGCGAATCTCAAAAAAGCCGGTCTCAGTTCGGATTGGGGTCTGCAACTCGACCCCATGAAGTCGGAGTTGCTAGTAA TCGCAGATCAGCATTGCTGCGGTGAATACGTTCCCCGGGCCTTGTACACACCGCCCGTCACGTCACGAAAGTCGGTA ACACCCGAAGCCGGTGGCCCAACCCCTTGTGGGAGGGAATCGTCGAAGGTGGGACTGGCGATTGGGACGAAGTCGT AACAAGGTAACCGTA

The PCR amplified 16S rDNA nucleotide sequences of S75-3

GGCGTGCTTACACATGCAAGTCGAACGATGAACCGGTTTCGGCCGGGGATTAGTGGCGAACGGGTGAGTAACACGT GGGCAATCTGCCCTGCACTCTGGGACAAGCCCTGGAAACGGGGTCTAATACCGGATATGACTGCCGACCGCATGGT CTGGTGGTGGAAAGCTCCGGCGGTGCAGGATGAGCCCGCGGCCTATCAGCTTGTTGGTGGGGTGATGGCCTACCAA GGCGACGACGGGTAGCCGGCCTGAGAGGGCGACCGGCCACACTGGGACTGAGACACGGCCCAGACTCCTACGGGAG GCAGCAGTGGGGAATATTGCACAATGGGCGCAAGCCTGATGCAGCGACGCCGCGTGAGGGATGACGGCCTTCGGGT TGTAAACCTCTTTCAGCAGGGAAGAAGCGCAAGTGACGGTACCTGCAGAAGAAGCGCCGGCTAACTACGTGCCAGC AGCCGCGGTAATACGTAGGGCGCAAGCGTTGTCCGGAATTATTGGGCGTAAAGAGCTCGTAGGCGGCTTGTCGCGT CGGATGTGAAAGCCCGGGGCTTAACTCCGGGTCTGCATTCGATACGGGCAGGCTAGAGTTCGGTAGGGGAGATCGG AATTCCTGGTGTAGCGGTGAAATGCGCAGATATCAGGAGGAACACCGGTGGCGAAGGCGGATCTCTGGGCCGATAC TGACGCTGAGGAGCGAAAGCGTGGGGAGCGAACAGGATTAGATACCCTGGTAGTCCACGCCGTAAACGTTGGGAAC TAGGTGTGGGCGACATTCCACGTTGTCCGTGCCGCAGCTAACGCATTAAGTTCCCCGCCTGGGGAGTACGGCCGCA AGGCTAAAACTCAAAGGAATTGACGGGGGGCCCGCACAAGCGGCGGAGCATGTGGCTTAATTCGACGCAACGCGAAG AACCTTACCAAGGCTTGACATACATCGGAAACCTCTGGAGACAGGGCCCCCCTTGTGGTCGGTGTACAGGTGGTGCA TGGCTGTCGTCAGCTCGTGTCGTGAGATGTTGGGTTAAGTCCCGCAACGAGCGCAACCCTTGTCCTGTGTTGCCAG CATGCCCTTTGGGGTGATGGGGAACTCACAGGAGCACTGCCGGGGATCAACTCGGAGGAAGGTGGGGCACGACGTC AAGTCATCATGCCCCTTATGTCTTGGGCTGCACACGTGCTACAATGGCCGGTACAATGAGCTGCGAAGCCGTGAGG TGGAGCGAATCTCAAAAAGCCGGTCTCAGTTCGGATTGGGGTCTGCAACTCGACCCCATGAAGTCGGAGTCGCTAG TAATCGCAGATCAGCATTGCTGCGGTGAATACGTTCCCGGGCCTTGTACACACCGCCCGTCACGTCACGAAAGTCG CAAGGTACCGTAAACT

