REFERENCES

- Holland, D. C.; Faul, K. C.; Roybal, J. E.; Murins, R. K. and Shimoda, W. Liquid chromatographic determination of chlortetracycline hydrochloride in ruminant and poultry/swine feeds. <u>J. Assoc. Off.</u> <u>Anal. Chem.</u> 74 (1991): 780-784.
- Blanchflower, W. J.; McCracken, R. J. and Rice, D. A. Determination of chlortetracycline residues in tissues using high-performance liquid chromatography with fluorescence detection. <u>Analyst</u> 114 (1989): 421-423.
- Zhang, X. R.; Baeyens, W. R. G.; Van der Borre, A; Van der Weken, G.; Calokerinos, A. C. and Sehulmani. S. G. Chemiluminescence determination of tetracycline based on their reactor with hydrogen peroxide catalyzed by the copper ion. <u>Analyst</u> 120 (1995): 463-466.
- Sabharwal, S.; Kishore, K. and Moorthy, P. N. Determination of tetracycline hydrochloride in presence of anhydrotetracycline by differential pulse poralograhy. J. Pharm. Sci. 77 (1988): 78-80.
- Caplis, M. E.; Ragheb, H. S. and Schall, E. D. Determination of tetracycline antibiotics by alternating-current polarography. <u>J. Pharm. Sci.</u> 54 (1965): 694-698.
- Dihuidi, K.; Kucharski, M. J.; Roets, E.; Hoogmartens, J. and Haeghe, H. V. Quantitative analysis of doxycycline and related substances by highperformance liquid chromatography. <u>J. Chromatogr.</u> 325 (1985): 413-424.

- Johnson, D. C. and Lacourse, W. R. Liquid chromatography with pulsed electrochemical detection at gold and platinum electrodes. <u>Anal.</u> <u>Chem.</u> 82 (1990): 598A-597A.
- 8. Christian, G. D. Analytical Chemistry. New York: John Wiley & Sons, 1986.
- Skoog, D. A.; West, D. M. and Holler, F. J. <u>Fundamental of Analytical</u> <u>Chemistry.</u> 6th Ed. New York: Saunders College Publishing, 1992.
- Braun, R. D. Introduction to Instrumental Analysis. Singapore: McGraw-Hill Book, 1987.
- Bard, A. J. and Feulkner, L. R. <u>Electrochemical Methods</u>. New York: John Wiley & Sons, 1980.
- Zyka, J. Instrumentation in Analytical Chemistry II. England: Ellis Horwood, 1994.
- Robinson, J. W. <u>Undergraduate Instrumental Analysis</u>. New York: Marcel Decker, 1970.
- 14. Wang, J. Analytical Electrochemistry. New York: John Wiley & Sons, 1994.
- Monk, P. M. S. <u>Fundamentals of Electroanalytical Chemistry</u>. New York: John Wiley & Sons, 2001.
- Sawyer, D. T. and Roberts, J. L. <u>Experimental Electrochemistry for Chemists</u>. New York: John Wiley & Sons, 1974.
- Lacourse, W. R. <u>Pulsed Electrochemical Detection in High-performance</u> <u>Liquid Chromatography.</u> New York: John Wiley & Sons, 1997.
- Lacoure, W. R. and Johnson, D. C. Optimization of waveform for pulsed amperometric detection of carbohydrates based on pulsed voltammetry. <u>Anal. Chem.</u> 85 (1993): 50-55.

