

การสังเคราะห์ฟิวโรฟิวแรนลิกแนนชนิดใหม่ผ่านการแทนที่ด้วยนิวคลีโอไฟล์ของซามีน



บทคัดย่อและแฟ้มข้อมูลฉบับเต็มของวิทยานิพนธ์ตั้งแต่ปีการศึกษา 2554 ที่ให้บริการในคลังปัญญาจุฬาฯ (CUIR)
เป็นแฟ้มข้อมูลของนิสิตเจ้าของวิทยานิพนธ์ ที่ส่งผ่านทางบัณฑิตวิทยาลัย

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วิทยานิพนธ์นี้เป็นส่วนหนึ่งของการศึกษาตามหลักสูตรปริญญาวิทยาศาสตรมหาบัณฑิต
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SYNTHESIS OF NEW FUROFURAN LIGNANS THROUGH NUCLEOPHILIC SUBSTITUTION
OF SAMIN

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A Thesis Submitted in Partial Fulfillment of the Requirements
for the Degree of Master of Science Program in Chemistry
Department of Chemistry
Faculty of Science
Chulalongkorn University
Academic Year 2016
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นันทพร สุรชัยชนวัฒน์ : การสังเคราะห์ฟิวโรฟิวแรนลิกแนนชนิดใหม่ผ่านการแทนที่ด้วยนิวคลีโอไฟล์ของซามิน (SYNTHESIS OF NEW FUROFURAN LIGNANS THROUGH NUCLEOPHILIC SUBSTITUTION OF SAMIN) อ.ที่ปรึกษาวิทยานิพนธ์หลัก: รศ. ดร.ปรีชา ภูวไพโรศิรศาล, อ.ที่ปรึกษาวิทยานิพนธ์ร่วม: รศ. ดร.วิวัฒน์ วชิรวงศ์กวิน, 137 หน้า.

การสังเคราะห์แบบ semi-synthesis ของ furofuran lignan ที่มีสูตรโครงสร้างแบบ [3.3.0]ไบไซคลิก เกือบ 50 อนุพันธ์เริ่มต้นจากสารที่พบได้ในธรรมชาติ sesamol ถูกอธิบาย วิธีการสังเคราะห์เกี่ยวข้องกับการ protonation ของ samini เพื่อให้เกิดเป็นสารมัธยันตร์ oxocarbenium ion ตามด้วยการเข้าทำปฏิกิริยาของ nucleophile ที่แตกต่างกัน 3 ชนิด ได้แก่ ฟีนอลิก ไทออล และ แอลกอฮอล์ วิธีการนี้ให้ผลิตภัณฑ์ที่เป็นคู่ diastereomer ยกเว้นจากการสังเคราะห์ด้วย samini กับ แอลกอฮอล์ กลไกการเกิดปฏิกิริยาแบบ stereoselective ถูกตรวจสอบโดยการติดตามปฏิกิริยาด้วย ^1H NMR และการคำนวณทางคอมพิวเตอร์ ผลการทดลองแสดงให้เห็นว่า ผลิตภัณฑ์ที่ได้น่าจะเกิดผ่านกลไกปฏิกิริยาแบบ $\text{S}_{\text{N}}1$ -like โดยการ protonation ของหมู่ hemiacetal ของ samini เพื่อเกิดเป็นสารมัธยันตร์ oxocarbenium ion ตามด้วยทำปฏิกิริยากับแอลกอฮอล์ เกิดผลิตภัณฑ์ *exo,exo*-alkoxysamin ทั้งโดยตรง หรือจากการ protonation ของ *endo,exo*-alkoxysamin ผ่าน $\text{S}_{\text{N}}2$ -like transition state

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5772028523 : MAJOR CHEMISTRY

KEYWORDS: FUROFURAN LIGNAN / SAMIN / NUCLEOPHILIC SUBSTITUTION

NANTAPORN SURACHAITANAWAT: SYNTHESIS OF NEW FUROFURAN LIGNANS THROUGH NUCLEOPHILIC SUBSTITUTION OF SAMIN. ADVISOR: ASSOC. PROF. PREECHA PHUWAPRAISIRISAN, Ph.D., CO-ADVISOR: ASSOC. PROF. VIWAT VCHIRAWONGKWIN, Ph.D., 137 pp.

The semi-synthesis of fifty furofuran lignans having a bicyclo[3.3.0]octane skeleton starting from the naturally available sesamol is described. Our methodology involved protonation of samin to generate oxocarbenium ion followed by the attack of three different nucleophiles, namely, phenolics (ArOH), thiols (RSH) and alcohols (ROH). This synthesis strategy provided diastereomeric products except for those synthesized from samin and alcohols. The mechanism of stereoselective formation was investigated by ^1H NMR monitoring technique and computational calculation. The results revealed that the products were presumably formed through S_N1 -like mechanism by protonation of the hemiacetal center of samin to generate the corresponding oxocarbenium ion as the intermediate. Subsequent reaction of this oxocarbenium ion with the alcohols then led to the observed *exo,exo*-alkoxysamin either directly, or alternatively, by protonation of the *endo,exo*-alkoxysamin through the S_N2 -like transition state.

Department: Chemistry

Field of Study: Chemistry

Academic Year: 2016

Student's Signature

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ACKNOWLEDGEMENTS

She would like to express my deepest gratitude to my advisor, Associate Professor Dr. Preecha Phuwapraisirisan and co-advisor, Associate Professor Dr. Viwat Vchirawongkwin for their generous assistance and providing me insight and expertise that greatly supported this research. Single-crystal X-ray diffraction facility was supported through Associate Professor Dr. Thammarat Aree.

She also gratefully acknowledges the committees, Associate Professor Dr. Vudhichai Parasuk, Assistant Professor Dr. Warinthron Chavasiri and Dr. Oraphin Chantarasriwong for their comments, guidance and extending cooperation over my presentation.

She is thankful to Dr. Wisuttaya Worawalai and Dr. Rico Ramadhan for their technical assistance and kindly suggestions. Also she thanks members of Natural Product Research group and all of her friends for their friendships she has had during her study.

She would like to express her gratitude to Department of Chemistry, Faculty of Science, Chulalongkorn University and the Science Achievement Scholarship of Thailand (SAST) for offering the chemicals and facilities throughout the course of study.

Last but not the least, she would like to thank her family for their spiritual supporting all over her life.

CONTENTS

	Page
THAI ABSTRACT	iv
ENGLISH ABSTRACT	v
ACKNOWLEDGEMENTS	vi
CONTENTS	vii
List of Tables	x
List of Figures.....	xi
List of Schemes.....	xii
List of Abrevations.....	xiii
CHAPTER I INTRODUCTION.....	1
CHAPTER II SYNTHESYS OF SAMIN.....	6
2.1 Isolation of sesamolin	6
2.2 Synthesis of samin	7
2.3 Experimental section.....	8
2.3.1 General experiment procedures.....	8
2.3.2 Chemical.....	8
2.3.3 Determining of saponification number.....	8
2.3.4 Isolation of sesamolin.....	9
2.3.4 Synthesis of samin.....	9
CHAPTER III SYNTHESIS OF NEW FUROFURAN LIGNANS FROM SAMIN.....	11
3.1 General procedure for the synthesis of new furofuran lignans.....	11
3.2 Synthesis of new furofuran lignans using phenolics as a nucleophile	11
3.2.1 Structural characterization of synthesized furofuran lignans (3.1 and <i>epi</i> -3.1).....	14

	Page
3.3 Synthesis of new furofuran lignans using thiols as a nucleophile	16
3.3.1 Structural characterization of synthesized furofuran lignans (3.2 and <i>epi</i> -3.2)	19
3.4 Synthesis of new furofuran lignans using alcohols as a nucleophile	21
3.4.1 Structural characterization of synthesized furofuran lignans (3.3 and <i>epi</i> -3.3)	24
3.5 Experimental section	25
3.5.1 General experiment procedures	25
3.5.2 Chemical	25
3.5.3 General procedure for synthesis of new furofuran lignans	25
3.5.4 X-ray Crystallographic Analysis of 3.2r and <i>epi</i> -3.2r	64
CHAPTER IV MECHANISTIC INVESTIGATION OF ALKOXYSAMIN (3.3) FORMATION	67
4.1 Mechanistic investigation on 3.3 formation	67
4.2 Mechanistic investigation on epimerization of alkoxyamin and <i>epi</i> - alkoxyamin	69
4.3 Computational calculations	71
4.4 Experimental section	74
4.4.1 Mechanistic investigation on 3.3 formation	74
4.4.2 Mechanistic investigation on epimerization	74
4.4.2.1 Synthesis of 4.1 and <i>epi</i> -4.1	74
4.4.2.2 ¹ H NMR monitoring section	75
4.4.3 Computational section	75
CHAPTER V CONCLUSION	76
REFERENCES	77

	Page
APPENDIX.....	78
VITA.....	137



List of Tables

Tables	Pages
3.1 Synthesis of furofuran lignans from samin (1.36) with phenolics (a - k)	13
3.2 Synthesis of furofuran lignans from samin (1.36) with thiols (l - z).....	18
3.3 Synthesis of furofuran lignans from samin with alcohols (A - N).....	23
3.4 Crystal data and refinement parameters of compounds 3.2r and <i>epi-3.2r</i> ..	65
4.1 Most important bond distances (in Å) and valence angle (in °) of transition state structure.....	72



List of Figures

Figures	Pages
1.1 Three subgroups of furofuran lignans	1
1.2 Structure of bioactive furofuran lignans	2
2.1 Isolation of sesamol (1.2) from sesame oil.....	7
3.1. Phenolics used in synthesis strategy.....	12
3.2. ¹ H NMR spectra of (i) 3.1b and (ii) <i>epi-3.1b</i>	15
3.3 Diagnostic NOESY and HMBC correlations of (i) 3.1b and (ii) <i>epi-3.1b</i>	16
3.4 Thiols used in synthesis strategy.....	17
3.5 ¹ H NMR spectra of (i) 3.2r and (ii) <i>epi-3.2r</i>	19
3.6 Selected HMBC correlations of 3.2r and <i>epi-3.2r</i>	20
3.7 ORTEP plots (20% probability level) of 3.2r and <i>epi-3.2r</i>	21
3.8 Alcohols used in synthesis strategy.....	21
3.9 ¹ H NMR spectra of (i) 3.3A and (ii) 3.2l	24
4.1 Overlaid ¹ H NMR spectra of methoxy-d ³ samin, 4.1 and <i>epi-4.1</i> (CD ₃ OD), occurring in the reaction, for different times.....	67
4.2 Time course of the evolution of samin (1.36), 4.1 , and <i>epi-4.1</i> in mole fraction by ¹ H NMR spectroscopy (400 MHz).....	67
4.3 Overlaid ¹ H NMR spectra of (i) <i>epi-4</i> and (ii) the treated <i>epi-4.1</i> with CD ₃ OD and TFA-d for 10 min.	70
4.4 Transition-state model for the epimerization reaction of <i>epi-4.1</i> . (Note ^a Atoms of TFA-d and ^b Atoms of CD ₃ OD)	71

List of Schemes

Schemes	Pages
1.1 Synthesis of 1-fluoro- <i>endo,exo</i> -furofurans 1.11-1.14	2
1.2 Synthesis of 1-fluoro- <i>endo,exo</i> -furofurans 1.18-1.32	3
1.3 Semi-synthesis of sesaminol (1.34) and alkyloxy samins (1.35) from sesamolin (1.2)	4
1.4 Synthetic strategy to furofuran lignans 3.1-3.3	5
2.1 Hydrolysis of sesamolin (1.2).....	7
3.1 Proposed mechanistic formation of 3.1a – 3.1k and epi-3.1a – epi-3.1k	14
3.2 Preparation of epi-3.2r-pBBE	20
4.1 Proposed mechanistic formation of 4.1 and epi-4.1	68
4.2 Epimerization of epi-4.1 under different conditions i) - iii).....	69
4.3 Proposed S _N 2 reaction between epi-4.1 and deuterated methanol (CD ₃ OD) with the present of deuterated-trifluoroacetic acid (CF ₃ CO ₂ D).....	70
4.4 A proposed mechanistic reaction of synthesis of stereoselective products (3.2 and 3.3).....	73

List of Abbreviations

^{13}C NMR	carbon-13 nuclear magnetic resonance
CDCl_3	deuterated chloroform
CD_3OD	deuterated methanol
d	doublet (NMR)
dd	doublet of doublet (NMR)
2D NMR	two dimensional nuclear magnetic resonance
ESIMS	electrospray ionization mass spectrometry
eq	equivalent (s)
g	gram (s)
^1H NMR	proton nuclear magnetic resonance
HMBC	heteronuclear multiple bond correlation experiment
Hz	hertz
HRESIMS	high resolution electrospray ionization mass spectrum
h	hour (s)
J	coupling constant
mg	milligram (s)
mL	milliliter (s)
mmol	millimole (s)
m/z	mass per charge
m	multiplet (NMR)
M.W.	molecular weight
M	molar
NOESY	nuclear overhauser enhancement spectroscopy
rt	room temperature
s	singlet (NMR)

TFA-d	deuterated-trifluoroacetic acid ($\text{CF}_3\text{CO}_2\text{D}$)
TLC	thin layer chromatography
UV	ultraviolet
δ	chemical shift
$^\circ\text{C}$	degree Celsius
%yield	percentage yield



CHAPTER I

INTRODUCTION

Furofurans are one of major subclasses of lignan family which originated from shikimic acid pathway followed by dimerization of phenyl propanoids (C_6-C_3) at center carbon (C-8) (Figure 1.1) [1]. Since their structures contain 2,6-diaryl substituted on 3,7-dioxabicyclo[3.3.0]octane skeleton, furofurans can be divided into three types; *endo-endo*, *endo-exo* and *exo-exo*, based on stereochemistry of substituents related to head bridge hydrogen (Figure 1.1)

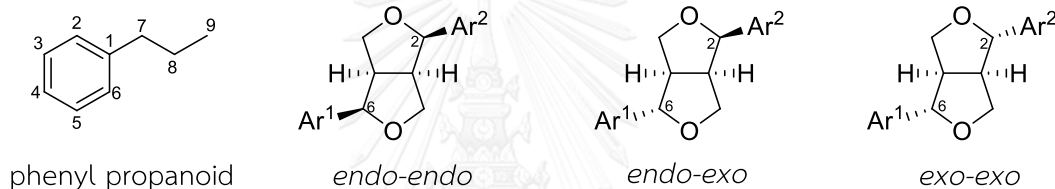


Figure 1.1 Structure of phenyl propanoid and three subgroups of furofuran lignans

According to variety of their structures, furofuran lignans are responsible for a wide range of bioactive compounds, such as antitumoral of syringaresinol (**1.1**) [2], antioxidant of sesamol (**1.2**) [3] and reducing serum and liver cholesterol levels in rat of sesamin (**1.3**) [4]. This has inspired organic chemists to develop synthetic methodologies for synthesis of this class of bioactive compounds and improve their chemical and pharmaceutical profiles.

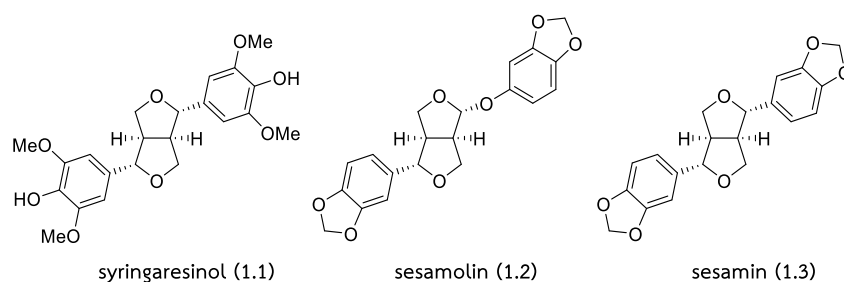
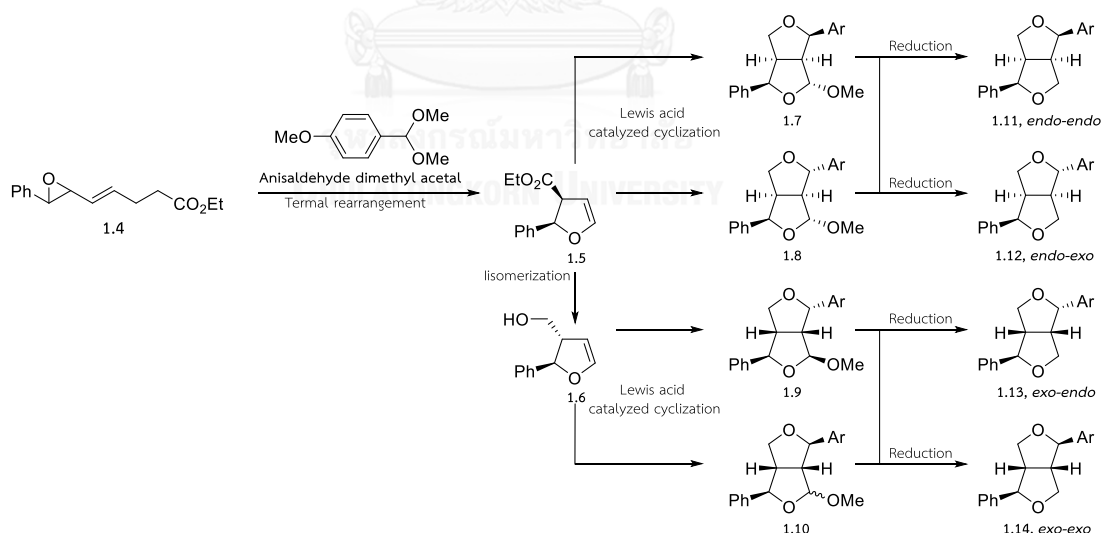


Figure 1.2 Structure of bioactive furofuran lignans.

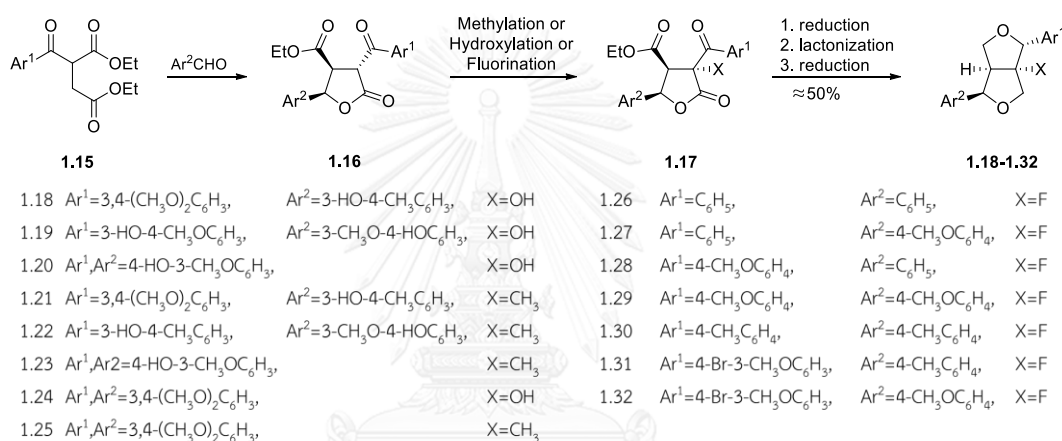
For instance in 2003, David [5] reported diastereoselective synthesis of furofuran lignans. Single vinyl epoxide (**1.4**) was used as a starting material followed by thermal rearrangement afforded *cis*-disubstituted dihydrofuryl ester (**1.5**). Subsequent lewis acid catalyzed cyclization gave either the *endo-endo* (**1.7**) or *endo-exo* furofuran acetal (**1.8**) depending on reaction temperatures. Then, isomerization of **1.5** provided the alternative *exo-endo* and *exo-exo* isomers (**1.9** and **1.10** respectively). Finally, acetal group was reduced to cyclic ether to complete the synthesis furofuran skeleton (**1.11-1.14**), as shown in Scheme 1.1.



Scheme 1.1 Synthesis of 1-fluoro-*endo,exo*-furofurans **1.11-1.14**

As well in 2005, Pohmakotr [6] synthesized 1-substituted *endo,exo*-2,6-diaryl-3,7-dioabicyclo[3.3.0]octanes (**1.18-1.24**) and (\pm)-gmelinol (**1.25-1.26**). Their investigation began with the synthesis of (2,3-*trans*)-(4,5-*cis*)- α -aroylparaconic esters

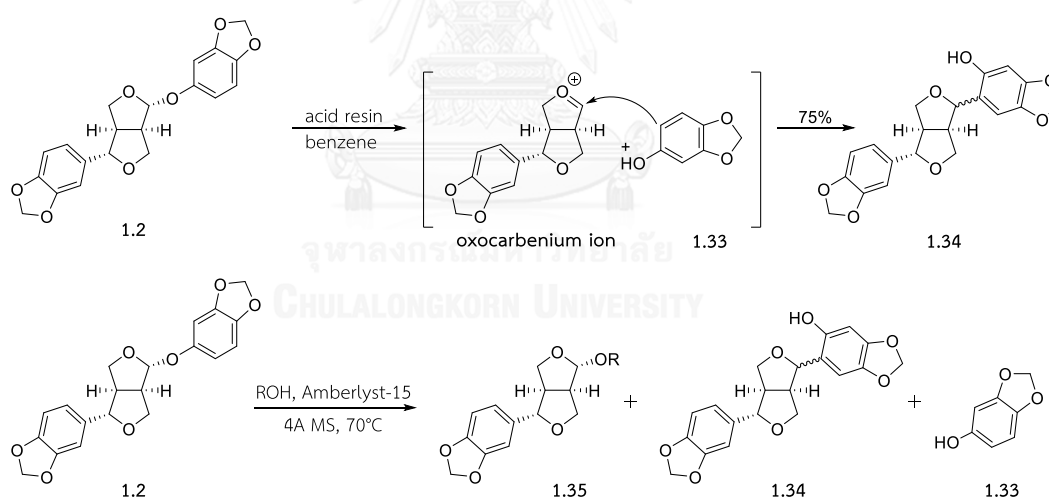
(1.16) by reacting vicinal dianions derived from the corresponding α -aroylsuccinic esters (1.15) with aromatic aldehydes. Subsequently, methylation or hydroxylation at carbon position 1 was carried out to produce 1-substituted lactone as core structure. They next achieved synthesis of furofurans 1.18-1.26 by three steps involving reductions and lactonization as shown in Scheme 1.1. Similarly in 2015, Punirun [7] reported the synthesis of 1-fluorine-substituted *exo,exo*-2,6-diaryl-3,7-dioxabicyclo[3.3.0]octanes (1.27-1.32) using aforementioned procedure (Scheme 1.2).



Scheme 1.2 Synthesis of 1-fluoro-*endo,exo*-furofurans 1.18-1.32

Although a number of developed synthetic methodologies and structure modifications of these bioactive compounds were explored, structural activity relationship studies (SARs) have not been reported due to the lack of a practical synthetic method of producing diverse furofuran lignans. As aforementioned synthetic strategies, furofurans were synthesized through many stepwise combinations of small building block leading to provide expected products in low yield ($\approx 50\%$). Moreover, a variety of furofuran lignans was also dropped because of the limited diversity of starting material. From this reasons, we would like to optimize a new synthetic approach to easily produce a huge series of furofuran lignans.

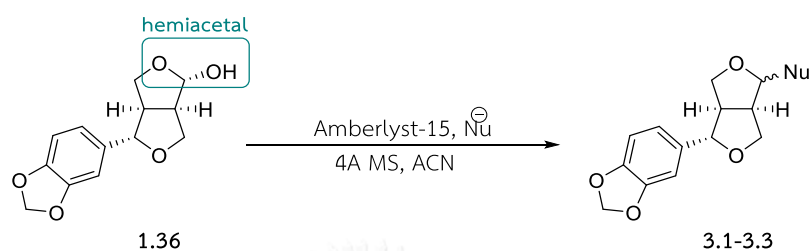
Recently, some researchers reported short route synthesis of furofuran lignans by modifying naturally available sesamol (1.2) to offer its derivatives. For example in 2012 [8], Haung synthesized sesaminol (1.34) from 1.2 in the presence of acidic resin using benzene as aprotic nonpolar solvent. Noteworthy, the key step in aforementioned synthesis involved acid-catalyzed formation of oxocarbenium ion. Nucleophilic substitution by carbon nucleophile such as sesamol (1.33) afforded 1.34. Identically in 2015, Mahamad [9] synthesized a series of alkyloxy samins (1.35) in one step by reaction of sesamol (1.2) with various alcohols. The products were generated with retention of configuration, which were different from mechanistic presumption involving oxocarbenium ion that would result in product mixtures. Moreover, this methodology also has limitation due to the occurrence of unexpected products sesaminol (1.34) and sesamol (1.33) (Scheme 1.3).



Scheme 1.3 Semi-synthesis of sesaminol (1.34) and alkyloxysamins (1.35) from sesamol (1.2)

From literature reviews, semi-synthesis seem to be a suitable approach to furnish diverse furofuran lignans. In this work, we applied above methodology using samin (1.36) as starter instead of 1.2 due to the fact that hemiacetal moiety in 1.36 is relatively more reactive than acetal group of 1.2 toward nucleophilic substitution.

Moreover, the released of H₂O generated upon protonation of hemiacetal could be trapped by molecular sieve readily in order to avoid regeneration of samin (**1.36**). With samin (**1.36**) in hand, we examined the wide application of this reaction with three types of nucleophiles, phenolics (C), thiols (S) and alcohols (O) as show in Scheme 1.4.



Scheme 1.4 Synthetic strategy to furofuran lignans **3.1-3.3**.

In addition, there is no research studying about mechanistic reaction of samin under acid condition; and the stereoselective outcome of previous report [9] still unclear, therefore the mechanism understanding this reaction was carefully investigated by a model reaction monitored by ¹H NMR and computational calculations. For computational study, the geometrical optimization and determination of product energy have been calculated using the density functional theory (DFT) with the popular hybrid method (B3LYP). The split valence with diffuse and polarization functions, 6-31+G(d,p), basis set is used in this study to propose the reaction mechanisms through minimum-energy geometry. All of these calculations will be performed by Gaussian 09 program [10]

CHAPTER II

SYNTHESIS OF SAMIN

2.1 Isolation of sesamol

The naturally available sesamol (**1.2**) utilized in this work was isolated from sesame seed oil which contains large amount of lignan together with sesamol 0.5-2 % w/w. According to existing of fatty acids in sesame seed oil, the purification by column chromatography directly is challenged. Therefore, the elimination of glyceride was performed in initial step. This approach is based on the fact that fatty acids can be saponified by KOH/MeOH, to give the corresponding potassium salts whereas **1.2** and other unsaponified materials remain unchanged. The slightly excess of KOH (~1.5 times of saponification number) was applied in order to ensure complete removal of triglyceride. After refluxed 3-5 h or completed saponification, MeOH was removed followed by liquid-liquid extraction (H₂O and EtOAc). The organic layer containing **1.2** was further purified using silica gel column chromatography, to yield sesamol (1 %w/w) as a white solid and its identities (¹H and ¹³C NMR) were found to be identical with those reported in the literature [11]. This isolation methodology is summarized in Figure 2.1.

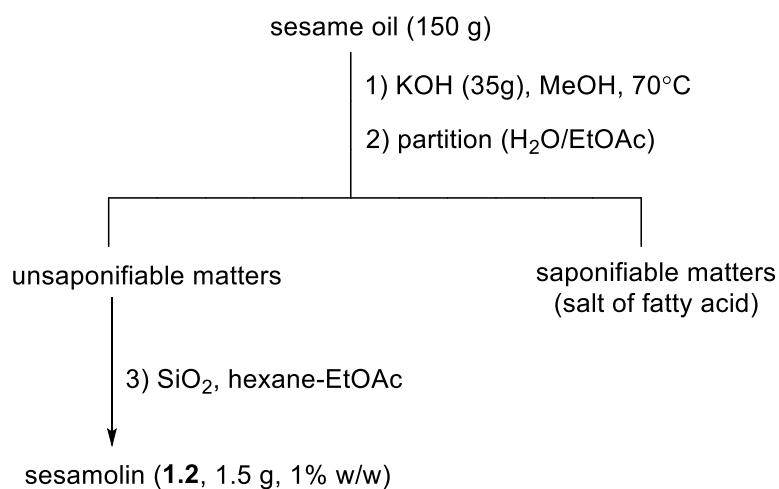
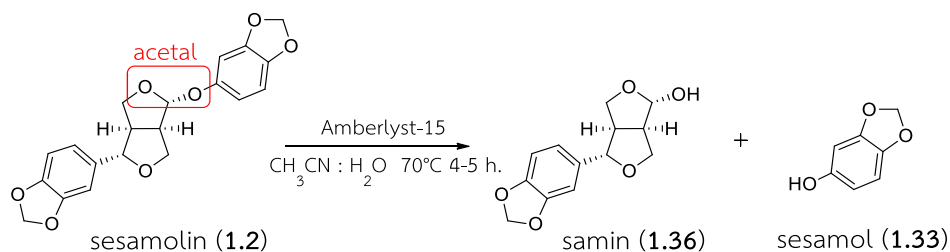


Figure 2.1 Isolation of sesamol (1.2) from sesame oil.

2.2 Synthesis of samin

According to predominant potentiality of hemiacetal, samin (1.36) was used as a starting material. The synthesis of samin was operated with the methodology described by Reshma [12] with nominal modification. Briefly, Amberlyst-15 was used for acid-catalyzed hydrolysis of sesamol (1.2) in 9:1 acetonitrile-water at 70 °C for 4-5 h. Sesamol (1.2) was protonated on an acetal group resulting in the loss of sesamol and formation of expected samin (1.36) as shown in Scheme 2.1. The identification of 1.36 was investigated after purification by SiO₂ column chromatography eluting with ethyl acetate and hexane.



Scheme 2.1 Hydrolysis of sesamol (1.2).

2.3 Experimental section

2.3.1 General experiment procedures

^1H and ^{13}C NMR were recorded (CDCl_3 as a solvent) at 400 and 100 MHz, respectively, Varian Mercury⁺ 400 NMR and a Bruker (Avance) 400 NMR spectrometer. The chemical shifts were reported in ppm downfield from TMS. Mass spectra were measured by HRESIMS obtaining from a micrOTOF Bruker mass spectrometer. Thin layer chromatography (TLC) was performed on pre-coated Merck silica gel 60 F₂₅₄ plates (0.25 mm thick layer) and visualized under 254 nm UV. Silica gel 60 Merk cat. No. 7729 was used for open column chromatography.

2.3.2 Chemical

Sesame seed oils was purchased from Soun-Pana (Bangkok Thailand). All reagents were obtained from Sigma-Aldrich and used without further purification.

2.3.3 Determining of saponification number

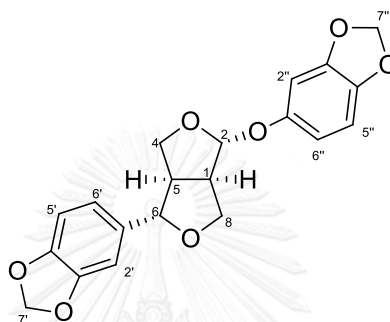
Saponification number was measured in triplicate by general procedure [13]. Briefly, a solution of 1 g sesame seed oil in 10 mL of 0.5M KOH/MeOH was stirred under 70°C for 3 h. After cooling, reaction was titrated against a standard solution of 0.5M HCl. The difference between test reaction and blank gave the volume of 0.5M HCl equivalence of KOH used in saponification 1 g sesame seed oil. Calculation of saponification number by below equation presented that sesame seed oil has S.N. around 156.6 mgKOH/oil.

$$\text{S.N. (mg KOH/oil)} = \frac{(V_{\text{HCl, blank}} - V_{\text{HCl test}}) \times [\text{HCl}] \times 56}{W_{\text{oil}}}$$

Note : 56 is the equivalent mass of KOH. W is exact

2.3.4 Isolation of sesamol

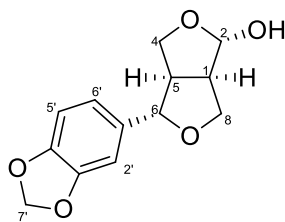
Sesame seed oil (150 g) dissolved in MeOH (150 mL) was saponified with KOH (34 g) for 5 h. After solvent removal, the resulting mixture was extracted with ethyl acetate and water. The organic layer was further purified by silica gel column chromatography using 1:9 ethylacetate/hexane to obtain sesamol (1.2) (1.5 g, 1 %w/w).



Sesamol (1.2) : as white solid; ¹H NMR (400 MHz, CDCl₃) δ 6.88 (s, 1H, H-2''), 6.84 – 6.77 (m, 2H, H-5'' and H-6''), 6.71 (d, J = 8.5 Hz, 1H, H-5'), 6.62 (d, J = 2.3 Hz, 1H, H-2'), 6.50 (dd, J = 8.4, 2.4 Hz, 1H, H-6'), 5.96 (s, 2H, H-7'), 5.92 (s, 2H, H-7''), 5.50 (s, 1H, H-2), 4.47 – 4.39 (m, 2H, H-6 and H-8), 4.13 (dd, J = 9.2, 6.1 Hz, 1H, H-4), 3.96 (d, J = 9.2 Hz, 1H, H-4), 3.64 (dd, J = 9.0, 7.6 Hz, 1H, H-8), 3.31 (dd, J = 16.7, 8.7 Hz, 1H, H-1), 2.95 (dd, J = 15.4, 6.6 Hz, 1H, H-5); ¹³C NMR (CDCl₃, 100 MHz) δ 151.8 (C-1''), 119.6 (C-6''), 108.9 (C-6'), 108.1 (C-5''), 108.0 (C-5'), 106.8 (C-2), 106.5 (C-2''), 101.2 (C-7''), 101.0 (C-7'), 100.1 (C-2'), 87.0 (C-6), 71.2 (C-8), 69.7 (C-4), 53.2 (C-1), 52.7 (C-5).

2.3.4 Synthesis of samin

To a solution of sesamol (1.2) (100 g, 0.39 mmol) in a mixture of acetonitrile/H₂O (9:1, 10 mL) was treated with acidic resin Amberlyst-15 (1 mg/0.005 mmol of sesamol (1.2)). After stirring at 70°C for 4-8 h, the reaction mixture was evaporated to dryness and purified by silica gel chromatography using 1:1 ethylacetate/hexane to give samin (1.36, 85%).



samin (**1.36**) : as brown crystal; ^1H NMR (CDCl_3 , 400 MHz) δ 6.85 (s, 1H, H-2'), 6.80-6.75 (m, 2H, H-5' and H-6'), 5.94 (s, 2H, OCH_2O), 5.36 (s, 1H, H-2), 4.35 (d, $J = 8.4$ Hz, 2H, H-6 and H-8), 4.16 (dd, $J = 9.2, 6.0$ Hz, 1H, H-4), 3.89 (d, $J = 9.2$ Hz, 1H, H-4), 3.56 (dd, $J = 8.8, 7.2$ Hz, 1H, H-8), 3.25 (brs, 1H, -OH), 3.05 (m, 1H, H-1), 2.86 (m, 1H, H-5); ^{13}C NMR (CDCl_3 , 100 MHz) δ 148.1, 147.4, 134.7, 119.7, 108.3, 106.7, 102.4, 101.2, 87.0, 71.4, 69.5, 53.7, 52.9.



CHAPTER III

SYNTHESIS OF NEW FUROFURAN LIGNANS FROM SAMIN

3.1 General procedure for the synthesis of new furofuran lignans

The synthesis of furofuran lignans from small starting materials through multi-step strategy (total-synthesis) does not serve the need of SAR study. Due to the less variety of substrates, few of furofurans were synthesized. Consequently, a semi-synthesis seems to be efficient over total-synthesis in generating variety of new furofuran lignans.

In this work, we exhibited short and efficient synthetic strategy to synthesize diversity of new furofuran lignans using samin (**1.36**) as a lead compound. Under acidic condition, samin was protonated to produce oxocarbenium ion along with release of water which could be trapped by 4^oA MS. Selected nucleophiles subsequently attacked on oxocarbenium ion yielding new furofuran lignans. Three types of nucleophile, phenolics (C), thiols (S) and alcohols (O), were used to synthesize these derivatives.

3.2 Synthesis of new furofuran lignans using phenolics as a nucleophile

Phenolics used in this experiment were classified into seven groups (**a-g**, Figure 3.1) which have the differentiation of number and orientation of electron donating groups on aromatic ring. Disappointingly, treatment of samin with anisole (**a**) failed to provide the desired furofuran lignan. The presence of one electron donating group such as OCH₃ is not enough to enhance the nucleophilicity of aromatic toward the reaction with samin. Therefore, disubstituted oxygenated benzenes (**b-d**) were introduced as stronger nucleophiles. Surprisingly, only the reactions of samin and 1,3-dioxygenated benzenes (**d** and **e**) afforded the target products **3.1d** and **3.1e** as

well as their epimers, respectively. This observation would be explained by the enhanced nucleophilicity reinforced by two electron donating groups at *meta* position. To test our hypothesis, the reaction of samin and *m*-cresol (**b**) was carried out. The products **3.1b** and *epi*-**3.1b** were obtained in moderate yields (45%). With the success of the reactions between samin and 1,3-dioxygenated benzenes, we further explored the reactions between samin and trioxygenated benzenes (**e-g**). The desired products were obtained in good yields (50 – 90%). Noticeably, all phenolics (**g-j**) used in the above reactions processed enhanced nucleophilicity caused by electron donating groups in a *meta*-relation to each other.

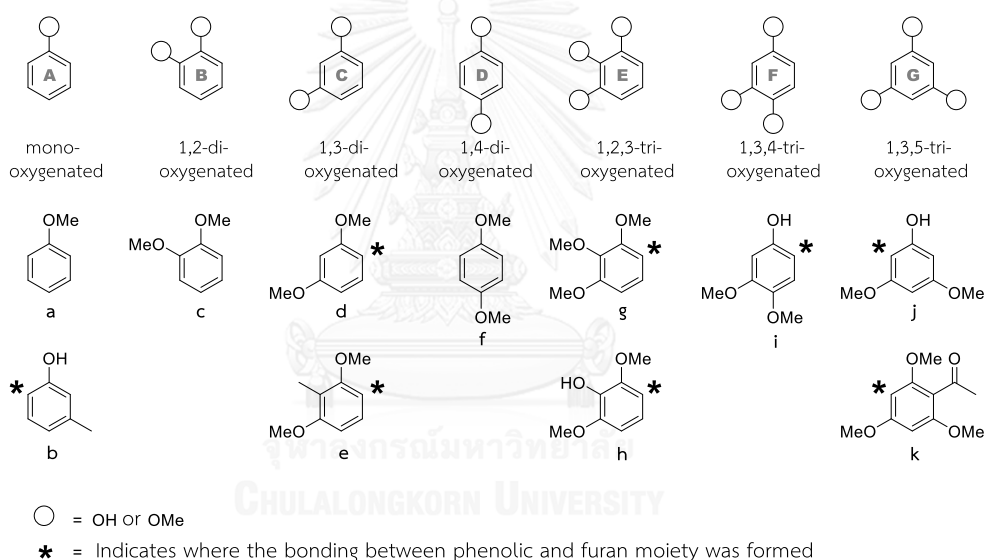
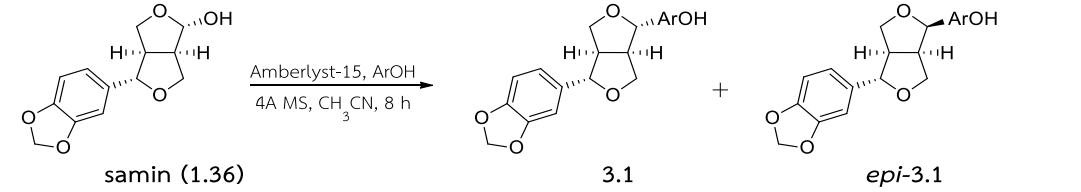


Figure 3.1. Phenolics used in synthesis strategy.

However, the presence of one electron withdrawing group such as acetyl group (COCH_3) decreased nucleophilicity of trioxygenated benzene **k**, resulting in lower yield (15%) of product **3.1k**. All of results are summarized in Table 3.1

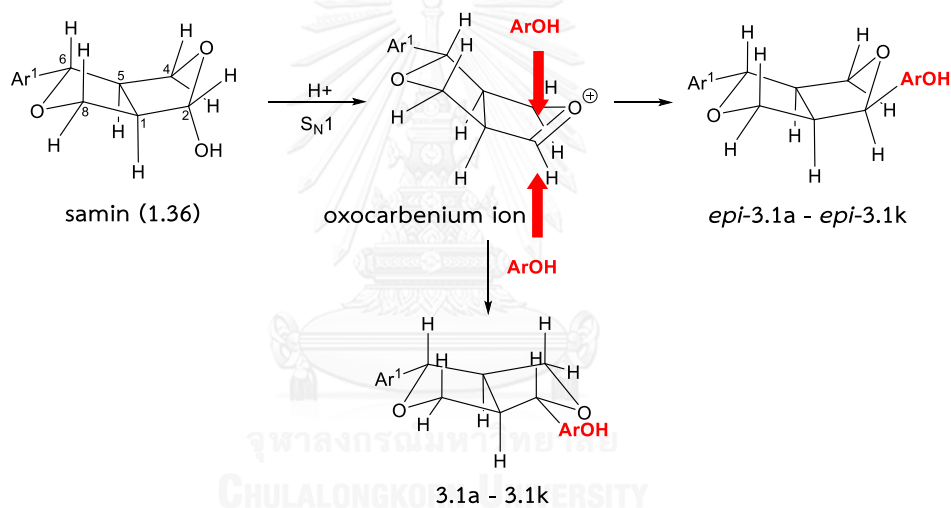
Table 3.1 Synthesis of furofuran lignans from samin (**1.36**) with phenolics (**a - k**)



Entry	ArOH	Isolated yield (%)	
		3.1	<i>epi</i> -3.1
1	a		NR
2	b	3.1b (30 %)	<i>epi</i> -3.1b (15 %)
3	c		NR
4	d	3.1d (55 %)	<i>epi</i> -3.1d (45 %)
5	e	3.1e (42 %)	<i>epi</i> -3.1e (31 %)
6	f		NR
7	g	3.1g (31 %)	<i>epi</i> -3.1g (22 %)
8	h	3.1h (27 %)	<i>epi</i> -3.1h (37 %)
9	i	3.1i (68 %)	<i>epi</i> -3.1i (17 %)
10	j	3.1j (47 %)	<i>epi</i> -3.1j (51 %)
11	k	3.1k (15 %)	<i>epi</i> -3.1k (trace)

NR = no reaction

Generally, the reactions between samin and phenolics would proceed through S_N1 mechanism involving oxocarbenium ion. The hydroxyl group at anomeric center (C-2) was initially protonated by strong acid to produce oxocarbenium ion (Scheme 3.1). Phenolics as carbon nucleophiles attacked on both sides of the planar intermediate to provide diastereomeric products in equal ratio. In terms of regioselectivity, the bonding between C-2 of samin and high electron density as well as least hindrance of phenolics by *ortho* group were generally observed. The structural identification and relative stereochemistries of other products were confirmed by 2D experiments and ^1H NMR patterns as predicated in next section.



Scheme 3.1 Proposed mechanistic formation of **3.1a – 3.1k** and ***epi*-3.1a – *epi*-3.1k**

3.2.1 Structural characterization of synthesized furofuran lignans (**3.1** and *epi*-**3.1**)

Structures of all synthesized compounds were characterized mainly by ^1H and ^{13}C NMR, along with 2D NMR for particular compounds. The ^1H and ^{13}C NMR spectra of **3.1** and *epi*-**3.1** showed closely related pattern, except for some signals from H atom of core structure. For instance, H-2 and H-6 in **3.1b** showed doublet signal with comparable coupling constants ($J \approx 4.0$ Hz) whereas those in *epi*-**3.1b** displayed

significantly distinct values; 5.6 Hz for H-2 and 8.0 Hz for H-6. A more strikingly significant observation is the splitting patterns of diastereomeric H-4, which appeared as expected as doublet of doublet. However, only H-4_{eq} of **epi-3.1b** showed an exceptional doublet signal caused solely by germinal coupling of H-4_{eq} and H-4_{ax}. These observations would account for a nearly 90° dihedral angle between H-4_{eq} and H-5 that gave rise to $J_{\text{H-4}_{\text{eq}},5} \approx 0$ Hz [14] (Figure. 3.2). The above evidence could be useful in readily distinguishing the identity of other synthesized lignans **3.1** and their epimers.

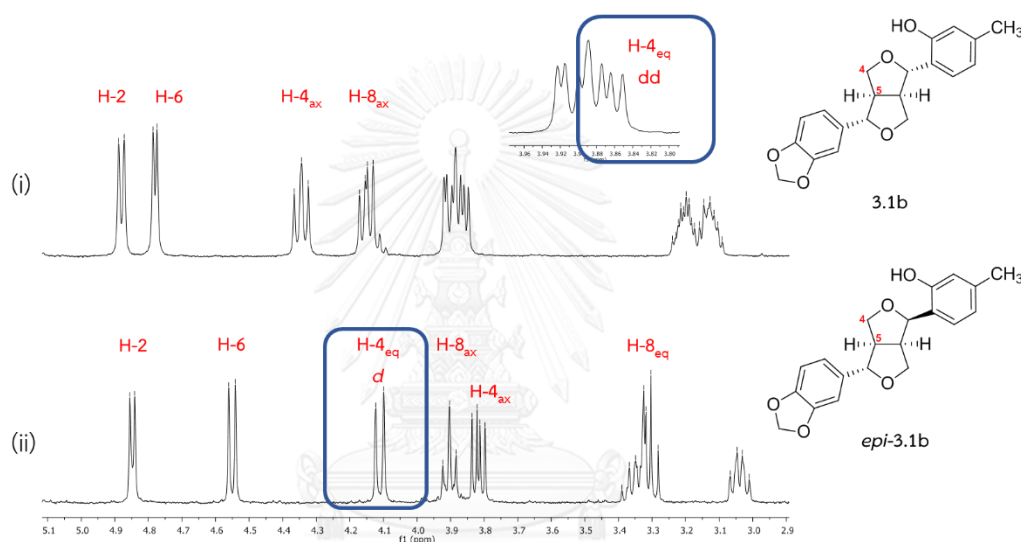


Figure 3.2. ¹H NMR spectra of (i) **3.1b** and (ii) **epi-3.1b**.

The relative configuration of **3.1b** and **epi-3.1b** at C-2 was further determined by NOESY data and coupling constant analysis. The NOESY spectrum of **3.1b** showed diagnostic correlations of H-2/H-8_{ax} and H-4_{ax}/H-6, suggesting that the occupation of two aryls are on the *exo-exo* face of the bicyclic core (Figure 3.2 (i)). This interpretation was corresponding with a chair-chair conformation of sesamin (**1.3**), which was verified by X-ray analysis [15]. On the other hand, **epi-3.1b** showed key NOESY correlations of H-2/H-4_{ax} and H-6/H-8_{ax}, which implied *endo,exo*-2,6-diarylfurofuran (Figure 3.2 (ii)). These observations correlated well with the chair-boat conformation of *epi*-sesamin or asarinin [15].

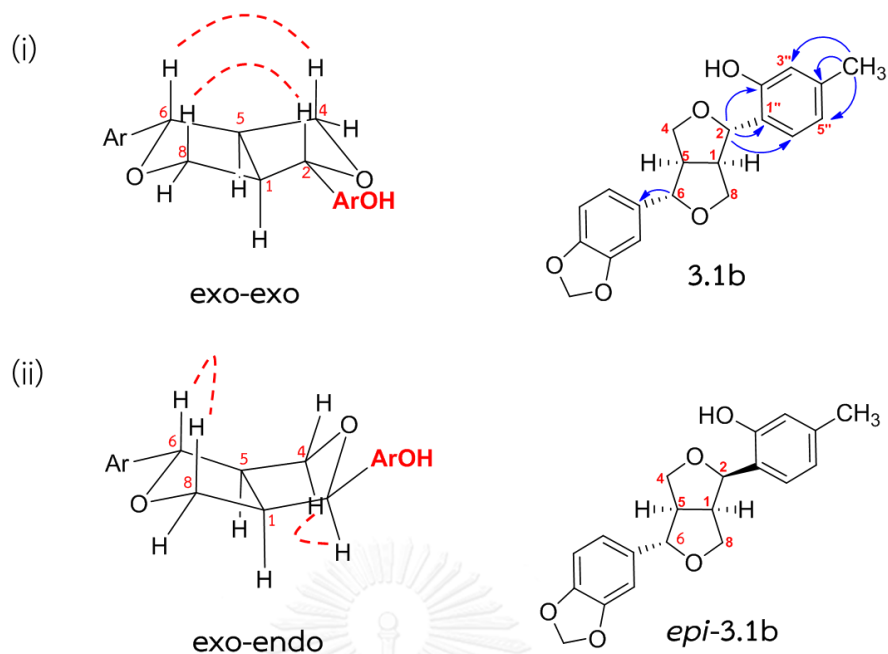


Figure 3.3 Diagnostic NOESY and HMBC correlations of (i) **3.1b** and (ii) **epi-3.1b**.

3.3 Synthesis of new furofuran lignans using thiols as a nucleophile

With the success in applying phenolic compounds, we further synthesized other furofuran lignans by using thiols as sulfur nucleophiles. Thiols used in this investigation are listed in Figure 3.4. They are divided into two categories; aliphatic thiols (sp^3 C-SH, **l-p**) and aromatic thiols (sp^2 C-SH, **q-z**).

A S atom is well recognized as a stronger nucleophile than C and O atoms. Therefore, the products from reaction of samin and thiols would be obtained in high to excellent yield (Table 3.2).

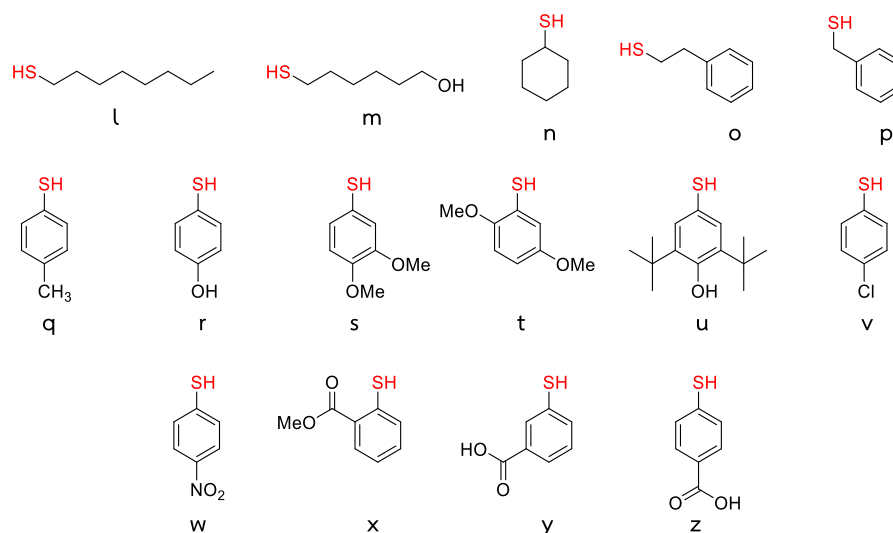
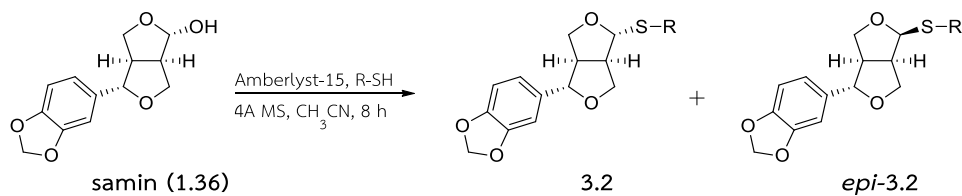


Figure 3.4 Thiols used in synthesis strategy.

