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## **APPENDICS**

## **APPENDIX A**

### **Instruments and chemical reagents**

#### **1. Instruments**

- Analytical balance: Mettler Toledo, model AG204, Switzerland.
- Autoclave: Tomy, model Avanti J25, USA; Eppendorf, model 5430, Germany; Sorvall, model RC-5C Plus, USA.
- Circulating Water Bath: Techre, model TE8 A, UK.
- Freeze Dryer: Savant, model Super Modulya 233, USA.
- Freezer: Sharp, model FC27 (-20°C), Japan and Deep Freezer: Revco, model ULT1790-7-V12 (-80°C), USA.
- Hot stirring plate: Thermolyne, model Crimarec2, USA.
- Incubator: Memmert, model BE500, Germany.
- Incubator shaker: New Brunswick Scientific, model innova 4300, USA
- Magnetic stirrer: Ika, model RO-10, Malaysia.
- Microwave: Sanyo, model EM-815FW, Japan.
- Oven: Memmert, model UE 600, Germany.
- pH Meter: Mettler Toledo, model CH-8603, Switzerland.
- Pipette man: Gilson, France.
- Precision balance: Mettler Toledo, model PB3002, Switzerland.
- Shaking Water Bath: Memmert, model WB22, Germany.
- Spectrophotometer: Sherwood Scientific, model 1259, Cambridge, UK.
- Vortex mixer: Barnstead/Thermolyne, model M37610-26, USA.

## 2. Chemicals (Analytical grade)

Chemicals	Company
Acetone	Merck
Di-Ammonium sulfate	Merck
L-arginine monohydrochloride	Fluka
Bovine serum albumin	Sigma
Carboxymethyl cellulose (CMC)	Merck
Cellulose powder	Merck
Chloroform	Mallinckrodt
Copper (II) sulfate pentahydrate	Sigma
Ethanol	Carlo Erba
Ethylene diamine tetraacetic acid (EDTA)	Merck
Ferric citrate	Merck
Ferric sulfate sevenhydrate	Carlo Erba
Folin-Ciocalteu's phenol	Merck
Hydrochloric acid	Merck
Magnesium sulfate heptahydrate	Sigma
Methanol	Merck
Phenol	Carlo Erba
Potassium chloride	Merck
Potassium hydrogen sulfate	Merck
Di-potassium tartate	Carlo Erba
Sodium chloride	Carlo Erba
Tri-sodium citrate dihydrate	Merck
Sodium dodecyl sulfate	Fluka
Sodium hydroxide	Merck
Sodium potassium tartate	Merck
Trichloroacetic acid	Merck
Trisma base	Merck
Tyrosine	Sigma
Xylan from oat spelt	Sigma

## **APPENDIX B**

### **Culture Media**

All media were dispensed and sterilized in autoclave at 120° C, 15 pounds/inch<sup>2</sup> pressure for 15 min except the media for acid production from carbon source testing which were sterilized at 110° C, 10 pounds/inch<sup>2</sup> pressure for 10 min.

#### **1. PY medium**

Polypeptone	5	g
Yeast extract	1	g
K <sub>2</sub> HPO <sub>4</sub>	4	g
MgSO <sub>4</sub> .7H <sub>2</sub> O	1	g
KCl	0.2	g
FeSO <sub>4</sub> .7H <sub>2</sub> O	0.02	g
(Agar	15	g)
Distilled water	1000	ml
Dissolve and adjust to pH 7.0		

#### **2. Cellulose powder (CPY) medium**

Cellulose powder	10	g
Peptone	5	g
Yeast extract	1	g
K <sub>2</sub> HPO <sub>4</sub>	4	g
MgSO <sub>4</sub> .7H <sub>2</sub> O	1	g
KCl	0.2	g
FeSO <sub>4</sub> .7H <sub>2</sub> O	0.02	g
(Agar	15	g)
Distilled water	1000	ml
Dissolve and adjust pH to 7.0		

### 3. Xylan (XPY) medium

Oat spelt xylan	10	g
Peptone	5	g
Yeast extract	1	g
K <sub>2</sub> HPO <sub>4</sub>	4	g
MgSO <sub>4</sub> .7H <sub>2</sub> O	1	g
KCl	0.2	g
FeSO <sub>4</sub> .7H <sub>2</sub> O	0.02	g
(Agar	15	g)
Distilled water	1000	ml

Dissolve and adjust pH to 7.0

### 4. Carboxymethyl cellulose (CMC) medium

CMC ( Carboxymethyl cellulose )	1	g
Peptone	5	g
Yeast extract	1	g
K <sub>2</sub> HPO <sub>4</sub>	4	g
MgSO <sub>4</sub> .7H <sub>2</sub> O	1	g
KCl	0.2	g
FeSO <sub>4</sub> .7H <sub>2</sub> O	0.02	g
(Agar	15	g)
Distilled water	1000	ml