- Kissinger, P. T. and Heineman Weineman W.R. <u>Laboratory Techniques in</u> <u>Electroanalytical Chemistry.</u> 2nd Ed. New York: Marcel Dekker, 1996.
- 20. Rocklin, R. D., Clarke, A. P. and Weitzhandler, M. Improved long-term reproducibility for pulsed amperometric detection of carbohydrates via a new quadruple-potential waveform. <u>Anal. Chem.</u> 70 (1998): 1496-1501.
- Ruzicka, J. and Hansen, E. H. <u>Flow Injection Analysis.</u> 2nd Ed. New York: John Wiley & Sons, 1988.
- Burke, L. D.; Buckley, D. T. and Morrissey, J. A. Novel view of the electrochemistry of gold. <u>Analyst</u> 119 (1994): 841-845.
- 23. Sultan, S. M.; Suliman, F. O.; Duffuaa, S. O. and Abu-Abdoun, I. I. Simplexoptimized and flow injection spectrophotometric assay of tetracycline antibiotics in drug formulations. <u>Analyst</u> 117 (1992): 1179-1183.
- 24. McCracken, R. J.; Blanchflower, W. J.; Haggan, S. A. and Kennedy, D. G. Simultaneous determination of oxytetracycline, tetracycline and chlortetracycline in animal tissues using liquid chromatography, postcolumn determination with aluminium, and fluorescence detection. <u>Analyst</u> 120 (1995): 1763-1766.
- 25. Oungpipat, W.; Southwell-Keely, P. and Alexander, P. W. Flow injection detection of tetracyclines by electrocatalytic oxidation at a nickelmodified glassy carbon electrode. <u>Analyst</u> 120 (1995): 1559-1565.
- 26. Houglum, J. E. and Larson, R. D. Assay of chlortetracycline in animal feeds by liquid chromatography with fluorescence detection. <u>J. AOAC Int.</u> 80 (1997): 961-965.

- 27. Webster, G. K.; Lugis, E. S.; Hearne, L. A.; Pomeroy, M. and Panozzo, L. A. Investigation of assay interference with microbiological determinations of chlortetracycline in feed-grade and premix samples. J. AOAC Int. 80 (1997): 298-301.
- Couto, C. M. C. M.; Lima, J. L. F. C.; Conceicao, M.; Montenegro, B. S. M. and Reis, S. Tetracycline, oxyteracycline and chlortetracycline determination by flow injection potentiometry. <u>J. Pharm. Biomed.</u> <u>Anal.</u> 18 (1998): 527-533.
- Zurhelle, G.; Pets, M.; Sietz, E. and Siewert, E. Metabolites of oxytetracycline, tetracycline, and their distribution in egg white, egg yolk, and hen plasma. J. Agric. Food Chem. 48 (2000): 6392-6396.
- 30. Zheng, X.; Mei, Y. and Zhujun, Z. Flow-injection chemiluminescence determination of tetracyclines with in situ electrogenerated bromine as the oxidant. <u>Anal. Chim. Acta</u> 440 (2001): 143-149.
- 31. Feng, P.; Shu, W. Q.; Huang, C. Z. and Li, Y. F. Total internal reflected resonance light scattering determination of chlortetracycline in body fluid with the complex cation of chlortetracycline-europium-trioctyl phosphine oxide at the water/tetrachloromethane interface. <u>Anal.</u> <u>Chem.</u> 73 (2001): 4307-4312.
- Jochsberger, T.; Cutie, A. and Mills, J. Differential pulsed polarography of tetracycline: Determination of complexing tendencies of tetracycline analogs in the presence of cations. <u>J. Pharm. Sci.</u> 68 (1979): 1061-1063.
- Sultan, S. M. Spectrophotometric determination of tetracycline with sodium molybdate. <u>Analyst</u> 111 (1986): 97-99.

- 34. Ji, H. and Wang, E. Flow injection amperometric detection based on ion transfer across a water-solidified nitrobenzene interface for the determination of tetracycline and terramycin. <u>Analyst</u> 113 (1988): 1541-1543.
- 35. Oka, H.; Ikai, Y.; Hayakawa, J.; Harada, K.; Asukabe, M.; Himei, R.; Horie, M.; Nakazawa, H. and MacNeil, J.D. Improvement of chemical analysis of antibiotics. 22. Identification of residual tetracyclines in honey by frit FAB/LC/MS using a volatile mobile phase. J. Agric. Food Chem. 42 (1994): 2215-2219.
- 36. Izquierdo, P.; Gomez-Hens, A. and Perez-Benedito, D. Study of the Eu (III)tetracycline-thenoyltrifluoroacetone system by using the stopped-flow mixing technique: Determination of tetracycline in serum. <u>Anal. Chim.</u> <u>Acta</u> 292 (1994): 133-139.
- 37. Tanase, I.; David, I.; Radu, G.; Lorgulesau, E. and Magearu, V. Optimized electroanalysis of tetracycline by alternating current polarography.
 <u>Analusis</u> 24 (1996): 281-284.
- 38. Savage, A. L.; Sarijo, S. H. and Baird, J. A novel screening method for tetracycline in milk combining sensitized-Eu (III) fluorescence and immunoaffinity techniques. <u>Anal. Chim. Acta</u> 375 (1998): 1-4.
- Wang, Y.; Liu, W.; Wang, K.; Shen, G. and Yu, R. Fluorescence optical fiber sensor for tetracycline. <u>Talanta</u> 47 (1998): 33-42.
- 40. Han, H.; He, Z. and Zeng, Y. Chemiluminescence determination of tetracycline using a tris (2,2'-bipyridine) ruthenium (II) and potassium permanganate system. <u>Anal. Sci.</u> 15 (1999): 467-470.