Under the same reaction condition, diastereomeric furofuran lignans (**3.2** and *epi-3.2*) were produced after sulfur nucleophiles attacked on an oxocarbenium intermediate. The diastereomeric ratios (dr) of products synthesized from samini and aliphatic thiols (**l-p**) are in range of 5:1 to 7:1. On the other hand, the dr values of products synthesized from samini and aromatic thiols (**q-z**) varied in broader range (2:1 to 12:1). The highly diastereomeric ratios of **3.2t** : *epi-3.2t* and **3.2x** : *epi-3.2x* were possibly contributed by steric hindrance of *ortho*-substituent (**t** and **x**). These observations were supported by the lower dr values of isomeric products. For example, treatment of samini with **y** and **z**, the *meta* and *para* isomer of **x**, provided products with lower diastereomeric ratios (6:1).

Table 3.2 Synthesis of furofuran lignans from samín (1.36) with thiols (l - z)

Entry	RSH	Isolated yield (%) ^a	Diastereomeric ratio ^b
			3.2 : epi-3.2
1	l	quant	7 : 1
2	m	65 %	5 : 1
3	n	79 %	Not determine
4	o	85.2 %	6 : 1
5	p	73 %	7 : 1
6	q	97 %	3 : 1
7	r	99 %	5 : 1
8	s	69 %	3 : 1
9	t	96 %	12 : 1
10	u	69 %	2 : 1
11	v	Quant	5 : 1
12	w	73.3 %	6 : 1
13	x	67 %	12 : 1
14	y	65 %	6 : 1
15	z	77 %	6 : 1

^a Isolated yield of diastereomeric mixture, ^b Ratios were determined by ¹H NMR analysis

Note: According to small amount of epimers cannot be separated purely by general technique, only pure spectra of their congeners were shown.

3.3.1 Structural characterization of synthesized furofuran lignans (3.2 and *epi*-3.2)

^1H NMR spectra of synthesized furofuran lignans **3.2** and *epi*-**3.2** have slightly different pattern with those obtained from samin and phenolics (**3.1a-3.1k**). However, careful inspection of ^1H NMR spectra indicated striking difference in splitting pattern of H-2. As shown in Figure 3.6, furofuran lignans **3.2** showed a singlet signal of H-2 whereas *epi*-**3.2** demonstrated doublet signal with $J \approx 6\text{-}7$ Hz. The unexpected singlet H-2 of **3.2** suggested a perpendicular dihedral angle ($\phi \approx 90^\circ$) between H-2 and H-1 (Figure 3.6 (iii)), indicating that **3.2** adopted *exo-exo* furofuran core structure the same as samin [16].

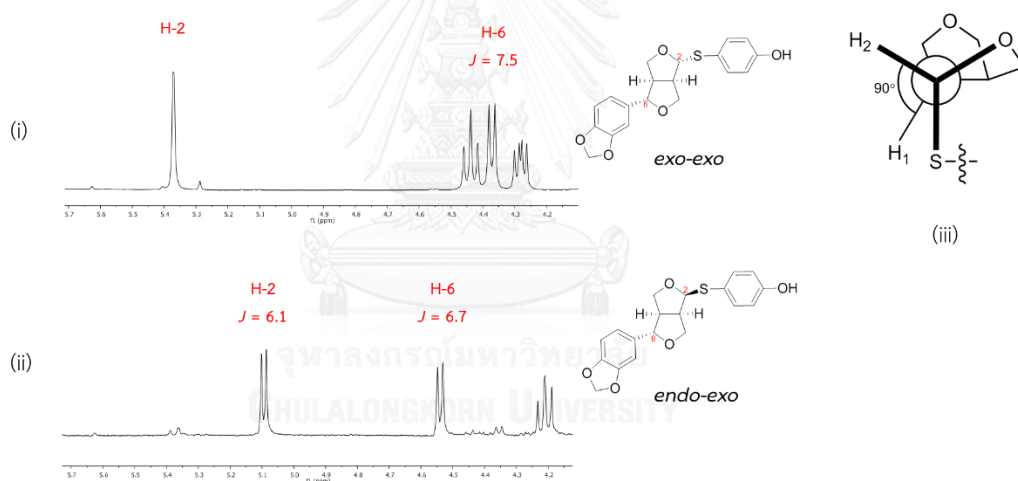


Figure 3.5 ^1H NMR spectra of (i) **3.2r** and (ii) *epi*-**3.2r**. Note that (iii) Newman projection of **3.2r** demonstrates dihedral angle nearly 90° between H-1 and H-2, thus resulting in singlet (s) signal rather than expected doublet (d).

Since furofuran lignans containing a S atom have never been reported, their structures were fully characterized by 2D NMR and X-ray analysis. All ^1H and ^{13}C signals of the representative lignans **3.2r** and *epi*-**3.2r** were first confirmed by HSQC and HMBC data, and selected HMBC correlations are shown in Figure 3.6.

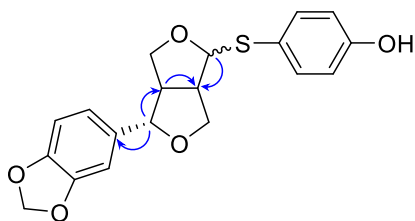
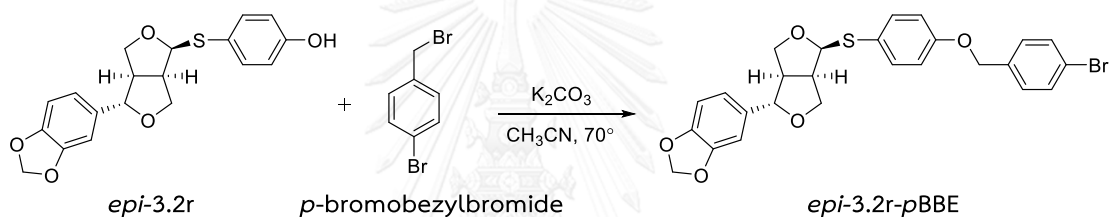


Figure 3.6 Selected HMBC correlations of **3.2r** and *epi*-**3.2r**.

To unambiguously elucidate furofuran core structure and orientation of the thiol moiety, X-ray crystallographic analysis was performed. Because a good single crystal of *epi*-**3.2r** could not be obtained, its *p*-bromobenzylether (*p*BBE) derivative (*epi*-**3.2r-pBBE**) was prepared (Scheme 3.2).



Scheme 3.2 Preparation of *epi*-**3.2r-pBBE**.

Crystal structures of the two isomers are shown in Figure 3.7. Thiol substituted at C-2 of **3.2r** was located at the *exo* position, and dihedral angle between H-1 and H-2 was approximately 96.14° , resulting in singlet signal of H-2 ($J = 0$ Hz). On the other hand, the thiol substituent at C-2 of *epi*-**3.2r-pBBE** was located at *endo* position, and dihedral angle between H-1 and H-2 was approximately 28.01° ($J = 6.1$ Hz).

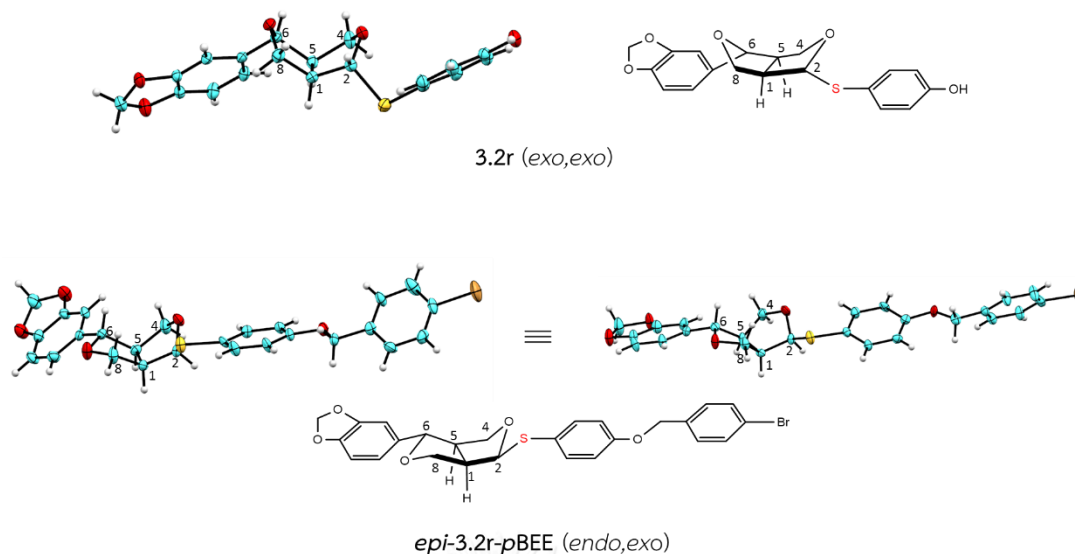


Figure 3.7 ORTEP plots (20% probability level) of **3.2r** and **epi-3.2r-pBEE** (color codes: C = cyan, S = yellow, Br = orange, O = red, H= white).

3.4 Synthesis of new furofuran lignans using alcohols as a nucleophile

A wide range of alcohols were also applied to demonstrate a usability of our designed synthetic method. Selected alcohols are varied in less steric alcohols (**A-F**), high steric alcohols (**G-J**) and allylic alcohols (**K-N**) (Figure 3.8).

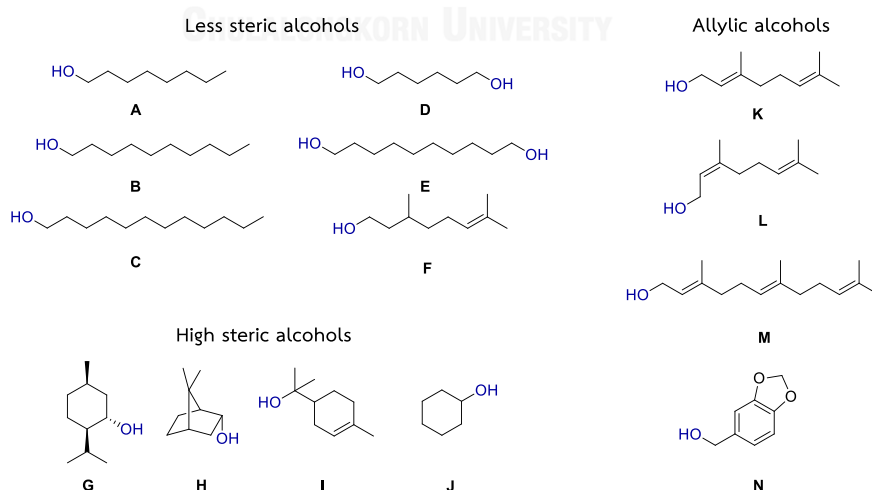


Figure 3.8 Alcohols used in synthesis strategy.

From this experiment, fourteen new furofuran lignans (**3.3A-3.3N**) were able to synthesize in high yield (69 % – quant, Table 3.3) although bulky or naturally allylic alcohols were used as nucleophiles. These results indicated that our methodology is compatible to all kinds of alcohols despite of alcohols **N**, which was susceptible to dehydration under strong acidic condition. Therefore, reaction between samini and **N** provided **3.3N** in moderated yield together with the byproduct generated from self-condensation of benzylic alcohol **N**.

Unexpectedly, each reaction provided single stereomeric products (**3.3A – 3.3N**) instead of mixture of two diastereomers as previously described in the reactions between samini and phenolics as well as thiols. The stereochemistry of **3.3** proved to be retention of configuration, which were elaborated in section 3.4.1.

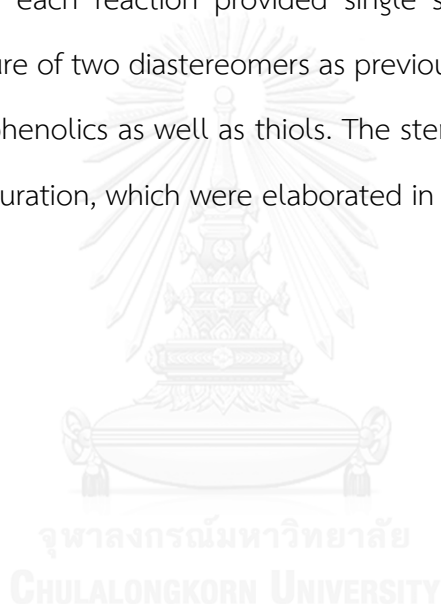
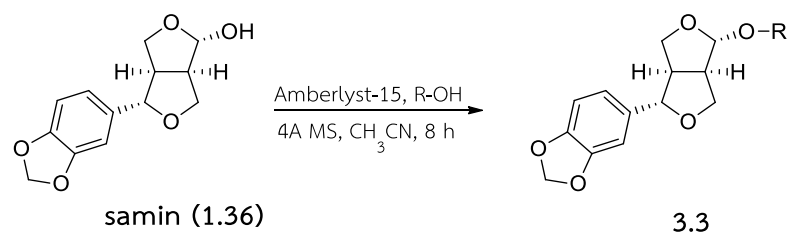


Table 3.3 Synthesis of furofuran lignans from samín with alcohols (A - N)

Entry	ROH	Isolated yield (%)
1	A	3.3A (78 %)
2	B	3.3B (quant)
3	C	3.3C (quant)
4	D	3.3D (quant)
5	E	3.3E (quant)
6	F	3.3F (88 %)
7	G	3.3G (quant)
8	H	3.3H (75 %)
9	I	3.3I (81 %)
10	J	3.3J (93 %)
11	K	3.3K (69 %)
12	L	3.3L (82 %)
13	M	3.3M (84 %)
14	N	3.3N (44.8 %)

3.4.1 Structural characterization of synthesized furofuran lignans (3.3 and *epi*-3.3)

Compounds **3.3A-3.3N** showed singlet signals of H-2 similar to those of **3.2**. For instant, **3.3A** and **3.2L** exhibited sharp singlet signal around 4-5.5 ppm. These observations indicated that their conformation adopted *exo-exo* furofuran type.

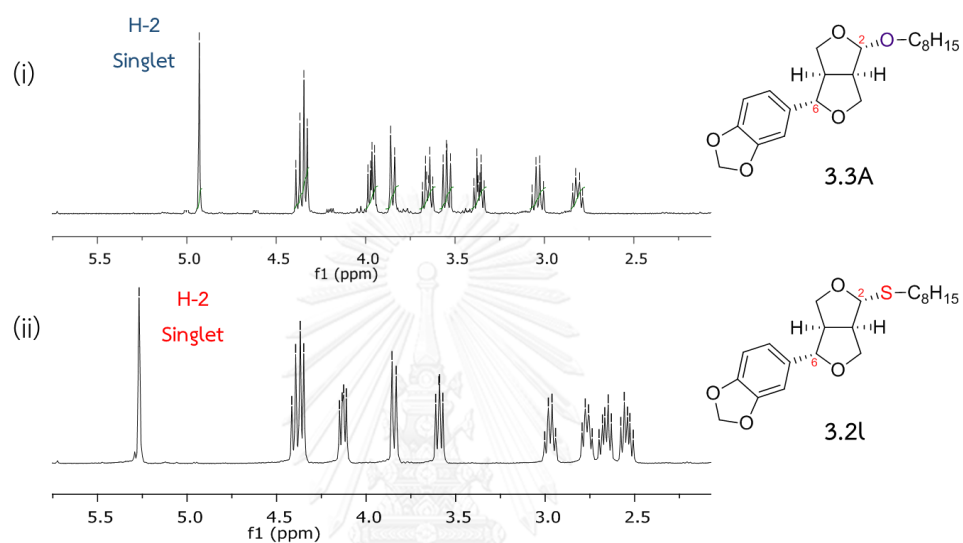


Figure 3.9 ^1H NMR spectra of (i) **3.3A** and (ii) **3.2L**

3.5 Experimental section

3.5.1 General experiment procedures

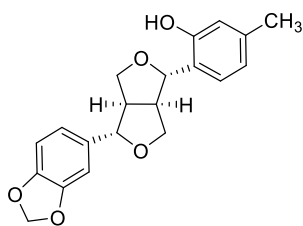
^1H and ^{13}C NMR were recorded (CDCl_3 as a solvent) at 400 and 100 MHz, respectively, Varian Mercury⁺ 400 NMR and a Bruker (Avance) 400 NMR spectrometer. The chemical shifts were reported in ppm downfield from TMS. Mass spectra were measured by HRESIMS obtaining from a micrOTOF Bruker mass spectrometer. Thin layer chromatography (TLC) and Preparative thin Layer Chromatography (Prep TLC) were operated on pre-coated Merck silica gel 60 F_{254} plates (0.25 and 0.50 mm thick layer, respectively) and visualized under 254 nm UV. Silica gel 60 Merk cat. No. 7729 was applied for open column chromatography.

3.5.2 Chemical

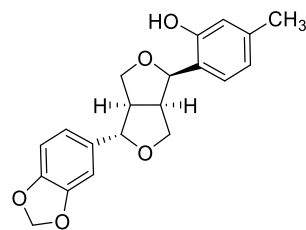
All reagents were obtained from Sigma-Aldrich and used without further purification.

3.5.3 General procedure for synthesis of new furofuran lignans

To a solution of samin (1 eq) in acetonitrile (1.0 mL/0.1 mmol of samin) was treated with selected nucleophile (2 eq), acidic resin amberlyst-15 (1 mg/0.005 mmol of samin) and molecular sieve, 4 Å. After stirring at 60°C for 8 h, the reaction mixture was evaporated to dryness and load onto silica gel column chromatography followed by Prep TLC in order to obtain purely desired products.



(1*R*, 2*R*, 5*S*, 6*S*)-2-(2-hydroxy-4-methylphenyl)-6-(3,4-methylenedioxyphenyl)-3,7-dioxabicyclo[3.3.0]octane (**3.1b**)

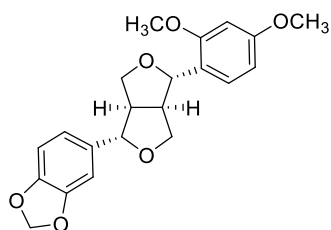


(1*R*, 2*S*, 5*S*, 6*S*)-2-(2-hydroxy-4-methylphenyl)-6-(3,4-methylenedioxyphenyl)-3,7-dioxabicyclo[3.3.0]octane (*epi*-**3.1b**)

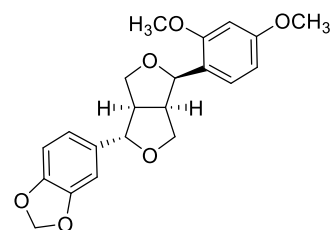
Following the general procedure, reaction of **1.36** (64.5 mg, 0.26 mmol), *m*-cresol (40 μ L, 0.39 mmol) in acetonitrile (2 mL) after 8 h yielded **3.1b** (27 mg, 30%) and *epi*-**3.1b** (13 mg, 15%) as white powder.

3.1b: $^1\text{H NMR}$ (CDCl_3 , 400 MHz) δ 7.89 (brs, 1H, -OH), 6.92 (d, $J = 7.6$ Hz, 1H, H-6''), 6.83-6.78 (m, 3H, H-2', H-5', and H-6'), 6.71 (s, 1H, H-3''), 6.67 (d, $J = 7.6$ Hz, 1H, H-5''), 5.95 (s, 2H, H-7'), 4.87 (d, $J = 6.5$ Hz, 1H, H-2), 4.78 (d, $J = 4.4$ Hz, 1H, H-6), 4.34 (dd, $J = 9.2, 7.6$ Hz, 1H, H-4), 4.15 (dd, $J = 9.2, 6.8$ Hz, 1H, H-8), 3.92-3.85 (m, 2H, H-4 and H-8), 3.21 (m, 1H, H-1), 3.14 (m, 1H, H-5), 2.29 (s, 3H, -CH₃); $^{13}\text{C NMR}$ (CDCl_3 , 400 MHz) δ 155.5, 148.2, 147.4, 139.8, 134.8, 126.8, 120.9, 120.9, 119.5, 117.9, 108.4, 106.7, 101.3, 86.7, 85.6, 72.5, 70.9, 53.6, 53.1, 21.2; HRMS m/z 363.1212 [$\text{M}+\text{Na}$]⁺ (calcd for C₂₀H₂₀NaO₅, 363.1208).

epi-**3.1b**: $^1\text{H NMR}$ (CDCl_3 , 400 MHz) δ 7.85 (brs, 1H, -OH), 6.92 (d, $J = 7.6$ Hz, 1H, H-6''), 6.86-6.80 (m, 3H, H-2', H-5', and H-6'), 6.71 (s, 1H, H-3''), 6.67 (d, $J = 7.6$ Hz, 1H, H-5''), 5.97 (s, 2H, H-7'), 4.85 (d, $J = 5.6$ Hz, 1H, H-6), 4.55 (d, $J = 7.9$ Hz, 1H, H-2), 4.11 (d, $J = 9.6$ Hz, 1H, H-8), 3.90 (dd, $J = 8.4, 7.6$ Hz, 1H, H-4), 3.82 (dd, $J = 9.6, 6.0$ Hz, 1H, H-8), 3.38-3.28 (m, 2H, H-1 and H-4), 3.04 (m, 1H, H-5), 2.29 (s, 3H, -CH₃); $^{13}\text{C NMR}$ (CDCl_3 , 400 MHz) δ 155.5, 147.9, 146.9, 139.8, 132.0, 126.9, 121.2, 120.8, 118.8, 118.0, 108.4, 106.5, 101.2, 88.6, 82.0, 70.7, 70.2, 53.4, 49.9, 21.3; HRMS m/z 363.1213 [$\text{M}+\text{Na}$]⁺ (calcd for C₂₀H₂₀NaO₅, 363.1208).



(1*R*, 2*R*, 5*S*, 6*S*)-2-(1,4-dimethoxyphenyl)-6-(3,4-methylenedioxyphenyl)-3,7-dioxabicyclo[3.3.0]octane (**3.1d**)

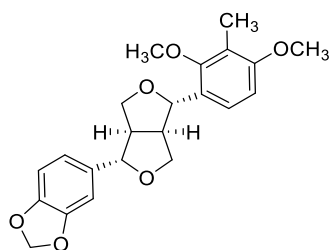


(1*R*, 2*S*, 5*S*, 6*S*)-2-(1,4-dimethoxyphenyl)-6-(3,4-methylenedioxyphenyl)-3,7-dioxabicyclo[3.3.0]octane (*epi*-**3.1d**)

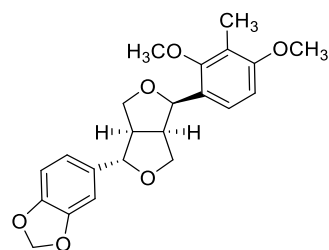
Following the general procedure, reaction of **1.36** (12.5 mg, 0.05 mmol), 1,3-dimethoxybenzene (12 mg, 0.075 mmol) in acetonitrile (0.5 mL) after 8 h yielded **3.1d** (10 mg, 55%) and *epi*-**3.1d** (8.2 mg, 45%) as colorless oil.

3.1d: $^1\text{H NMR}$ (CDCl_3 , 400 MHz) δ 7.25 (dd, $J = 8.0, 2.8$ Hz, 1H), 6.84 (s, 1H), 6.81-6.74 (m, 2H), 6.45 (d, $J = 7.2$ Hz, 1H), 6.44 (d, $J = 2.4$ Hz, 1H), 5.92 (s, 2H), 5.03 (d, $J = 4.8$ Hz, 1H), 4.64 (d, $J = 5.6$ Hz, 1H), 4.30 (dd, $J = 9.2, 7.6$ Hz, 1H), 4.19 (dd, $J = 8.8, 6.4$ Hz, 1H), 3.98 (dd, $J = 9.2, 5.2$ Hz, 1H), 3.90 (dd, $J = 9.2, 4.0$ Hz, 1H), 3.80 (s, 3H, $-\text{OCH}_3$), 3.79 (s, 3H, $-\text{OCH}_3$), 3.01 (m, 1H), 2.91 (m, 1H); $^{13}\text{C NMR}$ (CDCl_3 , 400 MHz) δ 160.2, 157.4, 147.9, 147.1, 135.4, 126.1, 122.8, 119.5, 108.1, 106.6, 103.8, 101.0, 98.6, 85.5, 82.0, 73.3, 71.2, 55.4, 55.3, 54.7, 53.7; HRMS m/z 393.1310 [$\text{M}+\text{Na}$] $^+$ (calcd for $\text{C}_{21}\text{H}_{22}\text{NaO}_6$, 393.1314).

epi-**3.1d**: $^1\text{H NMR}$ (CDCl_3 , 400 MHz) δ 7.44 (d, $J = 8.4$ Hz, 1H), 6.87 (s, 1H), 6.81-6.76 (m, 2H), 6.50 (dd, $J = 8.4, 2.4$ Hz, 1H), 6.44 (s, 1H), 5.94 (s, 2H), 4.91 (d, $J = 6.0$ Hz, 1H, H-6), 4.36 (d, $J = 8.0$ Hz, 1H, H-2), 4.09 (d, $J = 9.2$ Hz, 1H), 3.81 (s, 3H, $-\text{OCH}_3$), 3.80 (s, 3H, $-\text{OCH}_3$), 3.78-3.74 (m, 2H), 3.47 (m, 1H), 3.22 (dd, $J = 8.8, 8.8$ Hz, 1H), 2.84 (m, 1H); $^{13}\text{C NMR}$ (CDCl_3 , 400 MHz) δ 160.2, 156.6, 148.1, 147.3, 135.6, 127.3, 127.3, 119.7, 108.3, 106.8, 103.9, 101.1, 98.3, 87.6, 78.6, 70.5, 69.9, 55.5, 55.4, 54.9, 48.7; HRMS m/z 393.1310 [$\text{M}+\text{Na}$] $^+$ (calcd for $\text{C}_{21}\text{H}_{22}\text{NaO}_6$, 393.1314).



(1*R*, 2*R*, 5*S*, 6*S*)-2-(2,4-dimethoxy-3-methylphenyl)-6-(3,4-methylenedioxyphenyl)-3,7-dioxabicyclo[3.3.0]octane (**3.1e**)

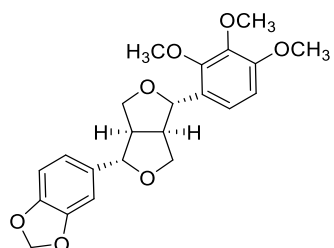


(1*R*, 2*S*, 5*S*, 6*S*)-2-(2,4-dimethoxy-3-methylphenyl)-6-(3,4-methylenedioxyphenyl)-3,7-dioxabicyclo[3.3.0]octane (*epi*-**3.1e**)

Following the general procedure, reaction of **1.36** (63.0 mg, 0.25 mmol), 2,6-dimethoxytoluene (74 μ L, 0.5 mmol) in acetonitrile (2 mL) after 8 h yielded **3.1e** (40 mg, 42%) and *epi*-**3.1e** (30 mg, 31%) as yellow oil.

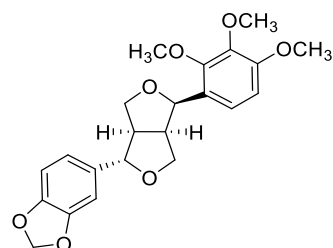
3.1e: ^1H NMR (CDCl_3 , 400 MHz) δ 7.16 (d, J = 8.5 Hz, 1H), 6.90–6.72 (m, 3H), 6.63 (d, J = 8.5 Hz, 1H), 5.95 (s, 2H), 5.06 (d, J = 4.0 Hz, 1H), 4.68 (d, J = 4.0 Hz, 1H), 4.30 (t, J = 8.2 Hz, 1H), 4.26–4.13 (m, 1H), 3.98 (dd, J = 9.0, 4.8 Hz, 1H), 3.90 (dd, J = 9.1, 3.9 Hz, 1H), 3.82 (s, 3H, $-\text{OCH}_3$), 3.75 (s, 3H, $-\text{OCH}_3$), 3.10 (m, 1H), 3.00 (m, 1H), 2.16 (s, 3H); ^{13}C NMR (CDCl_3 , 400 MHz) δ 158.6, 157.0, 148.1, 147.2, 135.5, 127.0, 123.8, 120.1, 119.6, 108.3, 106.7, 106.0, 101.2, 85.7, 82.3, 73.1, 71.5, 60.9, 55.8, 54.8, 54.1, 9.2; HRMS m/z 407.1469 $[\text{M}+\text{Na}]^+$ (calcd for $\text{C}_{22}\text{H}_{24}\text{NaO}_6$, 407.1471).

epi-**3.1e**: ^1H NMR (CDCl_3 , 400 MHz) δ 7.36 (d, J = 8.6 Hz, 1H), 6.90 – 6.74 (m, 3H), 6.66 (d, J = 8.5 Hz, 1H), 5.94 (s, 2H), 4.96 (d, J = 6.2 Hz, 1H, H-6), 4.38 (d, J = 7.4 Hz, 1H, H-2), 4.10 (d, J = 9.3 Hz, 1H), 3.83 (m, 4H), 3.74 (m, 4H), 3.46 (m, 1H), 3.24 (t, J = 8.6 Hz, 1H), 2.87 (m, 1H), 2.16 (s, 3H); ^{13}C NMR (CDCl_3 , 400 MHz) δ 158.3, 155.7, 148.1, 147.3, 135.6, 124.5, 124.2, 123.6, 119.8, 108.3, 106.78, 105.9, 101.2, 87.7, 78.8, 70.6, 69.9, 60.6, 55.8, 55.0, 49.3, 9.3; HRMS m/z 407.1470 $[\text{M}+\text{Na}]^+$ (calcd for $\text{C}_{22}\text{H}_{24}\text{NaO}_6$, 407.1471).

(1*R*, 2*R*, 5*S*, 6*S*)-2-(2,3,4-

trimethoxyphenyl)-

6-(3,4-methylenedioxyphenyl)-3,7-

dioxabicyclo[3.3.0]octane (**3.1g**)(1*R*, 2*S*, 5*S*, 6*S*)-2-(2,3,4-

trimethoxyphenyl)-

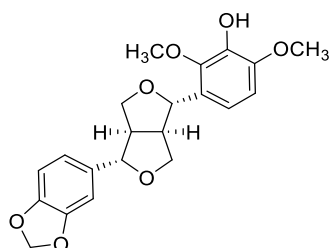
6-(3,4-methylenedioxyphenyl)-3,7-

dioxabicyclo[3.3.0]octane (*epi*-**3.1g**)

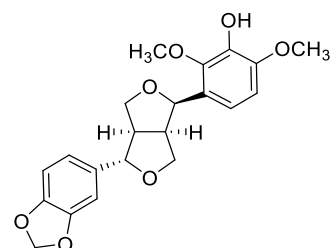
Following the general procedure, reaction of **1.36** (100 mg, 0.39 mmol), 1,2,3-trimethoxybenzene (70.3 mg, 0.58 mmol) in acetonitrile (4 mL) after 8 h yielded **3.1g** (73.3 mg, 31%) and *epi*-**3.1g** (51.7 mg, 22%) as brown oil.

3.1g: $^1\text{H NMR}$ (CDCl_3 , 400 MHz) δ 7.03 (d, J = 8.6 Hz, 1H), 6.90 – 6.73 (m, 3H), 6.64 (d, J = 8.6 Hz, 1H), 5.95 (s, 2H), 5.03 (d, J = 4.0 Hz, 1H), 4.67 (d, J = 4.0 Hz, 1H), 4.33 (dd, J = 8.9, 7.5 Hz, 1H), 4.21 (dd, J = 9.0, 6.6 Hz, 1H), 3.99 (dd, J = 8.0, 4.0 Hz, 1H), 3.92 (s, 3H, -OCH₃), 3.89 (m, 1H), 3.87 (s, 3H, -OCH₃), 3.85 (s, 3H, -OCH₃), 3.05 (m, 1H), 2.98 (m, 1H). $^{13}\text{C NMR}$ (CDCl_3 , 400 MHz) δ 153.4, 151.2, 148.1, 147.3, 142.4, 135.4, 128.0, 120.3, 119.6, 108.3, 107.1, 106.7, 101.2, 85.6, 82.4, 73.2, 71.5, 60.9, 60.9, 56.2, 54.8, 54.2; HRMS m/z 423.1431 [$\text{M}+\text{Na}$]⁺ (calcd for C₂₂H₂₄NaO₇, 423.1420).

epi-**3.1g**: $^1\text{H NMR}$ (CDCl_3 , 400 MHz) δ 7.21 (d, J = 8.6 Hz, 1H), 6.87-6.76 (m, 3H, H-2', H-5', and H-6'), 6.67 (d, J = 8.6 Hz, 1H), 5.94 (s, 2H, H-7'), 4.93 (d, J = 5.6 Hz, 1H, H-6), 4.37 (d, J = 8.0 Hz, 1H, H-2), 4.09 (d, J = 9.2 Hz, 1H, H-8), 3.92 (s, 3H, -OCH₃), 3.86 (s, 6H, -OCH₃ (x2)), 3.83-3.77 (m, 2H), 3.43 (m, 1H), 3.24 (m, 1H), 2.86 (m, 1H); $^{13}\text{C NMR}$ (CDCl_3 , 400 MHz) δ 153.2, 150.0, 148.1, 147.3, 141.8, 135.5, 124.4, 121.2, 119.7, 108.3, 107.0, 106.8, 101.2, 87.7, 78.6, 70.6, 69.9, 60.9, 60.8, 56.1, 55.0, 49.2; HRMS m/z 423.1431 [$\text{M}+\text{Na}$]⁺ (calcd for C₂₂H₂₄NaO₇, 423.1420).



(1*R*, 2*R*, 5*S*, 6*S*)-2-(3-hydroxy-2,4-dimethoxyphenyl)-6-(3,4-methylenedioxyphenyl)-3,7-dioxabicyclo[3.3.0]octane (**3.1h**)

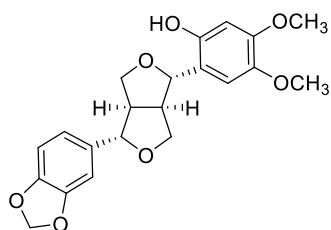


(1*R*, 2*S*, 5*S*, 6*S*)-2-(3-hydroxy-2,4-dimethoxyphenyl)-6-(3,4-methylenedioxyphenyl)-3,7-dioxabicyclo[3.3.0]octane (*epi*-**3.1h**)

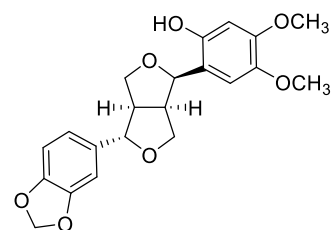
Following the general procedure, reaction of **1.36** (36.0 mg, 0.14 mmol), 2,6-dimethoxyphenol (43 mg, 0.28 mmol) in acetonitrile (2 mL) after 8 h yielded **3.1h** (15 mg, 27%) and *epi*-**3.1h** (20 mg, 37%) as yellow oil.

3.1h: ^1H NMR (CDCl_3 , 400 MHz) δ 6.89 – 6.73 (m, 4H), 6.62 (d, J = 8.6 Hz, 1H), 5.94 (s, 2H), 5.05 (d, J = 4.0 Hz, 1H), 4.68 (d, J = 4.0 Hz, 1H), 4.31 (dd, J = 9.1, 7.3 Hz, 1H), 4.22 (dd, J = 9.1, 6.6 Hz, 1H), 4.01 (dd, J = 9.2, 4.7 Hz, 1H), 3.92 (d, J = 4.3 Hz, 4H), 3.89 (d, J = 7.1 Hz, 4H), 3.10 – 3.02 (m, 1H), 3.01 – 2.93 (m, 1H); ^{13}C NMR (CDCl_3 , 400 MHz) δ 148.1, 147.4, 144.6, 138.7, 135.4, 128.3, 119.6, 115.9, 108.3, 106.7, 105.9, 101.2, 85.6, 82.4, 73.1, 71.6, 60.6, 56.4, 54.8, 54.2; HRMS m/z 409.1260 $[\text{M}+\text{Na}]^+$ (calcd for $\text{C}_{21}\text{H}_{22}\text{NaO}_7$, 409.1263).

epi-**3.1h**: ^1H NMR (CDCl_3 , 400 MHz) δ 7.02 (d, J = 8.5 Hz, 1H), 6.89–6.73 (m, 3H), 6.65 (d, J = 8.4 Hz, 1H), 5.95 (s, 2H), 4.95 (d, J = 5.9 Hz, 1H, H-6), 4.36 (d, J = 8.0 Hz, 1H, H-2), 4.09 (d, J = 9.4 Hz, 1H), 3.97–3.84 (m, 7H), 3.86–3.74 (m, 2H), 3.51–3.40 (m, 1H), 3.24 (t, J = 8.6 Hz, 1H), 2.86 (dd, J = 15.4, 7.2 Hz, 1H); ^{13}C NMR (CDCl_3 , 400 MHz) δ 148.1, 147.3, 147.2, 138.2, 135.5, 129.9, 124.6, 119.7, 116.8, 108.3, 106.8, 105.8, 101.2, 87.7, 78.7, 70.6, 69.9, 60.3, 56.4, 54.9, 49.2; HRMS m/z 409.1261 $[\text{M}+\text{Na}]^+$ (calcd for $\text{C}_{21}\text{H}_{22}\text{NaO}_7$, 409.1263).



(1*R*, 2*R*, 5*S*, 6*S*)-2-(2-hydroxy-4,5-dimethoxyphenyl)-6-(3,4-methylenedioxyphenyl)-3,7-dioxabicyclo[3.3.0]octane (**3.1i**)

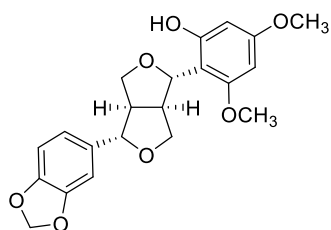


(1*R*, 2*S*, 5*S*, 6*S*)-2-(2-hydroxy-4,5-dimethoxyphenyl)-6-(3,4-methylenedioxyphenyl)-3,7-dioxabicyclo[3.3.0]octane (*epi*-**3.1i**)

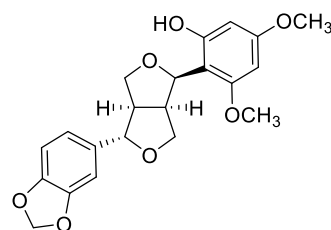
Following the general procedure, reaction of **1.36** (46.8 mg, 0.19 mmol), 3,4-dimethoxyphenol (58 mg, 0.37 mmol) in acetonitrile (2 mL) after 8 h yielded **3.1i** (49 mg, 68%) and *epi*-**3.1i** (15 mg, 17%) as white powder.

3.1i: ^1H NMR (CDCl_3 , 400 MHz) δ 7.71 (brs, 1H, -OH), 6.84-6.79 (m, 3H), 6.54 (s, 1H), 6.49 (s, 1H), 5.96 (s, 2H), 4.82 (d, $J = 4.0$ Hz, 1H), 4.78 (d, $J = 4.0$ Hz, 1H), 4.36 (dd, $J = 8.8, 7.2$ Hz, 1H), 4.16 (dd, $J = 9.6, 6.4$ Hz, 1H), 3.92-3.86 (m, 2H), 3.84 (s, 3H, -OCH₃), 3.82 (s, 3H, -OCH₃), 3.21-3.14 (m, 2H); ^{13}C NMR (CDCl_3 , 400 MHz) δ 150.3, 150.1, 148.2, 147.4, 142.6, 134.8, 125.2, 119.5, 111.2, 108.4, 106.7, 102.1, 101.3, 86.7, 85.6, 72.6, 70.8, 57.2, 56.1, 53.6, 53.2; HRMS m/z 409.1286 [$\text{M}+\text{Na}$]⁺ (calcd for C₂₁H₂₂NaO₇, 409.1263).

epi-**3.1i**: ^1H NMR (CDCl_3 , 400 MHz) δ 8.05 (brs, 1H, -OH), 6.87-6.77 (m, 3H), 6.46 (s, 1H), 6.42 (s, 1H), 5.95 (s, 2H), 5.01 (d, $J = 8.0$ Hz, 1H), 4.44 (d, $J = 6.8$ Hz, 1H), 4.19 (d, $J = 9.6$ Hz, 1H), 3.98 (t, $J = 8.8$ Hz, 1H), 3.88 (m, 1H), 3.85 (s, 3H, -OCH₃), 3.80 (s, 3H, -OCH₃), 3.49 (dd, $J = 8.4, 9.2$ Hz, 1H), 3.40 (m, 1H), 2.91 (m, 1H); ^{13}C NMR (CDCl_3 , 400 MHz) δ 150.1, 149.8, 148.2, 147.5, 142.7, 134.8, 125.2, 119.8, 110.5, 108.4, 106.7, 101.9, 101.2, 87.7, 84.6, 71.9, 70.1, 57.0, 56.0, 53.7, 50.8; HRMS m/z 409.1275 [$\text{M}+\text{Na}$]⁺ (calcd for C₂₁H₂₂NaO₇, 409.1263).



(1*R*, 2*R*, 5*S*, 6*S*)-2-(2-hydroxy-4,6-dimethoxyphenyl)-6-(3,4-methylenedioxyphenyl)-3,7-dioxabicyclo[3.3.0]octane (**3.1j**)

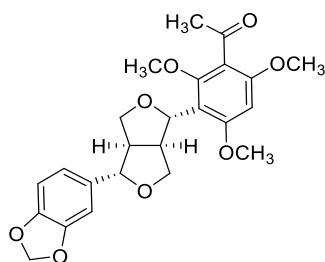


(1*R*, 2*S*, 5*S*, 6*S*)-2-(2-hydroxy-4,6-dimethoxyphenyl)-6-(3,4-methylenedioxyphenyl)-3,7-dioxabicyclo[3.3.0]octane (*epi*-**3.1j**)

Following the general procedure, reaction of **1.36** (56.5 mg, 0.22 mmol), 3,5-dimethoxyphenol (52 mg, 0.34 mmol) in acetonitrile (2 mL) after 8 h yielded **3.1j** (41 mg, 47%) and *epi*-**3.1j** (44 mg, 51%) as white powder.

3.1j: ^1H NMR (CDCl_3 , 400 MHz) δ 8.96 (brs, 1H, -OH), 6.82-6.77 (m, 3H), 6.06 (d, $J = 2.4$ Hz, 1H), 6.01 (d, $J = 2.4$ Hz, 1H), 5.95 (s, 2H), 5.21 (d, $J = 4.8$ Hz, 1H), 4.81 (d, $J = 4.0$ Hz, 1H), 4.47 (dd, $J = 9.2, 8.4$ Hz, 1H), 4.13 (dd, $J = 9.2, 2.8$ Hz, 1H), 4.03 (dd, $J = 9.2, 6.8$ Hz, 1H), 3.79 (m, 1H), 3.76 (s, 6H, $-\text{OCH}_3$ (x2)), 3.19 (m, 1H), 3.01 (m, 1H); ^{13}C NMR (CDCl_3 , 400 MHz) δ 161.0, 158.0, 157.6, 148.2, 147.3, 134.9, 119.5, 108.3, 106.7, 105.0, 101.2, 94.6, 91.0, 84.2, 84.2, 72.7, 71.0, 55.5, 55.5, 54.8, 53.7; HRMS m/z 387.1449 $[\text{M}+\text{H}]^+$ (calcd for $\text{C}_{21}\text{H}_{23}\text{O}_7$, 387.1444).

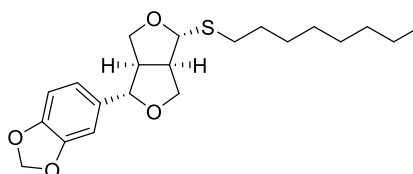
epi-**3.1j**: ^1H NMR (CDCl_3 , 400 MHz) δ 9.15 (brs, 1H, -OH), 6.87-6.77 (m, 3H), 6.07 (d, $J = 2.0$ Hz, 1H), 6.00 (d, $J = 2.4$ Hz, 1H), 5.95 (s, 2H), 5.17 (d, $J = 8.0$ Hz, 1H, H-2), 4.40 (d, $J = 6.8$ Hz, 1H, H-6), 4.17 (d, $J = 10.0$ Hz, 1H), 3.91 (dd, $J = 8.0, 8.0$ Hz, 1H), 3.81 (dd, $J = 9.6, 6.4$ Hz, 1H), 3.77 (s, 3H, $-\text{OCH}_3$), 3.76 (s, 3H, $-\text{OCH}_3$), 3.51-3.42 (m, 2H), 2.87 (m, 1H); ^{13}C NMR (CDCl_3 , 400 MHz) δ 160.9, 158.1, 157.4, 148.2, 147.5, 134.9, 119.8, 108.3, 106.8, 101.7, 101.2, 94.3, 90.8, 87.5, 81.9, 71.4, 70.3, 55.7, 55.4, 53.7, 49.6; HRMS m/z 409.1273 $[\text{M}+\text{Na}]^+$ (calcd for $\text{C}_{21}\text{H}_{22}\text{NaO}_7$, 409.1263).



(1*R*, 2*R*, 5*S*, 6*S*)-2-(3-acetyl-2,4,6-trimethoxyphenyl)-

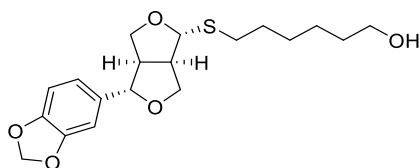
6-(3,4-methylenedioxyphenyl)-3,7-dioxabicyclo[3.3.0]octane (**3.1k**)

3.1k: Following the general procedure, reaction of **1.36** (100 mg, 0.39 mmol), 2,4,6-trimethoxyacetophenone (116.7 mg, 0.58 mmol) in acetonitrile (4 mL) after 8 h yielded **3.1k** (25.8 mg, 15%) as white powder; ^1H NMR (CDCl_3 , 400 MHz) δ 6.88 (m, 3H), 6.27 (s, 1H), 5.95 (s, 2H), 5.23 (d, $J = 6.0$ Hz, 1H), 4.74 (d, $J = 5.6$ Hz, 1H), 4.31 (dd, $J = 9.2, 7.2$ Hz, 1H), 4.13 (dd, $J = 8.8, 7.6$ Hz, 1H), 3.92-3.90 (m, 2H), 3.85 (s, 3H, $-\text{OCH}_3$), 3.83 (s, 3H, $-\text{OCH}_3$), 3.77 (s, 3H, $-\text{OCH}_3$), 3.44 (m, 1H) 3.12 (m, 1H), 2.49 (s, 3H, $-\text{CH}_3$); ^{13}C NMR (CDCl_3 , 400 MHz) δ 202.0, 160.6, 158.1, 157.8, 148.1, 147.2, 135.7, 119.7, 119.5, 108.3, 106.7, 101.2, 101.2, 92.0, 85.3, 79.1, 72.8, 72.6, 64.4, 56.0, 56.0, 55.9, 51.5, 29.8; HRMS m/z 465.1538 $[\text{M}+\text{Na}]^+$ (calcd for $\text{C}_{24}\text{H}_{26}\text{NaO}_8$, 465.1525).



(1*R*, 2*R*, 5*S*, 6*S*)-6-(3,4-methylenedioxyphenyl)-
2-(octylsulfanyl)-3,7-dioxabicyclo[3.3.0]octane (3.2l)

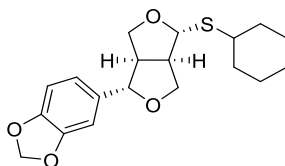
3.2l: Following the general procedure, reaction of **1.36** (80 mg, 0.32 mmol), 1-octanethiol (166 μ L, 0.96 mmol) in acetonitrile (3 mL) after 8 h yielded **3.2l** (107.52 mg, 89%) as a yellow oil; ^1H NMR (CDCl_3 , 400 MHz) δ 6.86 – 6.75 (m, 3H, H-2', H-5' and H-6'), 5.95 (s, 2H, H-7'), 5.38 (s, 1H, H-2), 4.38 (m, 2H, H-6 and H-8), 4.15 (dd, $J = 9.3, 5.9$ Hz, 1H, H-4), 3.83 (d, $J = 9.4$ Hz, 1H, H-4), 3.60 (m, 1H, H-8), 2.96 (m, 1H, H-1), 2.77 (m, 1H, H-5), 2.75 (m, 1H, H-1''), 2.55 (m, 1H, H-1''), 1.62 – 1.27 (m, 12H, H- CH_2), 0.88 (t, 3H, H- CH_3); ^{13}C NMR (CDCl_3 , 400 MHz) δ 148.0, 147.3, 134.6, 119.6, 108.2, 106.6, 101.1, 89.0, 86.9, 73.4, 68.3, 53.3, 53.2, 31.8, 31.1, 29.8, 29.2, 29.1, 29.0, 22.6, 14.0; HRMS m/z 401.1759 $[\text{M}+\text{Na}]^+$ (calcd for $\text{C}_{21}\text{H}_{30}\text{NaO}_4\text{S}$, 401.1763).



(1*R*, 2*R*, 5*S*, 6*S*)- 2-[(6-hydroxy)hexylsulfanyl]-

6-(3,4-methylenedioxyphenyl)-3,7-dioxabicyclo[3.3.0]octane (3.2m)

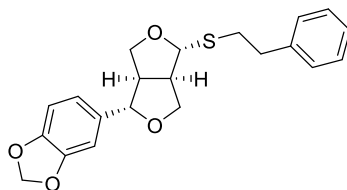
3.2m: Following the general procedure, reaction of **1.36** (43 mg, 0.172 mmol), 6-mercapto-1-hexanol (70 μ L, 0.515 mmol) in acetonitrile (2 mL) after 8 h yielded **3.2m** (40.90 mg, 65%) as a yellow oil; ^1H NMR (CDCl_3 , 400 MHz) δ 6.86 – 6.75 (m, 3H), 5.94 (s, 2H), 5.26 (s, 1H), 4.42 – 4.32 (m, 2H), 4.12 (dd, $J = 9.2, 6.0$ Hz, 1H), 3.84 (d, $J = 9.4$ Hz, 1H), 3.65 – 3.52 (m, 3H), 2.97 (m, 1H), 2.81 – 2.73 (m, 1H), 2.67 (m, 1H), 2.55 (m, 1H), 1.70 – 1.25 (m, 8H); ^{13}C NMR (CDCl_3 , 400 MHz) δ 148.0, 147.3, 134.5, 119.6, 108.2, 106.6, 101.1, 89.0, 86.9, 73.4, 68.3, 62.9, 53.3, 53.2, 32.6, 31.0, 29.7, 28.6, 25.3; HRMS m/z 389.1395 $[\text{M}+\text{Na}]^+$ (calcd for $\text{C}_{19}\text{H}_{26}\text{NaO}_5\text{S}$, 389.1399)



(1*R*, 2*R*, 5*S*, 6*S*)-2-(cyclohexylsulfanyl)-

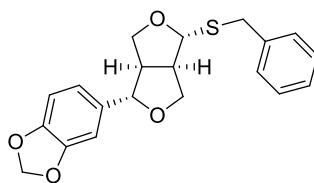
6-(3,4-methylenedioxyphenyl)-3,7-dioxabicyclo[3.3.0]octane (**3.2n**)

3.2n: Following the general procedure, reaction of **1.36** (20 mg, 0.08 mmol), cyclohexanethiol (29 μ L, 0.24 mmol) in acetonitrile (1 mL) after 8 h yielded **3.2n** (22 mg, 79%) as a yellow oil; ^1H NMR (CDCl_3 , 400 MHz) δ 6.86 – 6.75 (m, 3H), 5.95 (s, 2H), 5.38 (s, 1H), 4.42 – 4.35 (m, 2H), 4.15 (dd, $J = 9.3, 5.9$ Hz, 1H), 3.83 (d, $J = 9.4$ Hz, 1H), 3.60 (m, 1H), 2.96 (m, 1H), 2.76 (m, 1H), 2.12 – 1.17 (m, 11H); ^{13}C NMR (CDCl_3 , 400 MHz) δ 148.0, 147.3, 134.6, 119.6, 108.8, 106.6, 101.1, 87.9, 86.9, 73.5, 68.2, 53.4, 53.3, 43.4, 34.0, 33.8, 26.1, 26.0, 25.8; HRMS m/z 371.1297 $[\text{M}+\text{Na}]^+$ (calcd for $\text{C}_{19}\text{H}_{24}\text{NaO}_4\text{S}$, 371.1293).