Dissolve and adjust pH to 7.0

### 5. Carboxymethyl cellulose-basal (CMC-basal) medium

( NH <sub>4</sub> ) <sub>2</sub> SO <sub>4</sub>	1	g
CMC	5	g
Yeast extract	1	g
(Agar	10	g)
Distilled water	1000	ml

Dissolve and adjust pH to 7.0

**6. L-arginine agar medium**

Phenol red, 1.0% aq.solution	1.0	ml
L(+)arginine monohydrochloride	10.0	g
Agar	3.0	g
PY medium	1000	ml
Dissolve and adjust pH to 7.2		

**7. Aesculin broth**

Aesculin	1	g
Ferric citrate	0.5	g
PY medium	1000	ml

Dissolve and adjust pH to 7.4. Sterilization was performed at 110 °C for 10 min.

**8. Casein agar medium**

Skim milk	10	g
PY medium	1000	ml
Agar	15	g
Dissolve and adjust pH to 7.2		

**9. Gelatin agar medium**

Gelatin	10	g
PY medium	1000	ml
Agar	15	g
Dissolve and adjust pH to 7.2		

**10. Motility test medium**

Motility medium (Difco)	20	g
Distilled water	1000	ml
Dissolve and adjust pH to 7.2		

**11. Simmon Citrate agar**

Simon citrate agar (Difco)	24.2	g
Distilled water	1000	ml
Dissolve and adjust pH to 6.8		

**12. Starch agar medium**

Starch	10	g
PY medium	1000	ml
Agar	15	g
Dissolve and adjust pH to 7.2		

**13. Triple sugar iron agar medium**

Triple sugar iron agar (Difco)	60	g
Distilled water	1000	ml
Dissolve and adjust pH to 7.4		

**14. Tyrosine agar medium**

Tyrosine	50	g
PY medium	1000	ml
Agar	15	g
Dissolve and adjust pH to 7.2		

**15. Deoxyribonuclease (DNase) medium**

DNase test agar (Difco)	42	g
Distilled water	1000	ml
Adjust pH to 7.3 and heat to boiling to dissolve completely		

**16. Indole test medium**

Tryptone	10	g
Meat extract	3	g
Distilled water	1000	ml
Dissolve and adjust pH to 7.4		

**17. Nitrate broth**

Meat extract	3	g
Peptone	10	g
KNO <sub>3</sub>	1	g
Distilled water	1000	ml
Dissolve and adjust pH to 7.2		

**18. Tween 80 agar medium**

Tween 80	2	ml
PY medium	1000	ml
Agar	15	g
Dissolve and adjust pH to 7.2		

**19. Urea agar medium**

Urea	20	g
PY medium	1000	ml
Agar	15	g
Dissolve and adjust pH to 7.2		

**20. MR-VP broth**

MR-VP medium (Merck)	17	g
Distilled water	1000	ml
Dissolve and adjust pH to 6.9		

## APPENDIX C

### Reagents and Buffers

#### 1. Standard curve of glucose or xylose

Glucose or xylose solution (0, 20, 40, 60, 80, 100, 120, 140, 160, 180, 200 ug/ml) were prepared. The analytical reactions were carried out by a procedure described by Somogyi and Nelson (1952).

#### 2. Flagella staining reagent

Basic fuchisin	0.5	g
Tannic acid	0.2	g
Aluminium sulfate	0.5	g

Dissolve in solvent composed of 95% (v/v) ethanol (2ml), glycerol(0.5 ml), and Tris(hydroxymethyl)aminomethane(Tris) buffer (7.5ml).

#### 3. Kovac's reagent

$\rho$ -dimethylaminobenzaldehyde	5	g
Amyl alcohol	75	g
conc. HCl	25	ml

Dissolve the aldehyde in the alcohol by gently heating in a water bath (about 50-55 °C).

Cool then add the acid with care. Protect from light and store at 4 °C.

#### 4. Nitrate test reagent

Solution A (sulphanilic acid solution)

0.33% (w/v) sulphanilic acid in 5 N- acetic acid

Dissolve by gently heating

Solution B (N, N-dimethyl-1-naphthylamine)

0.6% (w/v) dimethyl- $\alpha$ -naphthylaminein 5 N-acetic acid

Dissolve by gently heating in a fume hood.

Add two drops of sulphanilic acid solution and three drops of *N,N*-dimethyl-l-naphthylamine into peptone nitrate broth, then inoculating with test microorganisms.

### 5. 6 N HCl

conc. HCl	60	ml
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Distilled water	60	ml
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Add conc. HCl into the distilled water

### 6. 2 N H<sub>2</sub>SO<sub>4</sub>

conc. H <sub>2</sub> SO <sub>4</sub>	2	ml
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Distilled water	34	ml
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Add conc. HCl into the distilled water

### 7. Ninhydrin solution

Ninhydrin	0.3	g
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1-Butanol	100	ml
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Glacial acetic acid	3	ml
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Dissolve ninhydrin in butanol containing acetic acid

### 8. Phenol : Chloroform (1:1 v/v)

Crystalline phenol was liquified in water bath at 65° C and mixed with chloroform in the ratio of 1:1 (v/v). The solution was stored in a light tight bottle.