- Kurittu, J.; Lonnberg, S.; Virta, M. and Karp, M. A group-specific microbiological test for the detection of tetracycline residues in raw milk. <u>J. Agric. Food Chem.</u> 48 (2000) 3372-3377.
- 42. Lindsey, M. E.; Meyer, M. and Thurman, E. M. Analysis of trace levels of sulfonamide and tetracycline antimicrobials in ground water using solid-phase extraction and liquid chromatography /mass spectrometry. <u>Anal. Chem.</u> 73 (2001): 4640-4646.
- Inoue, A.; Earley, R. L.; Lehmann, M. W. and Welch, L. E. A consideration of electrode polishing prior to application of pulsed amperometric detection. <u>Talanta</u> 46 (1998) 1507-1515.
- Olliff, C. J. and Chatten, L. G. AC polarography for tetracycline analysis. <u>J.</u> <u>Pharm. Sci.</u> 66 (1977): 1564-1566.
- 45. Bocker, R. Rapid analysis of doxycycline from biological samples by high-performance liquid chromatography. <u>J. Chromatogr.</u> 187 (1980): 439-441.
- 46. Bocker, R. Analysis and quantitation of a metabolite of doxycycline in mice, rat, and humans by high-performance liquid chromatography. <u>J.</u>
 <u>Chromatogr.</u> 274 (1983): 255-262.
- 47. Salinas, F.; Nevado, J. J. B. and Espinosa, A. Determination of oxytetracycline and doxycycline in pharmaceutical compounds, urine and honey by derivative spectrophotometry. <u>Analyst</u> 114 (1989): 1141-1145.
- 48. Naidong, W.; Verresen, K.; Busson, R.; Roets, E. and Hoogmartens, J. Isolation of doxycycline, 6-epidoxycycline and 2-acetyl-2decarboxamidometacycline from commercial methacycline by

preparative column liquid chromatography on silica gel. J. Chromatogr. 586 (1991): 67-72.

- Croubels, S. and Peteghem, C. V. Sensitive spectrofluorimetric determination of tetracycline residues in bovine milk. <u>Analyst</u> 119 (1994): 2713-2721.
- 50. Karlicek, R. and Solich, P. Flow-injection spectrophotometric determination of tetracycline antibiotics. <u>Anal. Chim. Acta</u> 285 (1994): 9-12.
- 51. Oka, H.; Ikai, Y.; Ito, Y.; Hayakawa, J.; Harada, K.; Suzuki, M.; Odani. H. and Maeda, K. Improvement of chemical analysis of antibiotics XXIII: Identification of residual tetracyclines in bovine tissues by electrospray high-performance liquid chromatography-tandem mass spectrometry. J. Chromatogr. B 693 (1997): 337-344.
- 52. Kazemifard, A. G. and Moore, D. E. Evaluation of amperometric detection for the liquid-chromatographic determination of tetracycline antibiotics and their common contaminants in pharmaceutical formulations. J. <u>Pharm. Biomed. Anal.</u> 16 (1997): 689-696.
- 53. Korpela, M. T.; Kurittu, J. S.; Karvinen, J. T. and Karp, M. T. A recombinant Escherichia coli sensor strains for the detection of tetracyclines. <u>Anal.</u> <u>Chem.</u> 70 (1998): 4457-4462.
- 54. Liu, W.; Wang, Y.; Tang, J.; Shen, G. and Yu, R. Optical fiber sensor for tetracycline antibiotics based on fluorescence quenching of covalently immobilized anthracene. <u>Analyst</u> 123 (1998): 365-369.
- 55. Choma, I.; Grenda, D.; Malinowska, I. and Suprynowicz, Z. Determination of flumequine and doxycycline in milk by a simple thin-layer chromatographic method. <u>J Chromatogr. B</u> 734 (1999): 7-14.