The PCR amplified 16S rDNA nucleotide sequences of S75-5

CGGCGTGCTTACCATGCAAGTCGAACGATGAAGCCCTTCGGGGTGGATTAGTGGCGAACGGGTGAGTAACACGTGG GCAATCTGCCCTTCACTCTGGGACAAGCCCTGGAAACGGGGTCTAATACCGGATACGACCCGCCGAGGCATCTCGG TGGGTGGAAAGCTCCGGCGGTGAAGGATGAGCCCGCGGCCTATCAGCTTGTTGGTGGGGTAACGGCCCACCAAGGC GACGACGGGTAGCCGGCCTGAGAGGGGGGGCGACCGGCCACACTGGGACTGAGACACGGCCCAGACTCCTACGGGAGGCA GCAGTGGGGAATATTGCACAATGGGCGAAAGCCTGATGCAGCGACGCCGCGTGAGGGATGACGGCCTTCGGGTTGT AAACCTCTTTCAGCAGGGAAGAAGCGCAAGTGACGGTACCTGCAGAAGAAGCGCCGGCTAACTACGTGCCAGCAGC CGCGGTAATACGTAGGGCGCAAGCGTTGTCCGGACATTATTGGGCGTAAAGAGCTCGCTAGGCGGCTTGTCACGTC GGGTGTGAAAGCCCGGGGCTTAACCCCGGGTCTGCATCCGATACGGGCAGGCTAGAGTGTGGTAGGGGAGATCGGA ATTCCTGGTGTAGCGGTGAAATGCGCAGATATCAGGAGGAACACCGGTGGCGAAGGCGGATCTCTGGGCCATTACT GACGCTGAGGAGCGAAAGCGTGGGGGAGCGAACAGGATTAGATACCCTGGTAGTCCACGCCGTAAACGTTGGGAACT AGGTGTTGGCGACATTCCACGTCGTCGGTGCCGCAGCTAACGCATTAAGTTCCCCGCCTGGGGAGTACGGCCGCAA GGCTAAAACTCAAAGGAATTGACGGGGGCCCGCACAAGCAGCGGAGCATGTGGCTTAATTCGACGCAACGCGAAGA ACCTTACCAAGGCTTGACATATACCGGAAACATCCAGAGATGGGTGCCCCCCTTGTGGTCGGTATACAGGTGGTGCA TGGCTGTCGTCAGCTCGTGTCGTGAGATGTTGGGTTAAGTCCCGCAACGAGCGCAACCCTTGTCCTGTGTTGCCAG CATGCCCTTCGGGGTGATGGGGACTCACAGGAGACCGCCGGGGTCAACTCGTGAGGAAGGTGGGGAACGCACGTCA AGTCATCATGCCCCTTATGTCTTGGGCTGCACACGTGCTACAATGGCCGGTACAAAGAGCTGCGATGCCGTGAGGC GGAGCGAATCTCAAAAAGCCGGTCTCAGTTCGGATTGGGGTCTGCAACTCGACCCCATGAAGTCGGAGTTGCTAGC AATCCGCAGATCAGCATTGCTGCGGGGGATACGTTCCCGGGCCTTGTACACACCGCCCGTCACGTCACGAAAGCGG