### 9. 0.5 M EDTA (pH 8.0)

Disodium ethylenediamine tetraacetate.2H<sub>2</sub>O (186.1 g) was added into 800 ml of distilled water and stirred vigorously on a magnetic stirrer. The pH was adjusted to 8.0 with NaOH (20 g of NaOH pellet). The volume was adjusted to 1 litre. The solution was dispensed into aliquot and sterilized by autoclaving at 15 lb/in<sup>2</sup> for 15 minutes.

**10. 2x PBS solution**

8 mM Na<sub>2</sub>HPO<sub>4</sub>

1.5 mM KH<sub>2</sub>PO<sub>4</sub>

137 mM NaCl

2.7 mM KCl

The ingredient of 2x PBS is shown above. The solution pH was adjusted to 7.0 with 1N NaOH or 1N HCL, then sterilized by autoclaving at 15 lb/in<sup>2</sup> for 15 minutes.

**11. 10 mg/ml Salmon sperm DNA**

A 10 mg of Salmon sperm DNA was dissolved in 1 ml of 10 mM TE buffer pH 7.6. Boiling for 10 minutes, immediately cooling in ice water and sonicating for 3 minutes.

**12. 3 M Sodium acetate, pH 5.2**

To 800 ml of distilled water, 408.1 g of sodium acetate was added and adjusted the pH to 5.2 with glacial acetic acid. The volume was adjusted to 1 litre. The solution was sterilized by autoclaving at 15 lb/in<sup>2</sup> for 15 minutes.

**13. 10% Sodium dodecyl sulphate (SDS)**

The solution was prepared by dissolving 10 g of sodium dodecyl sulphate in 100 ml sterile distilled water. Sterilization is not required.

**14. 20x SSC**

3 M NaCl

0.1 M Tri-sodium citrate

The ingredient of 20x SSC is shown above. The solution pH was adjusted to 7.0 with 1N NaOH, then sterilized by autoclaving at 15 lb/in<sup>2</sup> for 15 minutes

### **15. 1 M Tris-HCl (pH 8.0)**

The 1 M Tris was prepared by dissolving 121.1 g of Tris base in 800 ml of distilled water. The pH was adjusted to the desired value by adding conc. HCL (pH 8.0, 42 ml of HCl). The solution was cooled to room temperature before making final adjustment to the desired pH. The volume of the solution was adjusted to 1 litter with with distilled water and sterilized by autoclaving.

### **16. RNase A solution**

RNase A	20	mg
0.15 M NaCl	10	ml

Dissolve 20 mg of RNase A in 10 ml of 0.15 M NaCl and heat at 95° C for 5-10 minutes. Store at -20°C.

### **17. RNase T<sub>1</sub> solution**

RNase T <sub>1</sub>	80	μl
0.1 M Tris-HCl (pH 7.5)	10	ml

Add 80 μl of RNase T<sub>1</sub> into 10 ml of 0.1 M Tris-HCl (pH 7.5), then heat at 95°C for 5 minutes. Store at -20°C.

### **18. Proteinase K solution**

Proteinase K (Sigma)	4	mg
50 mM Tris-HCl (pH 7.5)	1	ml

Dissolve proteinase K in Tris-HCl buffer. Use freshly prepared solution.

**19. Nuclease P1 solution**

Nuclease P1	0.1	mg
40 mM CH <sub>3</sub> COONa+12 mM ZnSO <sub>4</sub> (pH5.3)	1	ml

Dissolve nuclease P1 in sodium acetate containing ZnSO<sub>4</sub>. Store at 4°C.

**20. Alkaline phosphatase solution**

Alkaline phosphatase	2.4	units
0.1 M Tris-HCl (pH 8.1)	1	ml

Dissolve alkaline phosphatase in Tris-HCl buffer. Store at -20°C.

**21. 0.1 M Tris-HCl buffer, pH 9**

Tris	1.21	mg
Distilled water	100	ml

Dissolve Tris base in distilled water, adjust the pH to 9 with conc.HCl.

**22. TE buffer (pH 8.0)**

10 mM Tris HCl (pH 8.0)
1 mM Na <sub>2</sub> -EDTA (pH 8.0)

The ingredient of TE buffer (pH 8.0) is shown above. Sterile by autoclaving at 15 lb/in<sup>2</sup> for 15 minutes, store at 4°C.

**23. TE buffer + RNase A**

Sterile TE buffer	960	ml
RNase A (2 mg/ml)	100	μl

The ingredient is shown above. Mix the ingredient together and store at 4°C.