- 56. McCormick, J. R. D.; Jensen, E. R.; Miller, P. A. and Doerschuk, A. P. The 6deoxytetracyclines. Further studies on the relationship between structure and antibacterial activity in the tetracycline series. J. Am. Chem. Soc. 82 (1960): 3381-3386.
- 57. Nilges, M. J.; Enochs, W. S. and Swartz, H. M. Identification and characterization of a tetracycline semiquinone formed during the oxidation of minocycline. J. Org. Chem. 56 (1991): 5623-5630.
- 58. Lacourse, W. R. and Johnson, D. C. Optimization of waveforms for pulsed amperometric detection (PAD) of carbohydrates following separation by liquid chromatography. <u>Carbohydr. Res.</u> 215 (1991): 159-178.
- 59. Fifield, F. W. and Kealey, D. <u>Principles and practice of analytical chemistry</u>. London: Chapman & Hall, 1995.
- Hargis, L. G. <u>Analytical chemistry: Principles and techniques.</u> London: Prentice Hall, 1988.
- 61. Kennedy, J. H. <u>Analytical chemistry: Principles.</u> 2nd Ed. New York: Saunders College Publishing, 1990.

APPENDICES

APPENDIX A



Rotating disk cyclic voltammetric results (pH dependence)

Figure A1 Rotating disk cyclic voltammograms of 1 mM tetracycline hydrochloride in 0.1 M KH₂PO₄ solution (pH 2) at Au RDE. The scan rate was varied from 10 to 300 mV s⁻¹. The rotation speed was 250 r.p.m.



Figure A2 Rotating disk cyclic voltammograms of 1 mM chlortetracycline hydrochloride in 0.1 M KH₂PO₄ solution (pH 2.5) at Au RDE. The scan rate was varied from 10 to 300 mV s⁻¹. The rotation speed was 250 r.p.m.



Figure A3 Rotating disk cyclic voltammograms of 1 mM doxycycline hydrochloride in 0.1 M KH₂PO₄ solution (pH 2) at Au RDE. The scan rate was varied from 10 to 300 mV s⁻¹. The rotation speed was 250 r.p.m.

APPENDIX B



Rotating disk cyclic voltammetric results (pH dependence)

Figure B1 Rotating disk cyclic voltammograms of 1 mM tetracycline hydrochloride in 0.1 M KH_2PO_4 solution (pH 2) at Au RDE. The rotation speed was varied from 200 to 600 r.p.m. The scan rate was 50 mV s⁻¹.



Figure B2 Rotating disk cyclic voltammograms of 1 mM chlortetracycline hydrochloride in 0.1 M KH_2PO_4 solution (pH 2.5) at Au RDE. The rotation speed was varied from 200 to 600 r.p.m. The scan rate was 50 mV s⁻¹.



Figure B3 Rotating disk cyclic voltammograms of 1 mM doxycycline hydrochloride in 0.1 M KH₂PO₄ solution (pH 2) at Au RDE. The rotation speed was varied from 200 to 600 r.p.m. The scan rate was 50 mV s⁻¹.

APPENDIX C

Flow injection with pulsed amperometric detection results

and calibration curves



Figure C1 Flow injection with pulsed amperometric detection results of tetracycline hydrochloride in $0.1 \text{ M KH}_2\text{PO}_4$ solution (pH 2) at gold disk electrode. The flow rate was 1 ml min⁻¹. The corresponding calibration curve is also shown (inset Figure).