(1*R*, 2*R*, 5*S*, 6*S*)- 6-(3,4-methylenedioxyphenyl)-
2-[(2-phenylethyl)sulfanyl]-3,7-dioxabicyclo[3.3.0]octane (**3.2o**)

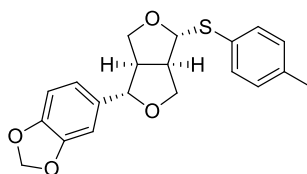
3.2o: Following the general procedure, reaction of **1.36** (65 mg, 0.26 mmol), phenylethylmercaptan (72 mg, 0.52 mmol) in acetonitrile (3 mL) after 8 h yielded **3.2o** (72.1 mg, 75%) as a yellow oil; ^1H NMR (CDCl_3 , 400 MHz) δ 7.34 – 7.20 (m, 5H), 6.87 – 6.76 (m, 3H), 5.95 (s, 2H), 5.23 (s, 1H), 4.40 – 4.35 (m, 2H), 4.12 (dd, $J = 9.4, 5.9$ Hz, 1H), 3.85 (d, $J = 9.4$ Hz, 1H), 3.56 (m, 1H), 3.01 – 2.73 (m, 5H); ^{13}C NMR (CDCl_3 , 400 MHz) δ 148.2, 147.5, 140.6, 134.7, 128.7, 128.6, 126.5, 119.8, 108.3, 106.7, 101.2, 89.3, 87.0, 73.6, 68.4, 53.5, 53.3, 36.6, 32.6; HRMS m/z 393.1149 $[\text{M}+\text{Na}]^+$ (calcd for $\text{C}_{21}\text{H}_{22}\text{NaO}_4\text{S}$, 393.1136).



(1*R*, 2*R*, 5*S*, 6*S*)- 2-benzylsulfanyl-

6-(3,4-methylenedioxyphenyl)-3,7-dioxabicyclo[3.3.0]octane (3.2p)

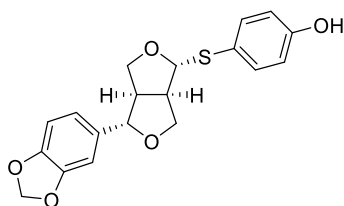
3.2p: Following the general procedure, reaction of **1.36** (68.8 mg, 0.27 mmol), benzylmercaptan (100 mg, 0.81 mmol) in acetonitrile (3 mL) after 8 h yielded **3.2p** (47.97 mg, 50%) as a yellow oil; ^1H NMR (CDCl_3 , 400 MHz) δ 7.37 – 7.24 (m, 5H), 6.86 – 6.76 (m, 3H), 5.95 (s, 2H), 5.12 (s, 1H), 4.37 – 4.31 (m, 2H), 4.18 (dd, $J = 9.4, 5.9$ Hz, 1H), 3.91 – 3.85 (m, 2H), 3.71 (d, $J = 13.5$ Hz, 1H), 3.51 (dd, $J = 8.9, 7.4$ Hz, 1H), 2.96 (m, 1H), 2.79 (m, 1H); ^{13}C NMR (CDCl_3 , 400 MHz) δ 148.0, 147.4, 138.2, 134.5, 128.9, 128.6, 127.0, 119.6, 108.2, 106.6, 101.1, 87.5, 86.9, 73.4, 68.4, 53.3, 52.8, 34.8; HRMS m/z 379.0974 $[\text{M}+\text{Na}]^+$ (calcd for $\text{C}_{20}\text{H}_{20}\text{NaO}_4\text{S}$, 379.0980).



(1*R*, 2*R*, 5*S*, 6*S*)-6-(3,4-methylenedioxyphenyl)-

2-(4-methylthiophenoxy)-3,7-dioxabicyclo[3.3.0]octane (3.2q)

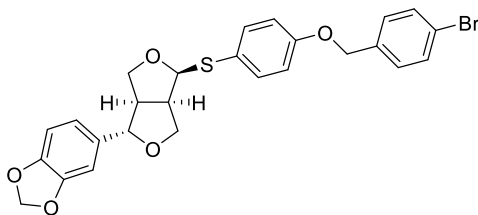
3.2q: Following the general procedure, reaction of **1.36** (50.8 mg, 0.203 mmol), 4-methylthiophenol (76 μ L, 0.609 mmol) in acetonitrile (2 mL) after 8 h yielded **3.2q** (50.53 mg, 69.8%) as a white powder; ^1H NMR (CDCl_3 , 400 MHz) δ 7.40 (d, $J = 7.7$ Hz, 2H), 7.13 (d, $J = 7.8$ Hz, 2H), 6.88 – 6.76 (m, 3H), 5.96 (s, 2H), 5.47 (s, 1H), 4.44 (t, $J = 8.7$ Hz, 1H), 4.36 (d, $J = 7.5$ Hz, 1H), 4.26 (dd, $J = 9.4, 5.9$ Hz, 1H), 3.92 (d, $J = 9.5$ Hz, 1H), 3.61 (m, 1H), 3.12 (m, 1H), 2.83 (m, 1H), 2.33 (s, 3H); ^{13}C NMR (CDCl_3 , 400 MHz) δ 148.2, 147.5, 137.7, 134.6, 132.3, 131.2, 129.8, 119.8, 108.3, 106.7, 101.2, 92.5, 87.0, 73.5, 68.7, 53.4, 53.3, 21.2; HRMS m/z 379.0987 $[\text{M}+\text{Na}]^+$ (calcd for $\text{C}_{20}\text{H}_{20}\text{NaO}_4\text{S}$, 379.0980).



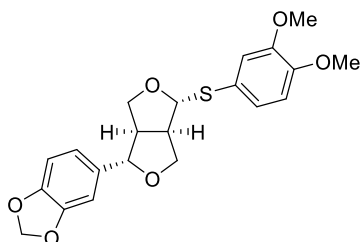
(1*R*, 2*R*, 5*S*, 6*S*)-2-(4-hydroxythiophenoxy)-

6-(3,4-methylenedioxyphenyl)-3,7-dioxabicyclo[3.3.0]octane (**3.2r**)

3.2r: Following the general procedure, reaction of **1.36** (70 mg, 0.28 mmol), 4-Hydroxythiophenol (71 mg, 0.56 mmol) in acetonitrile (3 mL) after 8 h yielded **3.2r** (71.65 mg, 71.4%) as a white powder; ^1H NMR (CDCl_3 , 400 MHz) δ 7.38 (d, J = 8.5 Hz, 2H), 6.87 – 6.71 (m, 5H), 5.95 (s, 2H), 5.37 (s, 1H), 4.44 (t, J = 8.7 Hz, 1H), 4.37 (d, J = 7.5 Hz, 1H), 4.28 (dd, J = 9.4, 5.9 Hz, 1H), 3.93 (d, J = 9.4 Hz, 1H), 3.61 (dd, J = 8.9, 7.1 Hz, 1H), 3.11 (m, 1H), 2.83 (m, 1H); ^{13}C NMR (CDCl_3 , 400 MHz) δ 156.1, 148.2, 147.6, 135.2, 134.4, 133.1, 119.9, 116.2, 108.4, 106.7, 101.2, 93.2, 87.0, 73.5, 68.7, 53.3, 53.0; HRMS m/z 381.0771 [$\text{M}+\text{Na}$] $^+$ (calcd for $\text{C}_{19}\text{H}_{18}\text{NaO}_5\text{S}$, 381.0773).



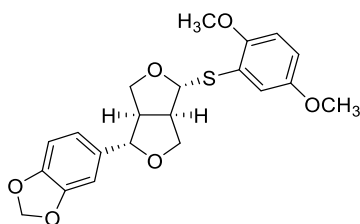
epi-3.2r-pBBE: Following the general procedure, reaction of **1.36** (100 mg, 0.40 mmol), 4-Hydroxythiophenol (100 mg, 0.80 mmol) in acetonitrile (4 mL) was carefully performed in room temperature for 1 h. Preparative TLC is used for purification of **epi-3.2r** (28.86 mg, 20%). A Solution of **epi-3.2r** in CH₃CN was further reacted with 4-bromobenzylbromide (30 mg, 0.12 mmol) under basic condition (K₂CO₃) for 4 h. Solvent was evaporated, water was added, and after the usual work up in EtOAc, the residue was purified by column chromatography. The white semisolid was recrystallized from MeOH to afford **epi-3.2r-pBBE** (50 mg, 80 %) as a white crystal; ¹H NMR (400 MHz, CDCl₃) δ 7.50 (dd, *J* = 11.3, 8.6 Hz, 4H, H-2''', H-3''', H-5''' and H-6''') 7.29 (d, *J* = 8.2 Hz, 2H, H-2'' and H-6'') 6.91 (d, *J* = 8.7 Hz, 2H, H-3'' and H-5'') 6.85 (s, 1H, H-2') 6.82 – 6.76 (m, 2H, H-5' and H-6') 5.95 (s, 2H, H-7') 5.11 (d, *J* = 6.1 Hz, 1H, H-2) 5.00 (s, 2H, H-7'') 4.54 (d, *J* = 6.7 Hz, 1H, H-6) 4.21 (t, *J* = 8.7 Hz, 1H, H-8) 4.02 (dd, *J* = 17.2, 9.7 Hz, 2H, H-4) 3.81 (dd, *J* = 9.4, 6.7 Hz, 1H, H-8) 3.38 (dd, *J* = 15.7, 8.0 Hz, 1H, H-1) 2.88 – 2.82 (m, 1H, H-5); HRMS *m/z* 549.0349, 551.0329 [M+Na]⁺ (calcd for C₂₆H₂₃⁷⁹BrNaO₅S, 549.0347 and C₂₆H₂₃⁸¹BrNaO₅S, 551.0327).



(1*R*, 2*R*, 5*S*, 6*S*)-2-(3,4- dimethoxythiophenoxy)-

6-(3,4-methylenedioxyphenyl)-3,7-dioxabicyclo[3.3.0]octane (3.2s)

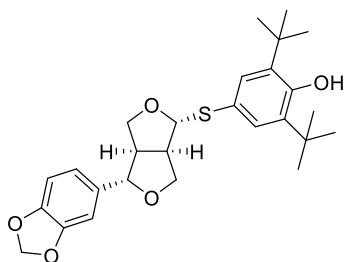
3.2s: Following the general procedure, reaction of **1.36** (107 mg, 0.427 mmol), 3,4-dimethoxythiophenol (123 μ L, 0.557 mmol) in acetonitrile (4 mL) after 8 h yielded **3.2s** (86.56 mg, 50.4%) as a colorless oil; ^1H NMR (CDCl_3 , 400 MHz) δ 7.09 (dd, $J = 8.3, 1.8$ Hz, 1H), 7.05 (d, $J = 1.6$ Hz, 1H) 6.81 (m, 4H), 5.95 (s, 2H), 5.42 (s, 1H), 4.44 (t, $J = 8.7$ Hz, 1H), 4.37 (d, $J = 7.5$ Hz, 1H), 4.27 (dd, $J = 9.4, 5.9$ Hz, 1H), 3.93 (d, $J = 9.5$ Hz, 1H), 3.89 (s, 3H, H-OCH₃), 3.87 (s, 3H, H-OCH₃), 3.62 (dd, $J = 8.9, 7.1$ Hz, 1H), 3.11 (m, 1H), 2.82 (m, 1H); ^{13}C NMR (CDCl_3 , 400 MHz) δ 149.3, 149.1, 148.0, 147.4, 134.4, 125.8, 125.5, 119.6, 116.1, 111.6, 108.2, 106.5, 101.9, 92.9, 86.8, 73.4, 68.6, 56.0, 56.0, 53.3, 53.0; HRMS m/z 425.1048 $[\text{M}+\text{Na}]^+$ (calcd for $\text{C}_{21}\text{H}_{22}\text{NaO}_6\text{S}$, 425.1035).



(1*R*, 2*R*, 5*S*, 6*S*)-2-(2,5- dimethoxythiophenoxy)-

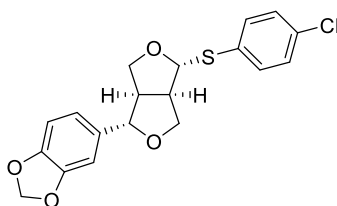
6-(3,4-methylenedioxyphenyl)-3,7-dioxabicyclo[3.3.0]octane (3.2t)

3.2t: Following the general procedure, reaction of **1.36** (77.7 mg, 0.310 mmol), 2,5-dimethoxythiophenol (93 μ L, 0.621 mmol) in acetonitrile (3 mL) after 8 h yielded **3.2t** (119.78 mg, 88.3%) as a colorless oil; ^1H NMR (CDCl_3 , 400 MHz) δ 7.17 (d, J = 2.6 Hz, 1H), 6.87 – 6.74 (m, 5H), 5.94 (s, 2H), 5.68 (s, 1H), 4.44 (t, J = 8.7 Hz, 1H), 4.37 (d, J = 7.6 Hz, 1H), 4.23 (dd, J = 9.5, 5.9 Hz, 1H), 3.91 (d, J = 9.5 Hz, 1H), 3.84 (s, 3H, H-OCH₃), 3.77 (s, 3H, H-OCH₃), 3.64 (dd, J = 8.9, 7.2 Hz, 1H), 3.17 (m, 1H), 2.84 (m, 1H); ^{13}C NMR (CDCl_3 , 400 MHz) δ 153.9, 152.1, 148.0, 147.4, 134.4, 124.0, 119.6, 118.0, 112.9, 111.7, 108.2, 106.6, 101.1, 89.8, 86.9, 73.4, 68.8, 56.5, 55.8, 53.3, 53.2; HRMS m/z 425.1048 [$\text{M}+\text{Na}$]⁺ (calcd for $\text{C}_{21}\text{H}_{22}\text{NaO}_6\text{S}$, 425.1035).



(1*R*, 2*R*, 5*S*, 6*S*)-2-(4-hydroxy-3,5-ditertiarybutylthiophenoxy)-
6-(3,4-methylenedioxyphenyl)-3,7-dioxabicyclo[3.3.0]octane (**3.2u**)

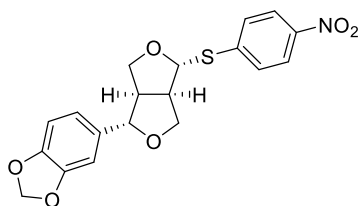
3.2u: Following the general procedure, reaction of **1.36** (50.8 mg, 0.209 mmol), 2,6-di-*tert*-butyl-4-mercaptophenol (96.78 mg, 0.406 mmol) in acetonitrile (2 mL) after 8 h yielded **3.2u** (45.9 mg, 48.3%) as a yellow oil; ^1H NMR (CDCl_3 , 400 MHz) δ 7.33 (s, 2H), 6.87 – 6.75 (m, 3H), 5.95 (s, 2H), 5.39 (s, 1H), 4.43 (t, $J = 8.7$ Hz, 1H), 4.37 (d, $J = 7.5$ Hz, 1H), 4.29 (dd, $J = 9.4, 5.9$ Hz, 1H), 3.93 (d, $J = 9.4$ Hz, 1H), 3.61 (m, 1H), 3.11 (m, 1H), 2.81 (m, 1H), 1.43 (s, 18H); ^{13}C NMR (CDCl_3 , 400 MHz) δ 154.2, 148.2, 147.5, 136.7, 130.4, 127.9, 119.8, 108.3, 106.7, 101.2, 93.1, 86.98, 73.5, 68.5, 53.5, 53.2, 34.5, 30.3; HRMS m/z 493.2024 $[\text{M}+\text{Na}]^+$ (calcd for $\text{C}_{27}\text{H}_{34}\text{NaO}_5\text{S}$, 493.2025).



(1*R*, 2*R*, 5*S*, 6*S*)-2-(4-chlorothiophenoxy)-

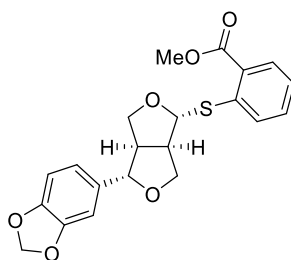
6-(3,4-methylenedioxyphenyl)-3,7-dioxabicyclo[3.3.0]octane (**3.2v**)

3.2v: Following the general procedure, reaction of **1.36** (28.9 mg, 0.115 mmol), 4-chlorothiophenol (50 mg, 0.35 mmol) in acetonitrile (2 mL) after 8 h yielded **3.2v** (36.27 mg, 83.7%) as a yellow oil; ^1H NMR (CDCl_3 , 400 MHz) δ 7.40 (d, $J = 8.4$ Hz, 2H), 7.25 (d, $J = 8.4$ Hz, 2H), 6.84 – 6.73 (m, 3H), 5.93 (s, 2H), 5.46 (s, 1H), 4.42 (t, $J = 8.7$ Hz, 1H), 4.34 (d, $J = 7.5$ Hz, 1H), 4.21 (dd, $J = 9.5, 5.9$ Hz, 1H), 3.91 (d, $J = 9.6$ Hz, 1H), 3.59 (dd, $J = 9.0, 7.1$ Hz, 1H), 3.09 (m, 1H), 2.81 (m, 1H); ^{13}C NMR (CDCl_3 , 400 MHz) δ 148.0, 147.4, 134.3, 133.5, 133.4, 132.8, 129.0, 119.6, 108.2, 106.5, 101.1, 92.0, 86.9, 73.3, 68.7, 53.2, 53.1.



(1*R*, 2*R*, 5*S*, 6*S*)-6-(3,4-methylenedioxyphenyl)-
2-(4-nitrothiophenoxy)-3,7-dioxabicyclo[3.3.0]octane (**3.2w**)

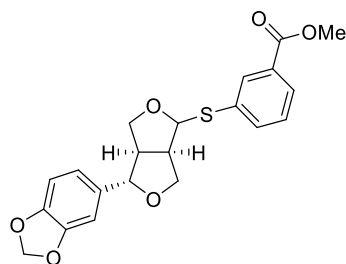
3.2w: Following the general procedure, reaction of **1.36** (65 mg, 0.26 mmol), 4-nitrothiophenol (81 mg, 0.52 mmol) in acetonitrile (3 mL) after 8 h yielded **3.2w** (70.5 mg, 70%) as a yellow powder; ^1H NMR (CDCl_3 , 400 MHz) δ 8.13 (d, $J = 8.8$ Hz, 2H), 7.59 (d, $J = 8.8$ Hz, 2H), 6.87 – 6.75 (m, 3H), 5.95 (s, 2H), 5.73 (s, 1H), 4.48 (t, $J = 8.8$ Hz, 1H), 4.41 (d, $J = 7.5$ Hz, 1H), 4.21 (dd, $J = 9.7, 5.8$ Hz, 1H), 3.99 (d, $J = 9.6$ Hz, 1H), 3.67 (dd, $J = 9.1, 7.0$ Hz, 1H), 3.16 (m, 1H), 2.88 (m, 1H); ^{13}C NMR (CDCl_3 , 400 MHz) δ 148.2, 147.6, 146.2, 145.8, 134.2, 128.8, 124.0, 119.8, 108.4, 106.6, 101.3, 90.6, 87.0, 73.4, 69.2, 53.3, 53.2.



(1*R*, 2*R*, 5*S*, 6*S*)-2-[(2-methoxycarbonyl)thiophenoxy]-

6-(3,4-methylenedioxyphenyl)-3,7-dioxabicyclo[3.3.0]octane (**3.2x**)

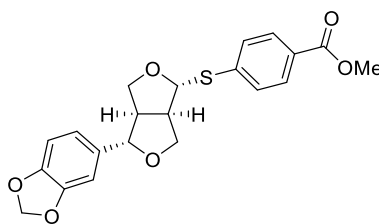
3.2x: Following the general procedure above, reaction of **1.36** (80 mg, 0.319 mmol), methylthiosalicylate (87.9 mg, 0.639 mmol) in acetonitrile (3 mL) after 8 h yielded **3.2x** (78.73 mg, 61.7%) as a white powder; $^1\text{H NMR}$ (CDCl_3 , 400 MHz) δ 7.93 (dd, $J = 7.8, 1.1$ Hz, 1H), 7.85 (d, $J = 8.1$ Hz, 1H), 7.47 (m, 1H), 7.23 (dd, $J = 14.2, 6.9$ Hz, 1H), 6.88 – 6.75 (m, 3H), 5.95 (s, 2H), 5.68 (s, 1H), 4.48 – 4.41 (m, 2H), 4.23 (dd, $J = 9.5, 5.8$ Hz, 1H), 3.99 – 3.89 (m, 4H), 3.70 (dd, $J = 9.1, 6.8$ Hz, 1H), 3.24 (m, 1H), 2.87 (m, 1H); $^{13}\text{C NMR}$ (CDCl_3 , 400 MHz) δ 167.0, 148.1, 147.4, 139.9, 134.3, 132.4, 130.9, 128.8, 128.4, 125.1, 119.6, 108.2, 106.5, 101.1, 89.8, 86.8, 73.5, 68.9, 53.3, 53.0, 52.2; HRMS m/z 423.0884 [$\text{M}+\text{Na}$] $^+$ (calcd for $\text{C}_{21}\text{H}_{20}\text{NaO}_6\text{S}$, 423.0878).



(1*R*, 2*R*, 5*S*, 6*S*)-2-[(3- methoxycarbonyl)thiophenoxy]-

6-(3,4-methylenedioxyphenyl)-3,7-dioxabicyclo[3.3.0]octane (**3.2y**)

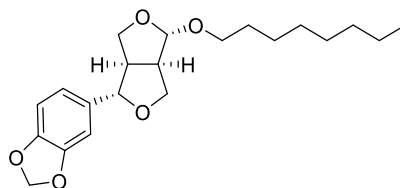
3.2y: To a solution of **1.36** (100 mg, 0.399 mmol) were treated with 3-mercaptopropionic acid (123 mg, 0.799 mmol), Amberlyst-15 (1 mg/0.005 mmol of **1.36**) and 4 Å molecular sieve. After stirring at room temperature for 8 h, the reaction mixture was evaporated to dryness. The crude reaction was then metelated [17] in order to make easily to separate by column chromatography, yielded **3.2y** (89.3 mg, 56%) as a white powder; ^1H NMR (CDCl_3 , 400 MHz) δ 8.16 (s, 1H), 7.92 (d, $J = 7.7$ Hz, 1H), 7.68 (d, $J = 7.8$ Hz, 1H), 7.38 (t, $J = 7.8$ Hz, 1H), 6.88 – 6.75 (m, 3H), 5.95 (s, 2H), 5.58 (s, 1H), 4.46 (t, $J = 8.7$ Hz, 1H), 4.38 (d, $J = 7.5$ Hz, 1H), 4.26 (dd, $J = 9.5, 5.9$ Hz, 1H), 3.96 (d, $J = 9.6$ Hz, 1H), 3.92 (s, 3H), 3.64 (dd, $J = 9.0, 7.1$ Hz, 1H), 3.14 (m, 1H), 2.85 (m, 1H); ^{13}C NMR (CDCl_3 , 400 MHz) δ 166.5, 148.1, 147.4, 135.7, 135.5, 134.3, 132.2, 130.9, 130.8, 128.9, 128.3, 119.6, 108.2, 106.5, 101.1, 91.8, 86.8, 73.3, 68.7, 53.2, 52.2; HRMS m/z 423.0884 [$\text{M}+\text{Na}$] $^+$ (calcd for $\text{C}_{21}\text{H}_{20}\text{NaO}_6\text{S}$, 423.0878).



(1*R*, 2*R*, 5*S*, 6*S*)-2-[(4-methoxycarbonyl)thiophenoxy]-

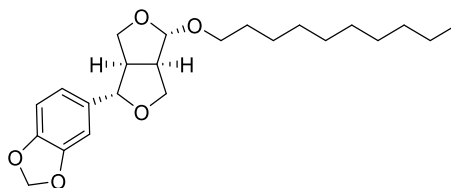
6-(3,4-methylenedioxyphenyl)-3,7-dioxabicyclo[3.3.0]octane (**3.2z**)

3.2z: To a solution of **1.36** (100 mg, 0.399 mmol) were treated with 4-mercatobenzoic acid (123 mg, 0.798 mmol), Amberlyst-15 (1 mg/0.005 mmol of **1.36**) and 4 Å molecular sieve. After stirring at room temperature for 8 h, the reaction mixture was evaporated to dryness. The crude reaction was then metelated [17] in order to make easily to sepearate via column cromatography, yielded **3.2z** (105.8 mg, 66.2%) as a white powder; ^1H NMR (CDCl_3 , 400 MHz) δ 7.95 (d, $J = 8.4$ Hz, 2H), 7.53 (d, $J = 8.4$ Hz, 2H), 6.88 – 6.76 (m, 3H), 5.96 (s, 2H), 5.68 (s, 1H), 4.47 (t, $J = 8.7$ Hz, 1H), 4.40 (d, $J = 7.5$ Hz, 1H), 4.23 (m, 1H), 3.97 (d, $J = 9.6$ Hz, 1H), 3.91 (s, 3H), 3.67 (dd, $J = 9.0, 7.1$ Hz, 1H), 3.15 (m, 1H), 2.86 (m, 1H); ^{13}C NMR (CDCl_3 , 400 MHz) δ 166.7, 148.1, 147.5, 142.1, 134.2, 129.9, 129.0, 128.8, 119.7, 108.2, 106.5, 101.1, 90.8, 86.9, 73.4, 68.9, 53.2, 53.1, 52.1; HRMS m/z 423.0884 $[\text{M}+\text{Na}]^+$ (calcd for $\text{C}_{21}\text{H}_{20}\text{NaO}_6\text{S}$, 423.0878).



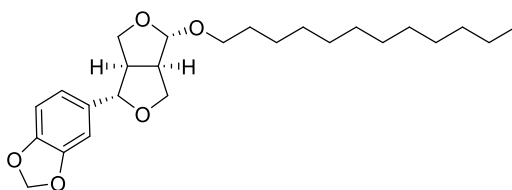
(1*R*, 2*R*, 5*S*, 6*S*)-6-(3,4-methylenedioxyphenyl)-
2-octyloxy-3,7-dioxabicyclo[3.3.0]octane (3.3A)

3.3A: Following the general procedure, reaction of **1.36** (16 mg, 0.06 mmol), 1-octanol (30 μ L, 0.18 mmol) in acetonitrile (0.6 mL) after 8 h yielded **3.3A** (18 mg, 78%) as a pale yellow oil; ^1H NMR (CDCl_3 , 400 MHz) δ 6.85 (d, $J = 0.8$ Hz, 1H), 6.80 (dd, $J = 8.0$, 1.2 Hz, 1H), 6.78 (d, $J = 7.6$ Hz, 1H), 5.94 (s, 2H), 4.93 (s, 1H), 4.38 (d, $J = 8.8$ Hz, 1H), 4.34 (t, $J = 7.2$ Hz, 1H), 3.97 (dd, $J = 8.8$, 6.0 Hz, 1H), 3.85 (d, $J = 8.8$ Hz, 1H), 3.65 (m, 1H), 3.55 (dd, $J = 8.8$, 7.6 Hz, 1H), 3.37 (m, 1H), 3.04 (q, $J = 8.8$ Hz, 1H), 2.81 (q, $J = 9.2$ Hz, 1H), 1.59-1.54 (m, 4H), 1.27 (overlap, 8H), 0.87 (t, $J = 6.8$ Hz, 3H); ^{13}C NMR (CDCl_3 , 400 MHz) δ 148.1, 147.4, 134.9, 119.8, 108.3, 107.6, 106.7, 101.2, 87.2, 71.6, 69.0, 67.6, 53.2, 52.9, 31.9, 29.8, 29.5, 29.4, 26.3, 22.8, 14.2; HRMS m/z 385.1997 $[\text{M}+\text{Na}]^+$ (calcd for $\text{C}_{21}\text{H}_{30}\text{NaO}_5$, 385.1991).



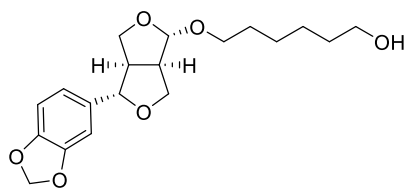
(1*R*, 2*R*, 5*S*, 6*S*)-2-decyloxy-6-(3,4-methylenedioxyphenyl)-
3,7-dioxabicyclo[3.3.0]octane (**3.3B**)

3.3B: Following the general procedure, reaction of **1.36** (11.3 mg, 0.05 mmol), 1-decanol (30 μ L, 0.15 mmol) in acetonitrile (0.5 mL) after 8 h yielded **3.3B** (20.4 mg, quantitative yield) as a colorless oil; ^1H NMR (CDCl_3 , 400 MHz) ^1H NMR (CDCl_3 , 400 MHz) δ 6.85 (d, $J = 1.6$ Hz, 1H), 6.81-6.75 (m, 2H), 5.94 (s, 2H), 4.93 (s, 1H), 4.38 (d, $J = 8.8$ Hz, 1H), 4.35 (t, $J = 7.6$ Hz, 1H), 3.97 (dd, $J = 9.2, 6.0$ Hz, 1H), 3.85 (d, $J = 8.4$ Hz, 1H), 3.65 (m, 1H), 3.55 (dd, $J = 8.8, 7.6$ Hz, 1H), 3.36 (m, 1H), 3.04 (q, $J = 8.8$ Hz, 1H), 2.81 (m, 1H), 1.60-1.52 (m, 4H), 1.26 (overlap, 12H), 0.87 (t, $J = 6.8$ Hz, 3H); ^{13}C NMR (CDCl_3 , 400 MHz) δ 148.1, 147.4, 134.9, 119.8, 108.3, 107.6, 106.7, 101.2, 87.2, 71.6, 69.0, 67.6, 53.3, 52.9, 32.0, 29.8, 29.7, 29.7, 29.6, 29.4, 26.3, 22.8, 14.2; HRMS m/z 413.2306 $[\text{M}+\text{Na}]^+$ (calcd for $\text{C}_{23}\text{H}_{34}\text{NaO}_5$, 413.2304).



(1*R*, 2*R*, 5*S*, 6*S*)-2-dodecyloxy-6-(3,4-methylenedioxyphenyl)-
3,7-dioxabicyclo[3.3.0]octane (**3.3C**)

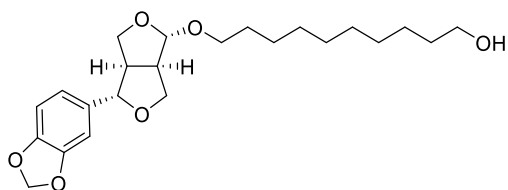
3.3C: Following the general procedure, reaction of **1.36** (13.6 mg, 0.05 mmol), 1-dodecanol (34 μ L, 0.15 mmol) in acetonitrile (0.5 mL) after 8 h yielded **3.3C** (20 mg, quantitative yield) as a colorless oil; ^1H NMR (CDCl_3 , 400 MHz) δ 6.85 (d, J = 1.2 Hz, 1H), 6.81-6.75 (m, 2H), 5.94 (s, 2H), 4.93 (s, 1H), 4.38 (d, J = 8.8 Hz, 1H), 4.34 (t, J = 7.6 Hz, 1H), 3.97 (dd, J = 9.2, 6.0 Hz, 1H), 3.85 (d, J = 8.8 Hz, 1H), 3.65 (m, 1H), 3.54 (dd, J = 8.8, 7.6 Hz, 1H), 3.36 (m, 1H), 3.04 (q, J = 8.4 Hz, 1H), 2.81 (m, 1H), 1.60-1.52 (m, 4H), 1.25 (overlap, 16H), 0.88 (t, J = 6.8 Hz, 3H); ^{13}C NMR (CDCl_3 , 400 MHz) δ 148.1, 147.4, 135.0, 119.8, 108.3, 107.6, 106.8, 101.2, 87.2, 71.6, 69.1, 67.6, 53.3, 53.0, 32.1, 29.8, 29.8, 29.8, 29.7, 29.6, 29.5, 26.4, 22.8, 14.2; HRMS m/z 441.2618 $[\text{M}+\text{Na}]^+$ (calcd for $\text{C}_{25}\text{H}_{38}\text{NaO}_5$, 441.2617).



(1*R*, 2*R*, 5*S*, 6*S*)-2-(6-hydroxy)hexyloxy-

6-(3,4-methylenedioxyphenyl)-3,7-dioxabicyclo[3.3.0]octane (**3.3D**)

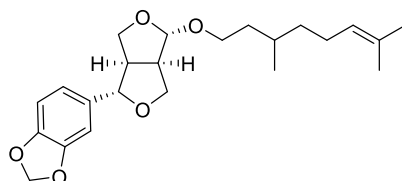
3.3D: Following the general procedure, reaction of **1.36** (6.5 mg, 0.03 mmol), 1,6-hexanediol (11 mg, 0.09 mmol) in acetonitrile (0.2 mL) after 8 h yielded **3.3D** (12 mg, quantitative yield) as a colorless oil; ^1H NMR (CDCl_3 , 400 MHz) δ 6.85 (s, 1H), 6.81-6.76 (m, 2H), 5.94 (s, 2H), 4.93 (s, 1H), 4.38 (d, $J = 8.8$ Hz, 1H), 4.35 (t, $J = 8.0$ Hz, 1H), 3.96 (dd, $J = 9.2, 6.0$ Hz, 1H), 3.85 (d, $J = 8.8$ Hz, 1H), 3.69-3.62 (m, 3H), 3.55 (dd, $J = 8.8, 8.0$ Hz, 1H), 3.38 (m, 1H), 3.03 (q, $J = 8.4$ Hz, 1H), 2.81 (q, $J = 8.8$ Hz, 1H), 1.59-1.56 (m, 2H), 1.43-1.33 (m, 6H); ^{13}C NMR (CDCl_3 , 400 MHz) δ 148.1, 147.4, 134.9, 119.8, 108.3, 107.6, 106.7, 101.2, 87.2, 71.5, 69.1, 67.4, 64.6, 53.2, 52.9, 32.8, 29.7, 28.7, 25.9; HRMS m/z 373.1630 $[\text{M}+\text{Na}]^+$ (calcd for $\text{C}_{19}\text{H}_{26}\text{NaO}_6$, 373.1627).



(1*R*, 2*R*, 5*S*, 6*S*)-2-(10-hydroxy)decyloxy-

6-(3,4-methylenedioxyphenyl)-3,7-dioxabicyclo[3.3.0]octane (**3.3E**)

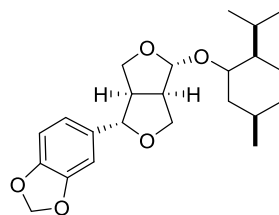
3.3E: Following the general procedure, reaction of **1.36** (7 mg, 0.03 mmol), 1,10-decanediol (16 mg, 0.09 mmol) in acetonitrile (0.3 mL) after 8 h yielded **3.3E** (13 mg, quantitative yield) as a colorless oil; ^1H NMR (CDCl_3 , 400 MHz) δ 6.85 (s, 1H), 6.81-6.75 (m, 2H), 5.94 (s, 2H), 4.93 (s, 1H), 4.38 (d, $J = 8.8$ Hz, 1H), 4.35 (t, $J = 7.6$ Hz, 1H), 3.97 (dd, $J = 8.8, 6.0$ Hz, 1H), 3.85 (d, $J = 8.8$ Hz, 1H), 3.68-3.62 (m, 3H), 3.55 (dd, $J = 8.8, 7.6$ Hz, 1H, H-8), 3.37 (m, 1H), 3.04 (q, $J = 8.4$ Hz, 1H), 2.82 (q, $J = 8.8$ Hz, 1H), 1.57-1.54 (m, 2H), 1.28-1.25 (m, 14H); ^{13}C NMR (CDCl_3 , 400 MHz) δ 148.1, 147.4, 134.9, 119.8, 108.3, 107.6, 106.7, 101.2, 87.2, 71.6, 69.1, 67.6, 63.2, 53.2, 52.9, 32.9, 29.8, 29.8, 29.7, 29.6, 29.5, 26.3, 25.9; HRMS m/z 429.2261 [$\text{M}+\text{Na}$] $^+$ (calcd for $\text{C}_{23}\text{H}_{34}\text{NaO}_6$, 429.2253).



(1*R*, 2*R*, 5*S*, 6*S*)-6-(3,4-methylenedioxyphenyl)-

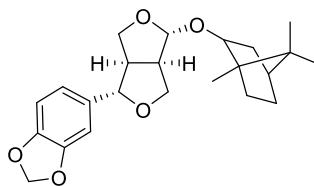
2-[3,7-dimethyloct-6-en-1-yloxy]-3,7-dioxabicyclo[3.3.0]octane (3.3F)

3.3F: Following the general procedure, reaction of **1.36** (27.8 mg, 0.11 mmol), β -citronellol (60 μ L, 0.33 mmol) in acetonitrile (1 mL) after 8 h yielded **3.3F** (38 mg, 88%) as a colorless oil; $^1\text{H NMR}$ (CDCl_3 , 400 MHz) δ 6.85 (s, 1H), 6.81-6.76 (m, 2H), 5.94 (s, 2H), 5.09 (t, $J = 6.8$ Hz, 1H), 4.93 (s, 1H), 4.38 (d, $J = 8.8$ Hz, 1H), 4.35 (t, $J = 7.2$ Hz, 1H), 3.97 (dd, $J = 9.8, 6.0$ Hz, 1H), 3.85 (d, $J = 8.8$ Hz, 1H), 3.70 (m, 1H), 3.55 (dd, $J = 8.8, 7.6$ Hz, 1H), 3.40 (m, 1H), 3.03 (q, $J = 9.2$ Hz, 1H), 2.81 (q, $J = 6.8$ Hz, 1H), 2.03-1.91 (m, 2H), 1.68 (s, 3H), 1.59 (s, 3H), 1.54 (m, 1H), 1.41-1.30 (m, 3H), 1.16 (m, 1H), 0.89 (d, $J = 6.4$ Hz, 3H); $^{13}\text{C NMR}$ (CDCl_3 , 400 MHz) δ 148.1, 147.4, 135.0, 131.3, 124.9, 119.8, 108.3, 107.5, 106.7, 101.2, 87.2, 71.6, 69.1, 65.9, 53.3, 53.0, 37.4, 36.8, 29.8, 25.8, 25.6, 19.8, 17.8; HRMS m/z 411.2155 $[\text{M}+\text{Na}]^+$ (calcd for $\text{C}_{23}\text{H}_{32}\text{NaO}_5$, 411.2147).



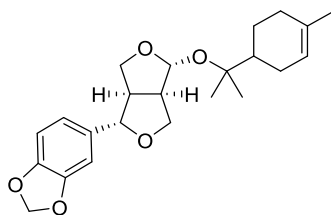
(1*R*, 2*R*, 5*S*, 6*S*)-2-[(1*R*, 2*S*, 5*R*)-2-isopropyl-5-methylcyclohexyloxy]-6-(3,4-methylenedioxyphenyl)-3,7-dioxabicyclo[3.3.0]octane (**3.3G**)

3.3G: Following the general procedure, reaction of **1.36** (12.6 mg, 0.05 mmol), menthol (23 mg, 0.15 mmol) in acetonitrile (0.5 mL) after 8 h yielded **3.3G** (20 mg, quantitative yield) as a colorless oil; ^1H NMR (CDCl_3 , 400 MHz) δ 6.84 (s, 1H), 6.81-6.75 (m, 2H), 5.94 (s, 2H), 5.01 (s, 1H), 4.38 (d, $J = 8.8$ Hz, 1H), 4.35 (dd, $J = 8.4, 6.4$ Hz, 1H), 4.05 (dd, $J = 9.2, 6.4$ Hz, 1H), 3.84 (d, $J = 8.8$ Hz, 1H), 3.54 (dd, $J = 9.2, 7.6$ Hz, 1H), 3.31 (ddd, $J = 10.4, 10.0, 4.4$ Hz, 1H), 3.04 (q, $J = 8.0$ Hz, 1H), 2.83 (q, $J = 8.8$ Hz, 1H), 2.13-2.04 (m, 2H), 1.63-1.59 (m, 3H), 1.40 (m, 1H), 1.17 (m, 1H), 1.01-0.94 (m, 2H), 0.91 (d, $J = 4.8$ Hz, 3H), 0.89 (d, $J = 4.4$ Hz, 3H), 0.78 (d, $J = 6.8$ Hz, 3H); ^{13}C NMR (CDCl_3 , 400 MHz) δ 148.1, 147.4, 135.1, 119.8, 109.4, 108.3, 106.7, 101.2, 87.3, 79.7, 71.7, 69.1, 53.4, 52.9, 48.9, 43.6, 34.5, 31.9, 25.9, 23.5, 22.4, 21.3, 16.5; HRMS m/z 411.2143 $[\text{M}+\text{Na}]^+$ (calcd for $\text{C}_{23}\text{H}_{32}\text{NaO}_5$, 411.2147).



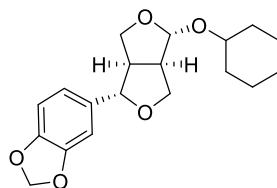
(1*R*, 2*R*, 5*S*, 6*S*)-2-(4,7,7-trimethylbicyclo[2.2.1]heptan-3-yloxy)-
6-(3,4-methylenedioxyphenyl)-3,7-dioxabicyclo[3.3.0]octane (**3.3H**)

3.3H: Following the general procedure, reaction of **1.36** (20 mg, 0.08 mmol), borneol (37 mg, 0.24 mmol) in acetonitrile (0.8 mL) after 8 h yielded **3.3H** (23 mg, 75%) as a colorless oil; ^1H NMR (CDCl_3 , 400 MHz) δ 6.85 (s, 1H), 6.82-6.76 (m, 2H), 5.95 (s, 2H), 4.95 (s, 1H), 4.38 (d, $J = 8.8$ Hz, 1H), 4.34 (dd, $J = 12.8, 8.0$ Hz, 1H), 3.98 (dd, $J = 8.8, 6.0$ Hz, 1H), 3.82 (d, $J = 8.8$ Hz, 1H), 3.76 (brd, $J = 8.0$ Hz, 1H), 3.53 (dd, $J = 8.8, 7.6$ Hz, 1H), 3.07 (q, $J = 8.4$ Hz, 1H), 2.84 (q, $J = 9.2$ Hz, 1H), 2.22 (m, 1H), 1.89 (m, 1H), 1.68-1.58 (m, 2H), 1.22-1.15 (m, 2H), 1.01 (dd, $J = 13.6, 3.2$ Hz, 1H), 0.85 (s, 3H), 0.84 (s, 3H), 0.83 (s, 3H); ^{13}C NMR (CDCl_3 , 400 MHz) δ 148.1, 147.4, 135.1, 119.8, 109.2, 108.3, 106.8, 101.2, 87.2, 84.0, 71.7, 69.0, 53.3, 53.1, 49.4, 47.5, 45.2, 37.9, 28.4, 26.9, 19.9, 18.9, 14.0; HRMS m/z 409.1992 $[\text{M}+\text{Na}]^+$ (calcd for $\text{C}_{23}\text{H}_{30}\text{NaO}_5$, 409.1991).



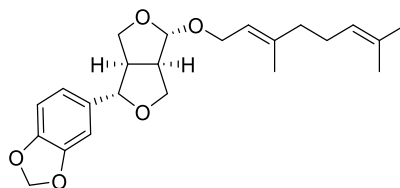
(1*R*, 2*R*, 5*S*, 6*S*)-2-[2-(4-methylcyclohex-3-en-1-yl)propan-2-yloxy]-6-(3,4-methylenedioxyphenyl)-3,7-dioxabicyclo[3.3.0]octane (**3.3I**)

3.3I: Following the general procedure, reaction of **1.36** (48 mg, 0.19 mmol), α -terpeneol (95 μ L, 0.57 mmol) in acetonitrile (2 mL) after 8 h yielded **3.3I** (60 mg, 81%) as a colorless oil; ^1H NMR (CDCl_3 , 400 MHz) δ 6.84 (d, $J = 1.6$ Hz, 1H), 6.80-6.77 (m, 2H), 5.94 (s, 2H), 5.37 (brs, 1H), 5.24 (s, 1H), 4.36 (d, $J = 5.6$ Hz, 1H), 4.33 (dd, $J = 6.0, 4.0$ Hz, 1H), 4.05 (dd, $J = 9.2, 6.0$ Hz, 1H), 3.80 (d, $J = 8.8$ Hz, 1H), 3.57 (dd, $J = 8.8, 7.2$ Hz, 1H), 2.95 (m, 1H), 2.83 (m, 1H), 2.04-1.96 (m, 4H), 1.83-1.81 (m, 3H), 1.64 (s, 3H), 1.57 (s, 3H), 1.19 (s, 3H); ^{13}C NMR (CDCl_3 , 400 MHz) δ 148.1, 147.4, 135.3, 134.1, 121.0, 119.7, 108.3, 106.7, 102.3, 101.2, 87.2, 78.5, 71.8, 69.1, 54.2, 53.2, 44.2, 31.2, 26.9, 24.3, 24.0, 23.6, 23.5; HRMS m/z 409.1992 $[\text{M}+\text{Na}]^+$ (calcd for $\text{C}_{23}\text{H}_{30}\text{NaO}_5$, 409.1991).



(1*R*, 2*R*, 5*S*, 6*S*)-2-cyclohexyloxy-6-(3,4-methylenedioxyphenyl)-
3,7-dioxabicyclo[3.3.0]octane (**3.3J**)

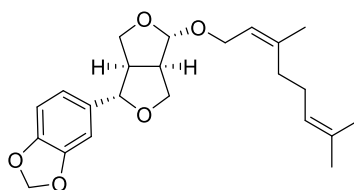
3.3J: Following the general procedure, reaction of **1.36** (12.9 mg, 0.05 mmol), cyclohexanol (16 μ L, 0.15 mmol) in acetonitrile (0.5 mL) after 8 h yielded **3.3J** (16 mg, 93%) as a colorless oil; ^1H NMR (CDCl_3 , 400 MHz) δ 6.85 (d, J = 0.8 Hz, 1H), 6.81-6.75 (m, 2H), 5.94 (s, 2H), 4.93 (s, 1H), 4.37 (d, J = 8.8 Hz, 1H), 4.34 (dd, J = 8.0, 5.6 Hz, 1H), 4.01 (dd, J = 8.8, 6.0 Hz, 1H), 3.83 (d, J = 8.8 Hz, 1H), 3.55 (m, 2H), 3.02 (q, J = 8.4 Hz, 1H), 2.83 (m, 1H), 1.89-1.52 (m, 10H); ^{13}C NMR (CDCl_3 , 400 MHz) δ 148.1, 147.4, 135.0, 119.8, 108.3, 106.8, 105.6, 101.2, 87.3, 74.9, 71.6, 69.0, 53.5, 53.0, 34.0, 32.0, 25.8, 24.5, 24.4; HRMS m/z 355.1527 $[\text{M}+\text{Na}]^+$ (calcd for $\text{C}_{19}\text{H}_{24}\text{NaO}_5$, 355.1521).



(1*R*, 2*R*, 5*S*, 6*S*)-2-[(2*E*)-3,7-dimethylocta-2,6-dien-1-yloxy]-

6-(3,4-methylenedioxyphenyl)-3,7-dioxabicyclo[3.3.0]octane (**3.3K**)

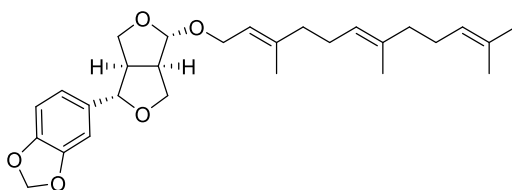
3.3K: Following the general procedure, reaction of **1.36** (17 mg, 0.07 mmol), geraniol (37 μ L, 0.21 mmol) in acetonitrile (0.7 mL) after 8 h yielded **3.3K** (18 mg, 69%) as a colorless oil; ^1H NMR (CDCl_3 , 400 MHz) δ 6.85 (s, 1H), 6.81-6.75 (m, 2H), 5.94 (s, 2H), 5.33 (t, $J = 6.4$ Hz, 1H), 5.09 (m, 1H), 4.99 (s, 1H), 4.38 (d, $J = 9.2$ Hz, 1H), 4.35 (dd, $J = 8.4, 7.2$ Hz, 1H), 4.17 (dd, $J = 12.0, 6.8$ Hz, 1H), 4.02-3.97 (m, 2H), 3.87 (d, $J = 8.8$ Hz, 1H), 3.55 (dd, $J = 9.2, 7.6$ Hz, 1H), 3.06 (m, 1H), 2.82 (m, 1H), 2.10-2.04 (m, 4H), 1.68 (s, 6H), 1.60 (s, 3H); ^{13}C NMR (CDCl_3 , 400 MHz) δ 148.1, 147.4, 141.1, 134.9, 131.8, 124.1, 120.2, 119.8, 108.3, 106.8, 106.7, 101.2, 87.2, 71.6, 69.1, 63.6, 53.3, 53.0, 39.8, 26.5, 25.8, 17.8, 16.6; HRMS m/z 409.1992 $[\text{M}+\text{Na}]^+$ (calcd for $\text{C}_{23}\text{H}_{30}\text{NaO}_5$, 409.1991).



(1*R*, 2*R*, 5*S*, 6*S*)-6-(3,4-methylenedioxyphenyl)-

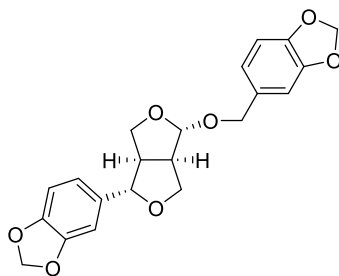
2-[(2*Z*)-3,7-dimethylocta-2,6-dien-1-yloxy]-3,7-dioxabicyclo[3.3.0]octane (**3.3L**)

3.3L: Following the general procedure, reaction of **1.36** (105.8 mg, 0.42 mmol), nerol (222 μ L, 1.26 mmol) in acetonitrile (4 mL) after 8 h yielded **3.3L** (134 mg, 82%) as a colorless oil; ^1H NMR (CDCl_3 , 400 MHz) δ 6.85 (s, 1H), 6.79-6.75 (m, 2H), 5.94 (s, 2H), 5.34 (m, 1H), 5.09 (brs, 1H), 4.98 (s, 1H), 4.38 (d, $J = 9.2$ Hz, 1H), 4.34 (dd, $J = 12.4, 7.6$ Hz, 1H), 4.16 (m, 1H), 4.01-3.94 (m, 2H), 3.86 (d, $J = 8.8$ Hz, 1H), 3.54 (m, 1H), 3.05 (m, 1H), 2.80 (m, 1H), 2.08 (brs, 4H), 1.75 (s, 3H), 1.68 (s, 3H), 1.60 (s, 3H); ^{13}C NMR (CDCl_3 , 400 MHz) δ 148.1, 147.4, 141.4, 134.9, 132.1, 123.9, 121.1, 119.8, 108.3, 106.7, 106.7, 101.2, 87.1, 71.5, 69.1, 63.3, 53.2, 53.0, 32.3, 26.9, 25.8, 23.7, 17.8; HRMS m/z 409.1997 $[\text{M}+\text{Na}]^+$ (calcd for $\text{C}_{23}\text{H}_{30}\text{NaO}_5$, 409.1991).