## **24. Saline-Na<sub>2</sub> EDTA**

0.1 M NaCl

50 mM EDTA.2Na (pH 8.0)

The ingredient is shown above. Mix the ingredient together, sterile by autoclaving at 15 lb/in<sup>2</sup> for 15 minutes. Store at room temperature.

## **25. Reagents and buffers for DNA-DNA hybridization**

### **25.1 Prehybridization solution**

100x Denhardt solution	5	ml
10 mg/ml Salmon sperm DNA	1	ml
Sterile 20x SSC	10	ml
Formamide	50	ml
Distilled water	34	ml

Sequentially dissolve the above ingredients in disstilled water.

### **25.2 Hybridization solution**

Prehybridization solution	100	ml
Dextran sulfate	5	g

Dissolve dextran sulphate in prehybridization solution, store at 4°C.

### **25.3 Solution I**

Bovine serum albumin (Fraction V)	0.25	g
Triton X-100	50	μl
PBS solution	50	ml

Dissolve Bovine serum albumin and Triton X-100 in PBS solution.

Store at 37°C.

**25.4 Solution II**

Streptavidin-POD	1	μl
Solution I	4	ml

Dissolve Streptavidin-POD in Solution I. Store at 4°C.

**25.5 Solution III**

3,3',5,5'-Tetramethylbenzidine (TMB)	100	μl
(10 mg/ml in DMFO)		
0.3% (v/v) H <sub>2</sub> O <sub>2</sub>	100	μl
0.4 M Citric acid + 0.2 M Na <sub>2</sub> HPO <sub>4</sub> buffer	100	μl

pH 6.2 in 10% (v/v) DMFO

Mix the above ingredient together. Store at 37°C.

**25.6 2 M H<sub>2</sub>SO<sub>4</sub>**

conc. H <sub>2</sub> SO <sub>4</sub>	22	ml
Distilled water	178	ml

Add conc. H<sub>2</sub>SO<sub>4</sub> into distilled water. The solution was sterilized by autoclaving at 15 lb/in<sup>2</sup> for 15 minutes. Store at room temperature.

**26. Fehling's solution**

Solution I:

Copper sulfate pentahydrate	34.64 g
Distilled water	500 ml

Solution II:

Sodium potassium tartate	173 g
Sodium hydroxide	50 g

110

Distilled water

500 ml

Mix solution I and II together. Store at room temperature.

## **APPENDIX D**

**Physiological and biochemical characteristics of isolates,  
primers, 16S rRNA gene sequences and DNA G+C contents**

Physiological and biochemical characteristics of isolates.

Isolate code.	Growth in (%NaCl)		Growth at pH					Growth at °C									Catalase test	Oxidase test	Anaerobic growth	Methyl red	Voges-Proskauer	DNAase	Urease	Indole production	Citrate	TSI	Nitrate reduction	Asculin	L-arginine	Hydrolysis			
	3	5	5	6	8	9	10	15	20	45	50	55	60																				
PA4-1	-	-	-	+	+	+	-	w	+	+	+	-	-	-	+	+	+	+	+	+	-	-	-	N/N	+	-	-	-	-	-			
PBS5	+	+	-	+	+	+	-	+	+	+	+	-	-	-	+	+	+	+	+	+	-	-	-	K/K	-	-	-	-	-	-			
T4-1	+	+	-	+	+	+	-	+	+	+	+	-	-	-	+	+	+	+	+	+	-	-	-	A/A	+	+	-	+	-	-			
T6-1	+	+	-	+	+	+	-	+	+	+	+	-	-	-	+	+	+	+	+	+	-	-	-	K/A	+	+	+	+	-	-			
T3-3	+	+	-	+	+	+	-	+	+	+	+	-	-	-	+	+	+	+	+	-	+	+	-	K/A	+	+	+	+	-	-			
T10-2	+	+	-	+	+	+	-	+	+	+	+	-	-	-	+	+	+	+	+	-	+	+	-	K/A	+	+	+	+	-	-			
PBS4	+	+	W	+	+	+	-	+	+	+	+	-	+	+	-	+	+	+	+	-	+	+	-	A/A	+	+	+	+	+	-			
PA3-3	+	+	-	+	+	+	-	+	+	+	+	-	+	+	-	+	+	-	-	+	-	-	-	A/A	+	+	+	+	-	-			
PA3-5	+	+	-	+	+	+	-	-	+	+	+	-	+	+	-	+	+	-	-	+	-	-	-	K/A	+	+	+	+	-	-			
PA4-3	+	+	+	+	+	+	-	+	+	+	+	-	+	+	-	+	+	-	-	+	-	-	-	A/A	+	+	+	-	-	-			
PA4-4	+	+	-	+	+	+	-	+	+	+	+	-	+	+	-	+	+	-	-	+	-	-	-	K/A	+	+	-	-	-	-			