The PCR amplified 16S rDNA nucleotide sequences of S76-1

CGGCGGGCTTACACATGCAAGTCGAACGGTGAAGCCCTTCGGGTGGTATCATGGCGAACGGGTGAGTAACACGTGG GCAATCTGCCCTGCACTCTGGGACAAGCCCTGGAAACGGGGTCTAATACCGGATATGACCTGGGACCGCATGGTCT GGGTGTAAAGCTCCGGCGGTGCAGGATGAGCCCGCGGCCTATCAGCTTGTTGGTGGGGTAATGGCCTACCAAGGCG ACGACGGGTAGCCGGCCTGAGAGGGGCGACCGGCCACACTGGGACTGAGACACGGCCCAGACTCCTACGGGAGGCAG CAGTGGGGAATATTGCACAATGGGCGCAAGCCTGATGCAGCGACGCCGCGTGAGGGATGACGGCCTTCGGGTTGTA AACCTCTTTCAGCAGGGAAGAAGCGCCAAGTGACGGTACCTGCAGAAGAAGCACCGGCTAACTACGTGCCAGCAGCC GCGGTAATACATAGGGTGCGAGCGTTATCCGGAATTATTGGGCGTAAAGAGCTCGCTAGGCGGCCTGTCGCGTCGG ATGTGAAAGCCCGGGGCTTAACCCCGGGTCTGCATTCGATACGGGCAGGCTAGAGTGTGGTAGGGGAGATCGGAAT TCCTGGTGTAGCGGTGAAATGCGCAGATATCAGGAGGAACACCGGTGGCGAAGGCGGATCTCTGGGCCATTACTGA CGCTGAGGAGCGAAAGCGTGGGGAGCGAACAGGATTAGATACCCTGGTAGTCCACGCCGTAAACGTTGGGAACTAG GTGTTGGTCACATTCCACGTGATCGGTGCCGCAGCTAACGCATTAAGTTCCCCGCCTGGGGAGTACGGCCGCAAGG CTAAAACTCAAAGGAATTGACGGGGGCCCGCACAAGCAGCGGAGCATGTGGCTTAATTCGACGCAACGCGAAGAAC CTTACCAAGGCTTGACATATGCCGGAAACACCTGGAGACAGGTGCCCCCTTGTGGTCGGTATACAGGTGGTGCATG GTTGTCGTCAGCTCGTGTCGTGAGATGTTGGGTTAAGTCCCGCAACGAGCGCAACCCTTGTTCTGTGTTGCCAGCA TGCCTTTCGGGGCTGATGGGGGATCTCACAGGCACACTGAAGGGGATCAACACGAGAGGAGGAGGGGGCACGACGTC AAATCATCATGCCCCTTATGTCTTGGGCTGCACACGTGCTACAATGGTCGGTACAAAGGGCTGCGATGCCGTGAGG CGGAGCGAATCCCAAAAAGCCGGCCTACAGTTCGGATTGGGGTCTGCAACTCGACCCCATGAAGTTGGAGTTGCTA GTAATCGCAGATCAGCATGCTGCGGTGAATACGTTCCCGGGCCTTGTACACCGCCCGTCACGTCACGAAAGTCG GTAACACCCGAAGCCGGTGGCCTAACCCTCTGGGATGGAGCCGTCAAGGTGGGACCAGCGATTGGGACGAAGTCGT AACAAGGTAACCGTA