(1*R*, 2*R*, 5*S*, 6*S*)-2-[(2*E*, 6*E*)-3,7,11-trimethyldodeca-2,6,10-trien-1-yloxy]-6-(3,4-methylenedioxyphenyl)-3,7-dioxabicyclo[3.3.0]octane (**3.3M**)

3.3M: Following the general procedure, reaction of **1.36** (77.2 mg, 0.31 mmol), *trans,trans*-farnesol (235 μ L, 0.93 mmol) in acetonitrile (3 mL) after 8 h yielded **3.3M** (118 mg, 84%) as a colorless oil; ^1H NMR (CDCl_3 , 400 MHz) δ 6.85 (s, 1H), 6.79-6.75 (m, 2H), 5.94 (s, 2H), 5.33 (m, 1H), 5.10-5.07 (m, 2H), 4.98 (s, 1H), 4.38 (d, $J = 8.8$ Hz, 1H), 4.34 (dd, $J = 9.6, 7.6$ Hz, 1H), 4.18 (m, 1H), 4.01-3.98 (m, 2H), 3.86 (d, $J = 8.8$ Hz, 1H), 3.54 (dd, $J = 8.8, 8.8$ Hz, 1H), 3.06 (m, 1H), 2.81 (m, 1H), 2.12-1.95 (m, 8H), 1.69 (s, 3H), 1.67 (s, 3H), 1.59 (s, 6H); ^{13}C NMR (CDCl_3 , 400 MHz) δ 148.1, 147.4, 141.1, 135.5, 134.9, 131.4, 124.5, 123.9, 120.2, 119.8, 108.3, 106.7, 106.7, 101.2, 87.1, 71.5, 69.1, 63.6, 53.2, 53.0, 39.8, 39.7, 26.9, 26.4, 25.8, 17.8, 16.6, 16.1; HRMS m/z 477.2617 [$\text{M}+\text{Na}$] $^+$ (calcd for $\text{C}_{28}\text{H}_{38}\text{NaO}_5$, 477.2617).



(1*R*, 2*R*, 5*S*, 6*S*)- 2-(3,4-(methylenedioxy)benzyloxy)-
6-(3,4-methylenedioxyphenyl)-3,7-dioxabicyclo[3.3.0]octane (**3.3N**)

3.3N: Following the general procedure, reaction of 1.36 (63.3 mg, 0.25 mmol), Piperonyl alcohol (235 μ L, 0.51 mmol) in acetonitrile (3 mL) after 8 h yielded **3.3N** (43.6 mg, 44.8%) as a white solid; ^1H NMR (400 MHz, CDCl_3) δ 6.80 (m, 6H), 5.95 (d, $J = 1.0$ Hz, 4H), 5.02 (s, 1H), 4.60 (d, $J = 11.4$ Hz, 1H), 4.42 – 4.31 (m, 3H), 4.03 (dd, $J = 9.1, 6.0$ Hz, 1H), 3.90 (d, $J = 9.0$ Hz, 1H), 3.57 – 3.51 (m, 1H), 3.09 (dd, $J = 16.9, 8.7$ Hz, 1H), 2.83 (dd, $J = 15.5, 6.7$ Hz, 1H); ^{13}C NMR (101 MHz, CDCl_3) δ 148.1, 134.8, 131.6, 121.9, 119.8, 108.9, 108.3, 106.7, 106.5, 101.2, 101.2, 87.2, 71.5, 69.3, 68.9, 53.2, 52.9.

3.5.4 X-ray Crystallographic Analysis of 3.2r and *epi*-3.2r

Single crystal X-ray diffraction data were collected at 296(2) K on a Bruker X8 PROSPECTOR KAPPA CCD diffractometer using an I μ S X-ray microfocus source with multilayer mirrors, yielding intense monochromatic Cu-K α radiation (λ = 1.54178 Å) for 3.2r and on a Bruker X8 APEX II KAPPA CCD diffractometer using graphite monochromatized Mo-K α radiation (λ = 0.71073 Å) for *epi*-3.2r. The structures were solved using SHELXTL XT 2013/1 [18], expanded using difference Fourier method, and refined using full-matrix least squares on F2 with SHELXTL XLMP 2014/7 [19]. Absolute configurations of the two compounds were ambiguously determined with the estimated Flack parameters (x's) [20] that are statistically close to zero; the corresponding respective values are 0.040(19) and 0.026(12). Details of crystal data and refinement parameters are listed in Table 3.4.

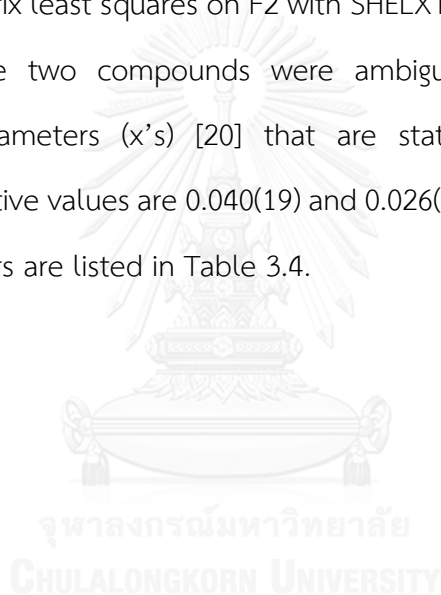


Table 3.4 Crystal data and refinement parameters of compounds **3.2r** and **epi-3.2r**.

	3.2r	epi-3.2r
Crystal habit	Thin plate, colorless	Thin plate, colorless
Crystal size (mm ³)	0.01×0.24×0.34	0.01×0.18×0.20
Empirical formula	C ₁₉ H ₁₈ O ₅ S	C ₂₆ H ₂₃ Br O ₅ S
<i>FW</i>	358.40	527.41
Crystal system	Monoclinic	Triclinic
Space group	<i>P</i> ₂ ₁ (no. 4)	<i>P</i> 1 (no. 1)
<i>a</i> [Å]	6.1485(2)	6.7501(13)
<i>b</i> [Å]	7.6475(2)	8.8663(18)
<i>c</i> [Å]	17.9288(4)	20.864(4)
α [°]	90	95.858(5)
β [°]	90.711(1)	94.023(5)
γ [°]	90	111.089(5)
<i>V</i> [Å ³]	842.96(4)	1151.3(4)
<i>Z</i>	2	2
ρ_{calcd} [Mg m ⁻³]	1.412	1.521
μ [mm ⁻¹]	1.948	1.912
<i>F</i> (000)	376	540
Diffractionmeter	Bruker X8 Prospector KAPPA CCD	Bruker X8 APEX II KAPPA CCD
Radiation (λ , Å)	CuK α (1.54178)	MoK α (0.71073)
Resolution (Å)	0.83	0.83
Temperature (K)	296(2)	296(2)
$2\theta_{\text{max}}$	136.8	50.6
Completeness (%)	99.6	99.7
Reflns collected/ unique/ > $2\sigma(I)$	6974 / 3010 / 2417	20477 / 8319 / 3229
<i>R</i> _{int}	0.0522	0.0943
Data / parameters	3010 / 227	8319 / 595
Goodness on fit	1.100	0.921
<i>R</i> ₁ , ^a <i>wR</i> ₂ ^b [<i>I</i> > $2\sigma(I)$]	0.0558, 0.1403	0.0584, 0.0827
<i>R</i> ₁ , <i>wR</i> ₂ [all data]	0.0672, 0.1535	0.2047, 0.1200
Flack parameter (<i>x</i>)	0.040(19)	0.026(12)
$\Delta\rho$ (e Å ⁻³)	-0.39, 0.21	-0.28, 0.28

^a $R = \sum ||F_o| - |F_c|| / \sum |F_o|$; ^b $wR = \sum \{w(F_o^2 - F_c^2)^2 / \sum w(F_o^2)^2\}^{1/2}$.

CHAPTER IV

MECHANISTIC INVESTIGATION OF ALKOXYSAMIN (3.3) FORMATION

Generally, hemiacetals are susceptible to protonation under strong acid condition to generate oxocarbenium ion [21]. The attack of nucleophile on both sides of oxocarbenium ion would produce a mixture of two diastereomers in an equal amount. The aforementioned phenomena were observed in the reaction between samin and phenolics. However, high stereoselectivity was noticed in reaction between samin and thiols. Moreover, the reaction of samin and alcohols merely provide a single stereomeric product. Therefore, the mechanism of unexpected of alkoxysamin formations (**3.3**) were carefully investigate by a model reaction monitored by ^1H NMR spectroscopy and computational calculation.

4.1 Mechanistic investigation on 3.3 formation

The model reaction monitored by ^1H NMR was set by treatment of samin (**1.36**) with CD_3OD as a representative nucleophile in the presence of deuterated-trifluoroacetic acid (TFA-d) (Figure 4.1). During time interval observation, we found that this reaction generated both **4.1** and *epi-4.1* with diverse ratios. The presence of compound **4.1** could be observed by singlet signal of H-2 at 4.88 ppm while its epimer (*epi-4.1*) showed H-2 as doublet signal at 4.95 ppm with $J = 5.4$ Hz. Noticeably, the depletion of starting samin could be monitored by signal integration at 5.29 ppm (Figure 4.1).

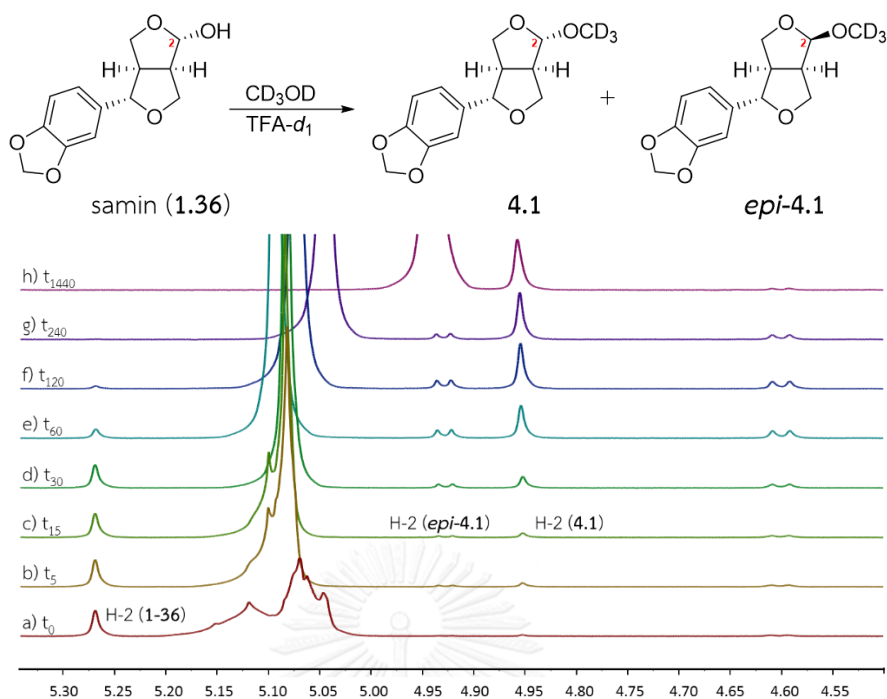


Figure 4.1 Overlaid ^1H NMR spectra of methoxy- d^3 samin, **4.1** and *epi*-**4.1** (CD_3OD), occurring in the reaction, for different times.

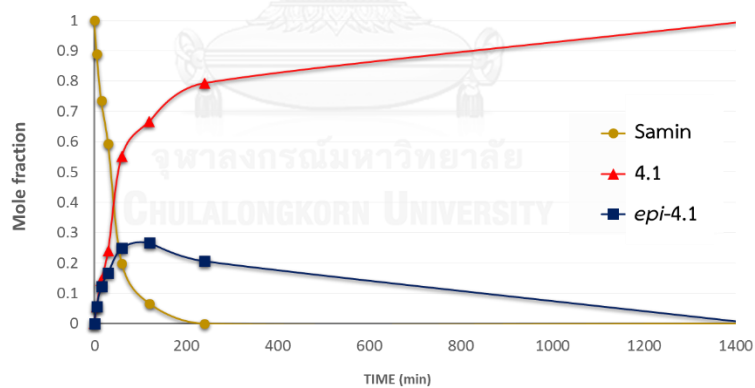
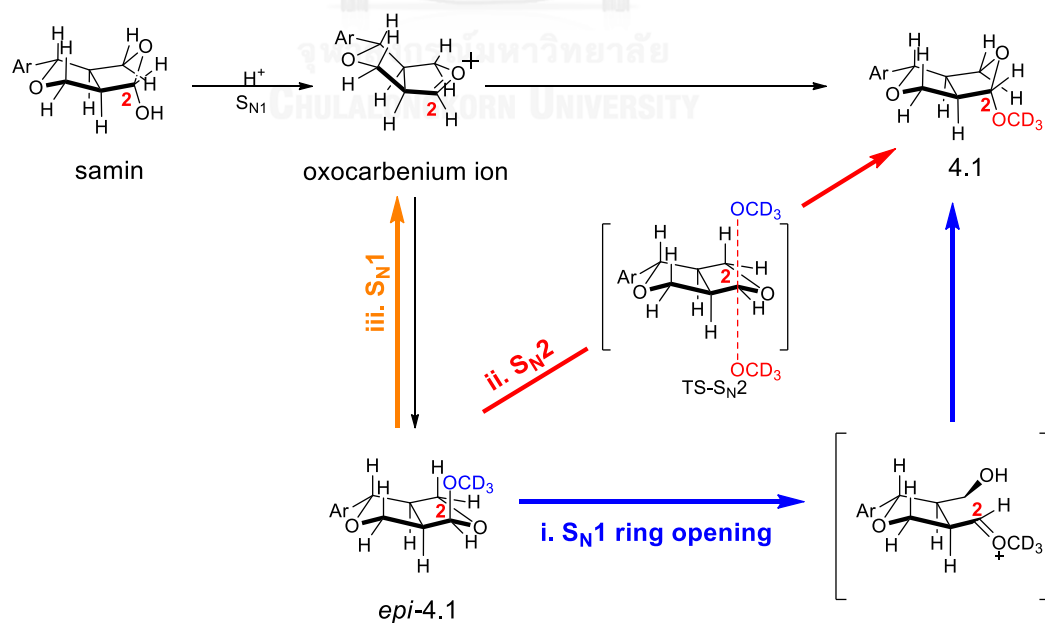


Figure 4.2 Time course of the evolution of samin (**1.36**), **4.1**, and *epi*-**4.1** in mole fraction by ^1H NMR spectroscopy (400 MHz).

Figure 4.2 showed time course of the evolution of **1.36**, **4.1**, and *epi*-**4.1** in mole fraction by integration of ^1H NMR signals. After 5 min of reaction, **4.1** and *epi*-**4.1** were first detected in the ratio of 1:1 (**4.1**:*epi*-**4.1**), suggesting that the reaction proceed through $\text{S}_{\text{N}}1$ -like mechanism. Noticeably, compound **4.1** was predominantly generated

after 60 min as observed in unequal ratio with **epi-4.1** (1.5:1 to 2:1). Although the starting samin was completely consumed after 240 min of reaction, the mole fraction of **4.1** still increased while **epi-4.1** gradually declined. We inevitably hypothesized that **epi-4.1** would be transformed to **4.1**. This result could explain the absence of *epi*-alkoxysamins from reaction between samin and alcohols (section 3.3.); however exact mechanism on transformation of **epi-4.1** to **4.1** remain unclear.

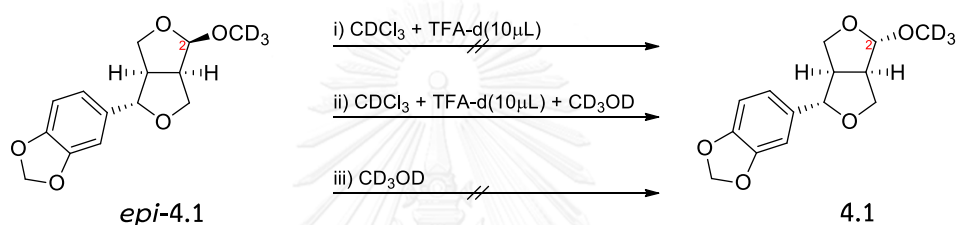
In fact, the epimerization of furofuran lignans has been rarely explained due to the complexity of their conformations. Mostly, isomerization on these compounds was claimed to occur through ring opening reaction [5, 22]. In this experiment, we proposed the mechanism for the transformation of **epi-4.1** to **4.1** as illustrated in Scheme 4.1. The first route involved the endocyclic C2–O cleavage facilitated by the electron-donating OCH₃ (Scheme 4.1 (i)). Alternatively, S_N2 mechanism using CD₃OD as nucleophile was also proposed (Scheme 4.1 (ii)). Lastly, compound **epi-4.1** might be reversed subsequently through oxocarbenium ion to form a stable compound **4.1** (Scheme 4.1 (iii)).



Scheme 4.1 Proposed mechanistic formation of **4.1** and **epi-4.1**

4.2 Mechanistic investigation on epimerization of alkoxyamin and *epi*-alkoxyamin

To investigate the reasonable epimerization mechanism from *epi*-**4.1** to **4.1**, we set the criteria to prove this mechanism using the diastereomeric pure *epi*-**4.1** as starting material (Scheme 4.2). After diastereomeric pure *epi*-**4.1** was gently produced and separated by high performance liquid chromatography (HPLC), it was dissolved in CDCl₃ in the presence of TFA-d (10μL) (Scheme 4.2).



Scheme 4.2 Epimerization of *epi*-**4.1** under different conditions i) - iii).

After ¹H NMR acquisition, there was neither change on *epi*-**4.1** nor presence of **4.1** formation under first condition. This observation suggested that epimerization through S_N1 ring opening (Scheme 4.1, route i) is impossible, and external nucleophile is presumably required. To verify this assumption, a small amount (20μL) of CD₃OD was added into the reaction NMR tube (Scheme 4.2, condition (ii)). After shaken well and left standing at room temperature for 10 min, the doublet signal H-2 of *epi*-**4.1** was completely disappeared while singlet signal H-2 of **4.1** was clearly observed. In addition, epimerization of *epi*-**4.1** to **4.1** could not proceed without the presence of strong acid as illustrate by dissolved *epi*-**4.1** in CD₃OD, and left it overnight without any change (Scheme 4.2, condition (iii)).

We further considered possible epimerization through route ii and iii. Protonation of *epi*-**4.1** to regenerate oxocarbenium ion (Scheme 4.1, route iii), which was subsequently attacked by nucleophile to produce **4.1**, is less likely to proceed

because there was no *epi-4.1* remained after reaction completed. In fact, if any reaction proceed with equilibrium, both starting material (*epi-4.1*) and product (**4.1**) would be detected in approximately equal amount.

- ii) *epi-4.1* with
TFA-d 10 μ L and CD₃OD 20 μ L
t = 10 min



- i) *epi-4.1*

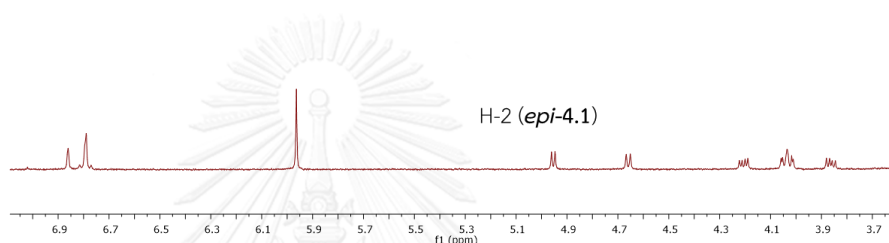
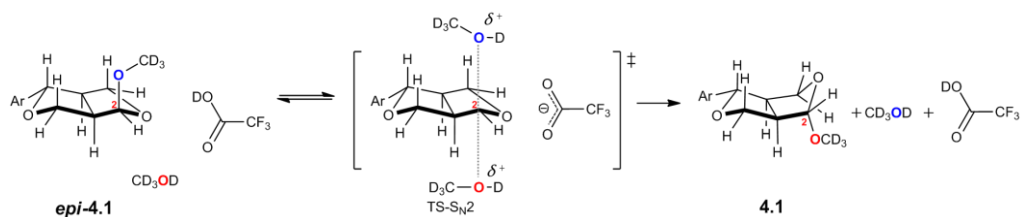


Figure 4.3 Overlaid ¹H NMR spectra of (i) *epi-4* and (ii) the treated *epi-4.1* with CD₃OD and TFA-d for 10 min.

From this result, the requiring of CD₃OD in the presence of TFA-d in epimerization reaction revealed that this reaction presumably undergo S_N2-like transition state contributed by acid-catalysis (Scheme 4.3).



Scheme 4.3 Proposed S_N2 reaction between *epi-4.1* and deuterated methanol (CD₃OD) with the present of deuterated-trifluoroacetic acid (CF₃CO₂D).

4.3 Computational calculations

The above experimental result was strikingly different with that of Johansson et al. [22], reported epimerization of furanosides by ring opening mechanism. Therefore, we then confirmed a transformational possibility of *epi-4.1* by bond distances observation of structures at transition state, using PCM/B3LYP/6-31+G(d,p) method to optimize the transition state structure (Figure 4.4). Table 4.1 lists major geometrical features of the transition state. Due to the nucleophile and leaving group are totally the same alcohols. C-O (breaking bond) and C-O^b (forming bond) bond distances at transition state were not significantly different. However, the slightly shorter new C-O^b (2.51 Å) bond inferred that there is more preferable bond formation than C-O bond. The short H^a-O bond (0.97 Å) also supported the -OCH₃ of *epi-4.1* prefer leaving to form free methanol molecule after receive proton from acid (TFA-d) rather than bonding with anomeric carbon (C-2). All illustrated evidences corroborated reasonably transformation of *epi-4.1* into **4.1** through S_N2-like transition state [10].

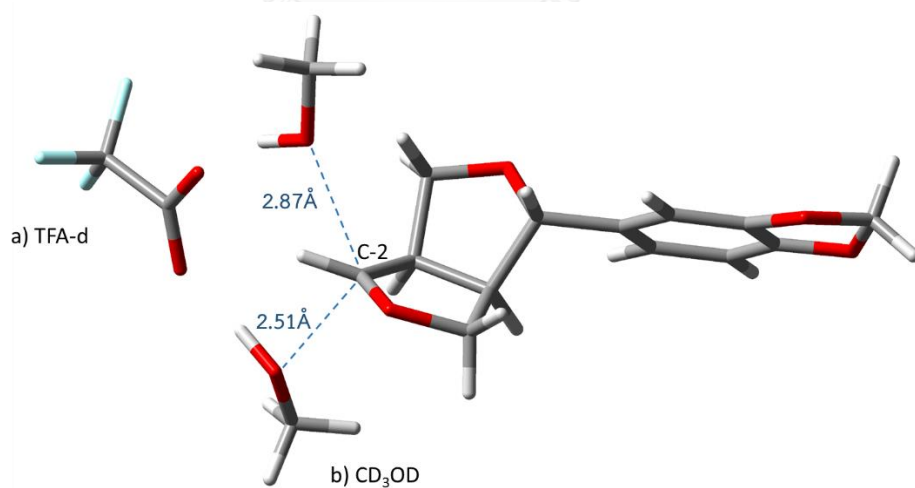


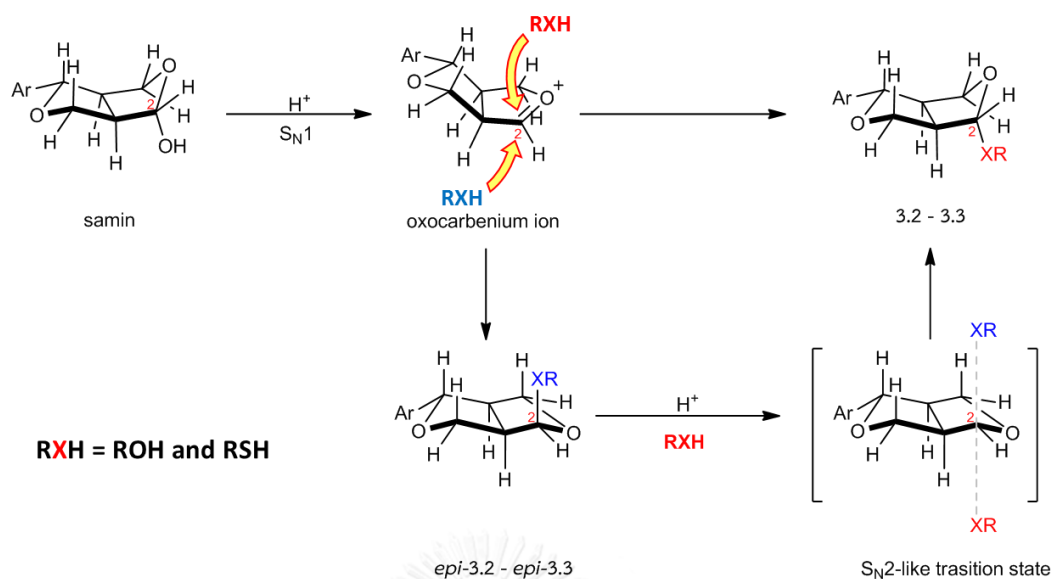
Figure 4.4 Transition-state model for the epimerization reaction of *epi-4.1*. (Note ^a Atoms of TFA-d and ^b Atoms of CD₃OD)

Table 4.1 Most important bond distances (in Å) and valence angle (in °) of transition state structure.

Coordinate/property		Transition state	
O ^a	H ^a	1.843	
H ^a	O	0.975	
O	C	2.868	
C	O ^b	2.517	
O ^b	H ^b	0.991	
H ^b	O ^a	1.715	
O	C	O ^b	123.1

^a Atoms of TFA-d ^b Atoms of CD₃OD

In summary, our stereoselective products from reaction between samin and alcohols as well as thiols were reasonably formed through S_N1-like mechanism by protonation of the hemiacetal center of samin to generate the corresponding oxocarbenium ion. Subsequent reaction of this oxocarbenium ion with the nucleophiles (**I-z** and **A-N**) then led to the observed only *exo,exo*-furofurans (**3.2-3.3**) either directly, or alternatively, by protonation of their epimers (*endo,exo*-furofurans) through the S_N2-like transition state contributed by acid-catalysis, likewise reaction sequence of model reaction monitoring which is exemplified in Scheme 4.4.



Scheme 4.4 A proposed mechanistic reaction of synthesis of stereoselective products (3.2 and 3.3).

4.4 Experimental section

4.4.1 Mechanistic investigation on 3.3 formation

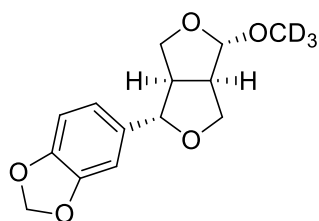
Solution of samin (7 mg, 0.027mmol), deuterated-trifluoroacetic acid (TFA-d, 20 μ L) and CD_3OD were sealed in NMR tube. At time interval, NMR proton spectra were recorded. Chemical shifts of all combinations in 1H NMR spectra obtained with Varian Mercury⁺ 400 were 5.29 ppm (1H, s) for samin, 4.95 ppm ($J= 5.4$, 1H, s) for *epi*-4.1 and 4.88 ppm (1H, s) for 4.1. Mole ratios were calculated from integration of NMR signals corresponding to samin, 4.1 and *epi*-4.1.

4.4.2 Mechanistic investigation on epimerization

4.4.2.1 Synthesis of 4.1 and *epi*-4.1

To a solution of samin (100 mg, 0.39 mmol) in acetonitrile (4 mL) was treated with methanol (0.80 mmol), acidic resin amberlyst-15 and molecular sieve, 4 Å. After stirring at 0°C for 1 h, the reaction was suddenly quenched, and evaporated to dryness.

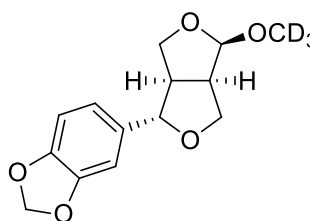
Reaction mixture was firstly purify by silica gel column chromatography followed by semi-preparative HPLC ($t_R = 45$ min, gradient of $\text{CH}_3\text{CN}/\text{H}_2\text{O}$).



(1*R*, 2*R*, 5*S*, 6*S*)-6-(3,4-

methylenedioxy-phenyl)-2-methyl- d_3 -
oxy-3,7-dioxabicyclo[3.3.0]octane

(4.1)



(1*R*, 2*S*, 5*S*, 6*S*)-6-(3,4-

methylenedioxy-phenyl)-2-methyl- d_3 -
oxy-3,7-dioxabicyclo[3.3.0]octane

(*epi*-4.1)

4.1; ^1H NMR (CDCl_3 , 400 MHz) δ 6.81 (d, $J = 17.9$ Hz, 3H), 5.96 (s, 2H), 4.95 (s, 1H), 4.44 (dd, $J = 21.2, 8.2$ Hz, 2H), 4.02 (dd, $J = 9.0, 5.9$ Hz, 1H), 3.95 (d, $J = 9.1$ Hz, 1H), 3.65 (dd, $J = 9.3, 7.6$ Hz, 1H), 3.11 (dd, $J = 16.5, 8.6$ Hz, 1H), 2.92 (dd, $J = 15.3, 7.2$ Hz, 1H).

***epi*-4.1;** ^1H NMR (CDCl_3 , 400 MHz) δ 6.80 (m, 3H), 5.94 (s, 2H), 4.93 (d, $J = 5.5$ Hz, 1H), 4.64 (d, $J = 6.5$ Hz, 1H), 4.18 (dd, $J = 8.9, 4.2$ Hz, 1H), 4.05 – 3.97 (m, 2H), 3.86 – 3.81 (m, 1H), 3.11 (m, 1H), 2.92 (m, 1H).

4.4.2.2 ^1H NMR monitoring section

Solution of compound ***epi*-4.1** (14 mM), in CDCl_3 were treated with two conditions, namely, deuterated-trifluoroacetic acid (TFA- d 10 μL) and TFA- d 10 μL along with deuterated methanol (CD_3OD , 20 μL), performed in NMR tube. ^1H NMR spectroscopy was applied to monitor the transformation under different condition of each compounds as well as ***epi*-4.1** in CD_3OD .

4.4.3 Computational section

The geometrical optimization and determination of product energy have been calculated using the density functional theory (DFT) with the popular hybrid method (B3LYP) with the 6-31+G(d,p) basis set. Methanol phase and the polarizable continuum model (PCM) calculations were carried out using the Gaussian09 package [23] with default convergence criteria.



CHAPTER V

CONCLUSION

Our synthesis approach is efficient to provide almost fifty furofuran lignans starting from naturally available sesamol. Key step of our methodology involved protonation of samidin to generate oxocarbenium ion, followed by nucleophilic addition. Three types of nucleophiles were used to investigate the feasibility of our designed synthetic method; namely, phenolics (ArOH), thiols (RSH), and alcohols (ROH). This synthesis strategy provided moderate to high yield of diastereomeric products except for those synthesized from samidin and alcohols which was further investigated by a time-course ^1H NMR technique. The result revealed that the stereoselective products were presumably obtained from nucleophilic addition of oxocarbenium ion, or alternatively, via protonation of their epimers through the $\text{S}_{\text{N}}2$ -like transition state which can be confirmed by structural energy and bond distances of optimized transition state structure, generated by Gaussian09.

This practical synthesis method of furofuran lignans suggested an opportunity of studying structural activity relationship would be improved. In addition, mechanistic insights make us easily controlled the producing of expected products in further research.