Symbols: +, positive; -, negative; w, weakly positive; N, neutral; K, alkaline; A, acid

Physiological and biochemical characteristics of isolates (Cont)

Isolate code.	Growth in (%NaCl)		Growth at pH					Growth at °C								Catalase test	Oxidase test	Anaerobic growth	Methyl red	Voges-Proskauer	DNAase	Urease	Indole production	Citrate	TSI	Nitrate reduction	Asciulin	L-arginine	Hydrolysis			
	3	5	5	6	8	9	10	15	20	45	50	55	60																			
T3-2	+	+	-	+	+	+	-	+	+	+	+	w	-	+	+	+	+	-	-	-	-	K/A	+	+	+	-	-	-	-			
T6-3	+	+	-	+	+	+	-	+	+	+	+	-	-	+	+	+	+	-	-	-	-	K/A	+	+	+	-	+	-	-			
N14-2	+	+	-	+	+	+	-	+	+	+	+	-	-	+	+	+	-	-	-	-	+	K/K	+	-	+	-	-	-	-			
PF2-1	+	-	-	+	+	+	-	+	+	+	+	-	-	+	+	+	-	+	-	-	+	K/K	+	-	+	-	-	-	-			
PD1-2	+	+	-	+	+	+	-	-	+	+	+	-	-	+	+	+	-	+	-	+	+	K/K	+	-	+	-	+	-	-			
T6-4	+	+	-	+	+	+	-	+	+	+	+	-	-	+	+	+	-	-	-	-	-	K/K	+	+	+	-	+	-	-			
N3-2	+	+	+	+	+	+	-	-	+	+	+	-	-	+	+	+	-	-	-	-	-	K/K	+	+	+	-	+	-	-			
N12	+	-	-	+	+	-	-	+	+	+	+	-	-	+	+	+	-	+	-	-	-	K/N	+	+	+	-	-	-	-			
N16-2	+	-	-	+	+	-	-	-	+	+	+	-	-	+	+	+	-	-	-	-	-	K/K	+	+	+	-	+	-	-			
PA1-4	+	+	+	+	+	+	-	+	+	+	+	-	-	+	+	+	-	+	-	-	-	K/A	+	-	-	-	-	-	-			
PA2-4	+	-	+	+	+	+	-	+	+	+	+	-	-	+	+	+	-	-	-	-	-	K/A	+	-	-	-	+	-	-			

Symbols: +, positive; -, negative; w, weakly positive; N, neutral; K, alkaline; A, acid

Physiological and biochemical characteristics of isolates (Cont)

Isolate code.	Growth in (%NaCl)		Growth at pH					Growth at °C								Catalase test	Oxidase test	Anaerobic growth	Methyl red	Voges-Proskauer	DNAase	Urease	Indole production	Citrate	TSI	Nitrate reduction	Asciulin	L-arginine	Hydrolysis			
	3	5	5	6	8	9	10	15	20	45	50	55	60													Casein	Gelatin	Starch	L-Tyrosine	Tween 80		
PA4-2	+	+	+	+	+	+	-	+	+	+	+	-	-	+	+	+	-	+	-	-	-	-	K/A	+	-	-	-	-				
PB11	+	+	+	+	+	+	-	+	+	+	+	+	-	+	+	+	+	+	-	-	-	-	K/A , gas <sup>*</sup>	+	+	-	-	-				
PBH2	+	+	+	+	+	+	-	-	+	+	+	-	-	+	+	+	+	+	-	-	-	-	K/K	+	-	+	-	+				
PBH4	+	+	+	+	+	+	-	-	+	+	+	+	-	+	+	+	+	+	+	+	+	-	K/A	+	+	+	-	-				
PBS3	+	+	+	+	+	+	-	-	+	+	+	+	-	+	+	+	+	+	+	+	-	+	K/A	+	+	+	-	-				
PT4-2	-	-	+	+	+	+	-	+	+	+	+	-	-	-	+	-	+	-	-	-	-	-	N/N	+	+	-	-	+				
PN8-3	-	-	+	+	+	-	-	+	+	+	+	-	-	+	-	+	-	+	-	+	-	-	N/N	+	+	-	+	-				
PN12-2	-	-	-	+	+	-	-	+	+	+	+	-	-	+	-	+	-	+	-	-	-	-	N/N	+	+	-	-	-				
PN12-3	-	-	-	+	+	-	-	+	+	+	+	-	-	+	-	+	-	+	-	-	-	-	N/N	+	+	-	-	-				
PT6-2	+	-	+	+	+	+	-	+	+	+	+	-	-	+	-	+	-	+	-	+	-	-	N/N	+	+	-	-	-				
PT6-3	-	-	+	+	+	+	-	+	+	+	+	+	-	+	-	+	-	+	-	+	-	-	N/N	+	+	-	+	-				

Symbols: +, positive; -, negative; w, weakly positive; N, neutral; K, alkaline; A, acid