Figure C2 Flow injection with pulsed amperometric detection results of chlortetracycline hydrochloride in $0.1 \text{ M KH}_2\text{PO}_4$ solution (pH 2.5) at gold disk electrode. The flow rate was 1 ml min⁻¹. The corresponding calibration curve is also shown (inset Figure).



Figure C3 Flow injection with pulsed amperometric detection results of doxycycline hydrochloride in $0.1 \text{ M KH}_2\text{PO}_4$ solution (pH 2) at gold disk electrode. The flow rate was 1 ml min⁻¹. The corresponding calibration curve is also shown (inset Figure).

APPENDIX D

Flow injection with pulsed amperometric detection results

of drug samples



Figure D1 Flow injection with pulsed amperometric detection results of tetracycline hydrochloride capsule in 0.1 M KH₂PO₄ solution (pH 2) with three replicated injections using the optimized PAD waveform. The flow rate was 1 ml min⁻¹. The corresponding calibration curve is also shown (inset Figure).



Figure D2 Flow injection with pulsed amperometric detection results of chlortetracycline hydrochloride capsule in 0.1 M KH₂PO₄ solution (pH 2.5) with three replicated injections using the optimized PAD waveform. The flow rate was 1 ml min⁻¹. The corresponding calibration curve is also shown (inset Figure).



Figure D3 Flow injection with pulsed amperometric detection results of doxycycline hydrochloride capsule in $0.1 \text{ M KH}_2\text{PO}_4$ solution (pH 2) with three replicated injections using the optimized PAD waveform. The flow rate was 1 ml min⁻¹. The corresponding calibration curve is also shown (inset Figure).

APPENDIX E

Description of analytical performance characteristics

Accuracy [8], [9]

Accuracy denotes that closeness of a measurement or set of measurements to the accepted value. Accuracy is normally reported in terms of error. Error is the difference between the accepted and measured values. There are several ways and units in which the accuracy can be expressed. Recovery is a term often used to describe accuracy, the equation for recovery is:

$$\% \text{Recovery} = \frac{\text{Measured value}}{\text{True value}} \times 100$$

Relative error is the another term that can be expressing the accuracy. The equation is shown below:

%error =
$$\frac{(\text{Measured value} - \text{True value})}{\text{True value}} \times 100$$

Precision [59], [60], [61]

Precision refers to the agreement between values in a set of data that have been carried out in exactly the same mode. It is a measure of the reproducibility of the analysis. Precision of the results can be ascertained through the use of replicate measurements. There are several popular ways to express the precision of data. Multiple injections of a homogeneous sample and calculation of the relative standard deviation (% RSD) do it. The equation for %RSD is shown below:

$$%RSD = \frac{\text{standard deviation}}{\text{Mean}} \times 100$$

Linearity (Linear range)

A linearity is the range where the analyte response is linearly proportional to concentration. The working sample concentration and samples tested for accuracy should be in the linear range.

Sensitivity

Sensitivity is the change in the analytical response divided by the corresponding change in the concentration of a standard (calibration) curve, i.e. the slope of the analytical calibration.

Limit of Detection (LoD)

The detection limit of a method is the lowest analyte concentration that can be determined to be different from an analyte blank. There are numerous way that detection limit have been defined. An example is the lowest analyte concentration that is above the noise level of the system, typically, three time the noise level (S/N = 3). This term is used to describe low analyte concentrations (< 10 μ M). For high analyte concentrations, the detection limit is defined as the lowest concentration that provides a signal to background ratio S/B of three. The equation of S/B ratio is shown below:

S/B ratio =
$$\frac{(\text{total signal} - \text{blank signal})}{\text{blank signal}}$$

APPENDIX F

The proposed method for determination of tetracycline antibiotics

F1 Chemicals and reagents

- F1.1 Tetracycline hydrochloride (Sigma)
- F1.2 Chlortetracycline hydrochloride (Sigma)
- F1.3 Doxycycline hydrochloride (Sigma)
- F1.4 Potassium dihydrogen orthophosphate (Merck)
- F1.5 Phosphoric acid (85% J.T. Baker)
- F1.6 Tetracycline hydrochloride capsule (TC Mycin 250 mg)
- F1.7 Cholrtetracycline hydrochloride capsule (Aureomycin 250 mg)
- F1.8 Doxycycline hydrochloride capsule (Medomycin 100 mg)

F2 Apparatus

F2.1 Gold disk electrode (0.07 cm², Bioanalytical System Inc.) pretreated by polishing with 0.05 micron of aluminum/water slurry

.