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APPENDIX

จุฬาลงกรณ์มหาวิทยาลัย
CHULALONGKORN UNIVERSITY

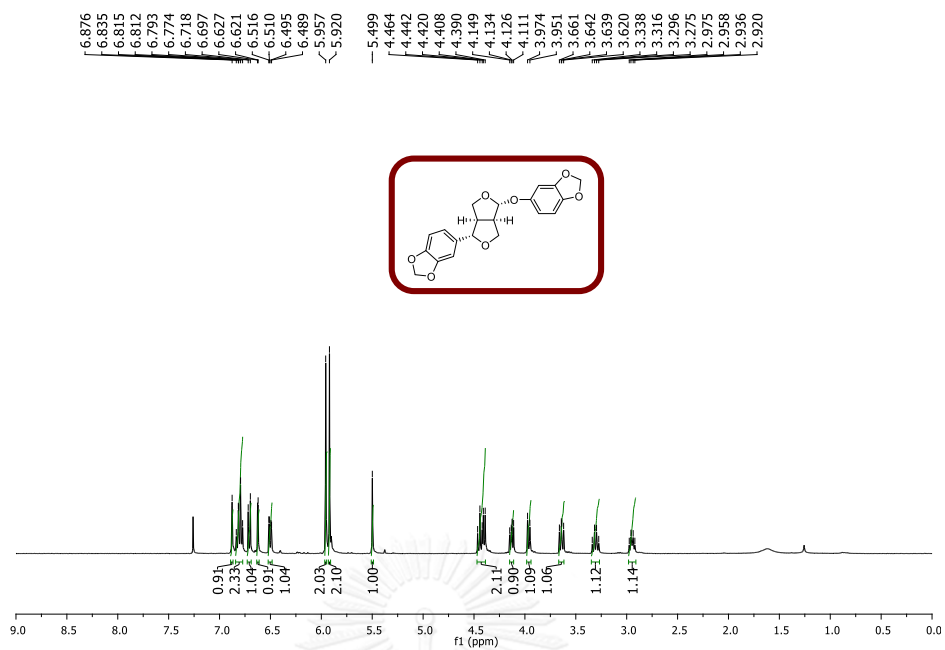


Figure 1. ^1H NMR spectrum of compound **1.2** in CDCl_3

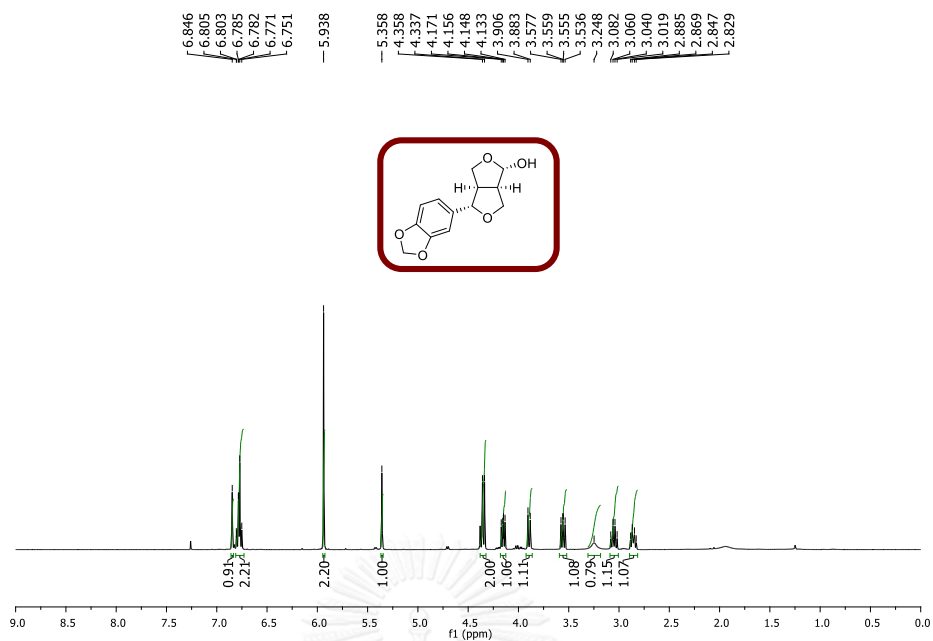


Figure 2. ^1H NMR spectrum of compound 1.36 in CDCl_3

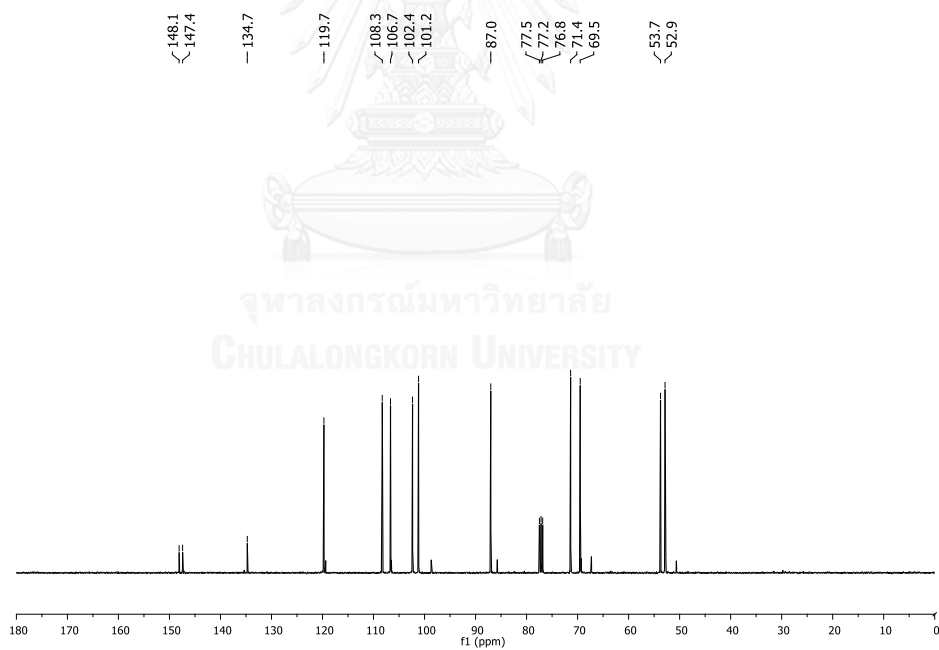


Figure 3. ^{13}C NMR spectrum of compound 1.36 in CDCl_3

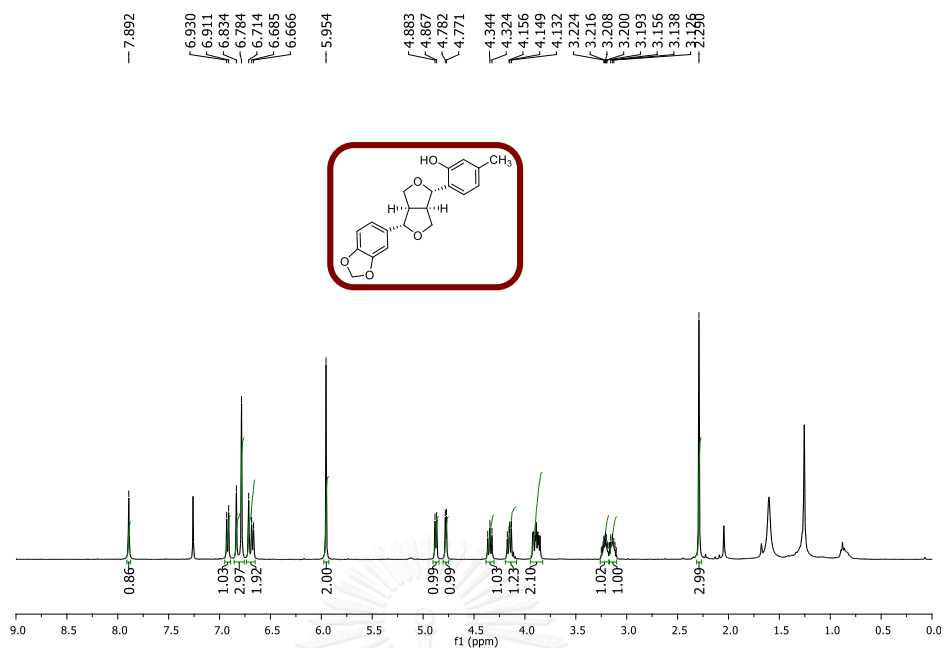


Figure 4. ^1H NMR spectrum of compound 3.1b in CDCl_3

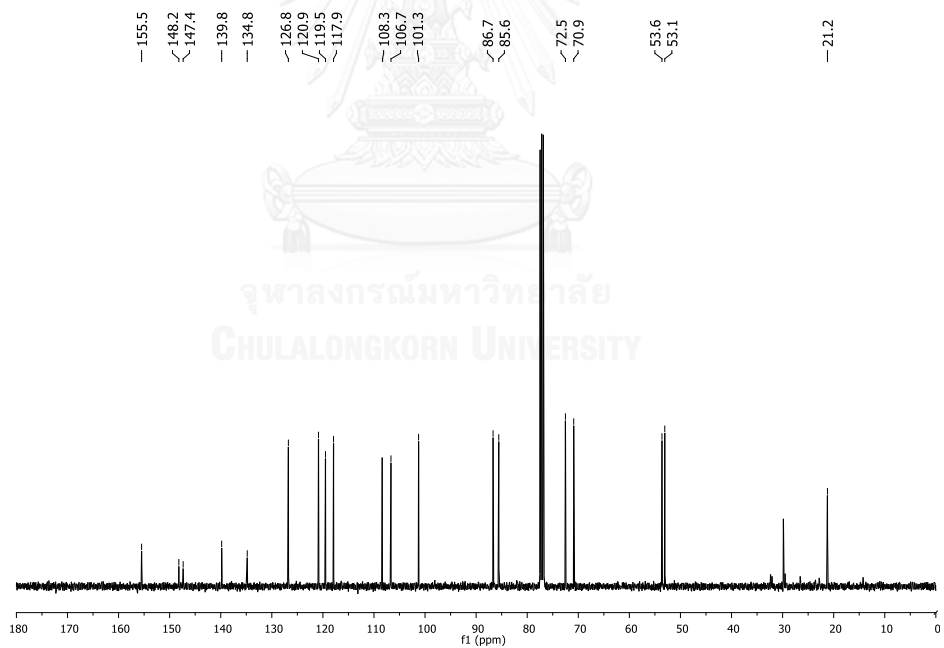


Figure 5. ^{13}C NMR spectrum of compound 3.1b in CDCl_3

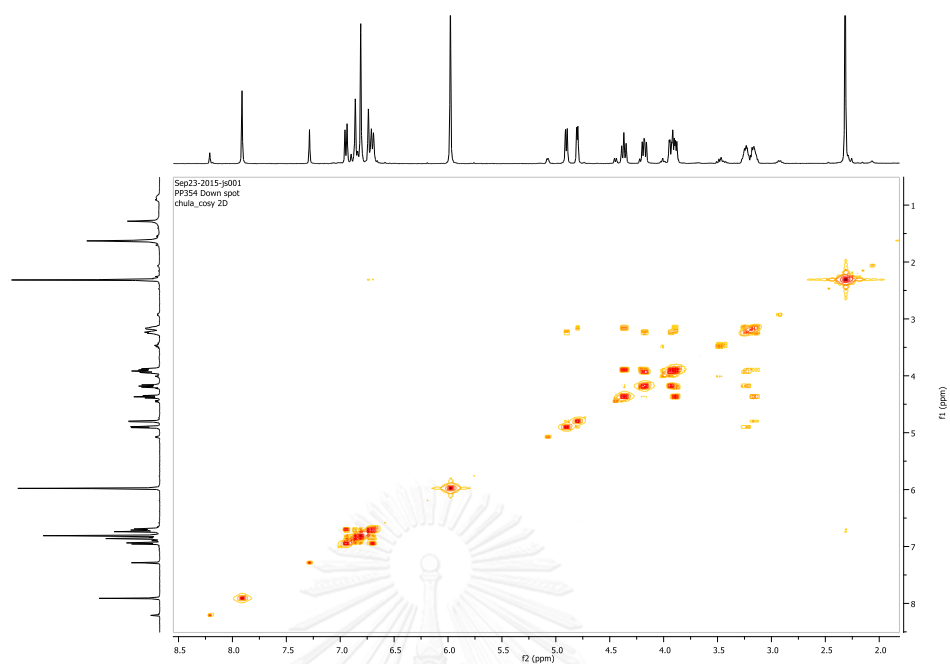


Figure 6. COSY experiment of **3.1b** in CDCl_3

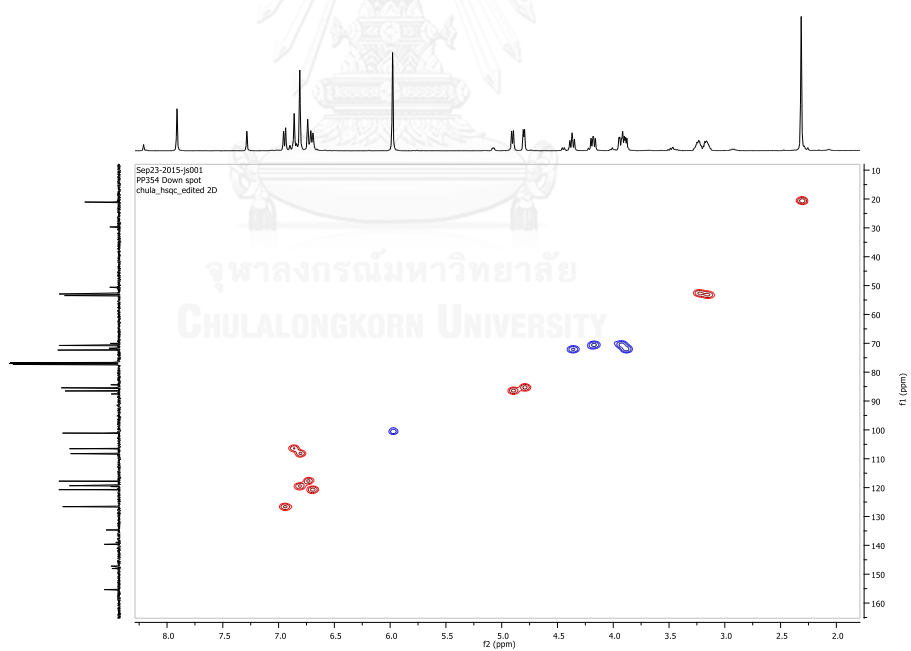


Figure 7. HSQC experiment of **3.1b** in CDCl_3

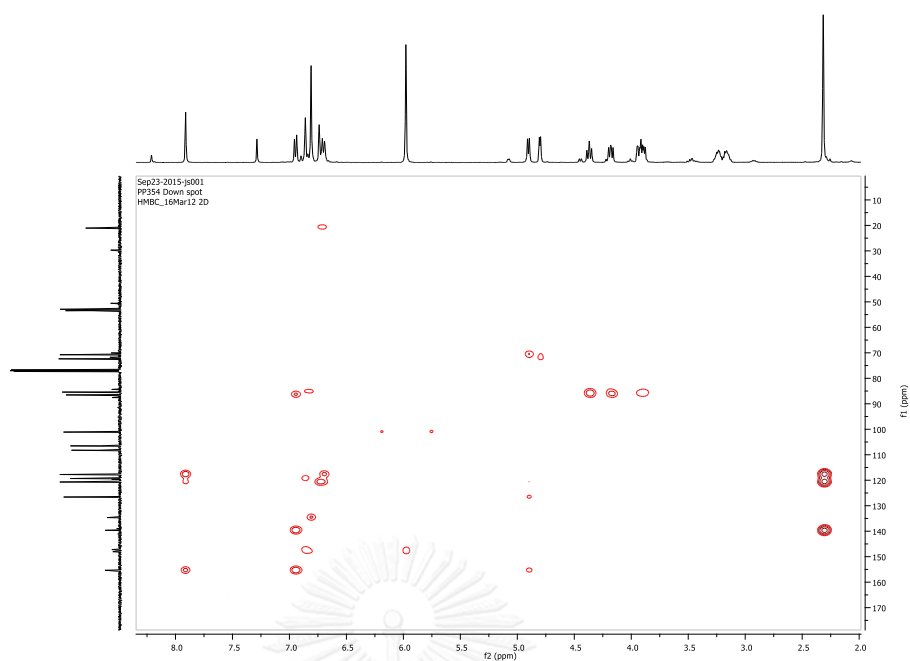


Figure 8. HMBC experiment of **3.1b** in CDCl_3

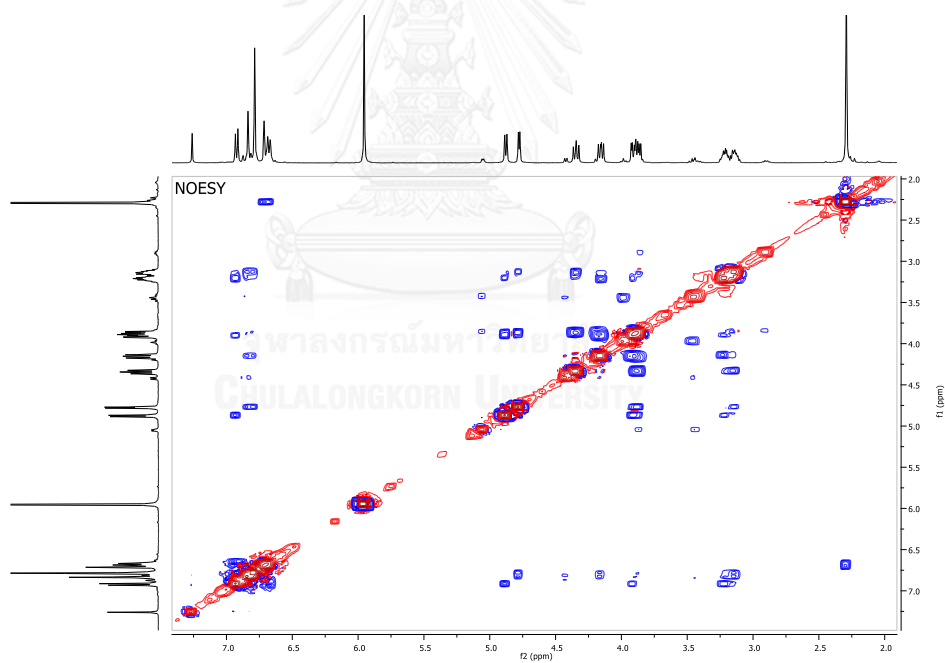


Figure 9. NOESY experiment of **3.1b** in CDCl_3

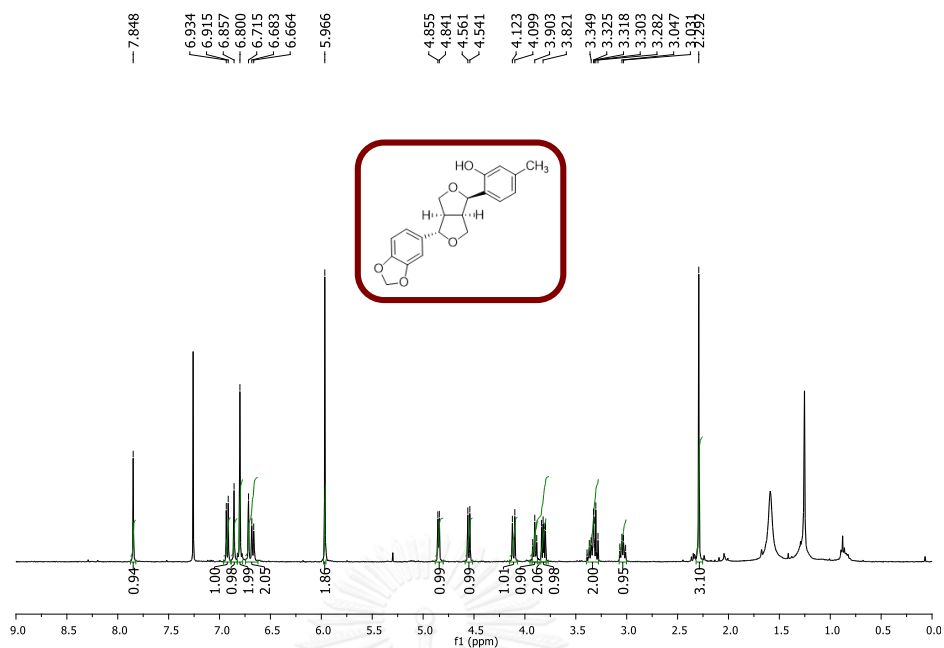


Figure 10. ^1H NMR spectrum of compound *epi-3.1b* in CDCl_3

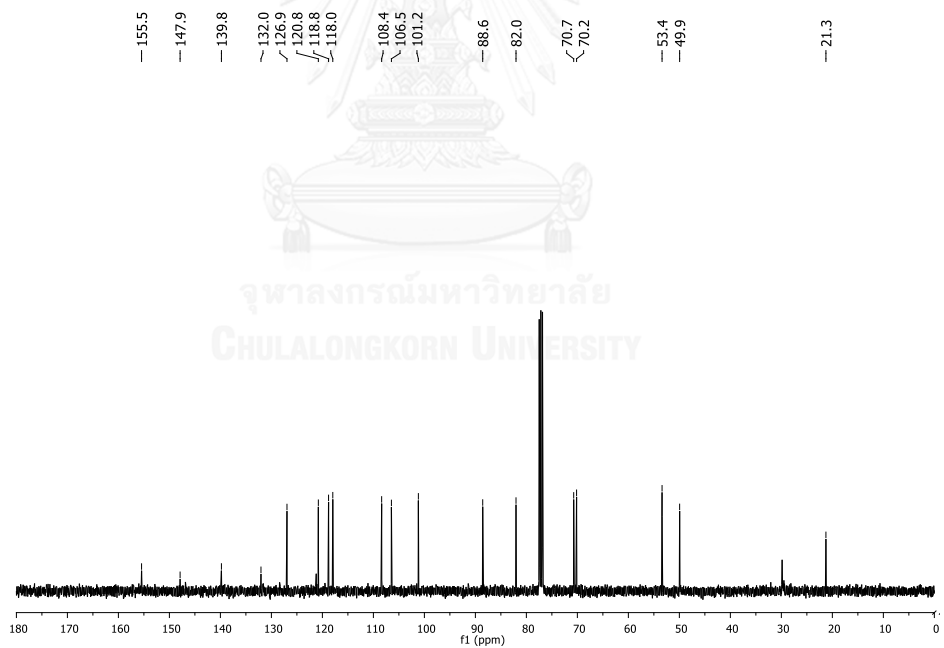


Figure 11. ^{13}C NMR spectrum of compound *epi-3.1b* in CDCl_3

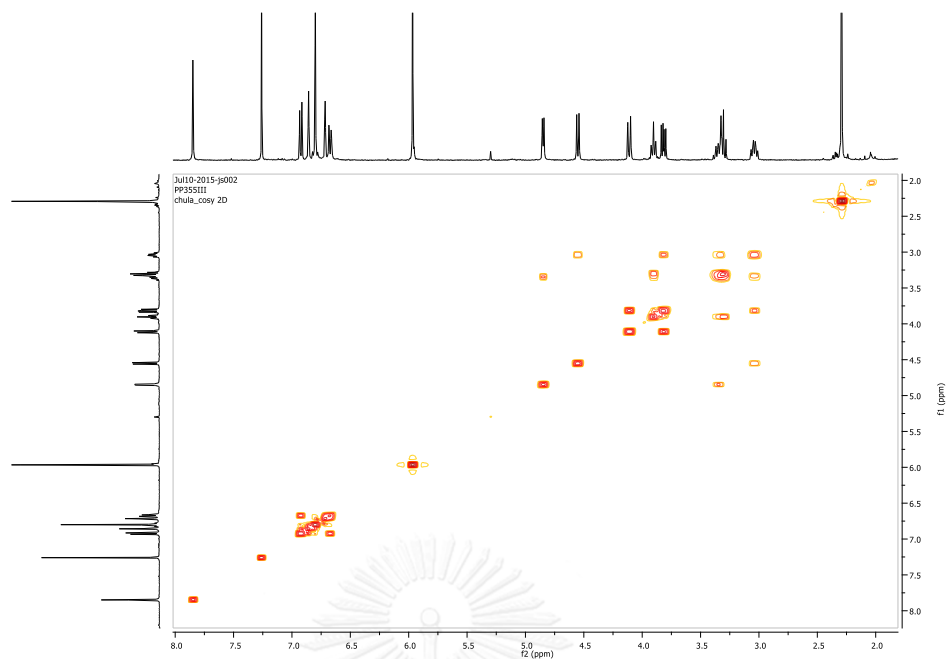


Figure 12. COSY experiment of *epi-3.1b* in CDCl_3

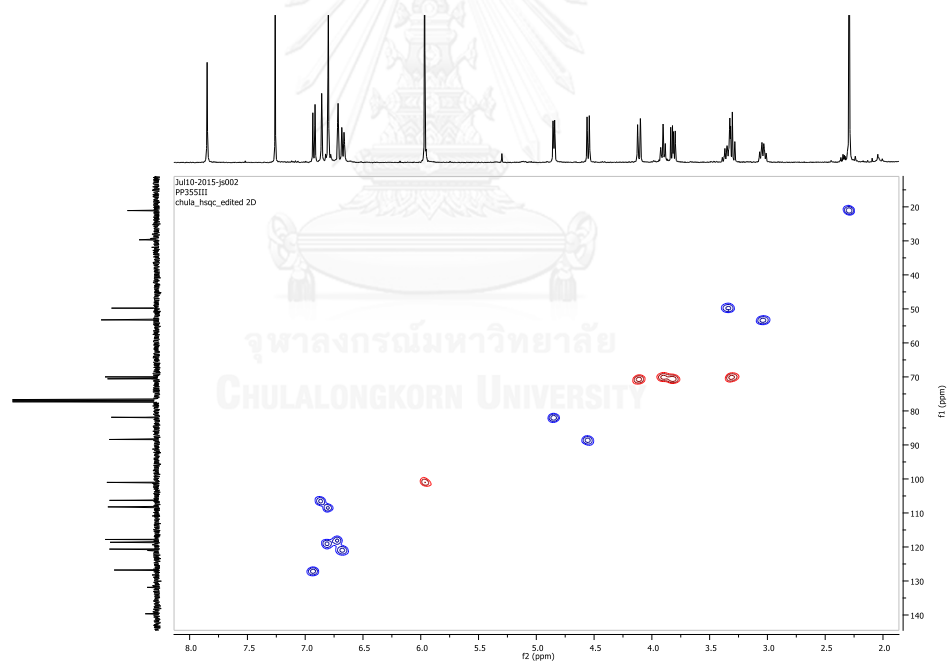


Figure 13. HSQC experiment of *epi-3.1b* in CDCl_3

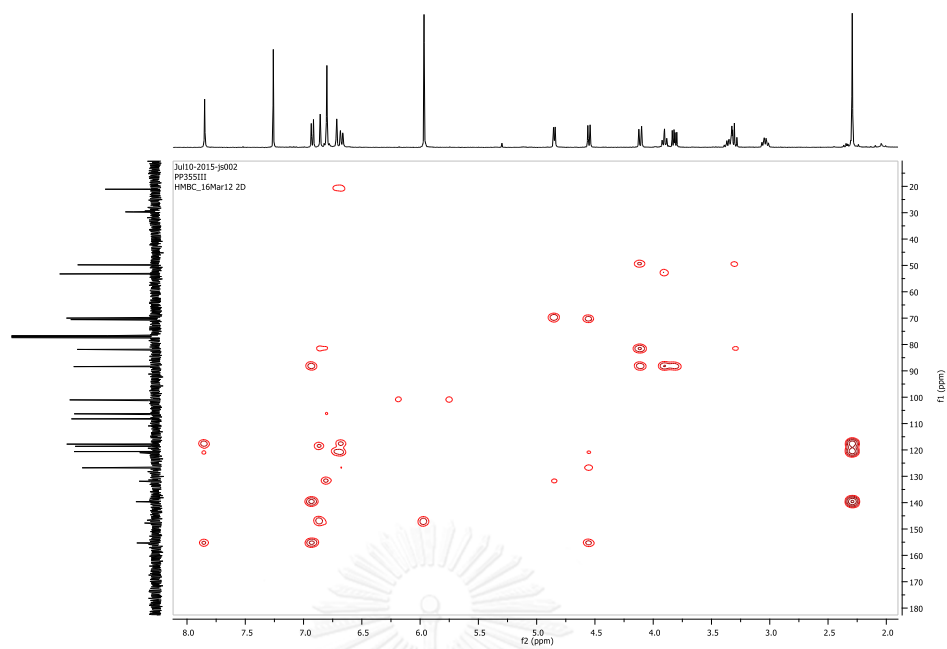


Figure 14. HMBC experiment of *epi-3.1b* in CDCl_3

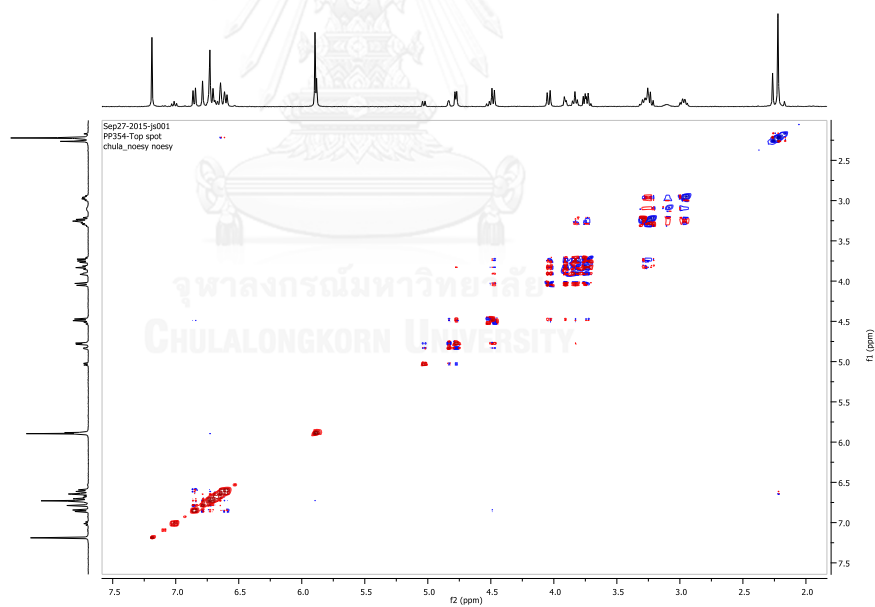


Figure 15. NOESY experiment of *epi-3.1b* in CDCl_3

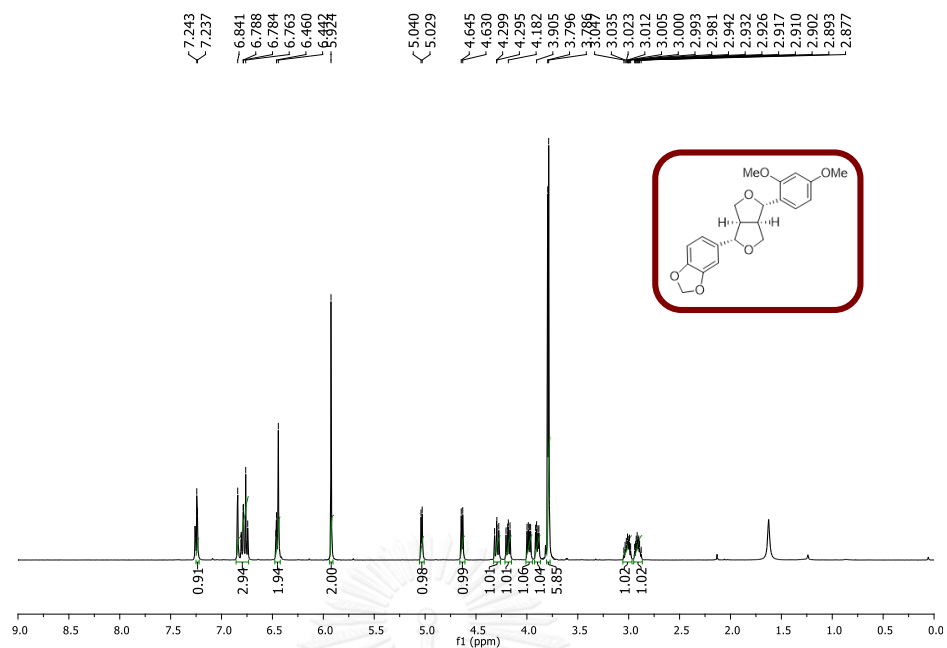


Figure 16. ^1H NMR spectrum of compound 3.1d in CDCl_3

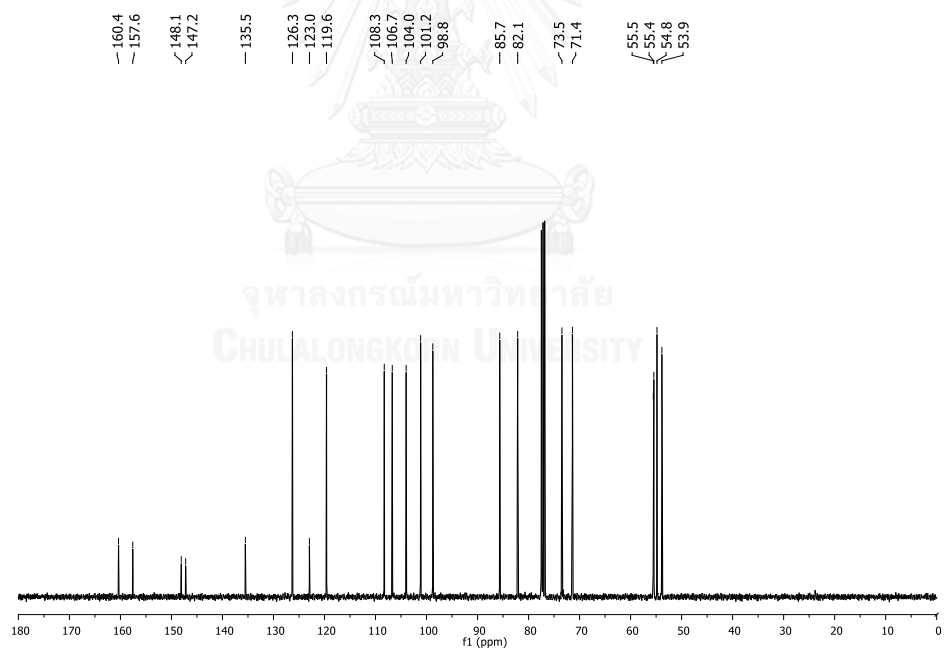


Figure 17. ^{13}C NMR spectrum of compound 3.1d in CDCl_3

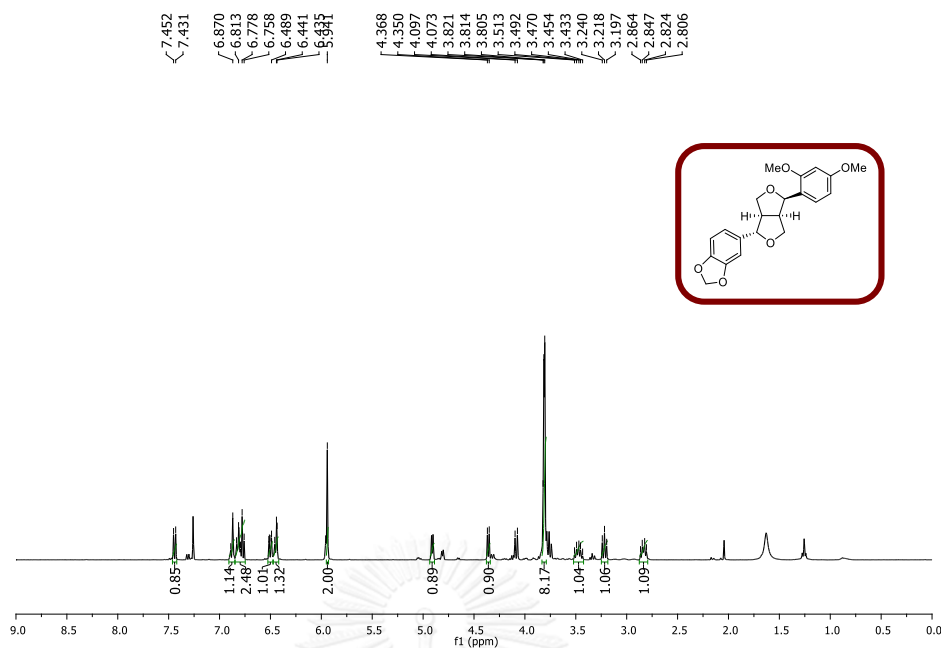


Figure 18. ^1H NMR spectrum of compound *epi-3.1d* in CDCl_3

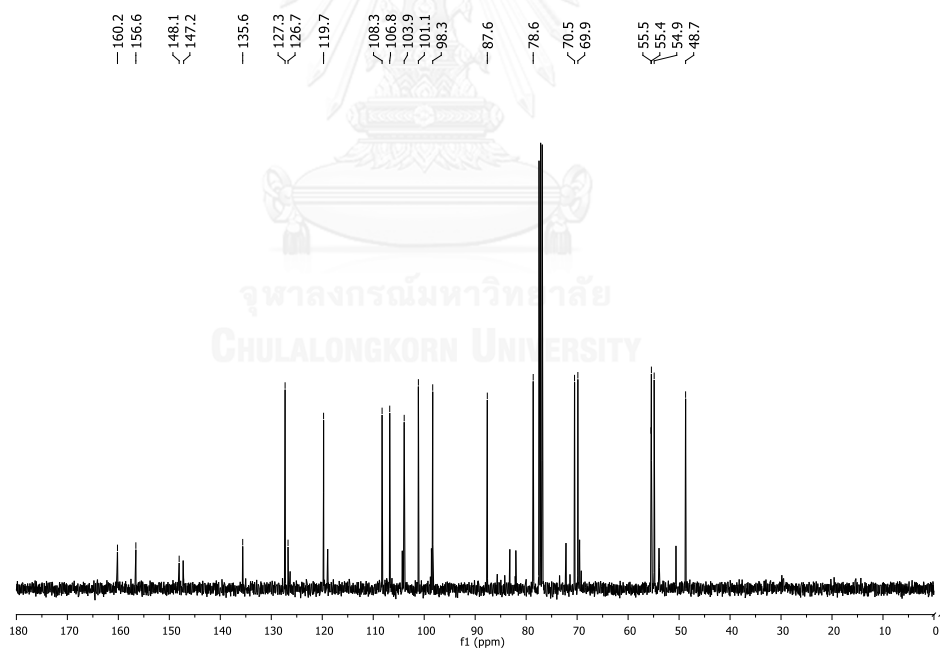


Figure 19. ^{13}C NMR spectrum of compound *epi-3.1d* in CDCl_3

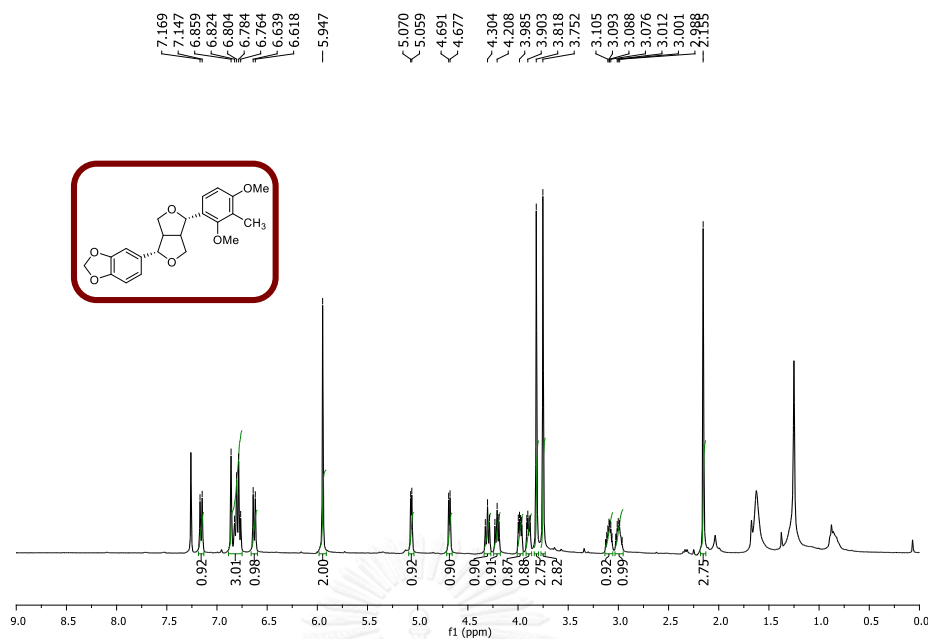


Figure 20. ^1H NMR spectrum of compound 3.1e in CDCl_3

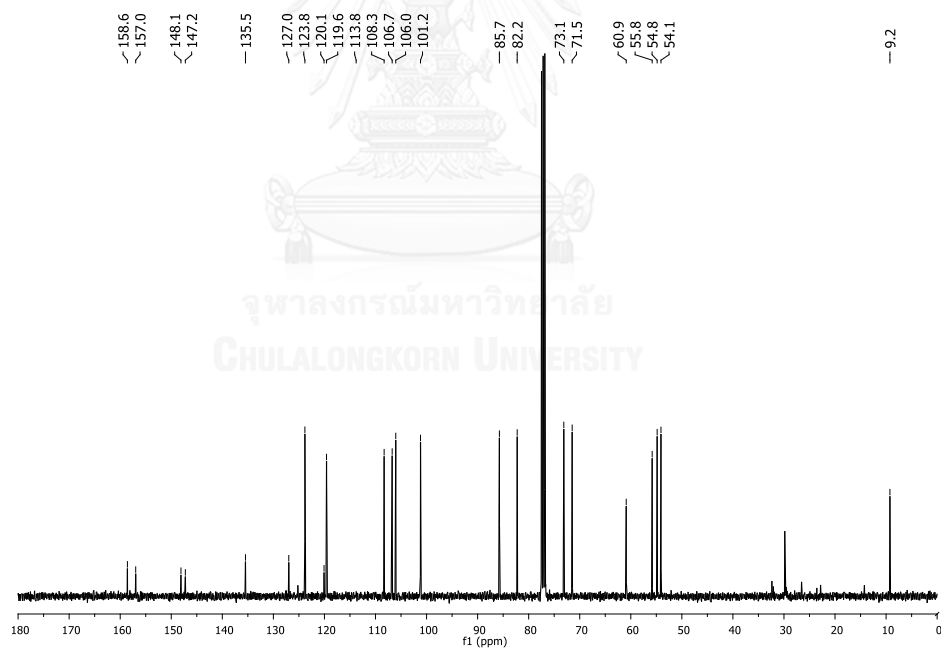


Figure 21. ^{13}C NMR spectrum of compound 3.1e in CDCl_3

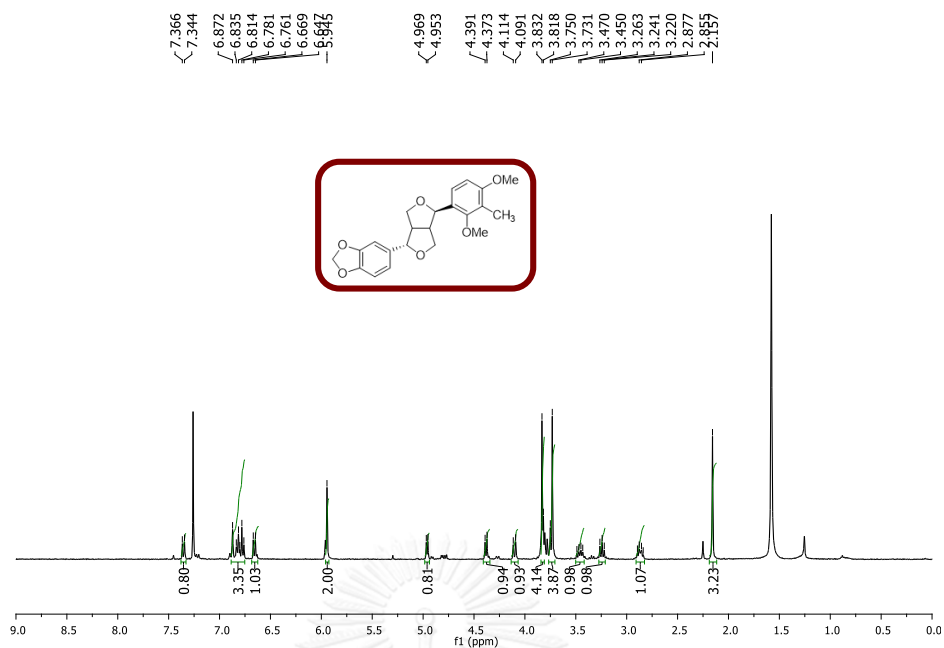


Figure 22. ^1H NMR spectrum of compound *epi-3.1e* in CDCl_3

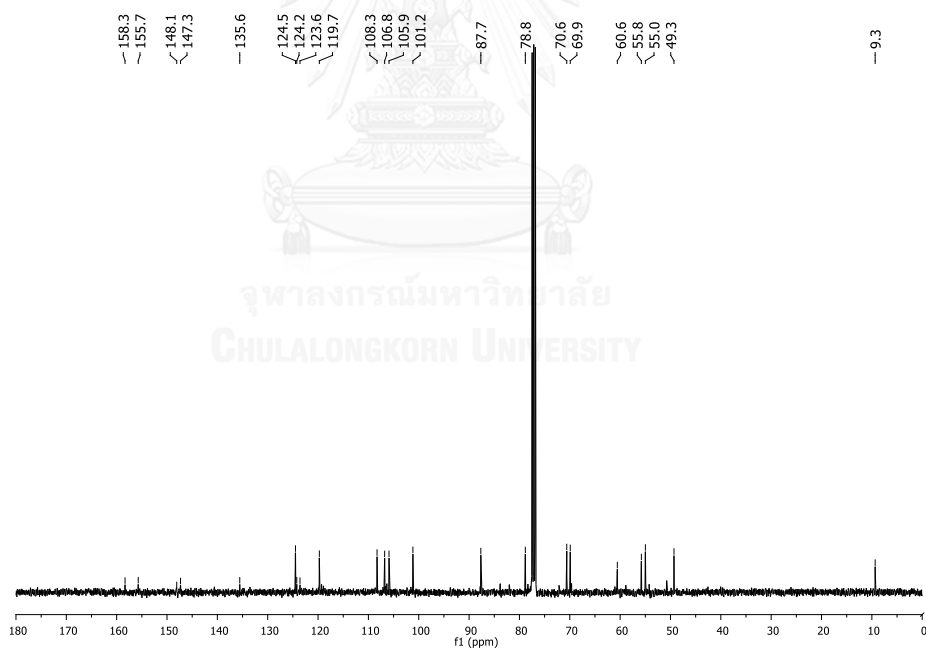


Figure 23. ^{13}C NMR spectrum of compound *epi-3.1e* in CDCl_3

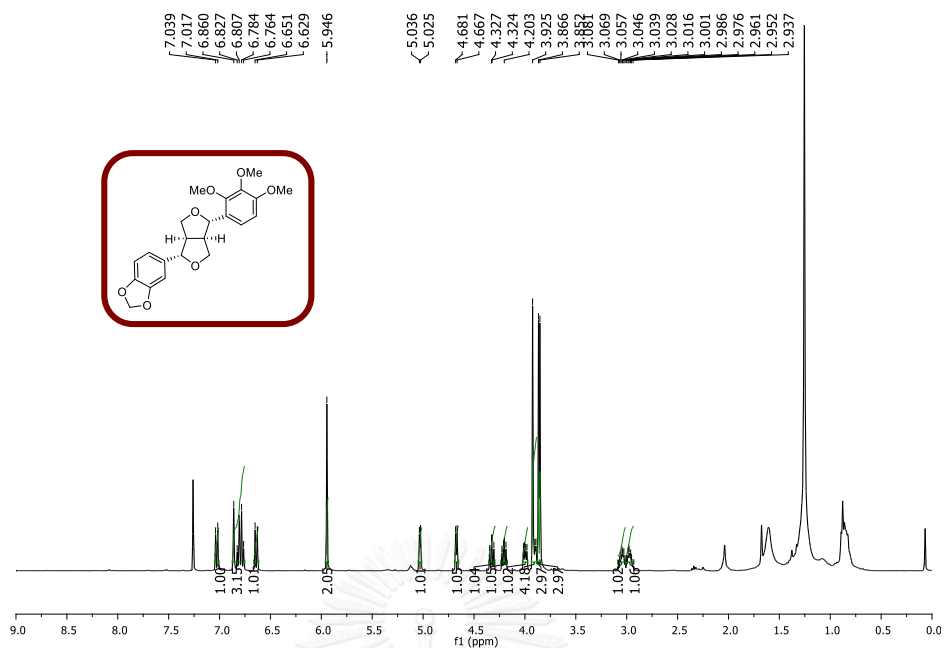


Figure 24. ^1H NMR spectrum of compound 3.1g in CDCl_3

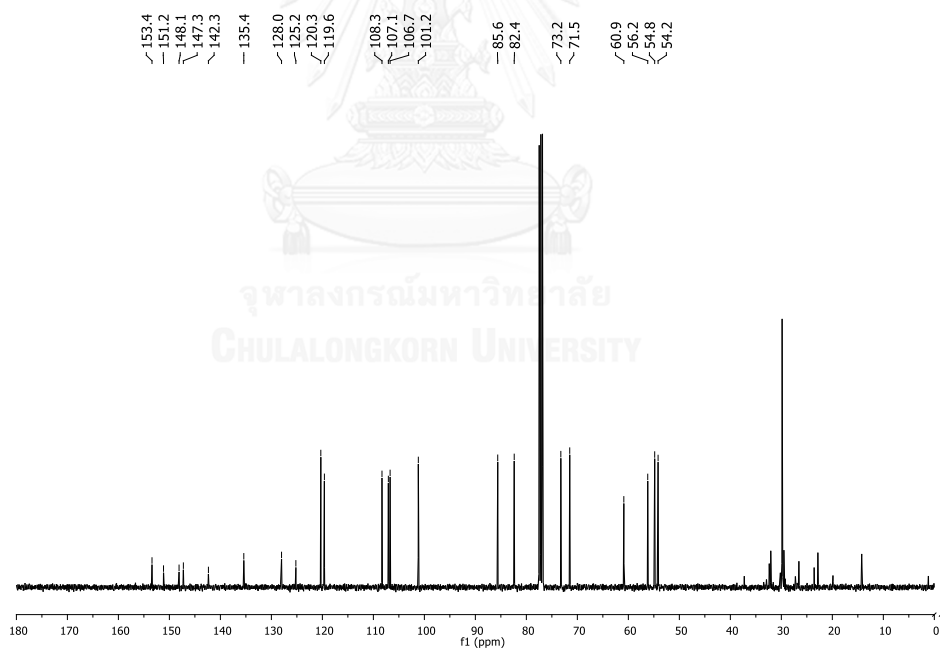


Figure 25. ^{13}C NMR spectrum of compound 3.1g in CDCl_3

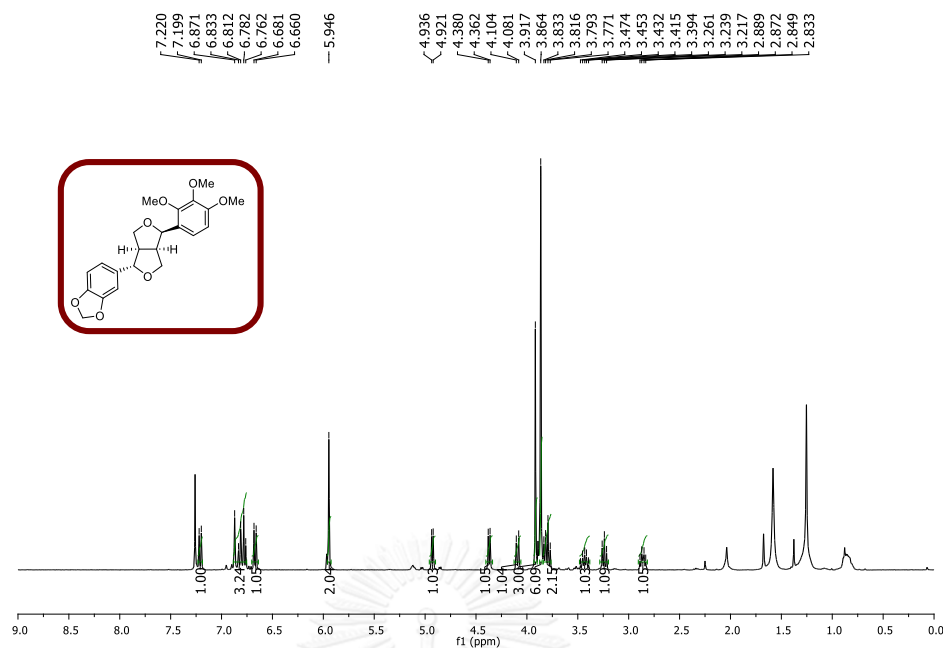


Figure 26. ^1H NMR spectrum of compound *epi-3.1g* in CDCl_3

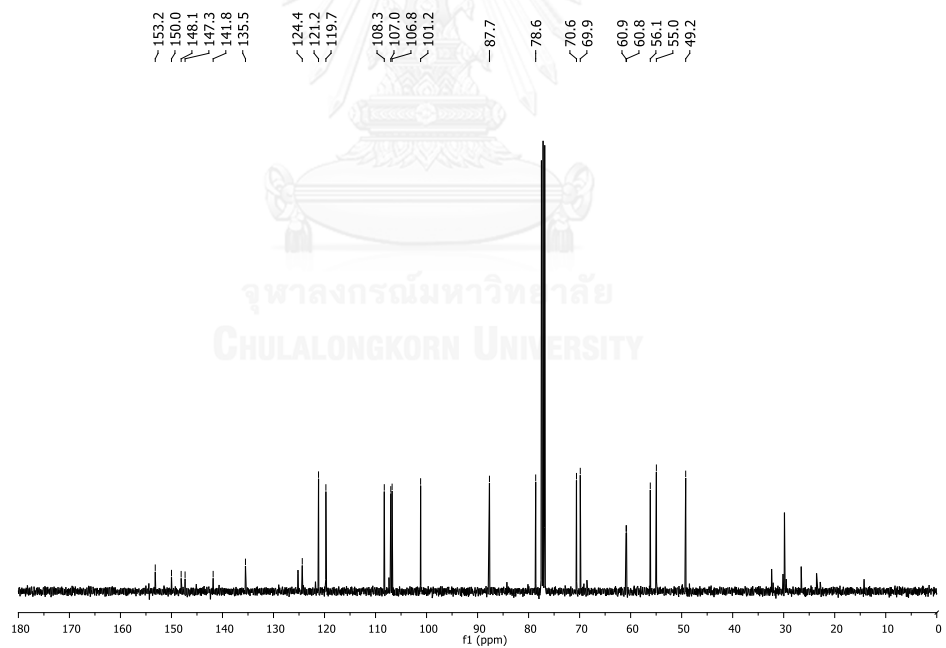


Figure 27. ^{13}C NMR spectrum of compound *epi-3.1g* in CDCl_3

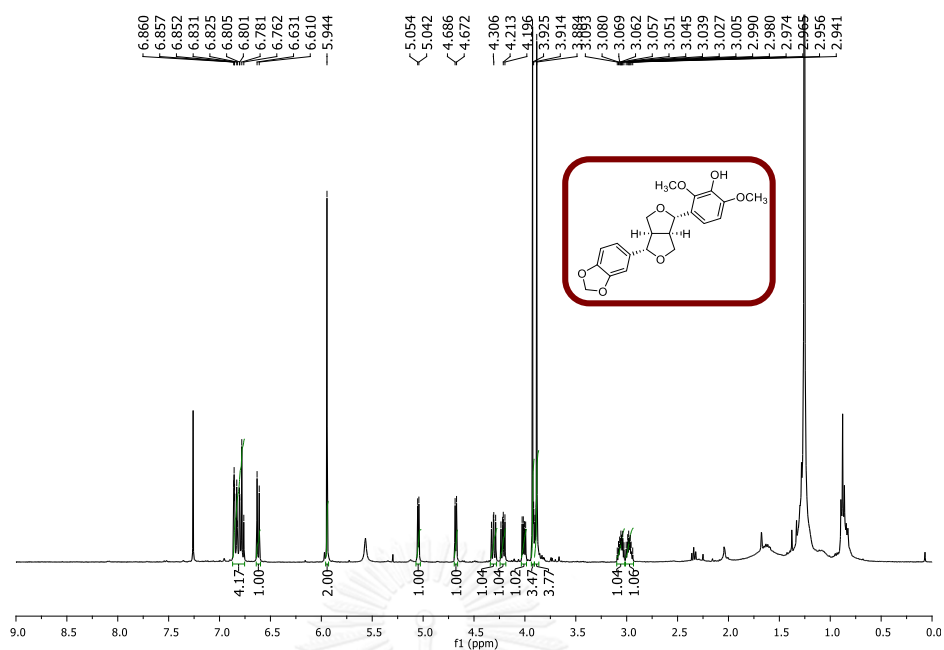


Figure 28. ^1H NMR spectrum of compound 3.1h in CDCl_3

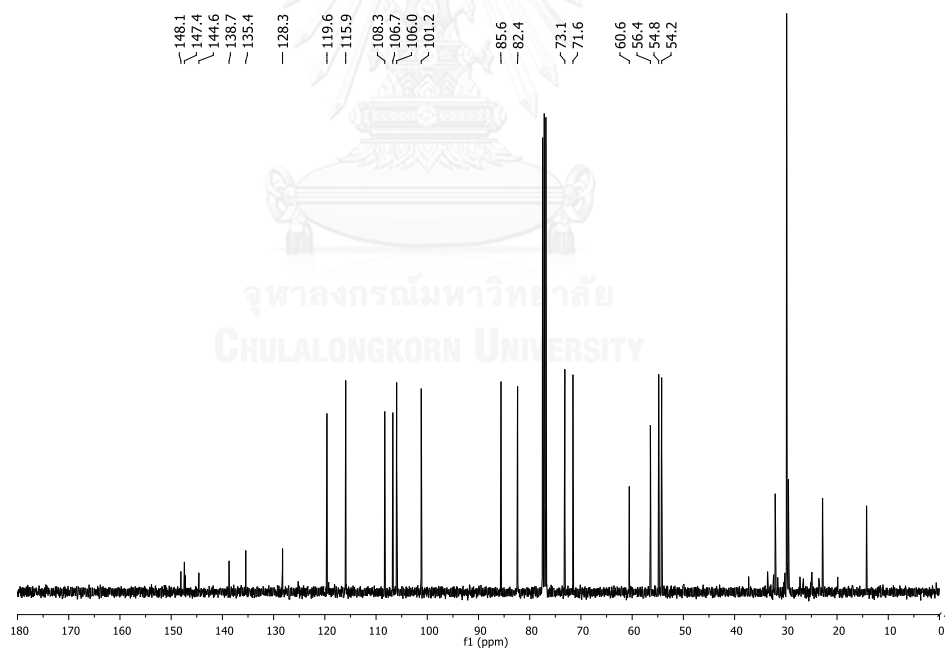


Figure 29. ^{13}C NMR spectrum of compound 3.1h in CDCl_3