Physiological and biochemical characteristics of isolates (Cont)

Isolate code.	Growth in (%NaCl)		Growth at pH					Growth at °C								Catalase test	Oxidase test	Anaerobic growth	Methyl red	Voges-Proskauer	DNAase	Urease	Indole production	Citrate	TSI	Nitrate reduction	Aesculin	L-arginine	Hydrolysis			
	3	5	5	6	8	9	10	15	20	45	50	55	60																			
PN13-1	+	-	-	-	+	+	-	-	+	+	+	-	-	+	+	-	-	-	-	-	-	N/N	+	+	-	-	+	+	-	-		
NS-3	-	-	+	+	+	-	-	-	+	+	+	-	-	+	+	+	-	-	-	-	-	K/N, gas*	+	+	+	-	+	-	-	-		
T3-2X	-	-	+	+	+	-	-	-	+	+	+	-	-	+	-	+	-	-	-	-	-	A/A	+	+	-	-	-	+	-	-		
PT2-3	-	-	-	+	+	-	-	-	+	+	+	-	-	+	-	-	-	-	-	-	-	N/N	+	+	-	-	+	+	-	-		
PN8-2	+	+	+	+	+	+	+	-	-	+	+	+	-	+	+	+	+	+	-	+	-	K/A	+	+	+	-	+	+	-	-		
PN1-2	+	+	+	+	+	+	+	-	+	+	+	+	-	+	+	+	-	-	-	-	-	K/N, H <sub>2</sub> S*	+	-	+	+	+	-	+	-		
TT2-2X	+	+	+	+	+	+	+	-	+	+	+	+	-	+	+	+	-	-	-	-	-	K/N	+	-	+	-	+	-	+	+		
T8-1X	+	+	+	+	+	+	-	-	+	+	+	+	-	+	+	+	-	-	-	-	-	K/N	+	-	+	-	-	-	-	+		
PN8-1	-	-	-	+	+	+	-	-	+	+	+	+	-	+	+	+	-	-	-	-	-	K/N	+	-	+	-	-	-	-	-		
PN9-3	+	-	+	+	+	+	-	+	+	+	+	-	-	+	-	-	-	-	-	-	-	K/N	-	-	+	-	+	-	+	+		
PN20-1	+	+	+	+	+	+	-	+	+	+	+	-	-	+	-	-	-	-	-	-	-	A/A	-	-	+	-	+	-	-	-		

Symbols: +, positive; -, negative; w, weakly positive; N, neutral; K, alkaline; A, acid

Physiological and biochemical characteristics of isolates (Cont)

Isolate code.	Growth in (%NaCl)		Growth at pH		Growth at °C										Catalase test	Oxidase test	Anaerobic growth	Methyl red	Voges-Proskauer	DNAase	Urease	Indole production	Citrate	TSI	Hydrolysis		
	3	5	5	6	8	9	10	15	20	45	50	55	60														
PT1-1	-	-	-	+	+	-	-	+	+	+	+	+	-	+	+	-	+	-	+	-	-	K/N	+	+	-	-	-
PT1-2	+	+	+	+	+	+	-	+	+	+	+	+	-	+	-	-	-	+	-	+	-	A/A	-	+	+	-	-
PT2-4	+	+	+	+	+	+	-	+	+	+	+	-	-	+	-	-	-	+	-	+	-	A/A	-	+	+	-	-
PT6-4	+	+	+	+	+	+	-	+	+	+	+	+	-	+	+	-	-	+	-	+	-	A/A	+	+	+	-	-
PN16-5	+	+	+	+	+	+	-	+	+	+	+	-	-	+	+	-	+	-	+	-	-	A/A	+	+	-	+	-
N5-1X	+	+	+	+	+	+	-	+	+	+	+	-	-	+	+	+	+	+	+	+	-	A/A	+	+	+	-	-
N9-2	+	+	+	+	+	+	-	w	+	+	+	w	-	+	+	+	+	+	-	+	+	A/A	+	-	+	-	-

Symbols: +, positive; -, negative; w, weakly positive; N, neutral; K, alkaline; A, acid