The PCR amplified 16S rDNA nucleotide sequences of SB3-2

GCGTGCTTACACATGCAAGTCGAACGATGATCCCGCTTCGGTGGGGGGTATTAGTGGCGCAACGGGTGAGTAACACG TGGGCAATCTGCCCTGTACTTTGGGATAACCTGGGAAACTGGGTCTAATACCGGATATGACCTTCCTCGCATGGGT TTGGTGAAACTCCGGCGGTACGGATGACCCGCGGCCTATCACTTGTTGGTGGGGTAATGGCCTACCAAGGCACACG GGTAGCCGGCTGAGAGGGTGACCGGCCACCTGGGACTGAACACGGCCCAACTCCTACGGGAGGCAGCAGTGGGGAA TATTGCACAATGGGCGCAAGCCTGATGCAGCGACCCGCGTGAGGGATGACGGCCTTCGGTTGTAAACCTCTTTCCC AGGGACAAGCGCAAGTGACGGTACCTGGATAAAAGCACCGGCTAACTACGTGCCAGCACCGCGGTAATACTAGGGT ACGCTTAACGTGGAGCGTGCGGGTCGTATACGGGCAGACTTGAGTTCGGTAGGGGAGACTGGAATTCCTGGTGTAG CGGTGAAATGCGCAGATATCAAGAGGAACACCGGTGGCAAAGGCGGCTCTCTGGGCCGATACTGACGCTGAGGAGC GAAAGCGTGGGGAGCGAACAGGATTAGATACCCTGGTAGTCCACGCTGTAAACGTTGGGCGCTAGGTGTGGGCGAC ATCCACGTTGTCCGTGCCGTAGCTAACGCATTAAGCGCCCCGCCTGGGGAGTACGGCCGCAAGGCTAAAACTCAAA GGAATTGACGGGGGCCCGCACAAGCGGCGGAGCATGTGGATTAATTCGATGCAACGCGAAGAACCTTACCTGGGCT TGACATGCGCCAGACATCCCTAGAGATAGGGCTTCCCTTGTGGTGGTGCACGGTGGCATGGCTGTCGTCAGC TCGTGTCGTGAGATGTTGGGTTAAGTCCCGCAACGAGCGCAACCCTTATCCTACGTTGCCAGCGCGTTATGGCGGG GACTCGTGGGAGACTGCCGGGGTCAACTCGGAGGAGGTGGGGATGACGTCAAGTCATCATGCCCCTTATGTCCAG GGCTTCACACATGCTACAATGGCTGGTACAGAGGGCTGCGATACCGCGAGGGTGGAGCGAATCCCTTAAAGCCGGT CTCAGTTCGGATCGCAGTCTGCAACTCGACTGCGTGAAGTCGGAGTCGCTAGTAATCGCAGATCAGCAACGCTGCG GTGAATACGTTCCCGGGCCTTGTACACCGCCCGTCACGTCATGAAAGTCGGTAACACCCGAAGCCCATGGCCCA ACCCGCAAGGGGGGGGGGGTGGTCGAAGGTGGGACTGGCGATTGGGACGAAGTCGTAACAAGGTAACCG

The PCR amplified 16S rDNA nucleotide sequences of SB7-3

GCGTGCTTACACATGCAAGTCGAACGCTGAACCGGTTTCGGCCGGGGATGAGTGGCGAACGGGTGAGTAACACGTG GGTAATCTGCCCTGTACTCTGGGATAAGCCTGGGAAACTGGGTCTAATACCGGATATGACCGCTACAGGCATCTGT GGTGGTGGAAAGTTCCGGCGGTATGGGATGAACCCGCGGCCTATCAGCTTGTTGGTGGGGTAATGGCCTACCAAGG CGACGACGGGTAGCCGGCCTGAGAGGGTGACCGGCCACACTGGGACTGAGACACGGCCCAGACTCCTACGGGAGGC AGCAGTGGGGAATATTGCACAATGGGCGCAAGCCTGATGCAGCGACGCCGCGTGAGGGATGACGGCCTTCGGGTTG TAAACCTCTTTCGCCAGGGACGAAGCGAAGAGTGACGGTACCTGGATAAGAAGCACCGGCTAACTACGTGCCAGCAG CCGCGGTAATACGTAGGGTGCGAGCGTTGTCCGGAATTATTGGGCGTAAAGAGCTCGTAGGCGGTTTGTCGCGTCG GCCGTGAAATCTCCACGCTTAACGTGGAGCGTGCGGTCGATACGGGCAGACTTGAGTTCGGCAGGGGAGACTGGAA TTCCTGGTGTAGCGGTGAAATGCGCAGATATCAGGAGGAACACCGGTGGCGAAGGCGGGTCTCTGGGCCGATACTG ACGCTGAGGAGCGAAAGCGTGGGGAGCGAACAGGATTAGATACCCTGGTAGTCCACGCTGTAAACGTTGGGCGCTA GGTGTGGGCGACATTCCACGTTGTCCGTGCCGTAGCTAACGCATTAAGCGCCCCGCCTGGGGAGTACGGCCGCAAG GCTAAAACTCAAAGGAATTGACGGGGGCCCGCACAAGCGGCGGAGCATGTGGATTAATTCGATGCAACGCGAAGAA GCTGTCGTCAGCTCGTGTCGTGAGATGTTGGGTTAAGTCCCGCAACGAGCGCAACCCTTATCCTACGTTGCCAGCG CGTTCATGGCGGGGGACTCGCTGGGAGACTGACCGGGGGTCAACATCGGAGGAAGGTGGGGATGACGTCAAGTCATC ATGCCCCTTATGTCCAGGGCTTCACACATGCTACAATGGCTGGTACAGAGGGCTGCGATACCGCGAGGTGGAGCGA ATCCCTTAAAAAGCCGGTCTCAGTTCCGGATCGCAGTCTGCAACTCGACTGCGTGAAGTCGGAGTCCCTAGTAATCG CAGATCAGCAACGCTGCGGAGAATACGTTCCCGGGCCTTGTACACACCGCCCGTCACGTCATGAAAGTCGGTAACA CCCGAAGCCCATGGCCCAACCCGCAAGGGAGGGAGTGGTCAAGGTGGGACTGGCGATTGGGACGAAGTCGTACAAG GTACCGTAAGTAACACC