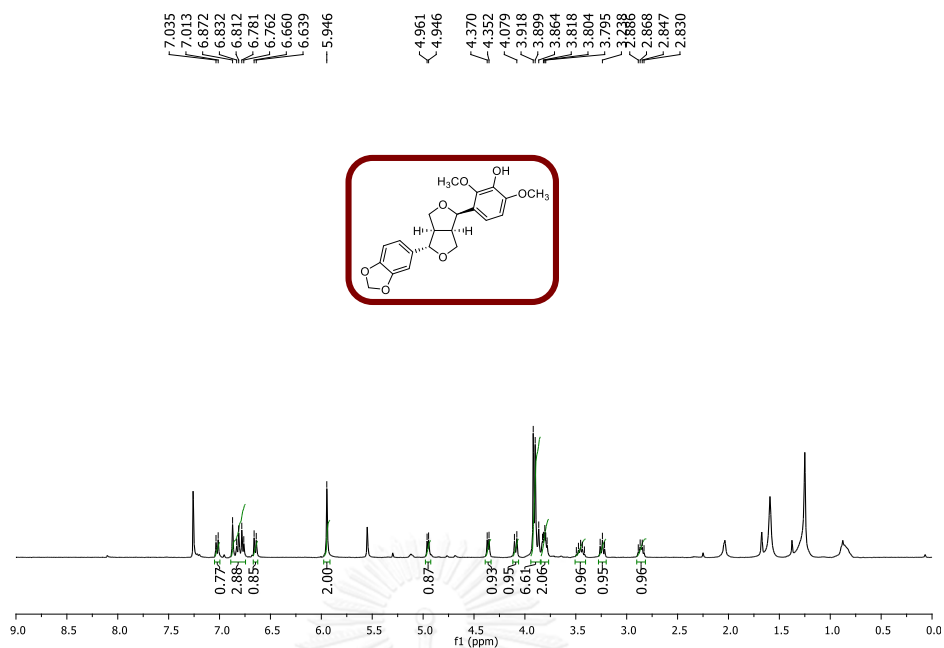


Figure 30. ^1H NMR spectrum of compound *epi-3.1h* in CDCl_3

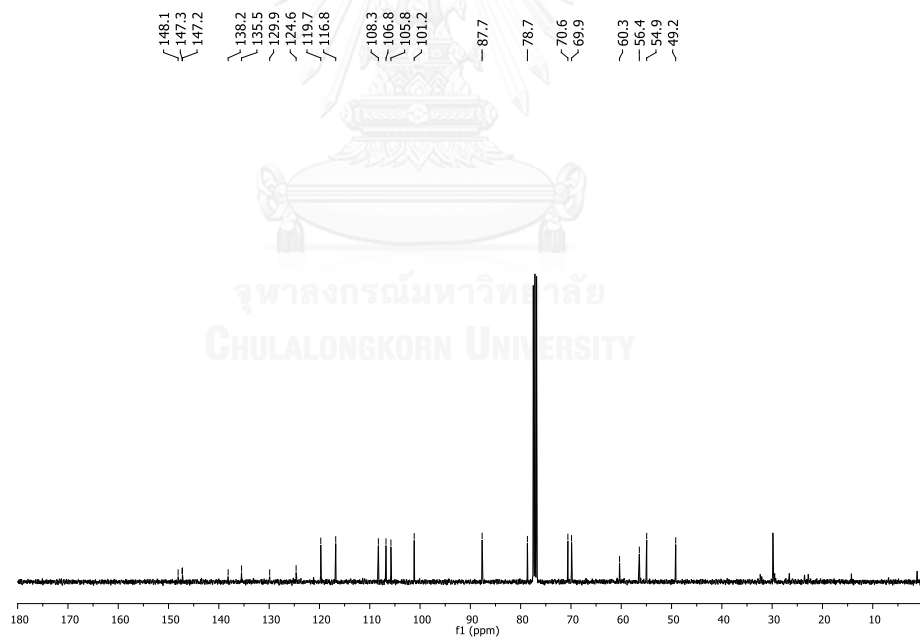


Figure 31. ^{13}C NMR spectrum of compound *epi-3.1h* in CDCl_3

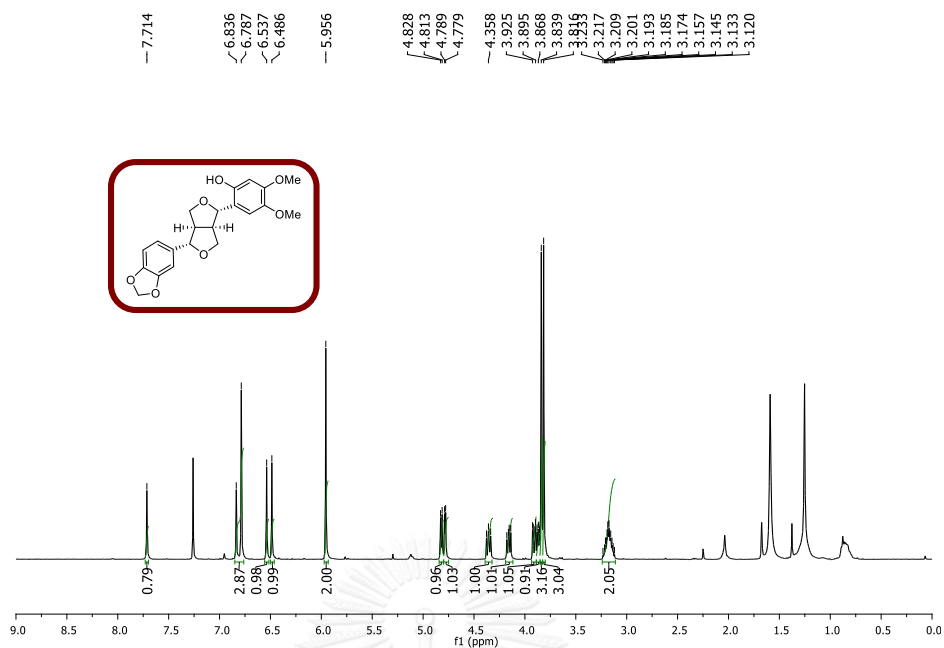


Figure 32. ^1H NMR spectrum of compound 3.1i in CDCl_3

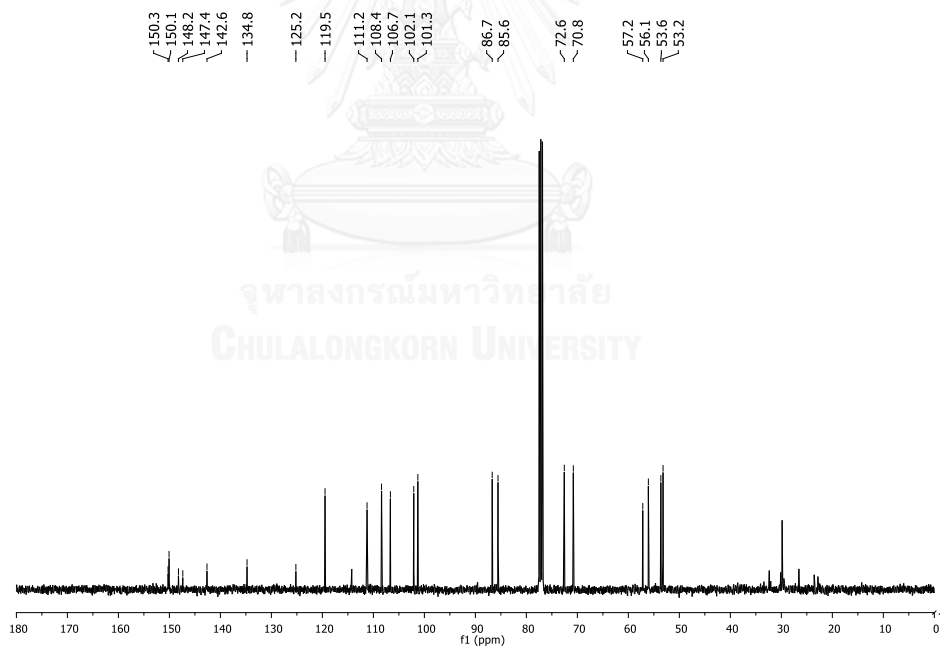


Figure 33. ^{13}C NMR spectrum of compound 3.1i in CDCl_3

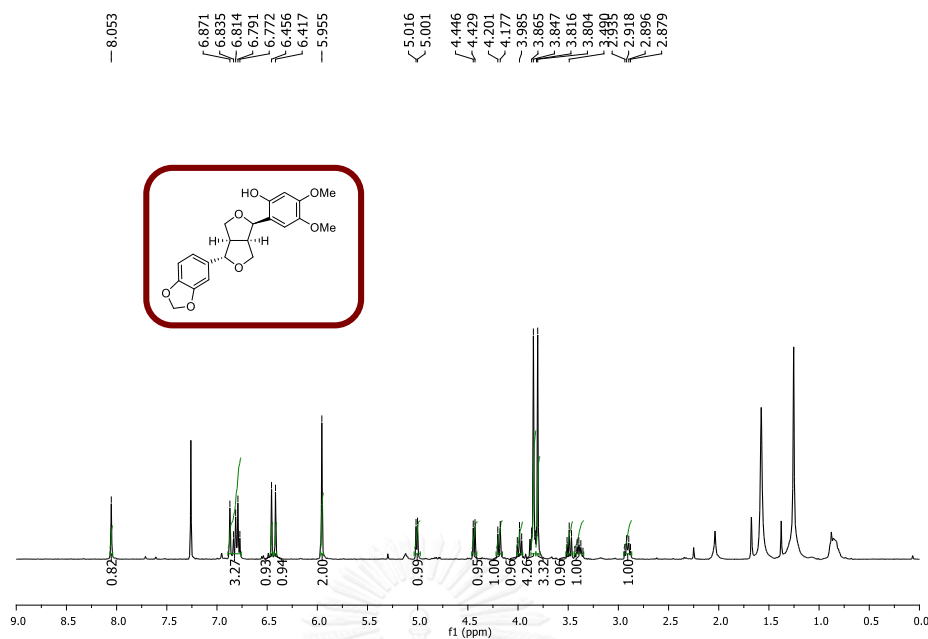


Figure 34. ^1H NMR spectrum of compound *epi-3.1i* in CDCl_3

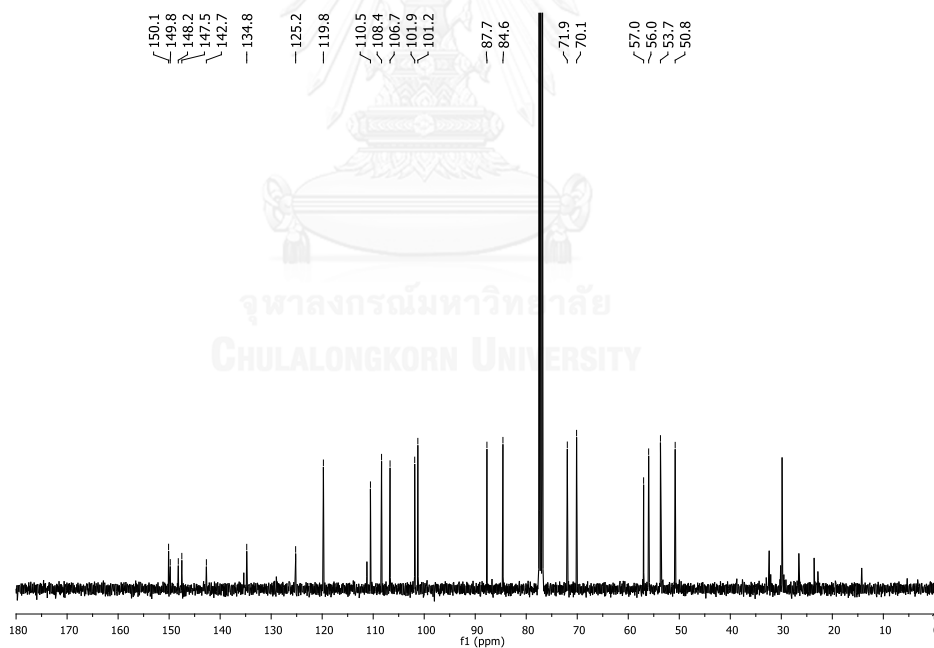


Figure 35. ^{13}C NMR spectrum of compound *epi-3.1i* in CDCl_3

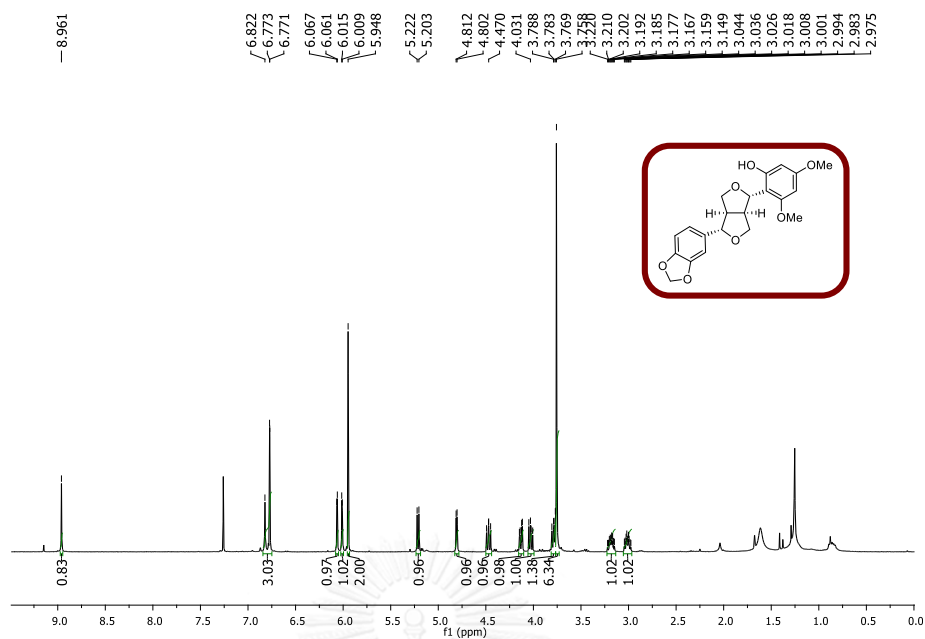


Figure 36. ^1H NMR spectrum of compound 3.1j in CDCl_3

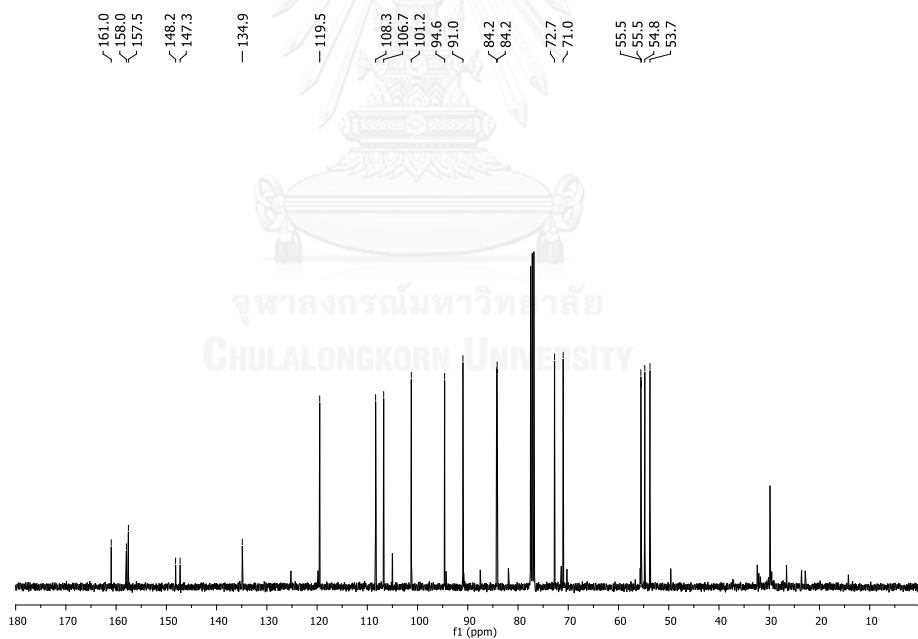


Figure 37. ^{13}C NMR spectrum of compound 3.1j in CDCl_3

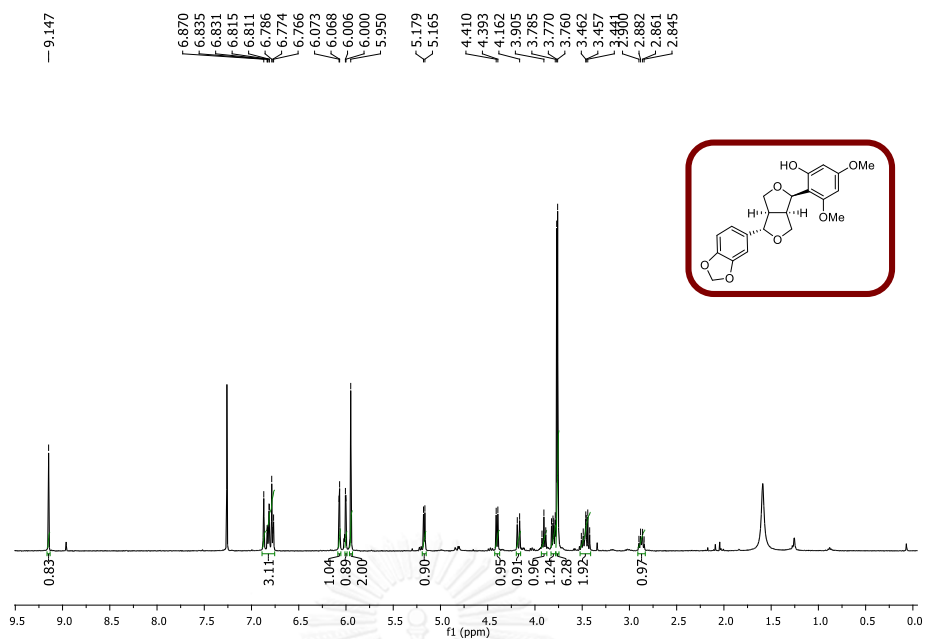


Figure 38. ^1H NMR spectrum of compound *epi-3.1j* in CDCl_3

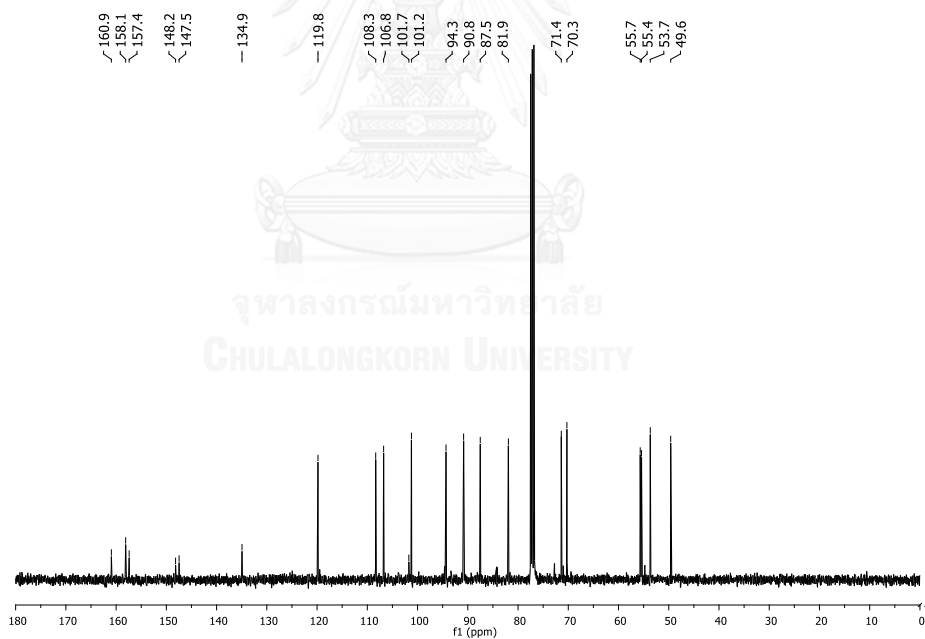


Figure 39. ^{13}C NMR spectrum of compound *epi-3.1j* in CDCl_3

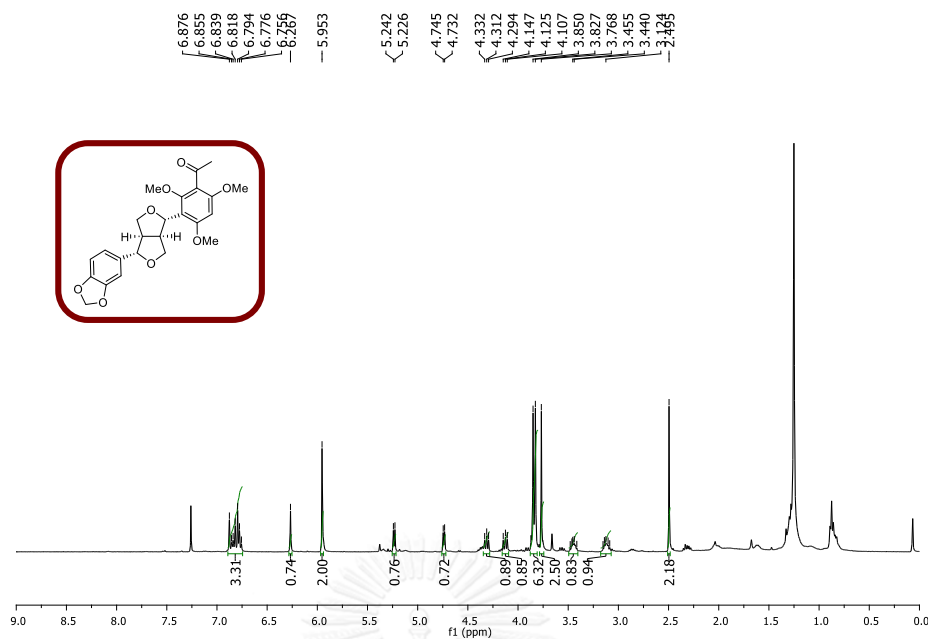


Figure 40. ^1H NMR spectrum of compound 3.1k in CDCl_3

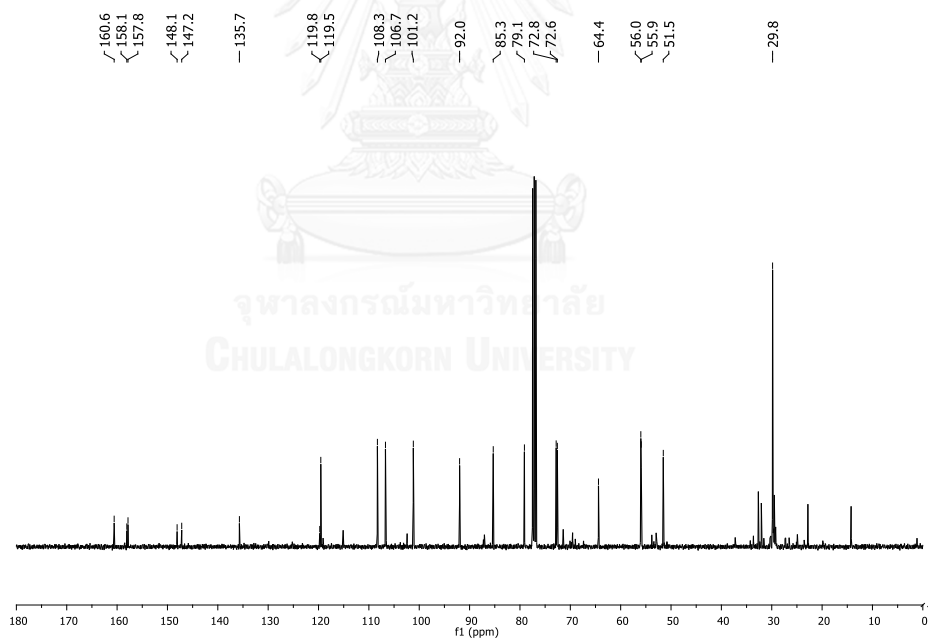


Figure 41. ^{13}C NMR spectrum of compound 3.1k in CDCl_3

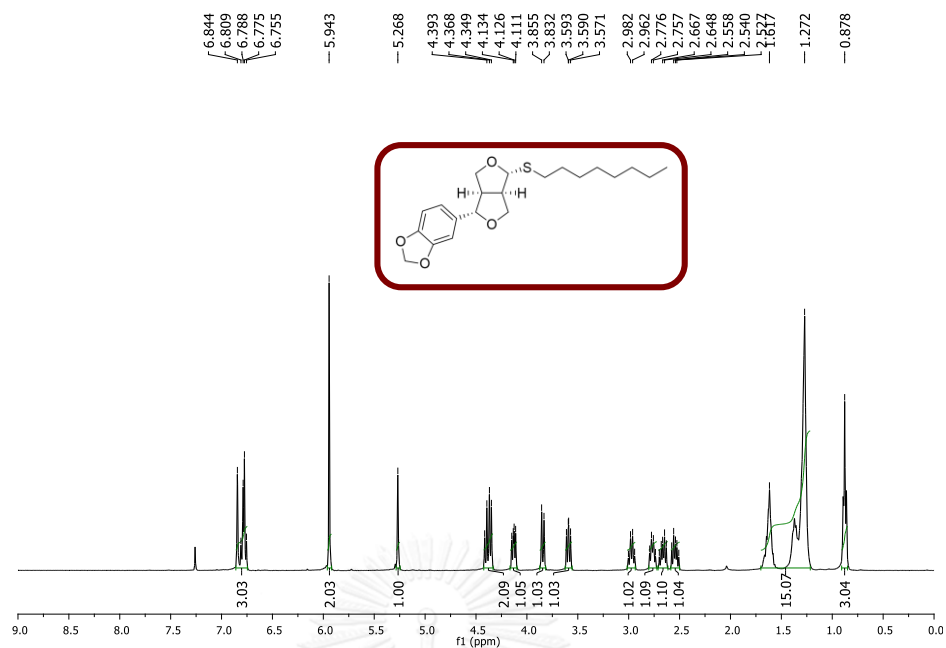


Figure 42. ^1H NMR spectrum of compound 3.2l in CDCl_3

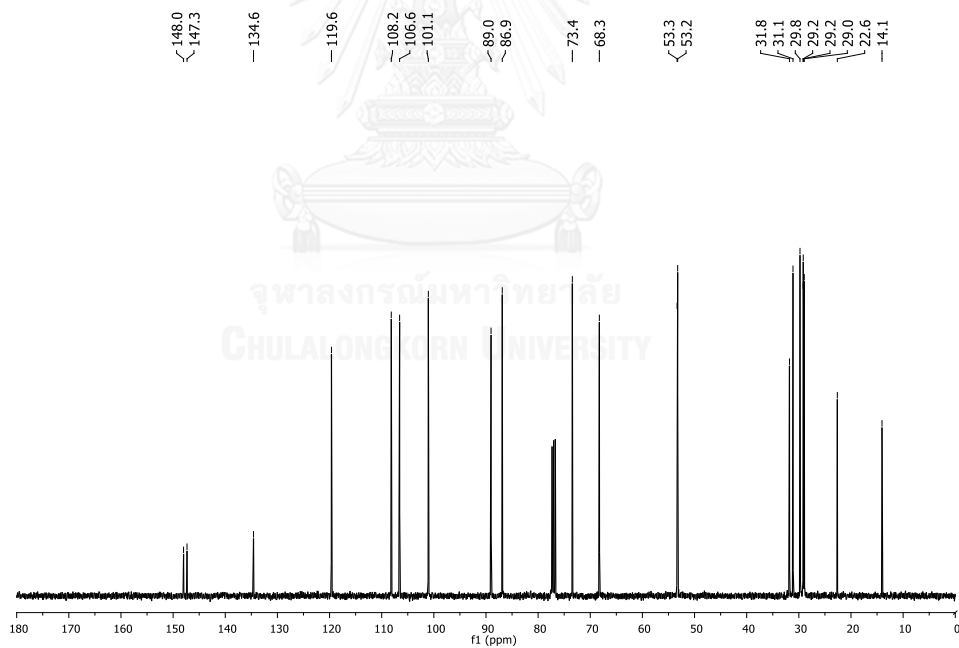


Figure 43. ^{13}C NMR spectrum of compound 3.2l in CDCl_3

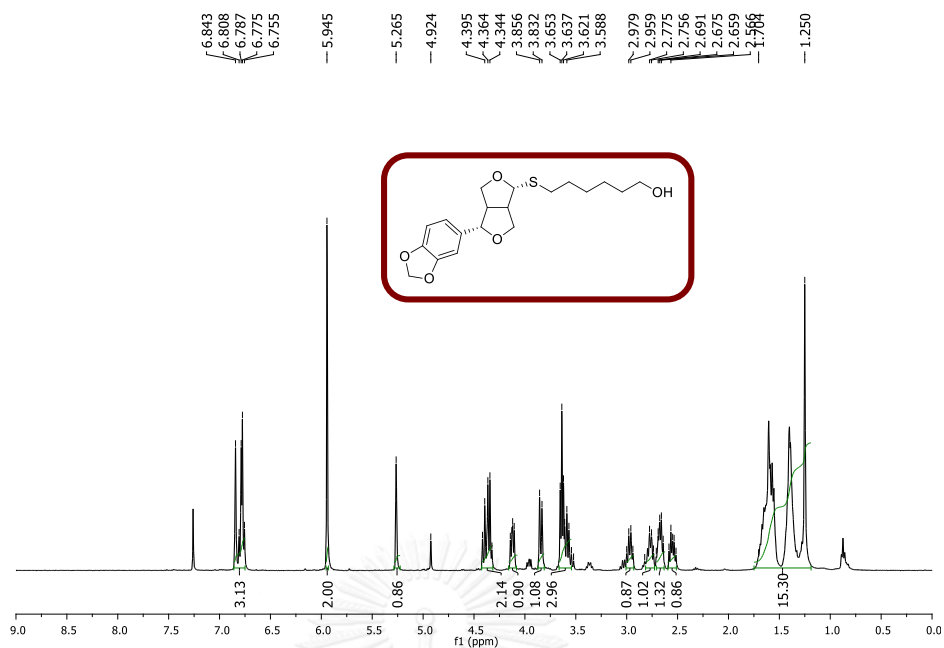


Figure 44. ^1H NMR spectrum of compound 3.2m in CDCl_3

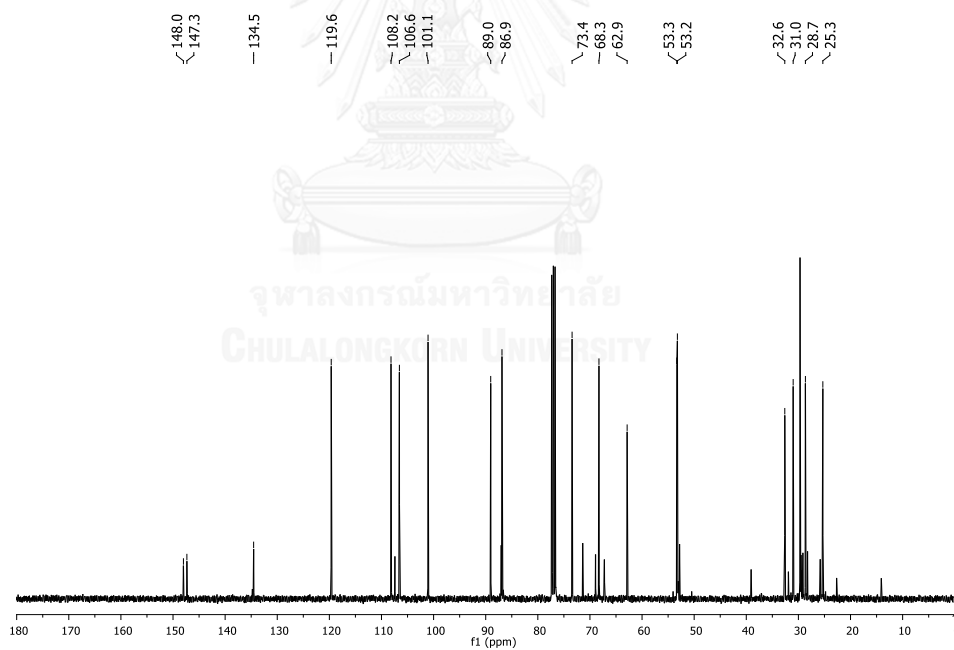


Figure 45. ^{13}C NMR spectrum of compound 3.2m in CDCl_3

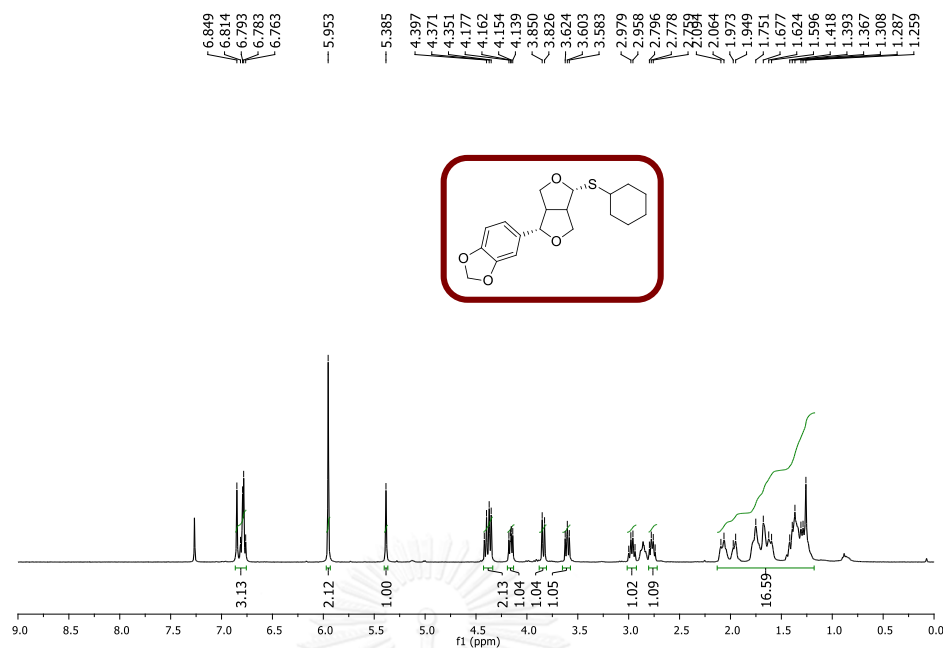


Figure 46. ^1H NMR spectrum of compound 3.2n in CDCl_3

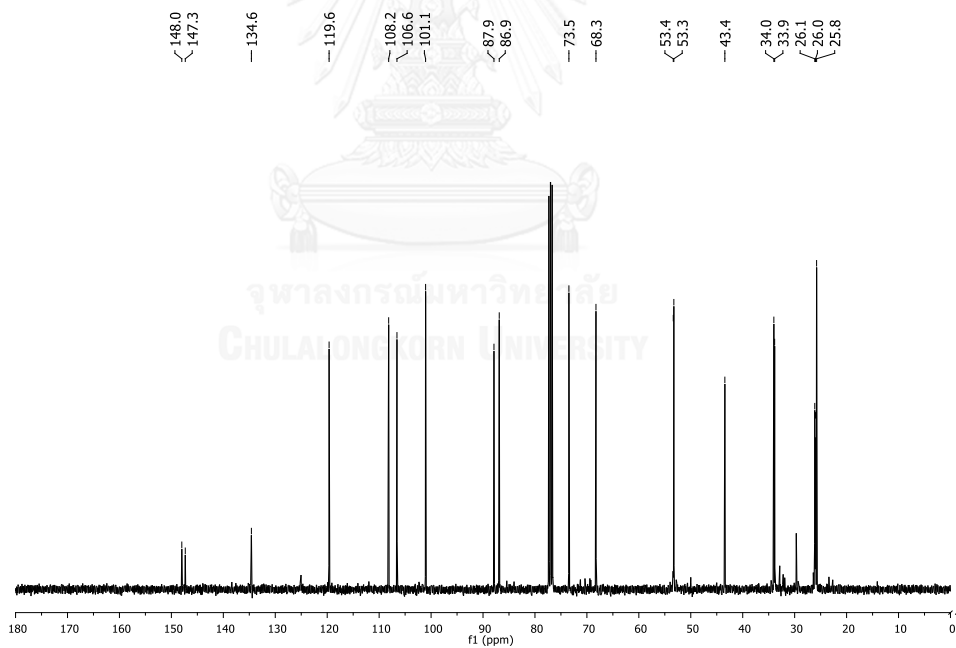


Figure 47. ^{13}C NMR spectrum of compound 3.2n in CDCl_3

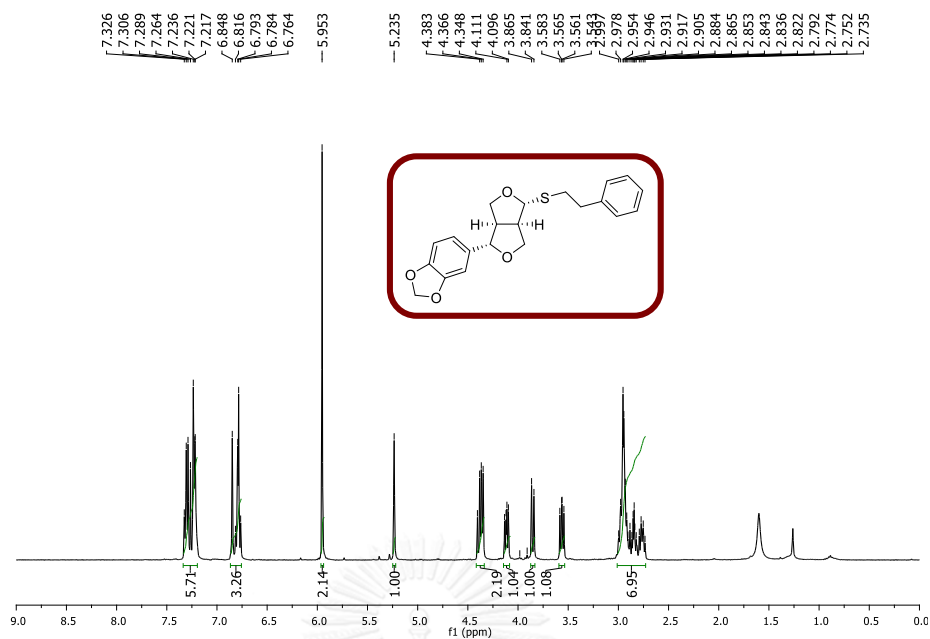


Figure 48. ^1H NMR spectrum of compound 3.2o in CDCl_3

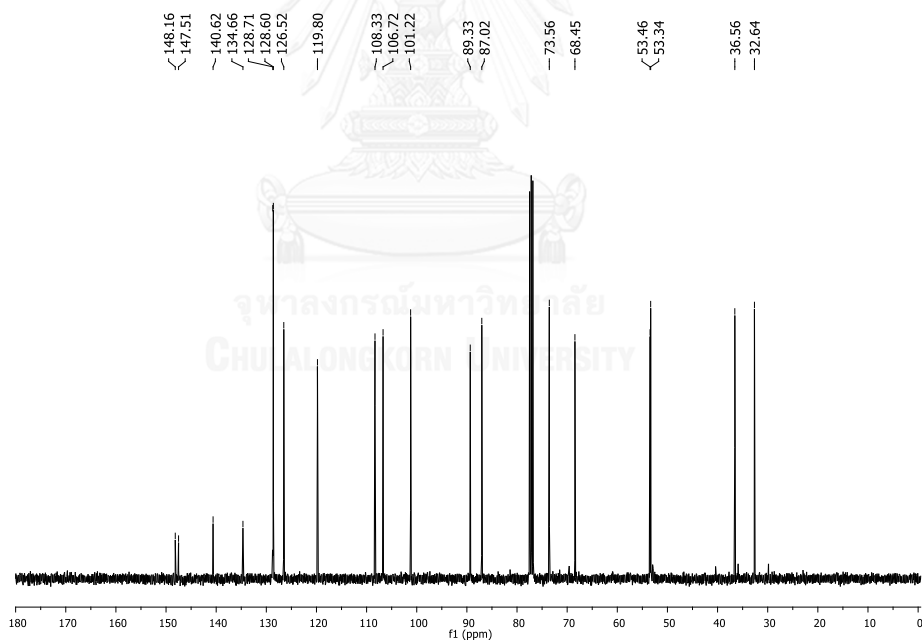


Figure 49. ^{13}C NMR spectrum of compound 3.2o in CDCl_3

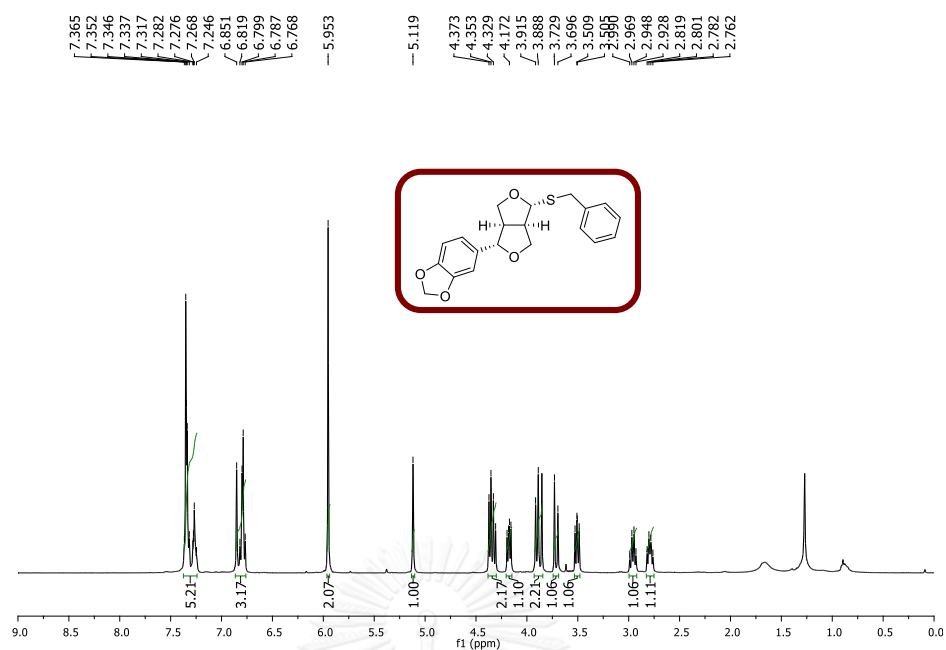


Figure 50. ^1H NMR spectrum of compound 3.2p in CDCl_3

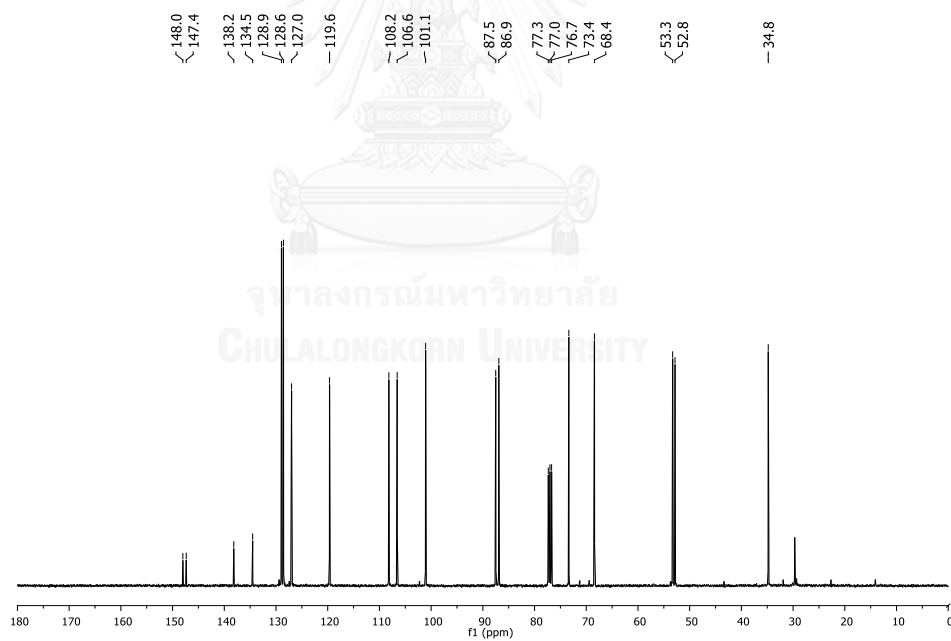


Figure 51. ^{13}C NMR spectrum of compound 3.2p in CDCl_3

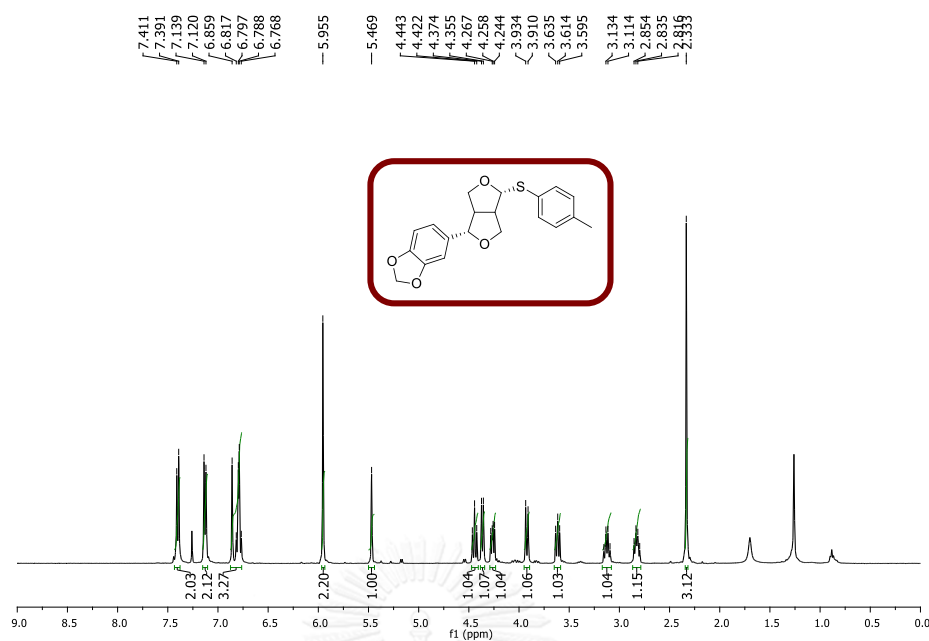


Figure 52. ^1H NMR spectrum of compound 3.2q in CDCl_3

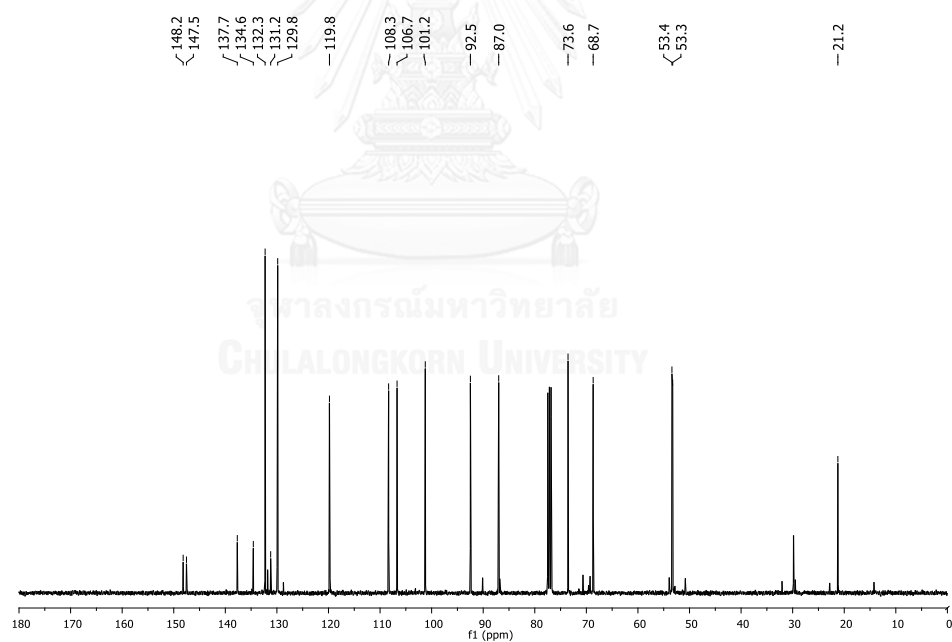


Figure 53. ^{13}C NMR spectrum of compound 3.2q in CDCl_3

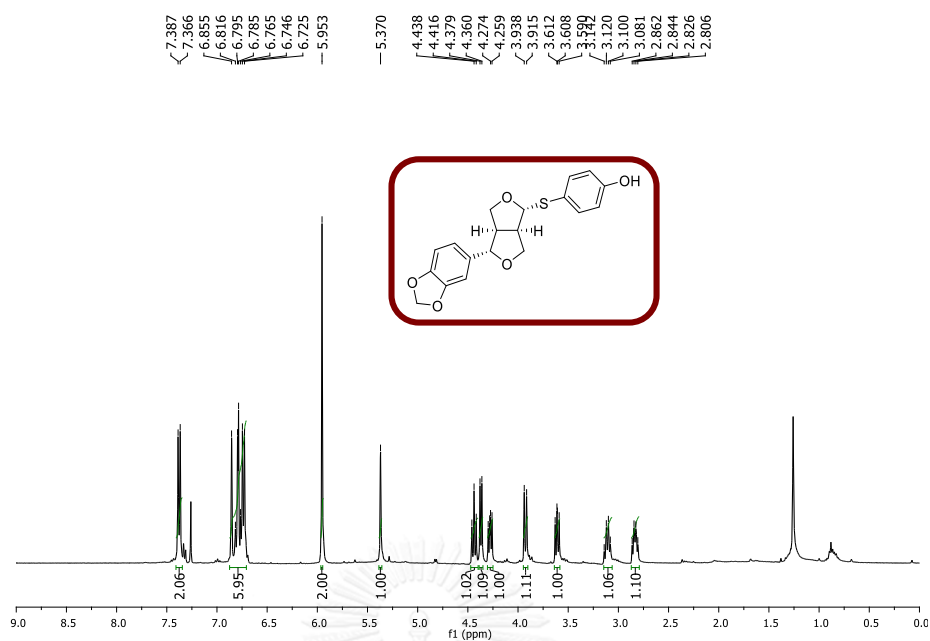


Figure 54. ^1H NMR spectrum of compound 3.2r in CDCl_3

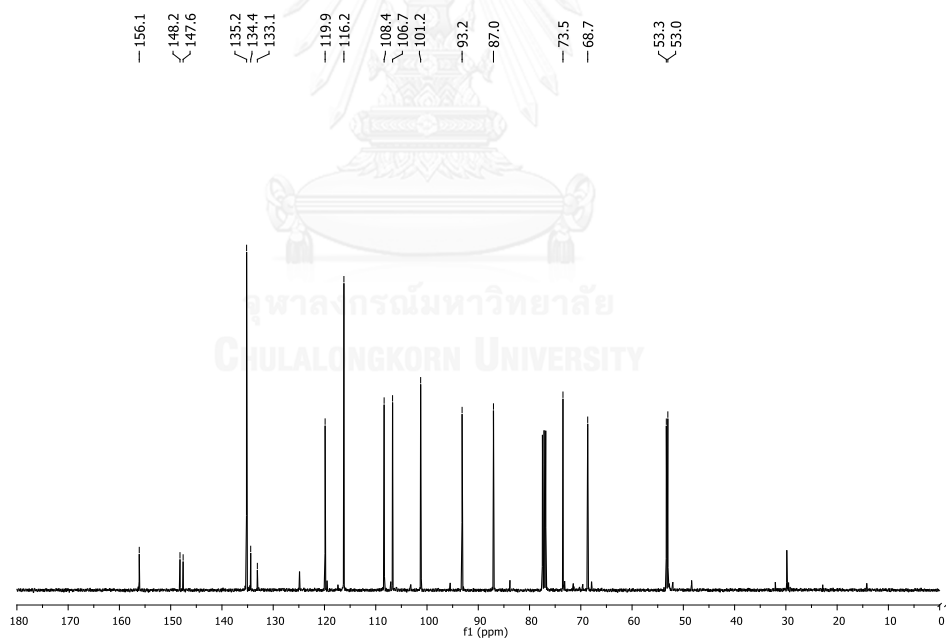
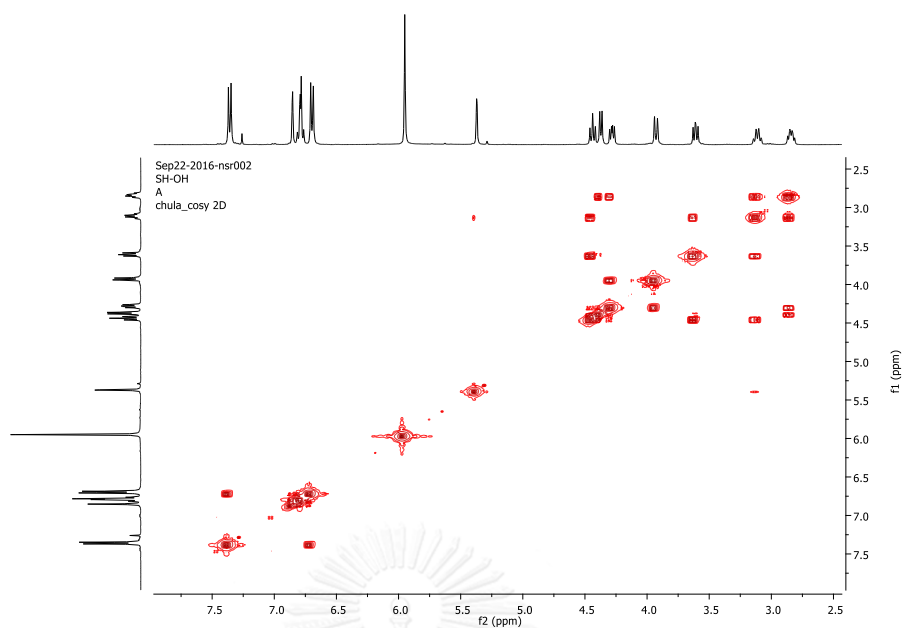
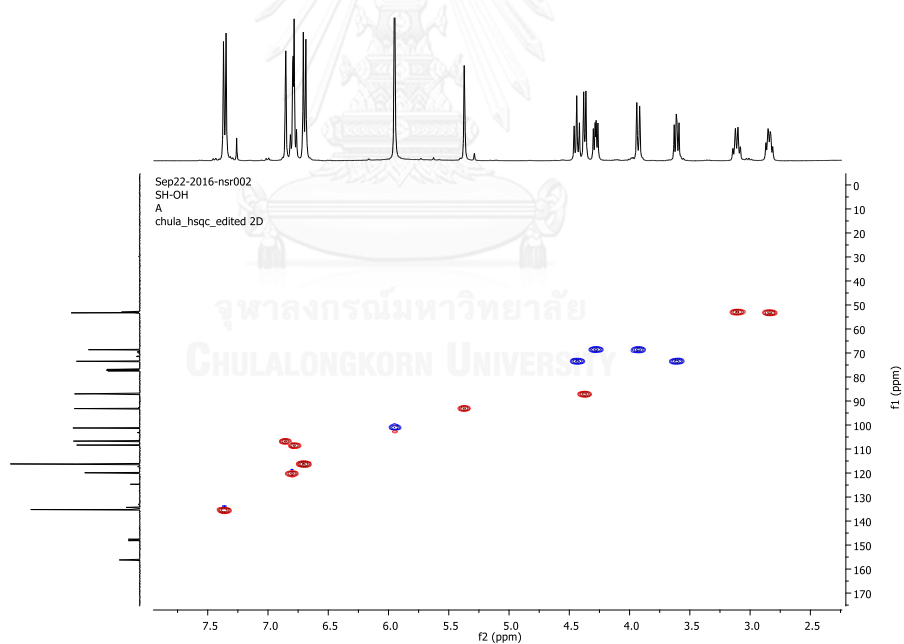
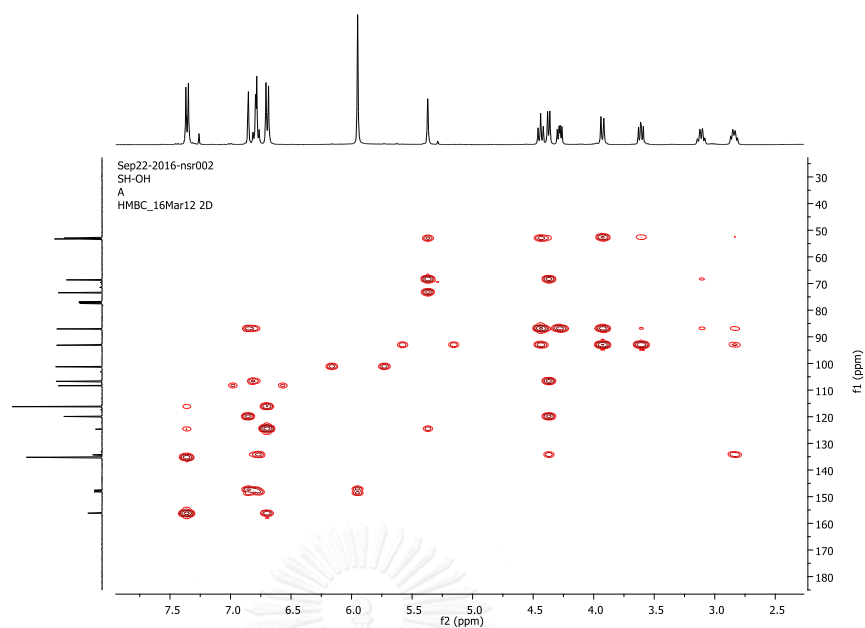
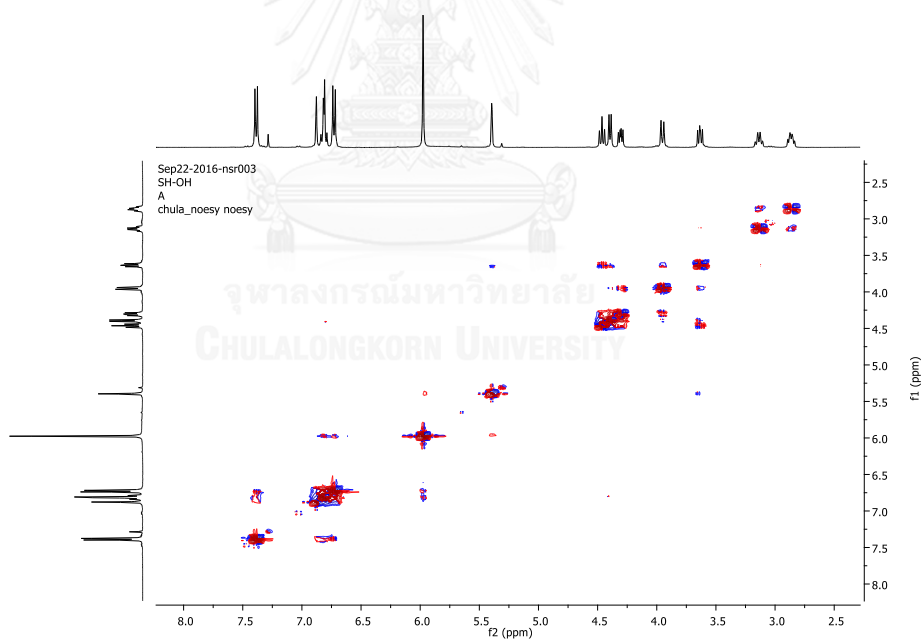


Figure 55. ^{13}C NMR spectrum of compound 3.2r in CDCl_3

Figure 56. NOESY experiment of **3.2r** in CDCl_3 Figure 57. HSQC experiment of **3.2r** in CDCl_3

Figure 58. HMBC experiment of **3.2r** in CDCl₃Figure 59. NOESY experiment of **3.2r** in CDCl₃