Acid production from carbohydrates

Isolate code.	D-Amygdalin	w L-Arabinose	+ D-Cellubiose	- D-Fructose	w D-Galactose	w D-Glucose	- Gluconate	- Glycerol	- Inositol	- Inulin	w + Lactose	- D-Maltose	- D-Mannitol	- D-Mannose	- D-Melibiose	- D-Melezitose	$\alpha$ - Methyl - D - glucoside	- Raffinose	- L-Rhamnose	- D-Ribose	- Salicin	w D-Sorbitol	- L-Sorbose	w Sucrose	- D-Trehalose	+ D-Xylose	
PA4-1	-	w	+ D-Amygdalin	- L-Arabinose	-	- D-Cellubiose	-	- D-Fructose	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
PBS5	-	w	+ D-Amygdalin	- L-Arabinose	-	- D-Cellubiose	-	- D-Fructose	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
T4-1	-	-	-	+ D-Amygdalin	- L-Arabinose	-	- D-Cellubiose	-	- D-Fructose	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
T6-1	-	-	-	+ D-Amygdalin	- L-Arabinose	-	- D-Cellubiose	-	- D-Fructose	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
T3-3	-	+	-	+ D-Amygdalin	- L-Arabinose	-	- D-Cellubiose	-	- D-Fructose	-	w D-Galactose	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
T10-2	+	+	-	+ D-Amygdalin	- L-Arabinose	-	- D-Cellubiose	-	- D-Fructose	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
PBS4	+	+	-	+ D-Amygdalin	- L-Arabinose	-	- D-Cellubiose	-	- D-Fructose	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
PA3-3	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
PA3-5	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
PA4-3	-	+	+	+ D-Amygdalin	- L-Arabinose	-	- D-Cellubiose	-	- D-Fructose	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
PA4-4	-	+	-	+ D-Amygdalin	- L-Arabinose	-	- D-Cellubiose	-	- D-Fructose	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	

Symbols: +, positive; -, negative; w, weakly positive

Acid production from carbohydrates (Cont)

Isolate code.	D-Amygdalin	L-Arabinose	D-Cellubiose	D-Fructose	D-Galactose	D-Glucose	Gluconate	Glycerol	Inositol	Inulin	Lactose	D-Maltose	D-Mannitol	D-Mannose	D-Melibiose	D-Melezitose	$\alpha$ - Methyl - D - glucoside	Rafinose	L-Rhamnose	D-Ribose	Salicin	D-Sorbitol	L-Sorbitose	Sucrose	D-Trehalose	D-Xylose
T3-2	-	-	+ D-Amygdalin	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
T6-3	+	-	+	+	-	+	w	w	-	-	-	-	w	+	-	-	-	-	-	-	w	-	-	-	-	-
N14-2	+	-	+	+	-	+	-	w	-	-	-	-	+	+	-	-	-	-	-	-	-	-	-	-	-	-
PF2-1	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
PD1-2	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
T6-4	+	-	+	+	-	+	-	-	-	-	-	+	+	+	-	-	-	-	-	-	-	-	-	-	-	-
N3-2	+	-	+	+	-	+	-	w	-	-	-	-	+	+	-	-	-	-	-	-	-	-	-	-	-	-
N12	+	-	-	-	-	-	-	-	+	-	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-
N16-2	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
PA1-4	+	-	+	+	-	+	-	+	-	-	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-
PA2-4	w	-	+	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-

Symbols: +, positive; -, negative; w, weakly positive

Acid production from carbohydrates (Cont)

Isolate code.	D-Amygdalin	L-Arabinose	D-Cellobiose	D-Fructose	D-Galactose	D-Glucose	Gluconate	Glycerol	Inositol	Inulin	Lactose	D-Maltose	D-Mannitol	D-Mannose	D-Melibiose	$\alpha$ -Methyl-D-glucoside	Raffinose	L-Rhamnose	D-Ribose	Salicin	D-Sorbitol	L-Sorbose	Sucrose	D-Trehalose	D-Xylose		
PA4-2	+	-	+ D-Amygdalin	- L-Arabinose	+ D-Cellobiose	- D-Fructose	- D-Galactose	- D-Glucose	- Gluconate	- Glycerol	- Inositol	- Inulin	- Lactose	- D-Maltose	- D-Mannitol	- D-Mannose	- D-Melibiose	- $\alpha$ -Methyl-D-glucoside	- Raffinose	- L-Rhamnose	- D-Ribose	- Salicin	- D-Sorbitol	- L-Sorbose	- Sucrose	- D-Trehalose	- D-Xylose
PB11	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
PBH2	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
PBH4	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
PBS3	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
PT4-2	+	+	+	+	+	+	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
PN8-3	+	+	+	+	+	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
PN12-2	+	+	+	+	+	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
PN12-3	w	w	+	w	w	w	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
PT6-2	+	+	+	+	+	+	-	w	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
PT6-3	+	+	+	+	+	+	-	w	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	

Symbols: +, positive; -, negative; w, weakly positive

Acid production from carbohydrates (Cont)