- F2.2 Ag/AgCl electrode
- F2.3 Autolab Potentiostat (PGSTAT 30, Metrohm)
- F2.4 Peristaltic pump (Eyela, SMP-23)
- F2.5 Rheodyne injection valve, Model 7225 (Altech)
- F2.6 Thin layer flow cell (Bioanalytical System Inc.
- F2.7 0.2 µM Nylon membrane filter (Altech)
- F2.8 0.45 μM Nylon membrane syringe filter with polypropylene (PP) housing (Orange Scientific filter)

F3 The preparation of supporting electrolyte and standard solutions

F3.1 0.1 M Potassium dihydrogen orthophosphate (KH₂PO₄)

 KH_2PO_4 13.60 g was dissolved in 1.0 L of deionized water and then adjusted with 85 % phosphoric acid to the required pH (pH 2 and 2.5).

F3.2 Tetracycline hydrochloride solution

The 1 mM tetracycline hydrochloride solution was prepared by weighing 0.0481 g tetracycline hydrochloride powder and transferring into 100 ml volumetric flask. The 0.1 M KH_2PO_4 solution (pH 2) was used for diluting this aliquot to the mark.

F3.3 Chlortetracycline hydrochloride solution

The 0.5 mM chlortetracycline hydrochloride solution was prepared by weighing 0.0258 g tetracycline hydrochloride powder and transferring into 100 ml volumetric flask. The 0.1 M KH_2PO_4 solution (pH 2.5) was used for diluting this aliquot to the mark.

F3.4 Doxycycline hydrochloride solution

The 0.5 mM doxycycline hydrochloride solution was prepared by weighing 0.0240 g tetracycline hydrochloride powder and transferring into 100 ml volumetric flask. The 0.1 M KH_2PO_4 solution (pH 2.) was used for diluting this aliquot to the mark.

F4 Procedure

The FI-PAD apparatus consisted of a thin-layer flow-through cell, a 20 μ l sample injection loop, a peristaltic pump and an electrochemical detector. The flow rate was 1 ml min⁻¹. The optimized waveform parameters of each analyte were shown in Table F1, F2, and F3.

Table F1 Optimal waveform parameters for pulsed amperometric detection of tetracycline hydrochloride at an Au working electrode

Potential (V vs. Ag/AgCl)		Time (ms)	
parameter	optimal	parameter	optimal
E _{det}	1.15	t _{del}	500
		t _{int}	100
E _{oxd}	1.6	t _{oxd}	130
E _{red}	0.1	t _{red}	300

Table F2 Optimal waveform parameters for pulsed amperometric detection of chlortetracycline hydrochloride at an Au working electrode

Potentia! (V vs. Ag/AgCl)		Time (ms)	
parameter	optimal	parameter	optimal
E _{det}	1.05	t _{del}	200
		t _{int}	100
E _{oxd}	1.3	t _{oxd}	70
E _{red}	0.25	t _{red}	400

Potential (V vs. Ag/AgCl)		Time (ms)	
parameter	optimal	parameter	optimal
E _{det}	1.15	t _{del}	150
		t _{int}	70
E_{oxd}	1.5	t _{oxd}	70
E_{red}	0.25	t _{red}	400

Table F3 Optimal waveform parameters for pulsed amperometric detection of doxycycline hydrochloride at an Au working electrode

F4.1 Tetracycline hydrochloride

The stock standard solution (1 mM) volume of 0.5, 1.0, 2.0, and 3.0 ml were pipetted into each 10 ml volumetric flask and diluted with 0.1 M KH₂PO₄ solution (pH 2). The final concentrations of the standard solutions were 24.05, 48.10, 96.12, and 144.17 μ g ml⁻¹, respectively. Each concentration was injected for three replicates. The calibration curve was obtained from the plotting between the averaged peak currents and the varied concentrations.