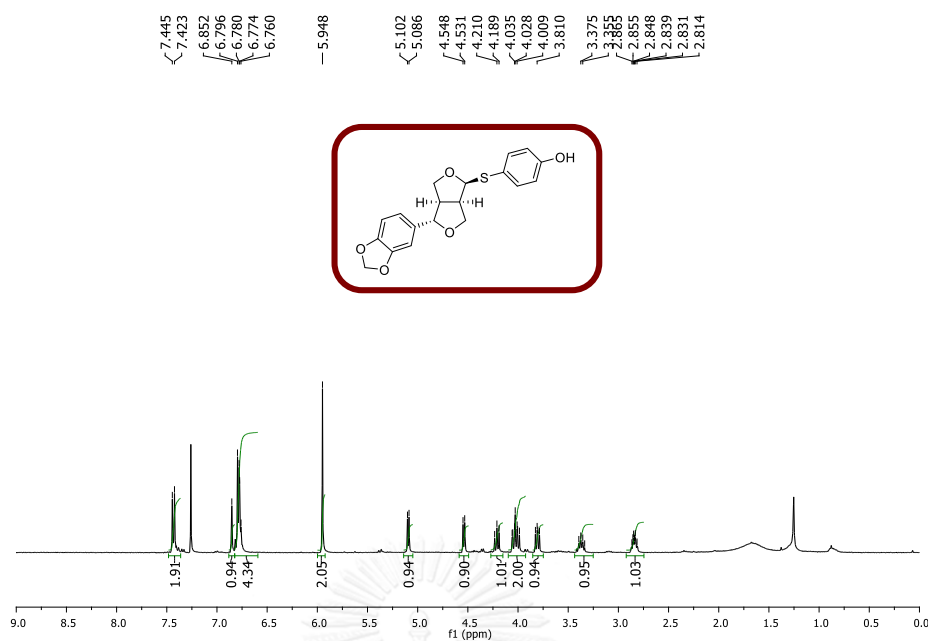


Figure 60. ^{13}C NMR spectrum of compound *epi-3.2r* in CDCl_3

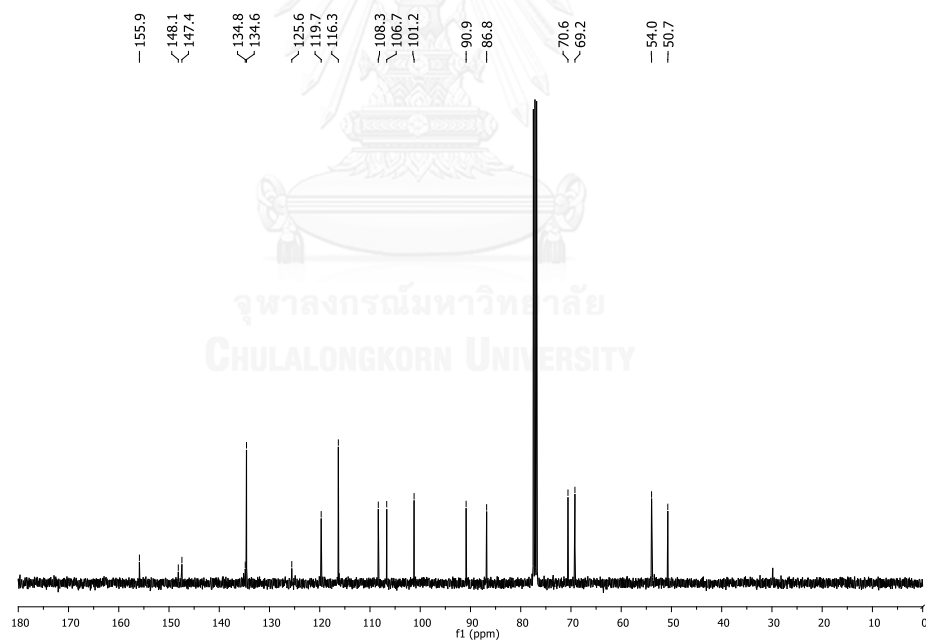


Figure 61. ^{13}C NMR spectrum of compound *epi-3.2r* in CDCl_3

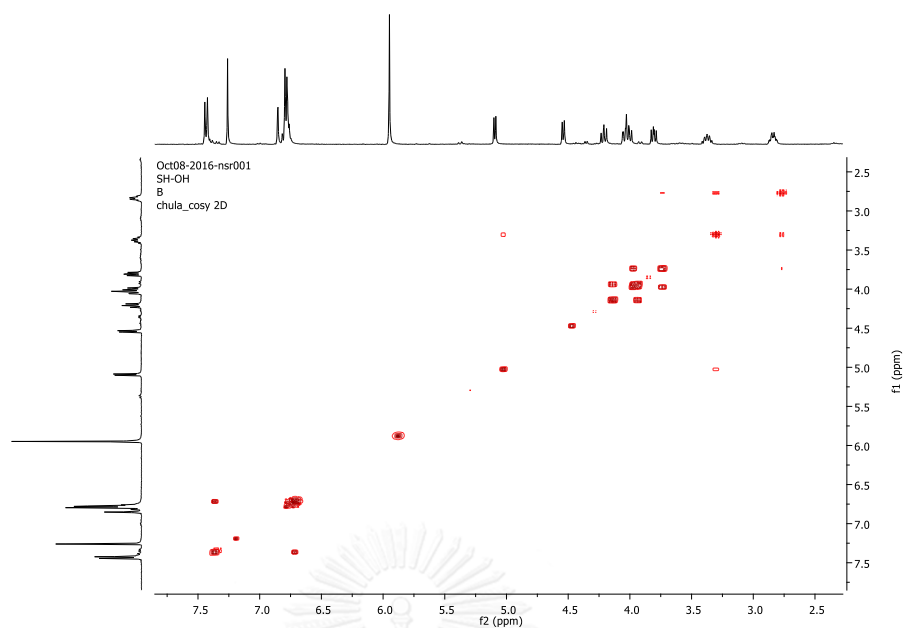


Figure 62. COSY experiment of *epi-3.2r* in CDCl₃

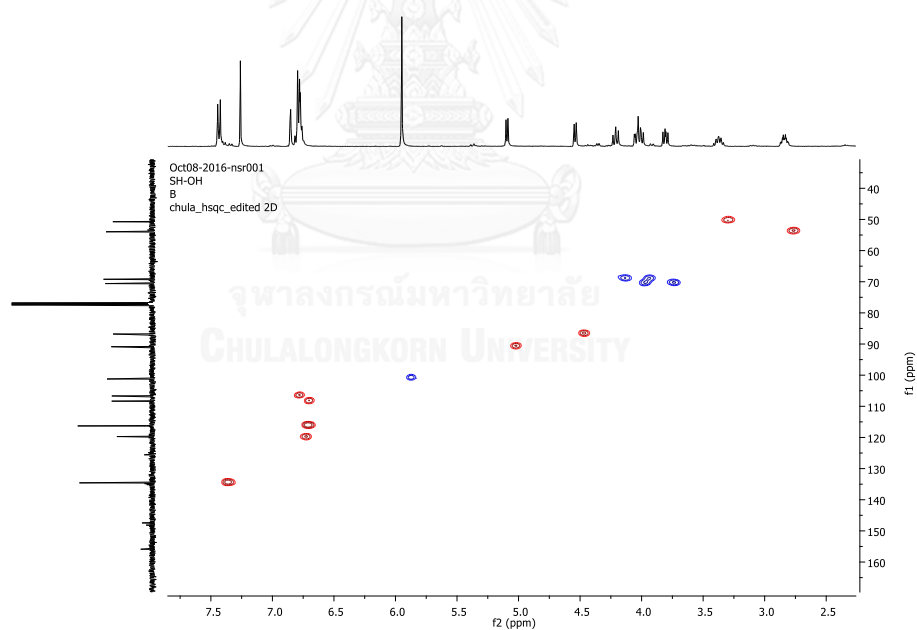


Figure 63. HSQC experiment of *epi-3.2r* in CDCl₃

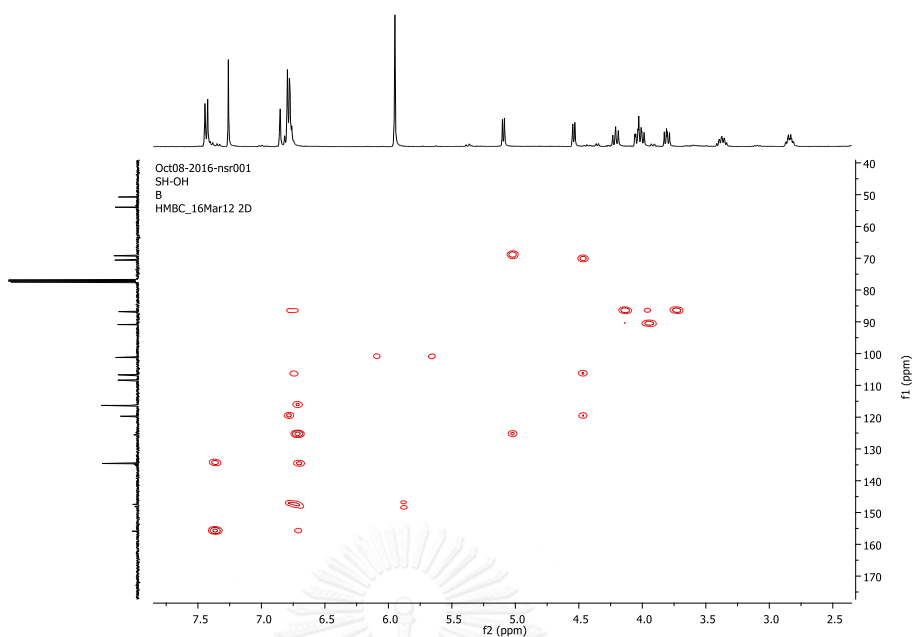


Figure 64. HMBC experiment of *epi-3.2r* in CDCl₃

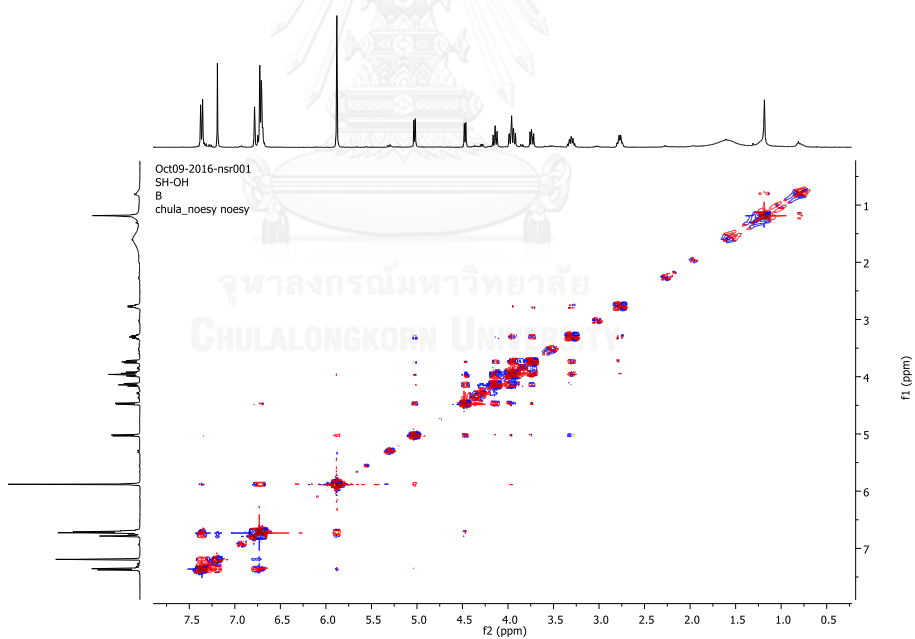


Figure 65. NOESY experiment of *epi-3.2r* in CDCl₃

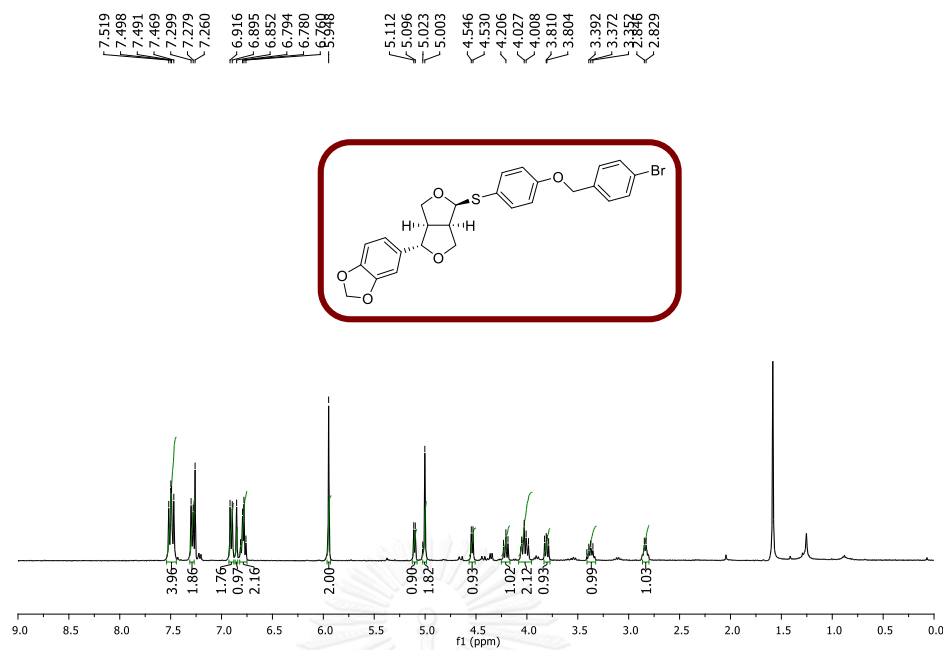


Figure 66. ^{13}C NMR spectrum of compound *epi-3.2r-pBBE* in CDCl_3

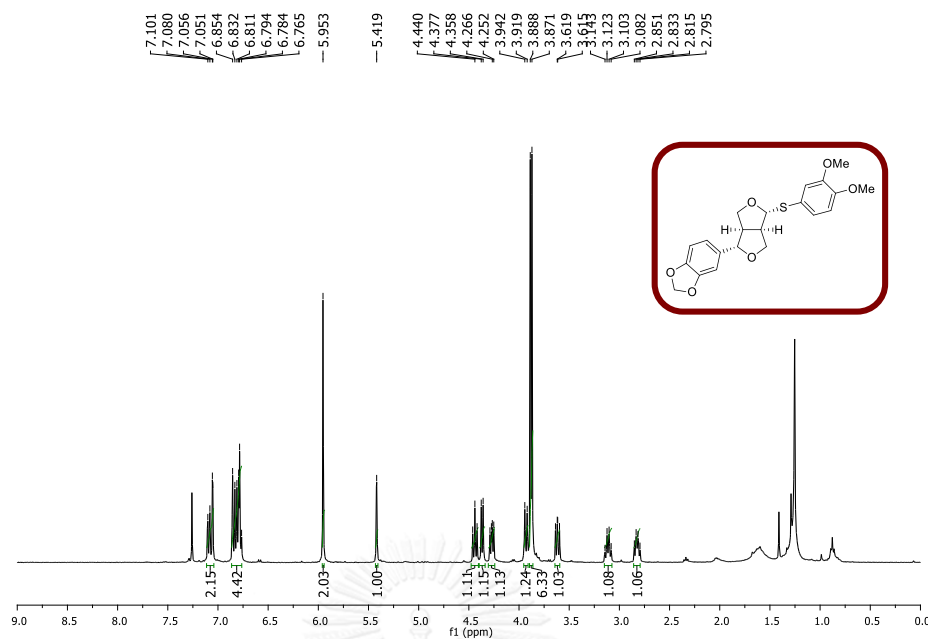


Figure 67. ^1H NMR spectrum of compound 3.2s in CDCl_3

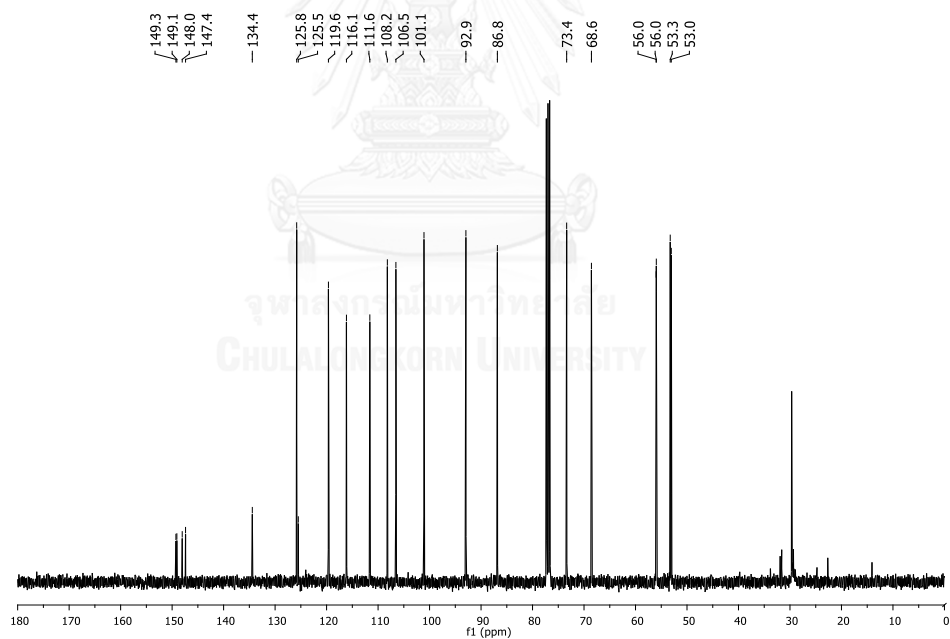


Figure 68. ^{13}C NMR spectrum of compound 3.2s in CDCl_3

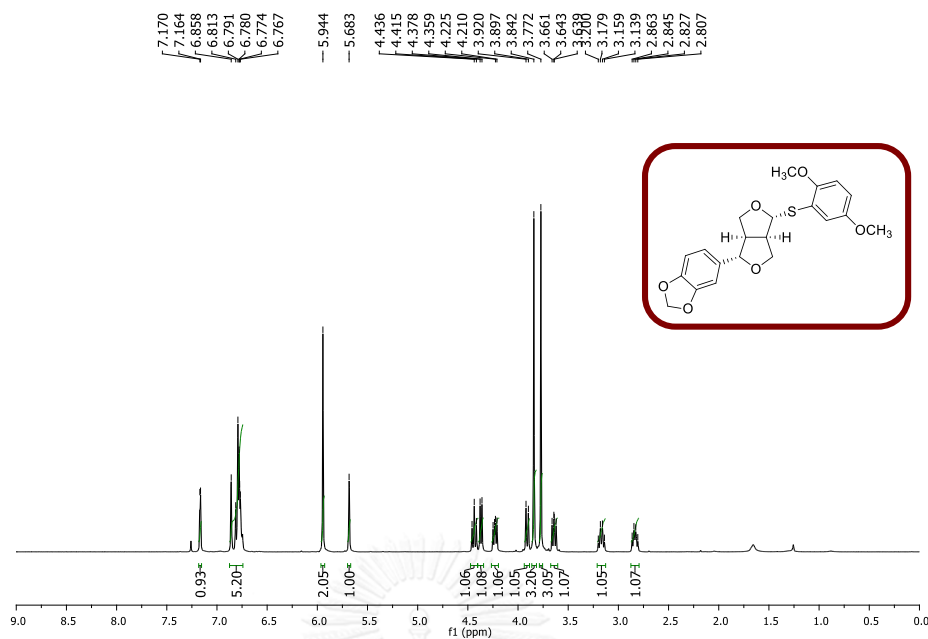


Figure 69. ^1H NMR spectrum of compound 3.2t in CDCl_3

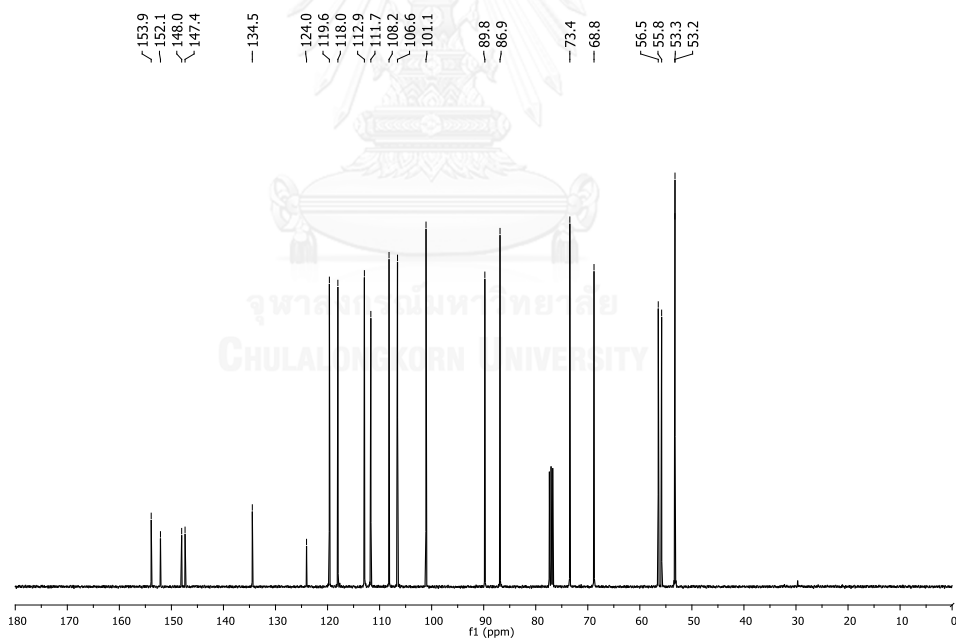


Figure 70. ^{13}C NMR spectrum of compound 3.2t in CDCl_3

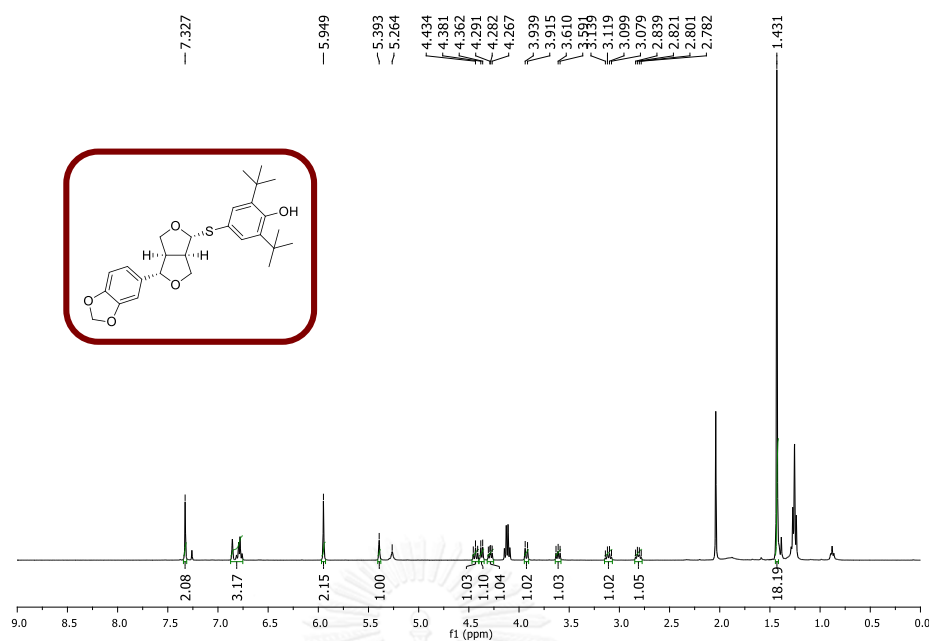


Figure 71. ^1H NMR spectrum of compound 3.2u in CDCl_3

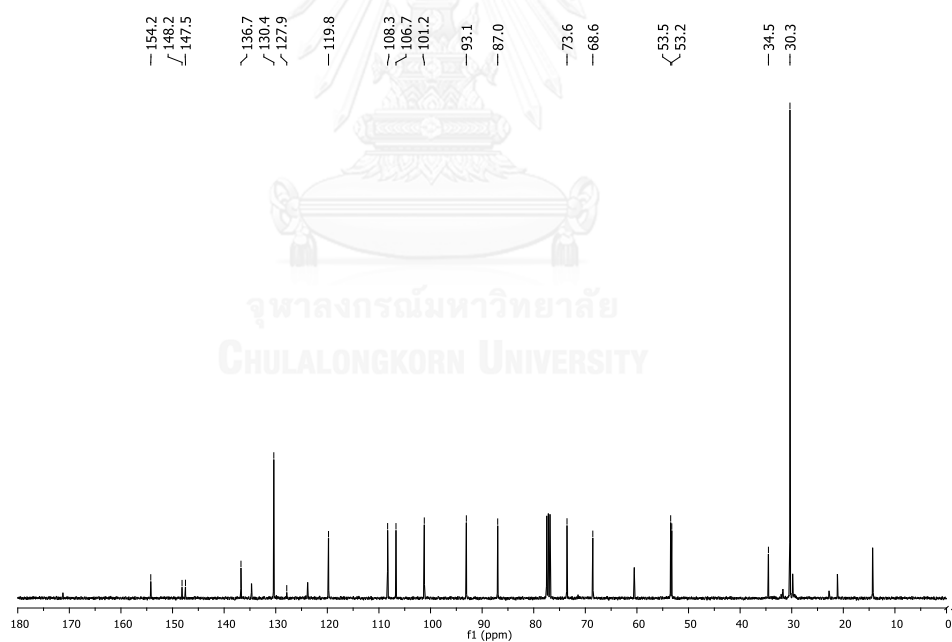


Figure 72. ^{13}C NMR spectrum of compound 3.2u in CDCl_3

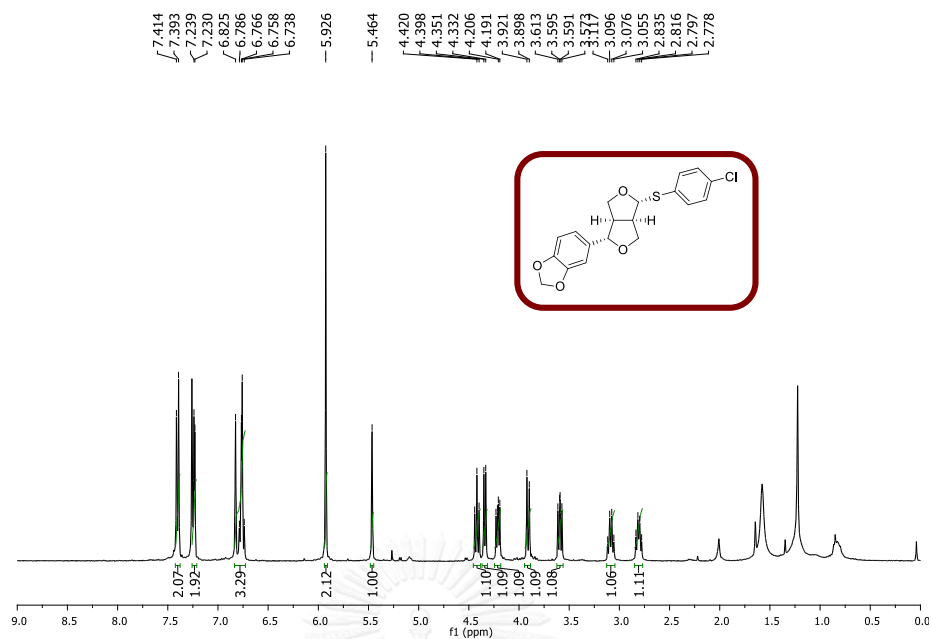


Figure 73. ^1H NMR spectrum of compound 3.2v in CDCl_3

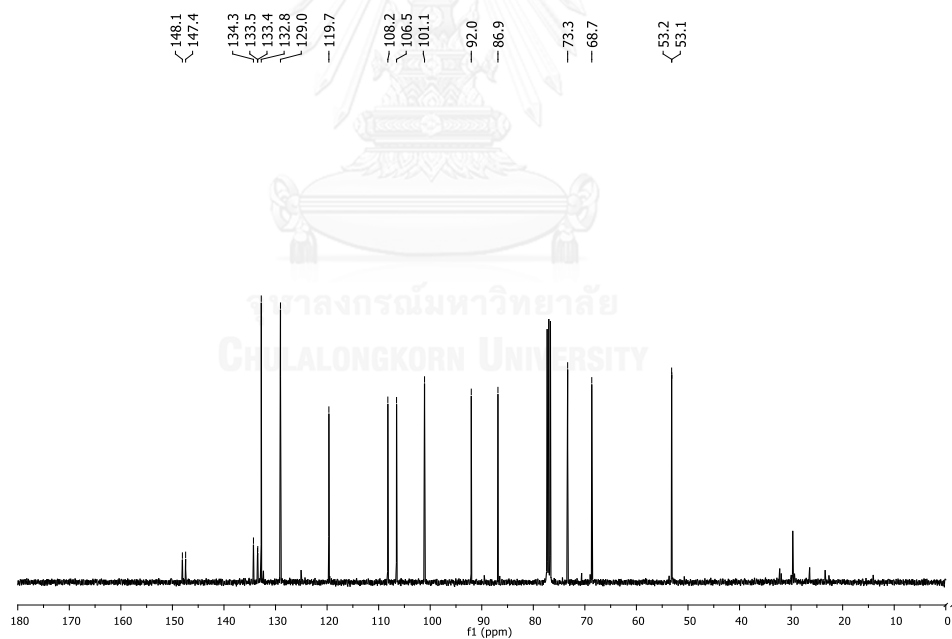


Figure 74. ^{13}C NMR spectrum of compound 3.2v in CDCl_3

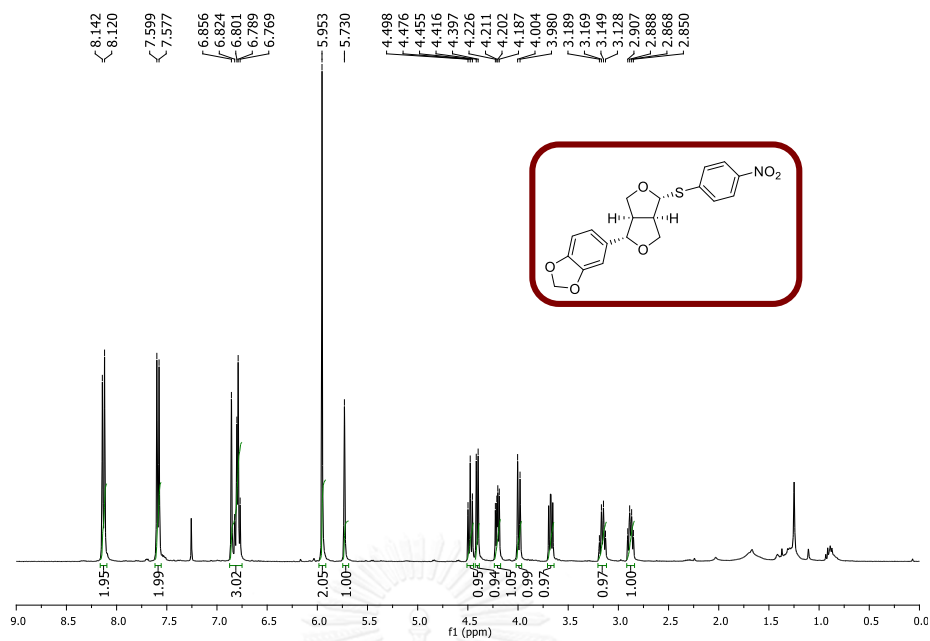


Figure 75. ^1H NMR spectrum of compound 3.2w in CDCl_3

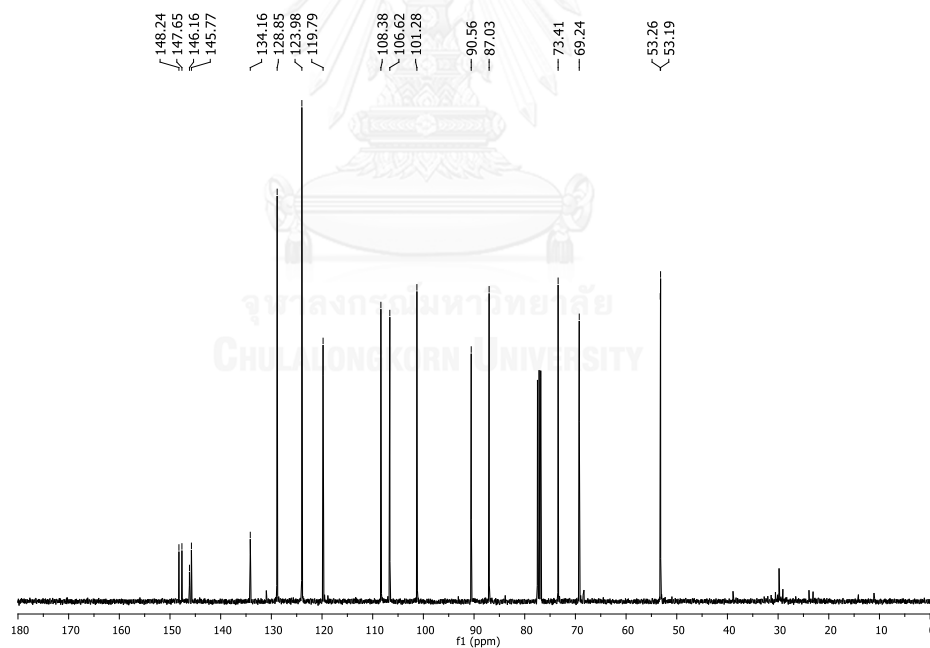


Figure 76. ^{13}C NMR spectrum of compound 3.2w in CDCl_3

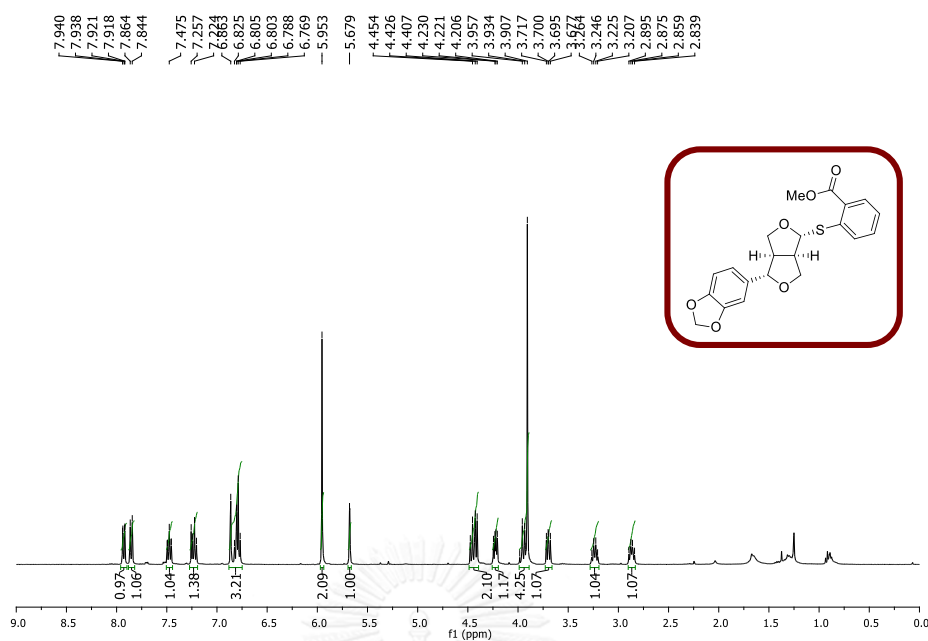


Figure 77. ^1H NMR spectrum of compound 3.2x in CDCl_3

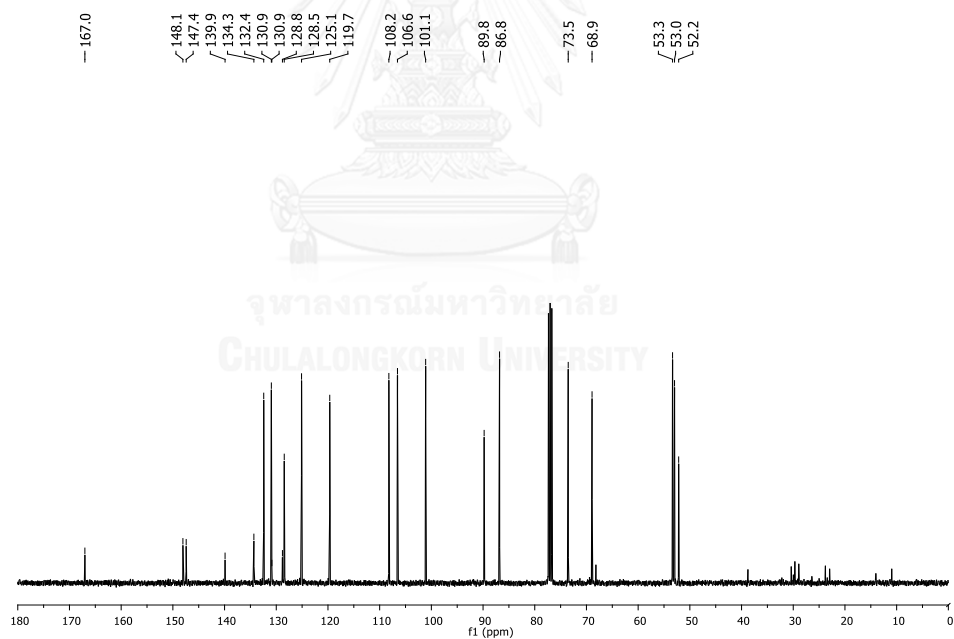


Figure 78. ^{13}C NMR spectrum of compound 3.2x in CDCl_3

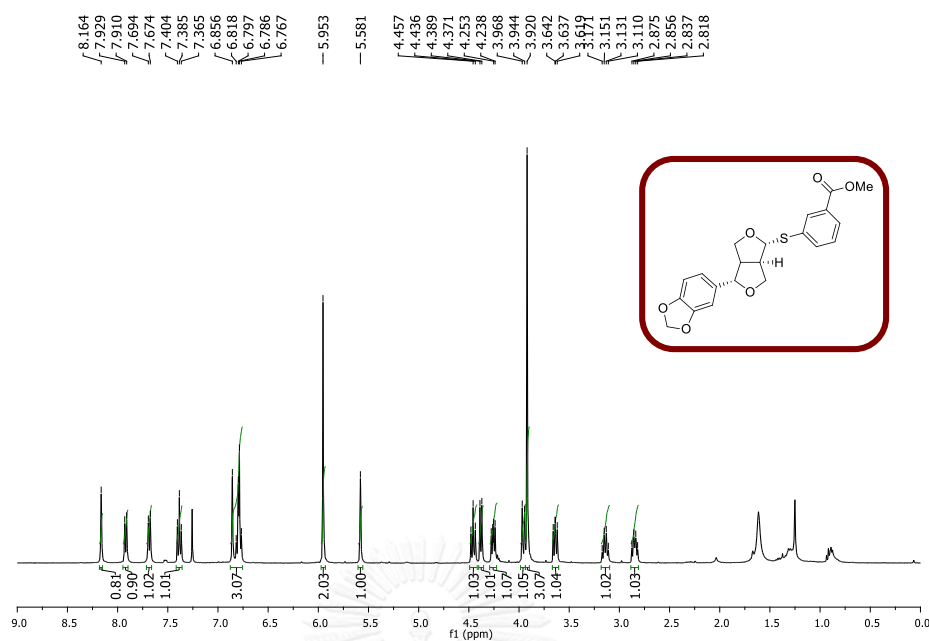


Figure 79. ^1H NMR spectrum of compound 3.2y in CDCl_3

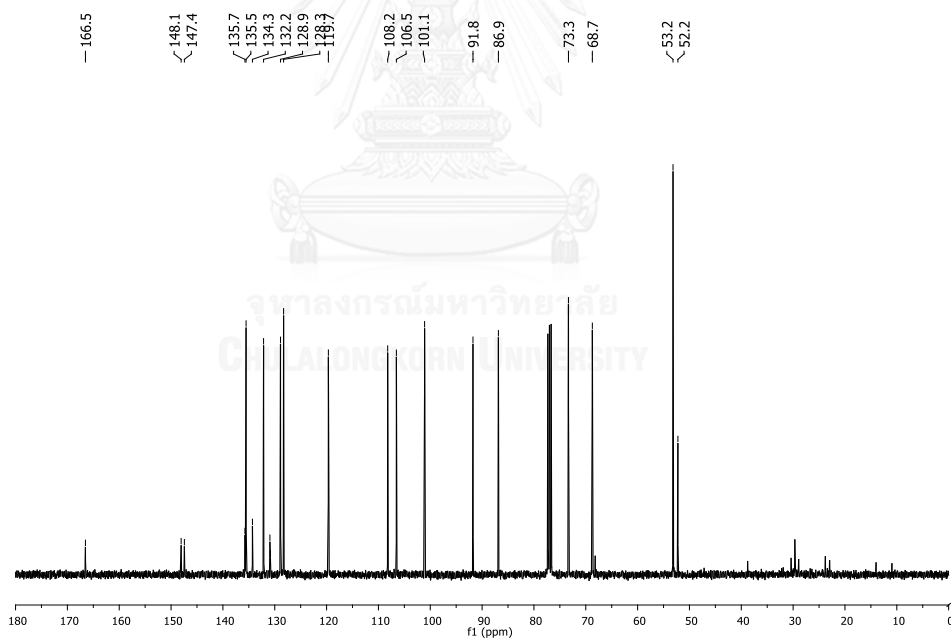


Figure 80. ^{13}C NMR spectrum of compound 3.2y in CDCl_3

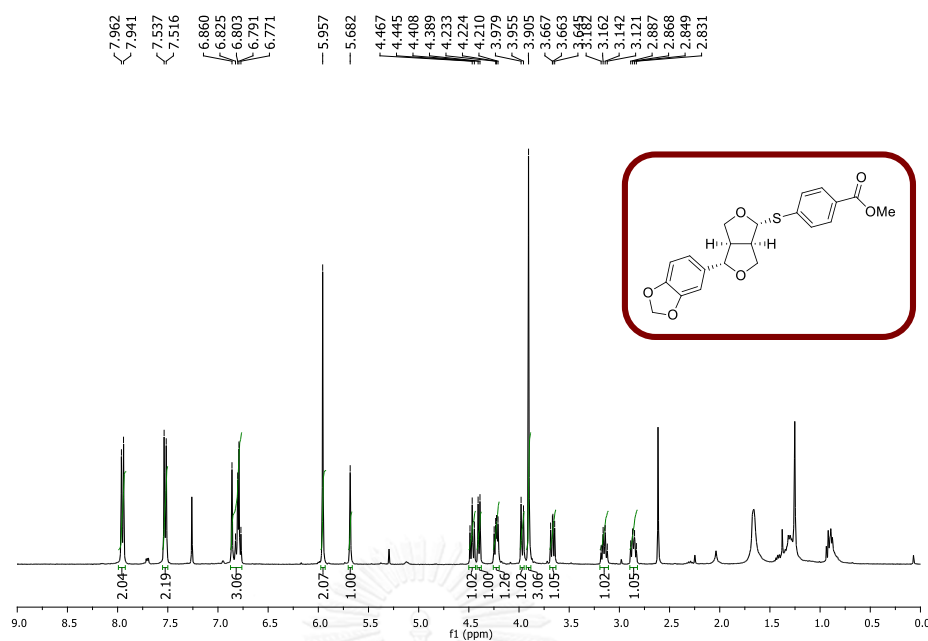


Figure 81. ^1H NMR spectrum of compound 3.2z in CDCl_3

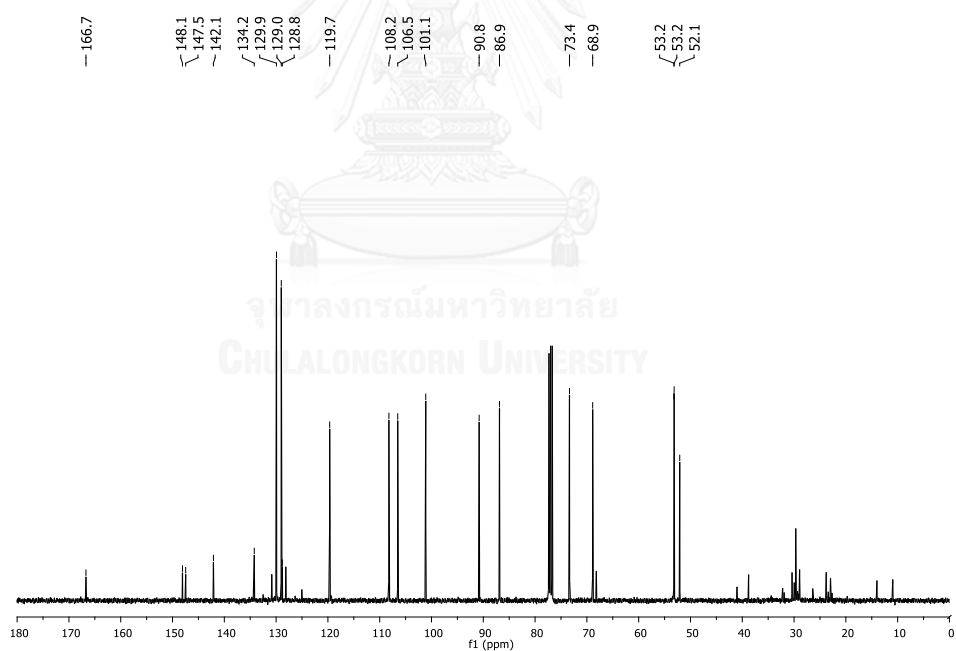


Figure 82. ^{13}C NMR spectrum of compound 3.2z in CDCl_3

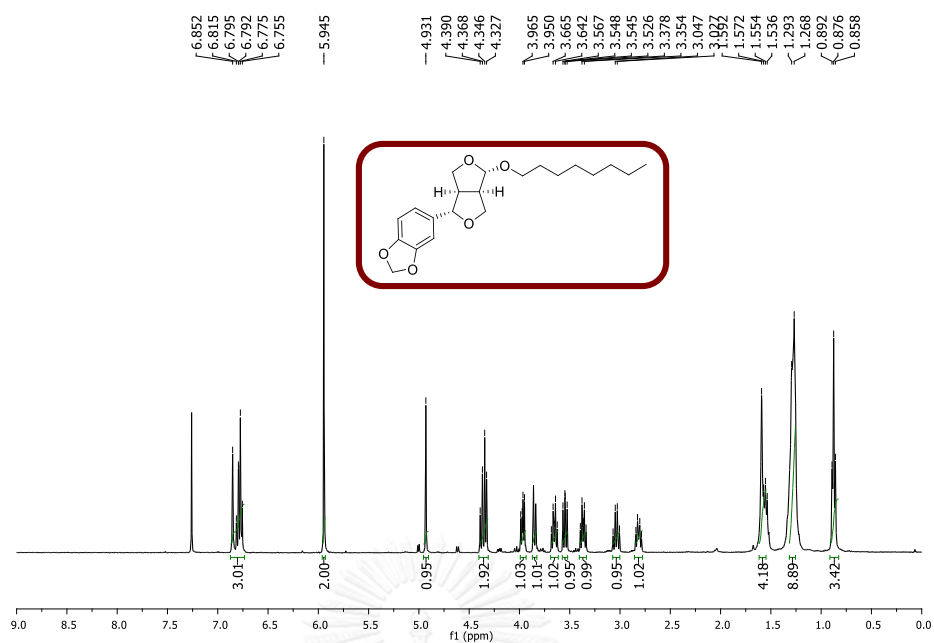


Figure 83. ^1H NMR spectrum of compound 3.3A in CDCl_3

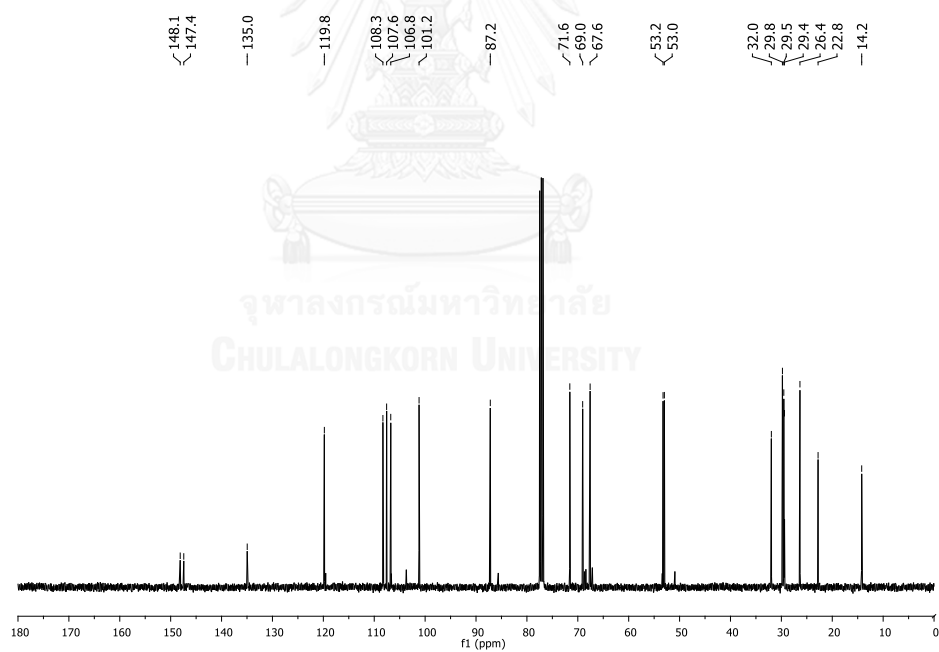


Figure 84. ^{13}C NMR spectrum of compound 3.3A in CDCl_3

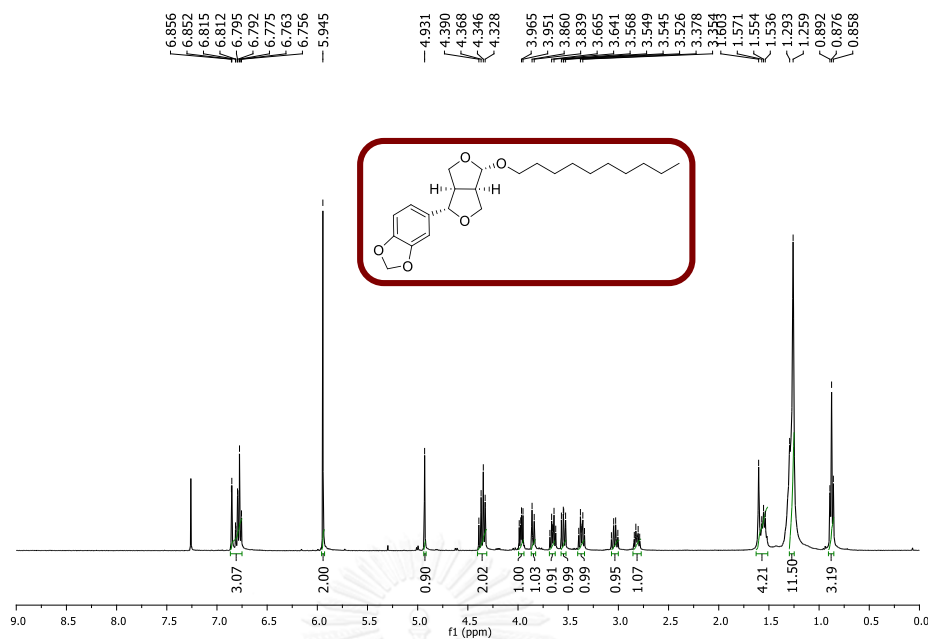


Figure 85. ^1H NMR spectrum of compound 3.3B in CDCl_3

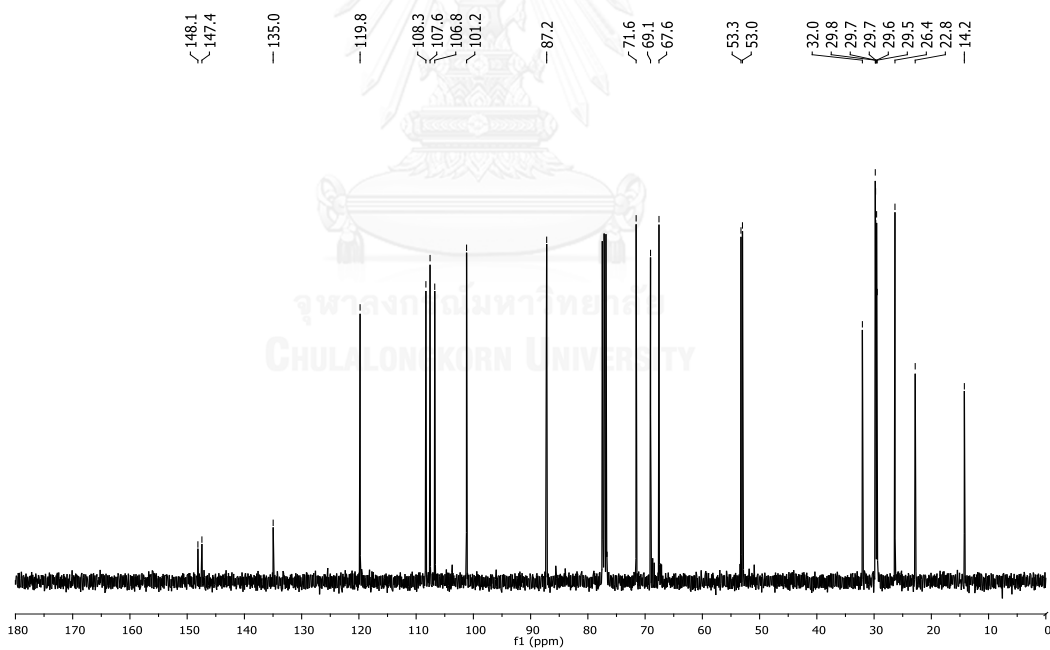


Figure 86. ^{13}C NMR spectrum of compound 3.3B in CDCl_3

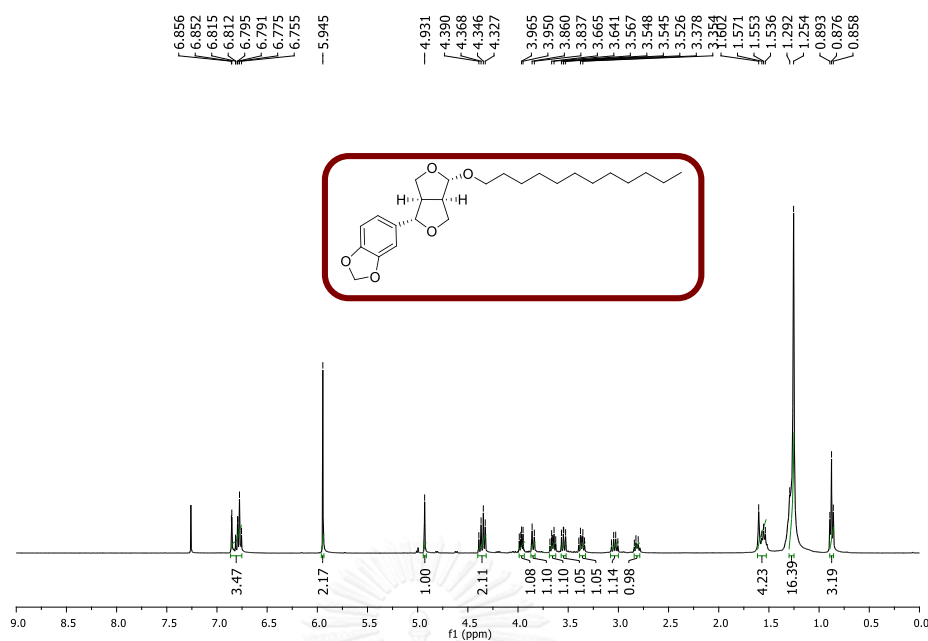


Figure 87. ^1H NMR spectrum of compound 3.3C in CDCl_3

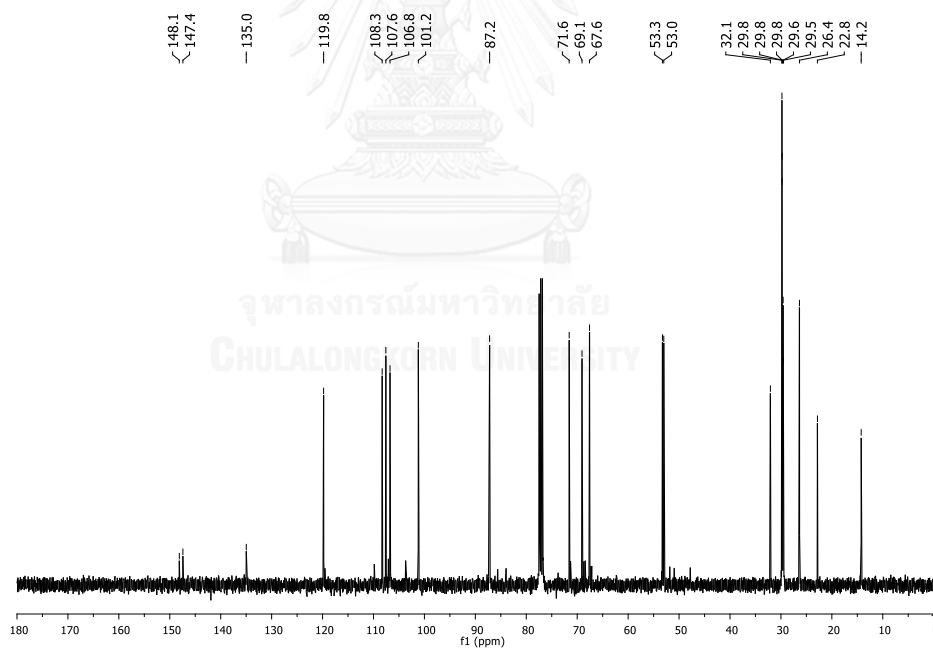


Figure 88. ^{13}C NMR spectrum of compound 3.3C in CDCl_3

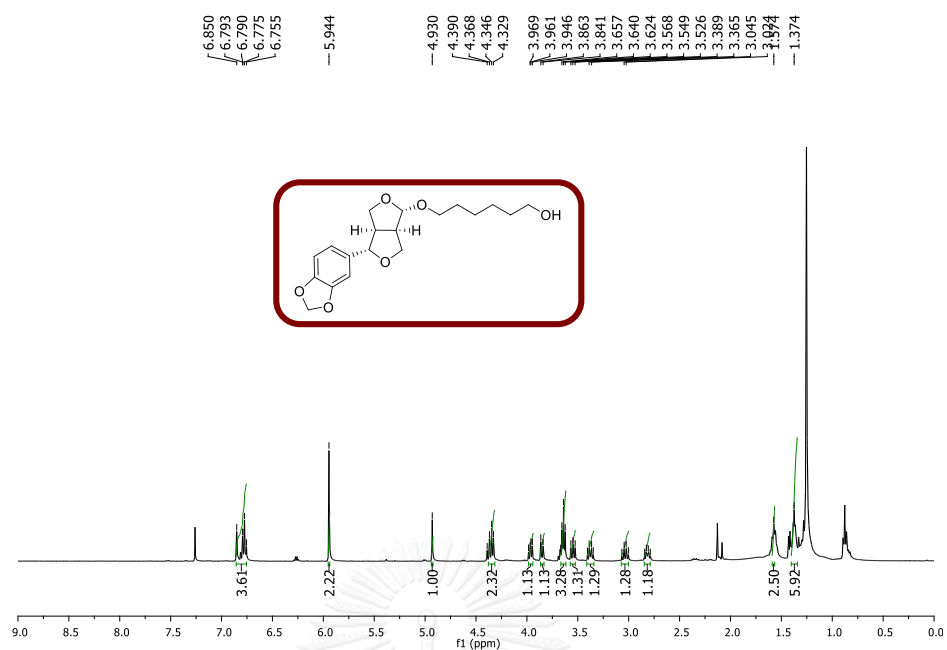


Figure 89. ^1H NMR spectrum of compound 3.3D in CDCl_3

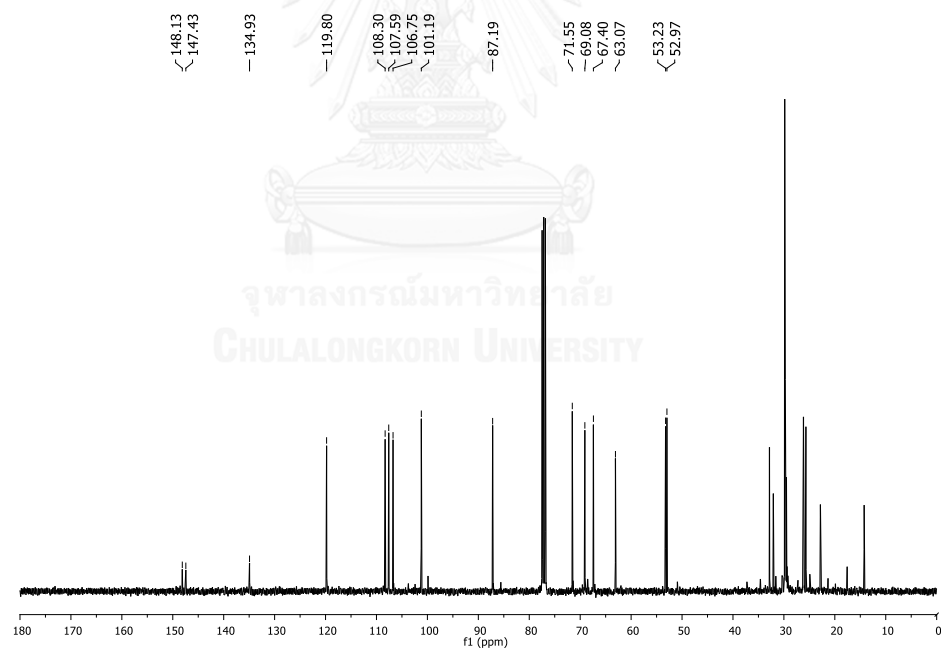


Figure 90. ^{13}C NMR spectrum of compound 3.3D in CDCl_3

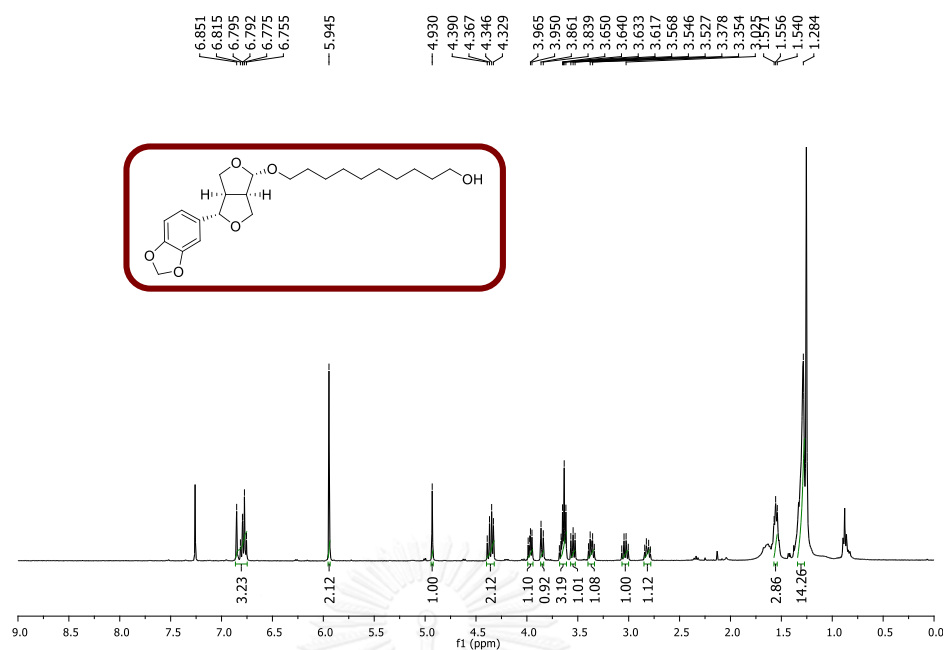