The PCR amplified 16S rDNA nucleotide sequences of S39-7

TGCTTACACATGCAAGTCGAACGCTGAACCACTTTCGGGTGGGGATGAGTGGCGAACGGGTGAGTAACACGTGGGT AATCTGCCCTGCACTCTGGGATAAGCCTTGGAAACGAGGTCTAATACCGGATATCACTCCTTCGCATGGAAGATGT TGAAAGCTCCGGCGGTGCAGGATGAACCCGCGGCCTATCAGCTTGTTGGTGGGGTAGTGGCCTACCAAGGCGACGA CGGGTAGCCGGCCTGAGAGGGTGACCGGCCACACTGGGACTGAGACACGGCCCAGACTCCTACGGGAGGCAGCAGT GGGGAATATTGCACAATGGGCGCAAGCCTGATGCAGCGACGCCGCGTGAGGGATGACGGCCTTCGGGTTGTAAACC TCTTTCGCCAGGGACGAAGCGCAAGTGACGGTACCTGGATAAGAAGCACCGGCTAACTACGTGCCAGCAGCCGCGG TAATACGTAGGGTGCGAGCGTTGTCCGGATTTATTGGGCGTAAAGAGCTCGTAGGCGGTTTGTCGCGTCGGCCGTG AAAATCTCCACGCTTAACGTGGAGCGTGCGGTCGATACGGGCAGACTTGAGTTCGGTAGGGGAGACTGGAATTCCT GGTGTAGCGGTGAAATGCGCAGATATCAGGAGGAACACCGGTGGCGAAGGCGGGTCTCTGGGCCGATACTGACGCT GAGGAGCGAAAGCGTGGGGAGCGAACAGGATTAGATACCCTGGTAGTCCACGCTGTAAACGTTGGGCGCTAGGTGT GGGCGACATCCACGTTGTCCGTGCCGTAGCTAACGCATTAAGCGCCCCGCCTGGGGAGTACGGCCGCAAGGCTAAA ACTCAAAGGAATTGACGGGGGGCCCGCACAAGCGGCGGAGCATGTGGATTAATTCGATGCAACGCGAAGAACCTTAC CGTCAGCTCGTGTCGTGAGATGTTGGGTTAAGTCCCGCAACGAGCGCAACCCTTATCCTACGTTGCCAGCGCGTTA TGTCCAGGGTCTTCACACATGCTACAATGGCTGGTACAGAGGGCTGCGATACCGCGAGGTGGAGCGAATCCCTTAA AGCCGGTCTCAGTTCGGATCGCAGTCTGCAACTCGACTGCGTGAAGTCGGAGTCGCTAGTAATCGCAAATCAGCAA CGCTGCGGTGAATACGTTCCCGGGCCTTGTACACACCGCCCGTCACGTCATGAAAGTCGGTAACACCCGAAGCCCA