For real drug capsule (TC Mycin, 250 mg per capsule), a mass of powder of ten capsules was transferred to a 1000 ml volumetric flask and dissolved in 0.1 M KH₂PO₄ solution (pH 2), filtrated solution through a 0.45 μ M Nylon membrane syringe filter. The filtrated was further diluted with 0.1 M KH₂PO₄ solution (pH 2) to obtain a final concentration of 50.01 μ g ml⁻¹ (0.104 mM). The working sample solution was further injected for three replicates. The amount of tetracycline hydrochloride in drug capsule was calculated from the calibration curve. For other real samples such as milk and body fluid, the separation of tetracycline hydrochloride from other interference by HPLC was suggested. The obtained analyte portion was injected for three replicates. If the concentration was not in the calibration range, the dilution of concentration was performed.

F4.2 Chlortetracycline hydrochlorid:

For real drug capsule (Aureomycin, 250 mg), a mass of powder of ten capsules was transferred to a 1000 ml volumetric flask and dissolved in 0.1 M KH₂PO₄ solution (pH 2.5), filtrated through a 0.45 μ M Nylon membrane syringe filter. Then, the filtrated solution was further diluted with 0.1 M KH₂PO₄ solution to obtain a final concentration of 257.65 μ g ml⁻¹ (0.5 mM).

2.5 ml of sample solution (0.5 mM) was pipetted in each 10 ml volumetric flask and then 0, 1.0, 2.0, 3.0 and 4.0 ml of a standard solution was added to give final concentration of the standard solutions was 0, 25.77, 51.53, 103.06, and 206.12 μ g ml⁻¹, respectively. Each working solution was injected for three replicates. The amount of chlortetracycline hydrochloride in drug capsule was calculated from the calibration curve.

For other real samples such as milk and body fluid, the obtained analyte portion, which separated from other interference by HPLC, was injected for three replicates. If the concentration was more or less than 0.5 mM (compare to the peak height of standard at the same concentration), the dilution of concentration was performed. The working sample solutions were prepared as mentioned above and injected three replicates for each concentration. F4.3 Doxycycline hydrochloride

For real drug capsule (Medomycin, 100 mg), a mass of powder of ten capsules was transferred to a 1000 ml volumetric flask and dissolved in 0.1 M KH₂PO₄ solution (pH 2), filtrated through a 0.45 μ M Nylon membrane syringe filter. Then, the filtrated solution was further diluted with 0.1 M KH₂PO₄ solution to obtain a final concentration of 240.45 μ g ml⁻¹ (0.5 mM).

2.5 ml of sample solution (0.5 mM) was pipetted in each 10 ml volumetric flask and then 0, 1.0, 2.0, 3.0 and 4.0 ml of a standard solution was added to give final concentration of the standard solutions 0, 24.05, 48.09, 144.27, and 192.36 μ g ml⁻¹, respectively. Each working solution was injected for three replicates. The amount of chlortetracycline hydrochloride in drug capsule was calculated from the calibration curve.

For other real samples such as milk and body fluid, the obtained analyte portion, which separated from other interference by HPLC, was injected for three replicates. If the concentration was more or less than 0.5 mM (compare to the peak height of standard at the same concentration), the dilution of concentration was performed. The working sample solutions were prepared as mentioned above and injected three replicates for each concentration.

CURRICULUM VITAE



Name:		Sanit Palaharn	A Staunsain Time
Date and place of birth:		20 December 1975, Prachinburi, THAILAND	
Gender:		Male	
Education:	2002-2000	Chulalongkorn University, postgraduate study	
		leading to a Master of Science degree in	
		Analytical chemistry	
1999-1998		Chulalongkorn University,	
		Bachelor of Science	
1997-1995		The Institute of Analytical Chemistry Training	
		(the affiliated institute	e of Chulalongkorn
		University) Diploma o	of Analytical Chemistry
		Training	
1993-1991		Prachantarat bumroong school, Prachinburi,	
		High school	
	1990-1988	Wat Phromprasit scho	ool, Prachinburi,
		Secondary school	
	1987-1982	Wat Makokkeaw scho	ool, Prachinburi,
		Primary school	