Figure 91. ^1H NMR spectrum of compound 3.3E in CDCl_3

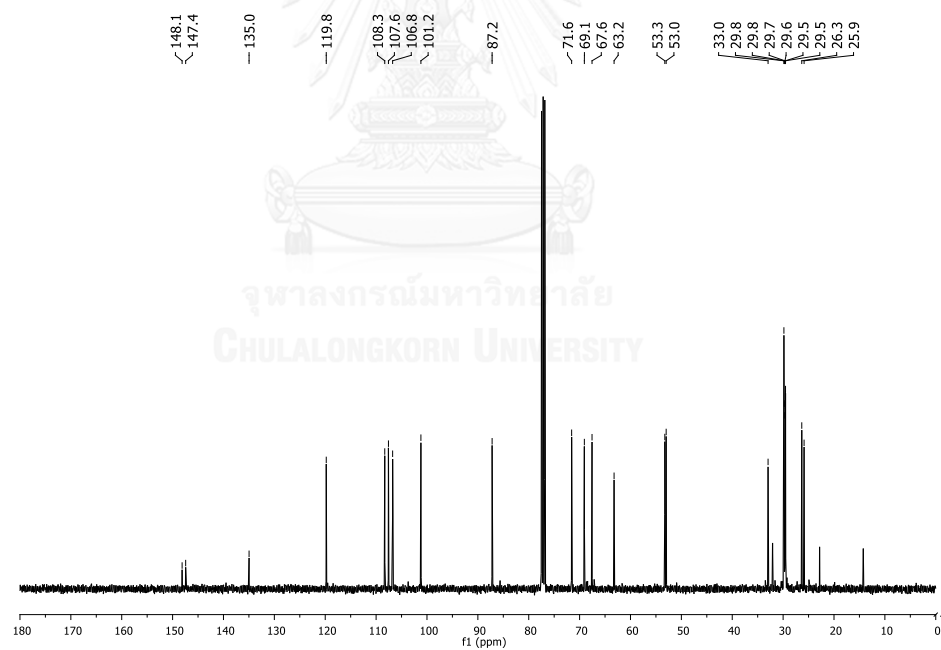


Figure 92. ^{13}C NMR spectrum of compound 3.3E in CDCl_3

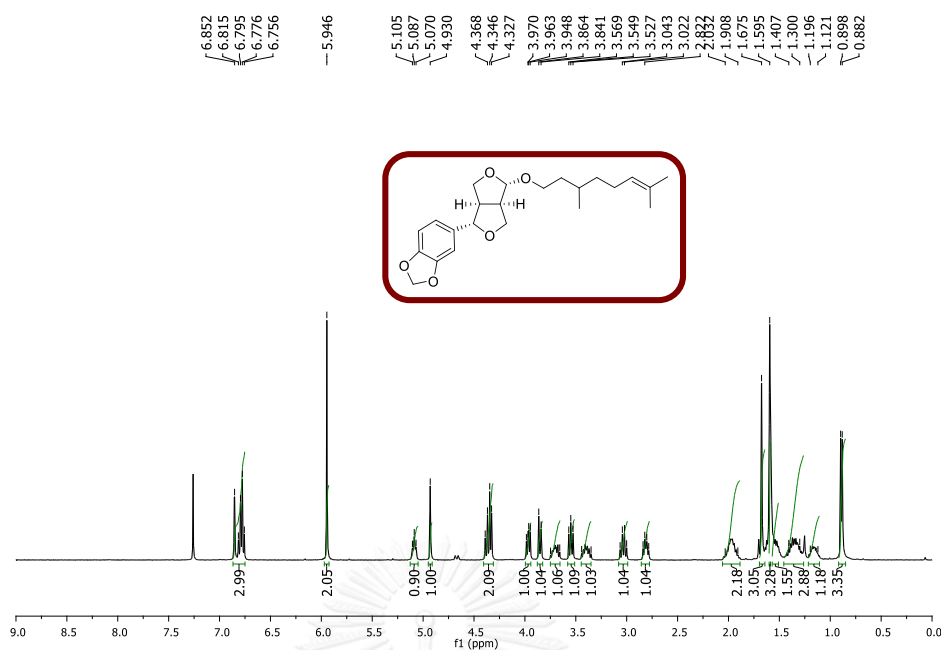


Figure 93. ^1H NMR spectrum of compound 3.3F in CDCl_3

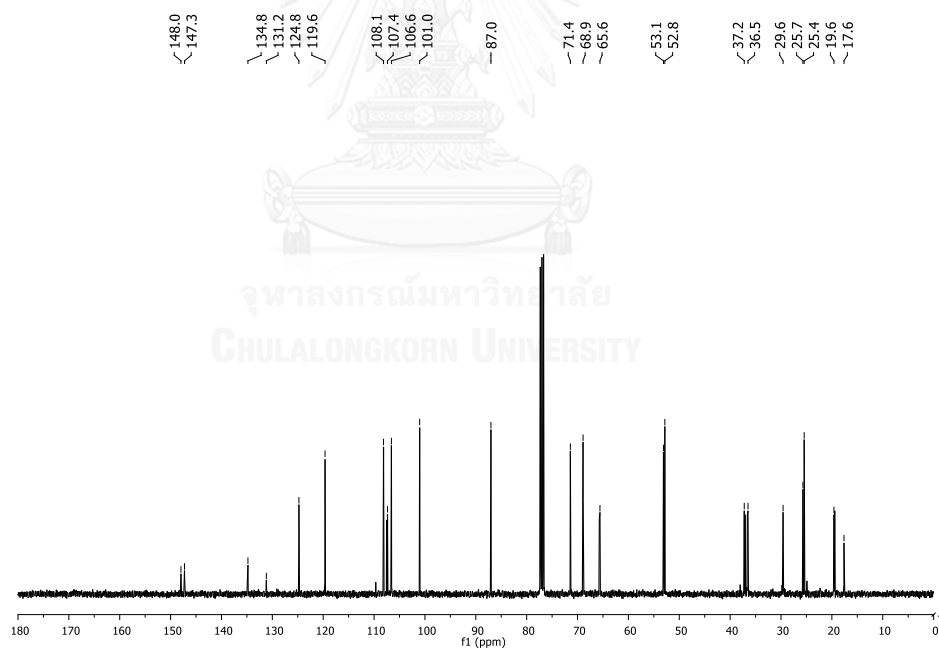


Figure 94. ^{13}C NMR spectrum of compound 3.3F in CDCl_3

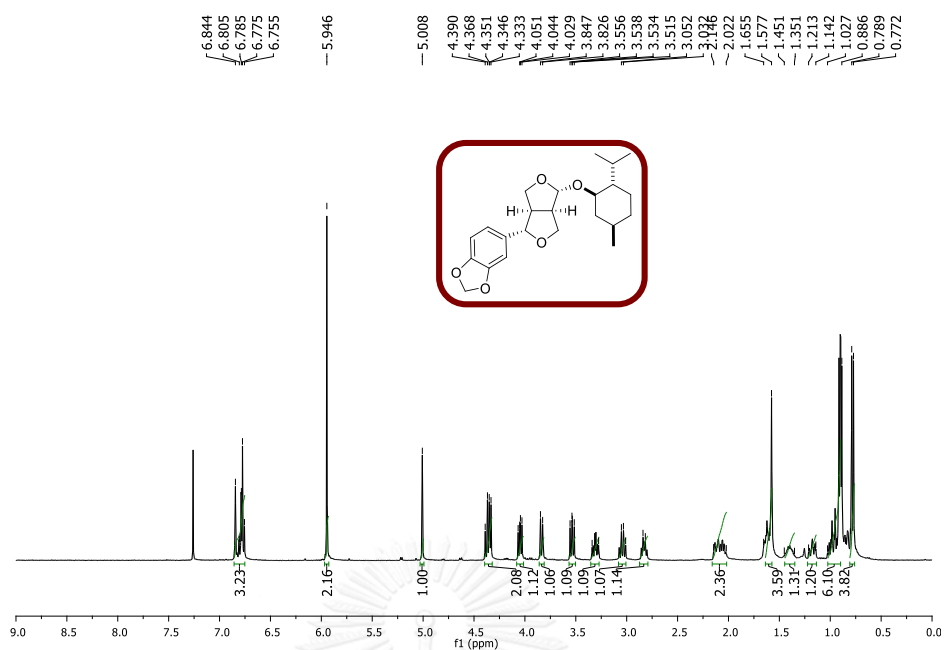


Figure 95. ^1H NMR spectrum of compound 3.3G in CDCl_3

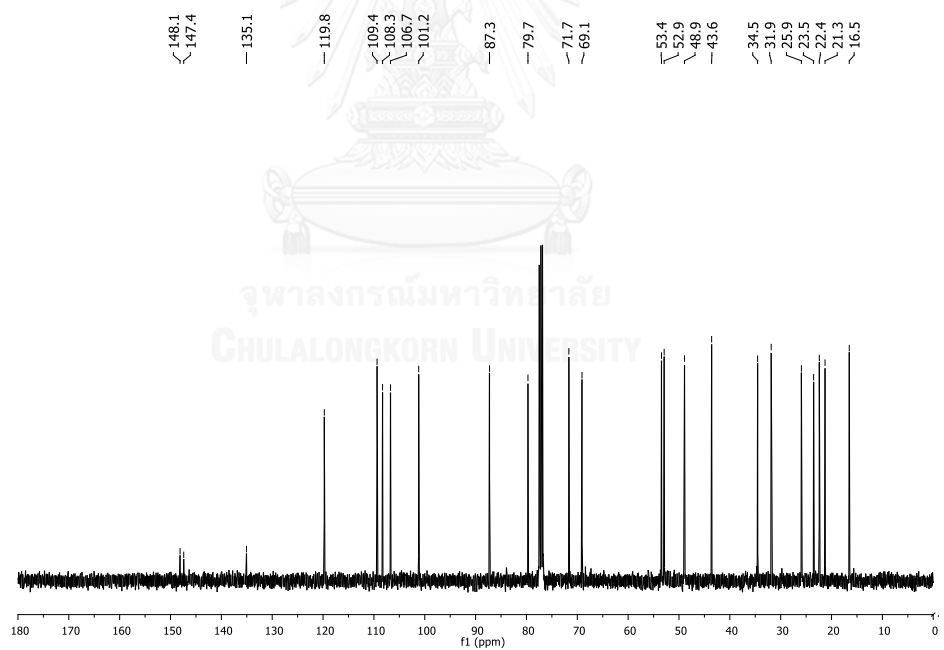


Figure 96. ^{13}C NMR spectrum of compound 3.3G in CDCl_3

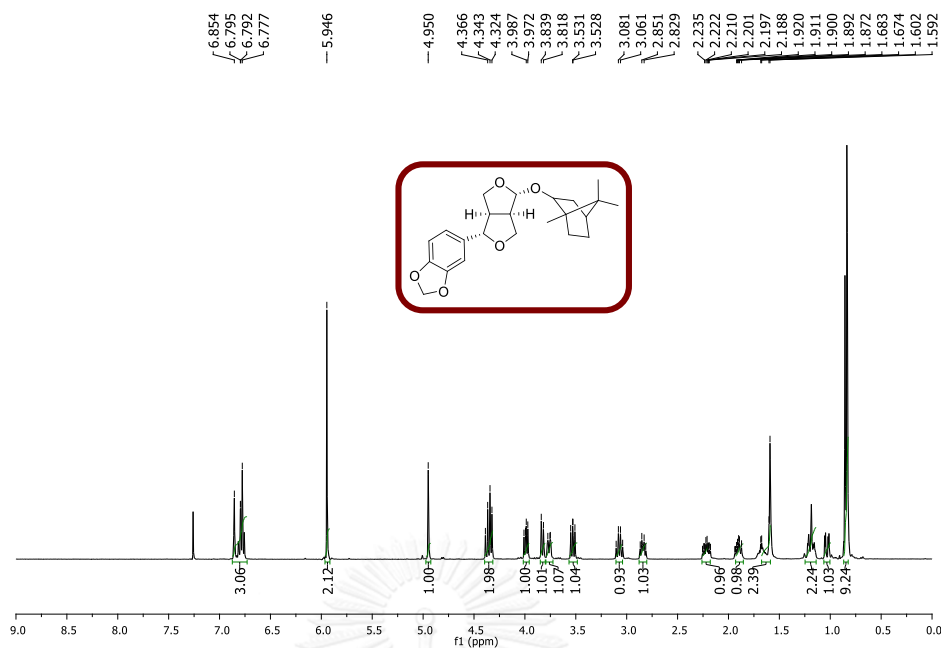


Figure 97. ^1H NMR spectrum of compound 3.3H in CDCl_3

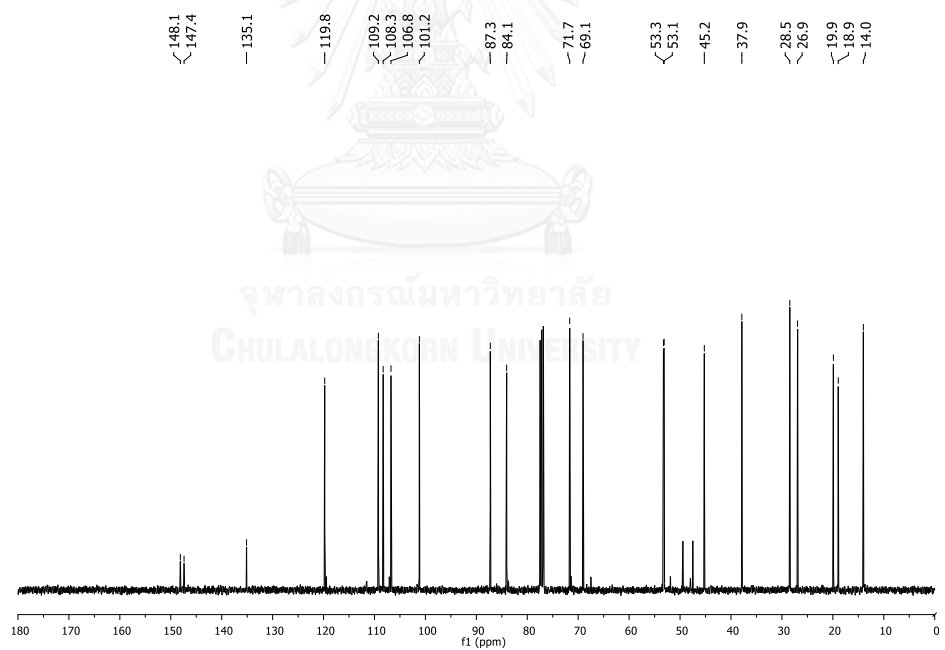


Figure 98. ^{13}C NMR spectrum of compound 3.3H in CDCl_3

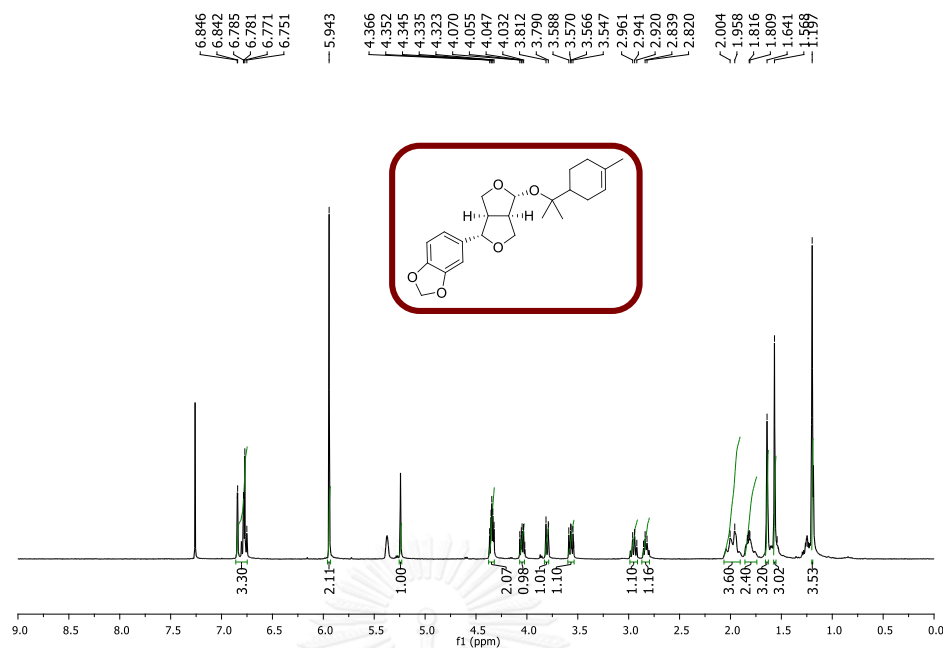


Figure 99. ^1H NMR spectrum of compound 3.31 in CDCl_3

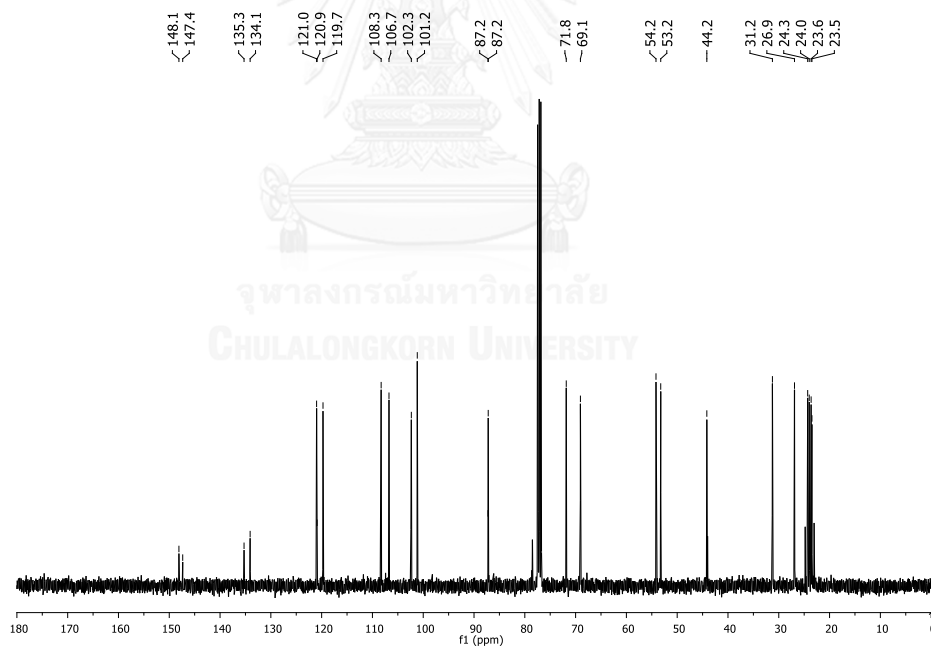


Figure 100. ^{13}C NMR spectrum of compound 3.31 in CDCl_3

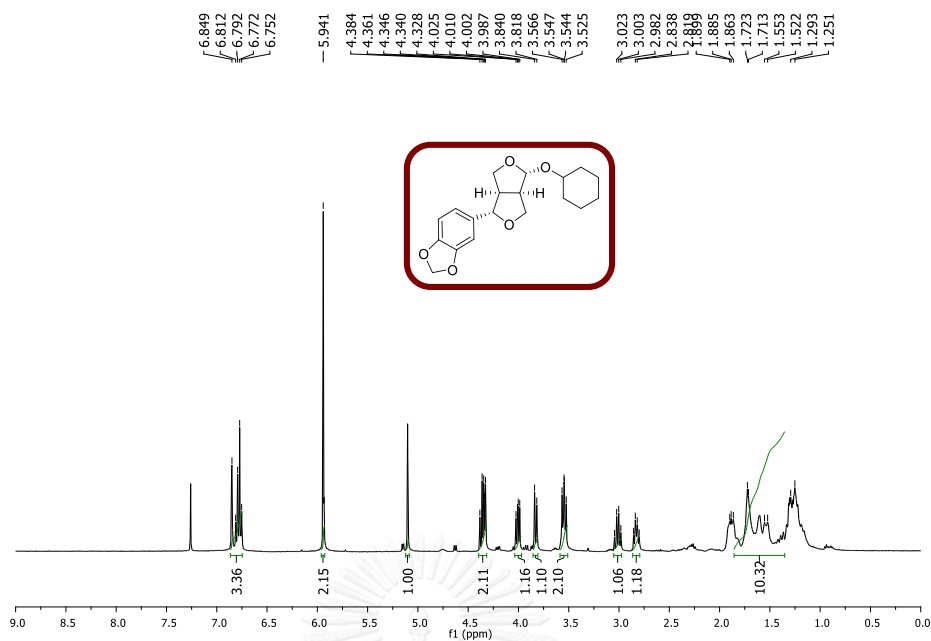


Figure 101. ^1H NMR spectrum of compound 3.3J in CDCl_3

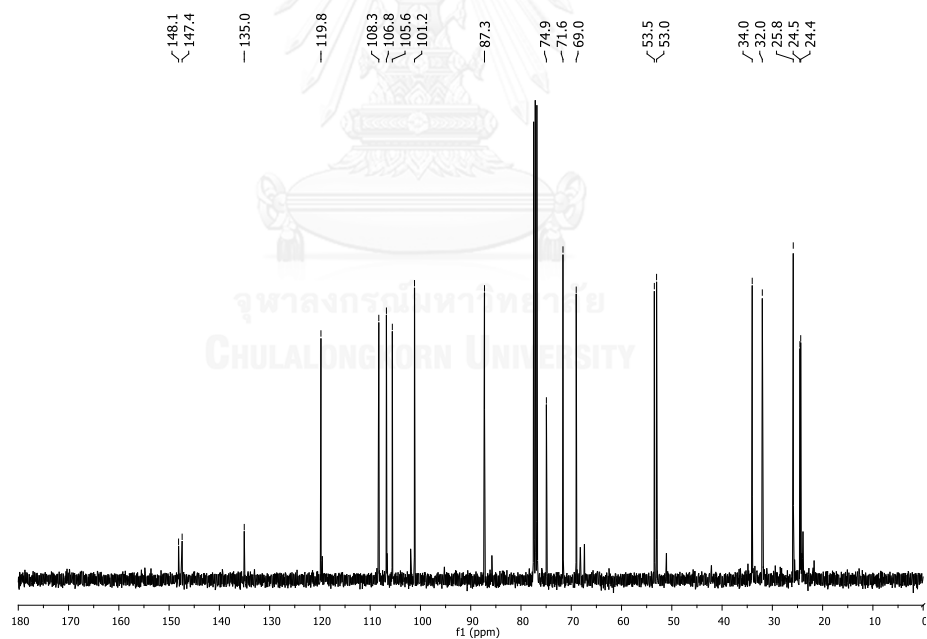


Figure 102. ^{13}C NMR spectrum of compound 3.3J in CDCl_3

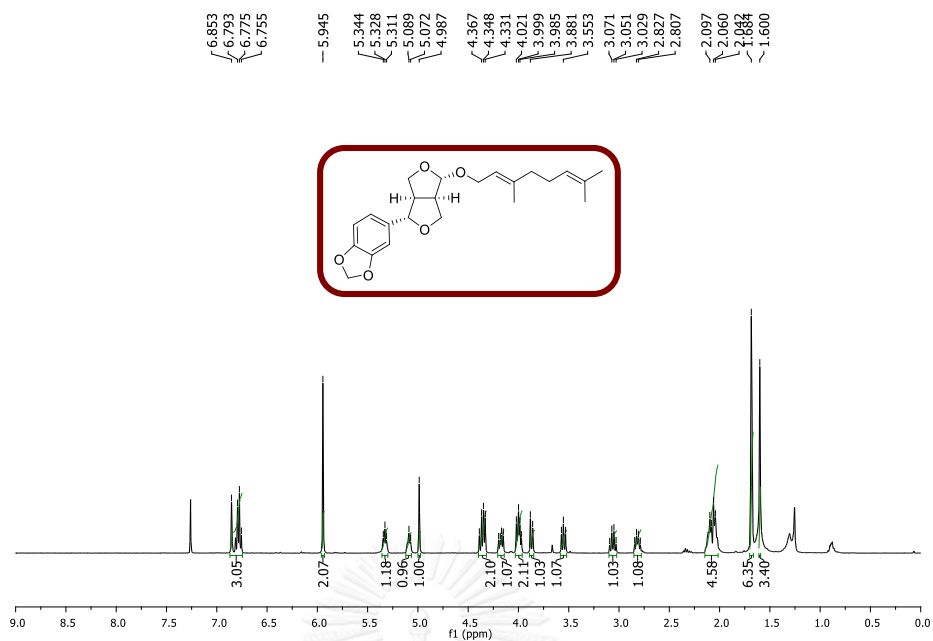


Figure 103. ^1H NMR spectrum of compound 3.3K in CDCl_3

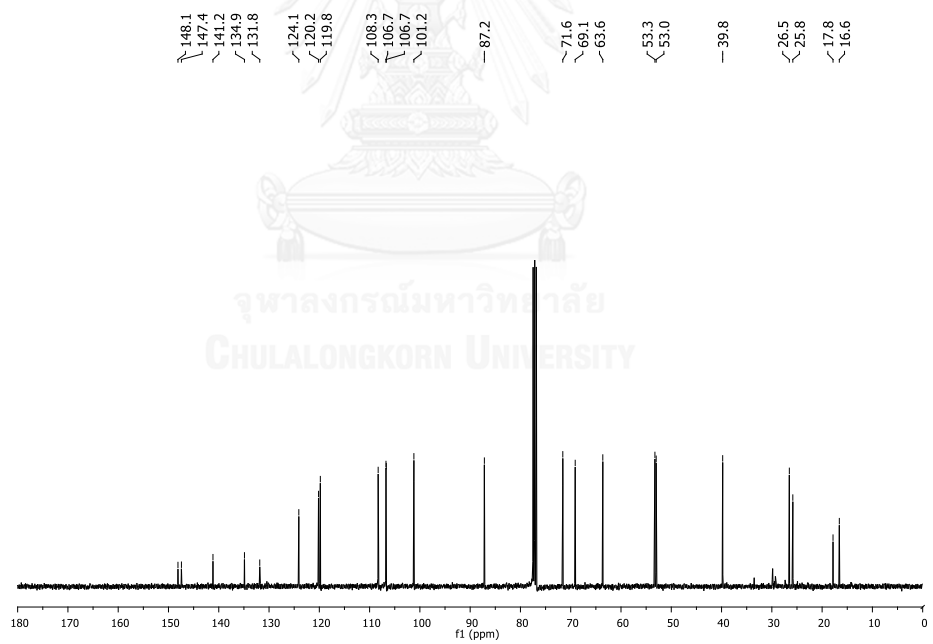


Figure 104. ^{13}C NMR spectrum of compound 3.3K in CDCl_3

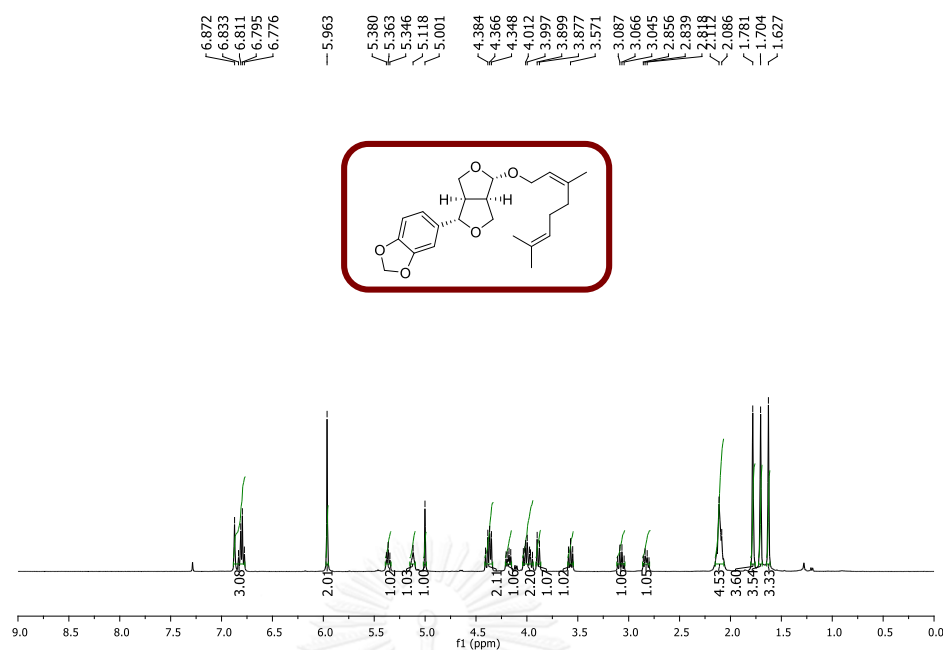


Figure 105. ¹H NMR spectrum of compound 3.3L in CDCl₃

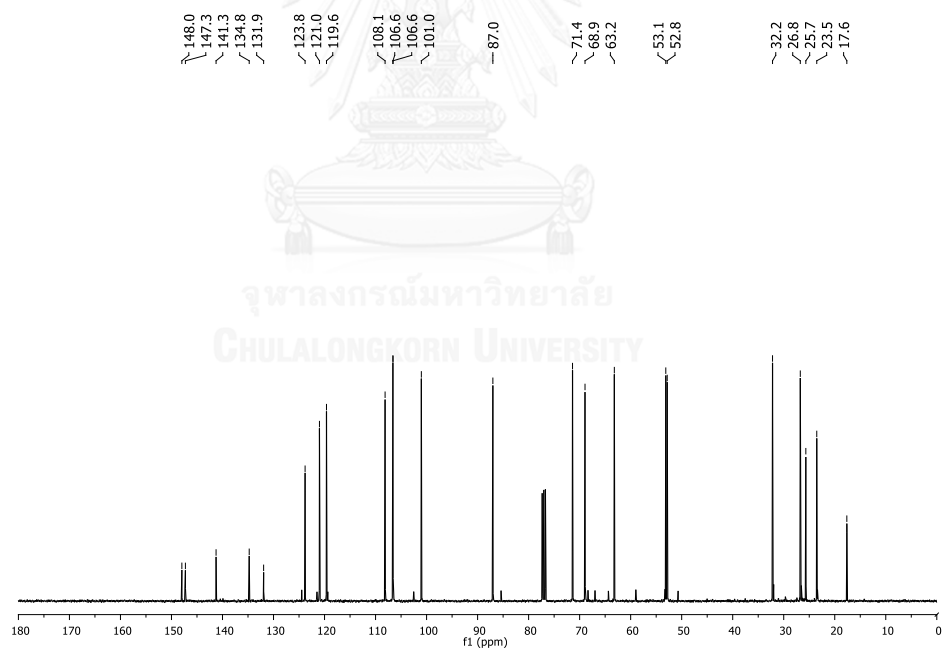
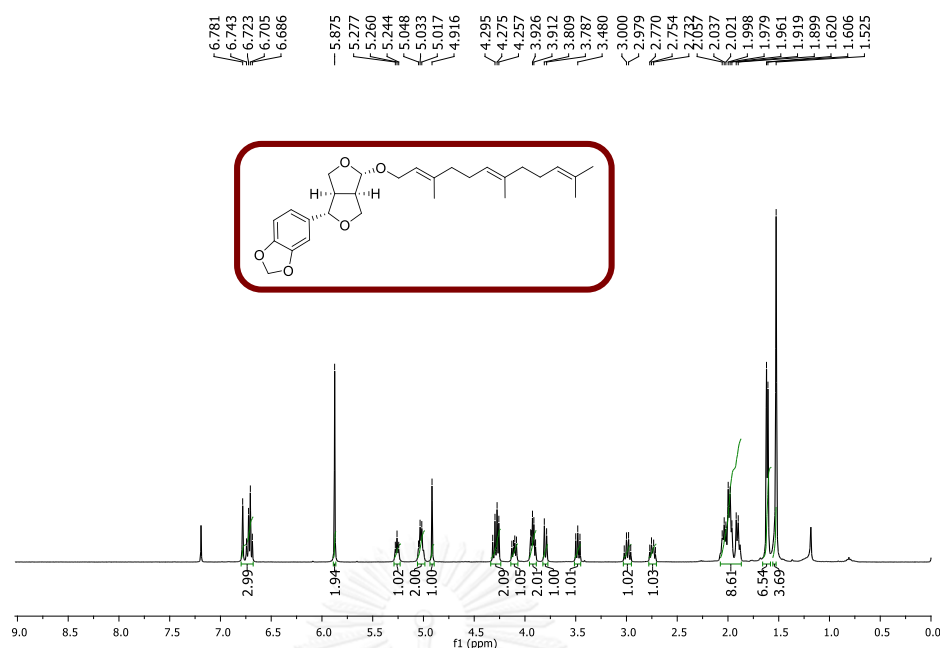
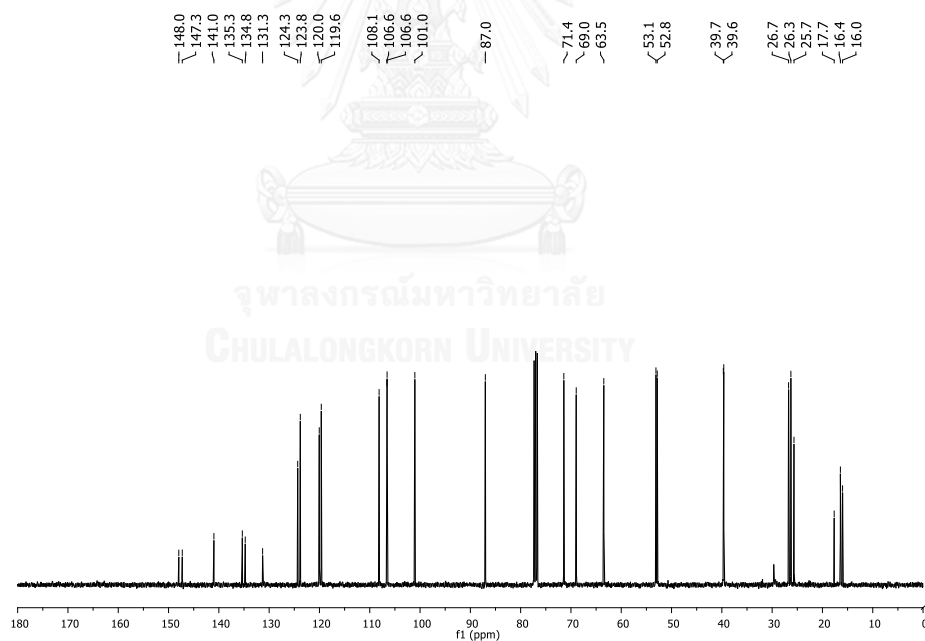


Figure 106. ¹³C NMR spectrum of compound 3.3L in CDCl₃

Figure 107. ¹H NMR spectrum of compound 3.3M in CDCl₃Figure 108. ¹³C NMR spectrum of compound 3.3M in CDCl₃

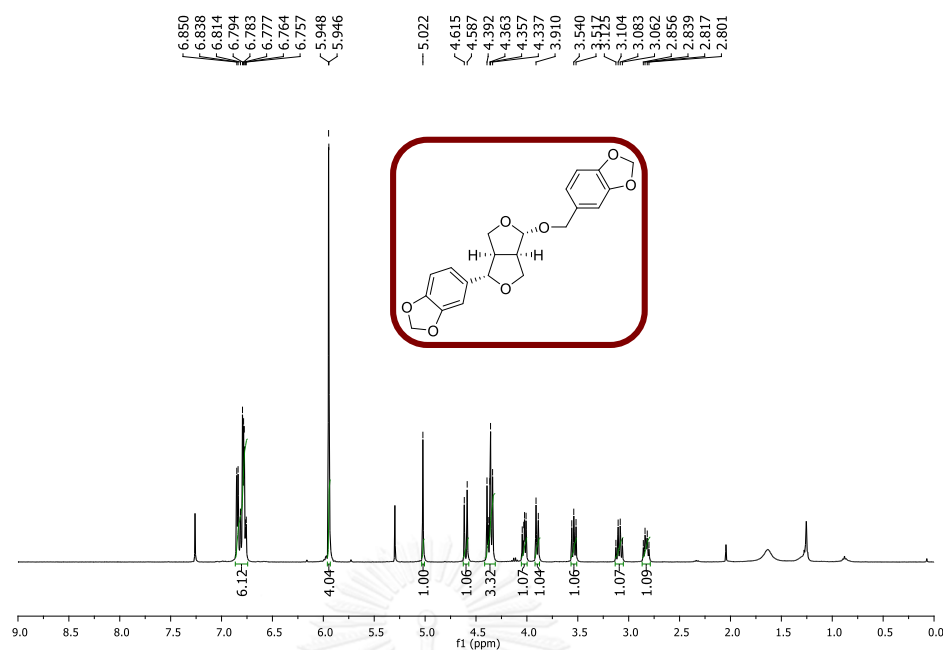


Figure 109. ^1H NMR spectrum of compound 3.3N in CDCl_3

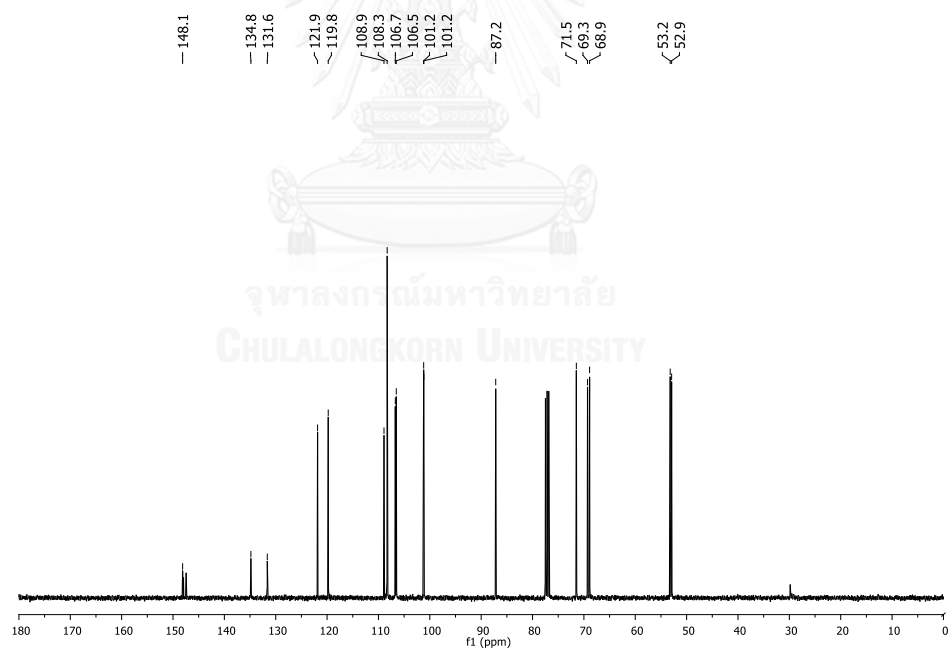


Figure 110. ^{13}C NMR spectrum of compound 3.3N in CDCl_3

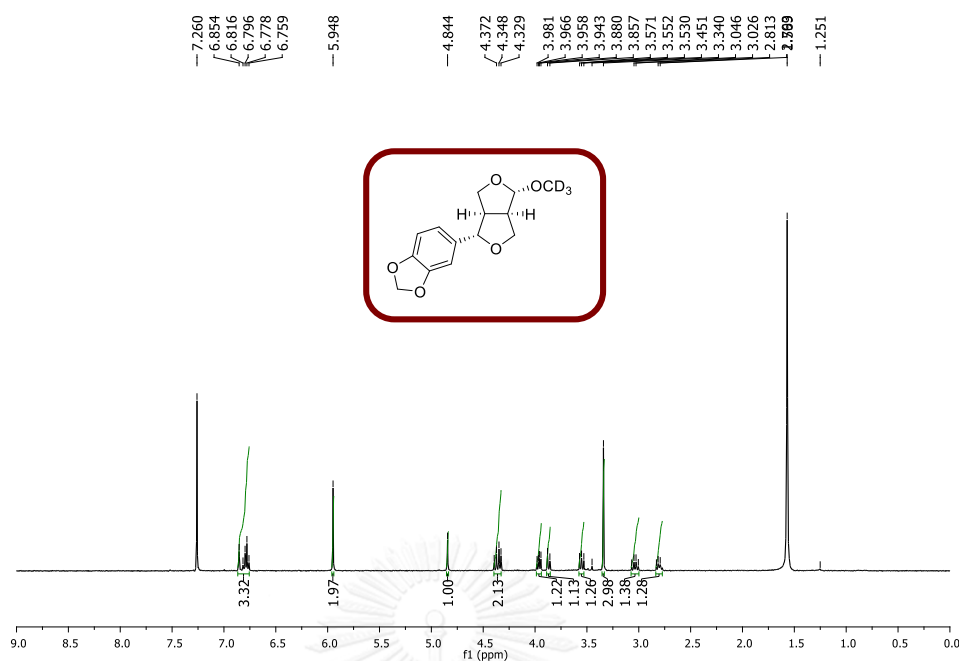


Figure 111. ^1H NMR spectrum of compound 4.1 in CDCl_3

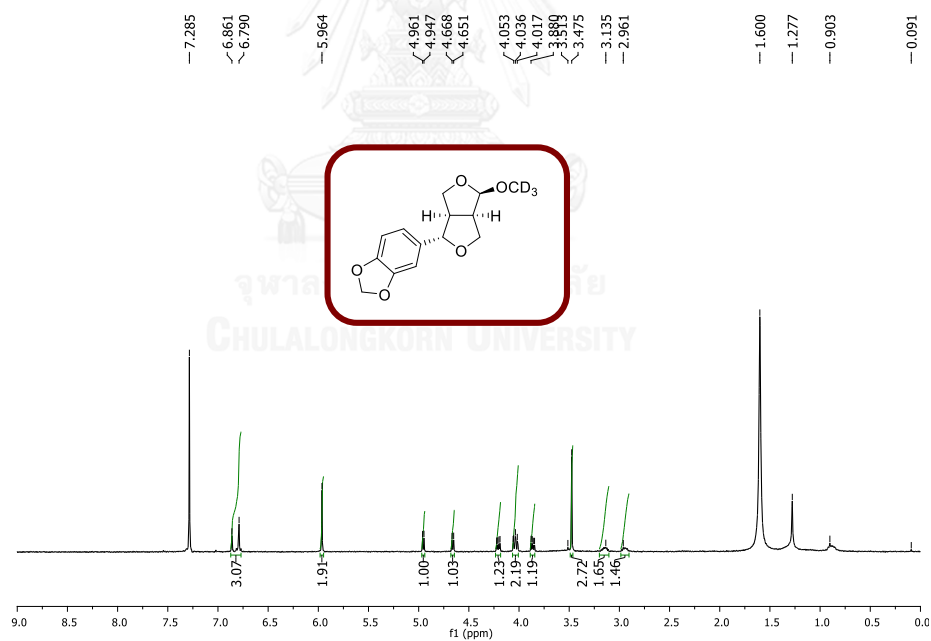
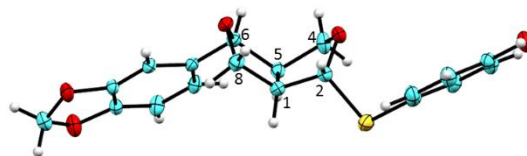


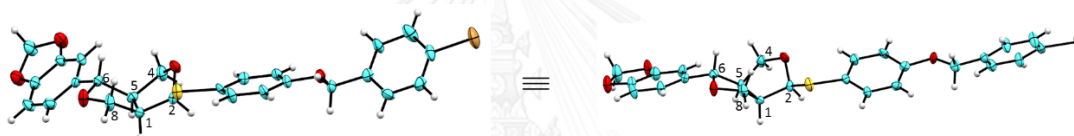
Figure 112. ^1H NMR spectrum of compound *epi*-4.1 in CDCl_3

Figure 113. ORTEP plot of **3.2r**Bond angles of **3.2r**

Number	Atom1	Atom2	Atom3	Angle
1	C2	C1	C8	113.33
2	C4	C5	C6	117.56

Torsion angles of **3.2r**

Atom1	Atom2	Atom3	Atom4	Torsion
H2	C2	C1	H1	-96.14

Figure 114. ORTEP plot of *epi*-**3.2r**-pBBEBond angles of *epi*-**3.2r**-pBBE

Number	Atom1	Atom2	Atom3	Angle
1	C2	C1	C8	114.01
2	C4	C5	C6	116.04

Torsion angles of *epi*-**3.2r**-pBBE

Atom1	Atom2	Atom3	Atom4	Torsion
H2	C2	C1	H1	28.01

VITA

Ms. Nantaporn Surachaitanawat was born on May 4, 1992 in Bangkok Thailand. She graduated with Bachelor's degree of Science, major in Chemistry from King Mongkut's University of Technology Thonburi in 2013 under supervised Assoc. Prof. Dr. Ploenpit Boochathum. She continued her master degree at department of chemistry, Chulalongkorn University under supervised Assoc. Prof. Dr. Preecha Phuwapraisirisan.

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