The PCR amplified 16S rDNA nucleotide sequences of KC19-1

GCGGGCTTACACATGCAAGTCGAACGCATGTAACCACTATTCGGGGTGGGGTTAATGGTCGAAGGCATGTAGTAAC ACGTGGGGTAATCTGCCCTGCACTCTGGGGATAACCTTGAAAACGAGGTCTAATACCGGTATATCACTGCTCTCGC ATGGGGAATGTTGAAAGCTCCGTGCGGCTGCAGGTTGAACCCGCGGCCTATCCTTGTTGGTGGGGTAGTGGCCTAC CAAGCGACGACGGGTAGCCGCCTGACAGGGTGACCGGCCACACTGGGACTGAACACGGCCCATACTCCTACTGGAG GCGATGGGGAATATTGCACATGGGGCAAGCCTGATGCAACCACCCCCGCGTGAGGGATGACTGCCTTCGGGTTGTAA ACCTCTTTCCCAGGGACAAGCGAGTGACGGTACCTGGATAAGAAGACCGGCTAACTACGTGCCAGCAGCCGCGGTA ATACTAGGTGCGAGCGTTGTCCGGGATTTATTGGGCGTAAACAGCTCGTAAGCGGTTTGTCGCGTTCGGCCGTGAAA ATCTCCACGCTTAACGTGGAACGTGCGGTCATACGGGCAGACTTGAGTTCGGTAGGGAGACTGGAATTCCTGGTGT AGCGGTGAAATGCCAAATATCAGGAGGAACACCGGTGGCAAGGCGGTCTCTGGCCGATACTGACCTAGGACGAAAG CTGGGGAGCGAACAGGATTAATACCCTGGTATCCACCTGTAAACTTGGGCGCTAGGTGTGGGCGACATCCACTTGT CCGTGCCGTACTAACGCATTAACGCCCCGCCTGGGAGTACGGCCCAAGGTAAAACTCAAAGGAATTGACGGGGGGCC CGCACAAGCGGCGACATGTGGATTAATTCATGCAACGCGAAGAACCTTACCTGGCTTGACATGCGCCAGACATCCC CAGAATGGGGCTTCCCTTGTGGTGGTGGTGCACGGTGGCATGGCTGTCGTCGTCGTGGGGTGTGGGGTT AAGTCCCGCAACGAGCGCAACCCTTATCCTACGTTGCCAGCGCGTTATGGCGGGGGACTCGTGGGGAGATCTGCCGGG ATCAACTCGAAGAAGTTGTGGGGGCATGAACATCAAGATCATGATCCATGCCCCATTATGTCCAGGGTCTTCACACATGCT ACAATGGGCTGGCTACAGAGGGCTGCGATACCCCGAGGTGGAGCGAATCCCTTAAAGCCGGTCTCAGTTCGAATCG CAGTCTGCACTCCACTGCTGAAGTCGGAGTCCCTAGTAATCGCAAATCAGCACCGCTGCGGTGAATATTCCCGGGC CTTGTACACCGCCCGTCACGTCATGAATGAAGTAACACCCCGAACCCATGGCCACCCTTAGAGGGAGTGGTCGAA GGTGGGACTGGCGATTGGACCAAATCGTGCAAGGTATCCGTA

The PCR amplified 16S rDNA nucleotide sequences of KC20-1

The PCR amplified 16S rDNA nucleotide sequences of K57-1

VITA

Miss Piyapat Sripairoj was born on July 30, 1980 in Khonkaen Province, Thailand. She received her Bachelor Degree of Science in Pharmacy (second class honors) in 2003 from the faculty of Pharmaceutical Sciences, Chulalongkorn University, Thailand.