Isolate code.	D-Amygdalin	L-Arabinose	D-Cellobiose	D-Fructose	D-Galactose	D-Glucose	Gluconate	Glycerol	Inositol	Inulin	L-lactose	D-Maltose	D-Mannitol	D-Mannose	D-Melibiose	D-Melezitose	$\alpha$ -Methyl-D-glucoside	Raffinose	L-Rhamnose	D-ribose	Salicin	D-Sorbitol	L-Sorbose	Sucrose	D-Trehalose	D-Xylose
PN13-1	+	+	-	w	+	+	+	w	w	-	-	-	-	-	-	-	w	-	w	-	w	-	-	w	-	-
N5-3	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	w	-	-	-	w	-	-
T3-2X	+	+	-	w	+	-	-	+	-	+	+	+	+	+	+	+	-	-	-	-	-	-	-	-	w	-
PT2-3	-	w	w	w	w	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
PN8-2	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
PN1-2	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
TT2-2X	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
T8-1X	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
PN8-1	w	-	w	w	-	-	-	+	-	-	w	w	+	-	w	w	-	-	w	w	-	-	-	-	w	-
PN9-3	-	-	-	-	+	-	-	-	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-
PN20-1	-	-	-	-	+	-	-	-	-	-	-	-	-	+	-	-	w	-	-	-	-	-	-	-	-	-

Symbols: +, positive; -, negative; w, weakly positive

Acid production from carbohydrates (Cont)

Isolate no.	D-Amygdalin	L-Arabinose	D-Cellobiose	D-Fructose	D-Galactose	D-Glucose	Gluconate	Glycerol	Inositol	Inulin	Lactose	D-Maltose	D-Mannitol	D-Mannose	D-Melibiose	D-Melezitose	$\alpha$ -Methyl-D-glucoside	Rafinose	L-Rhamnose	D-Ribose	Salicin	D-Sorbitol	L-Sorbose	Sucrose	D-Trehalose	D-Xylose
PT1-1	-	-	+	-	+	+	-	-	-	+	+	+	-	+	+	-	-	-	-	-	-	-	-	-	-	-
PT1-2	+	-	+	+	+	+	-	+	+	+	+	+	-	+	+	+	+	+	+	+	+	+	+	+	+	+
PT2-4	+	-	+	+	+	+	-	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
PT6-4	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
PN16-5	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
N5-1X	+	-	+	+	+	+	-	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
N9-2	-	+	-	+	+	+	+	+	-	-	+	+	+	+	+	-	-	-	-	-	-	-	-	-	-	+

Symbols: +, positive; -, negative; w, weakly positive

## 1. Primers for 16S rRNA gene amplification and sequencing

<b>9F</b>	5'-GAGTTT GATCCTGGCTCAG-3'
<b>357R</b>	5'-CTGCTGCCTCCCGTAG-3'
<b>802R</b>	5'-TACCAGGGTATCTAATCCC-3'
<b>1115R</b>	5'-AGGGTT GCGCTCGTTG-3'
<b>1541R</b>	5'-AAGGAGGT GATCCAGCC-3'

## 2. 16S rRNA gene sequences

### 2.1 Strain PA4-1 (1080 bp)

TTTAGTTAACGCTGGCTCAGGACGAACGCTGGCGGTGCTTAATACATGCAAGTCGAGCGGATCTCAAGGGAGCTGCTTCCTGAGAAG  
 GTTAGCGGCCGAAACGGGGT GAGTAACACGTTAGGCAACCTGCCCTCAAGACGGGATAACATTGGAAACGAATGCTAAGACGGGATACGCCAA  
 GGAGGAGGCATCTCTCTGGGAAACA CGCGCAAGCTGTGGCTTGAGGATGGGCTGCGCGCATTAGCTAGTTGGGGGGTAA CGGCCACC  
 AAGGCACGATGCGTAGCCGACCTGAGAGGGT GAA CGGCCACACCTGGGATTGAGACACGGCCAGACTCTACGGGAGGCAGCAGTAGGGA  
 ATCTTCCACAATGGCGCCAAGCCTGATGGAGCAACCGCGTGTAGTGAGTGAGGAAGCCCTCGGGCTGTAAGCTCTGTTGCCAGGGAGAATAAG  
 AGCCAGTTAACCTGCTGGTTCGATGACGGTACCTGAGAAGAAAGCCCCGCTAACTAOGTGCCAGCAGCGCGGTAATACGTAGGGGCAAGCGTT  
 GTCCGGAAATTGGCGTAAAGCGCGCGCAGCGGTTCTTAAGTCTGGTTTAAGTGGGGGCTCAACCCCGTGA CG CACTGGAAACTGGGA  
 GACTTGAGTGCGAGAGGGAGAGCGGAATTCCACGTGAGCGGTAAATCGCTAGAGATGTGGAGGAACCCAGTGGCAAGGCCCTCTG  
 ACTGTAAGTGACGCTGAGGCGCGAAAGCGTGGGGAGCAAACAGGATTAGATAACCTGGTAGTCCA CGCGTAAACGATGAGTGCTAGGTGTTGG  
 GGGGGTCCACCCCTCGGTGCGAAGTTAACACATTAAGCACTCCGCTGGGAGTACGGTOGCAAGACTGAAACTCAAAGGAATTGACGGGGAC  
 CGCACAAGCAGTGGAGTATGTGGTTAACGCAAGCAACGCGAAGAACCTTACCAAGGTCTGACATCCCTCTGAATCGTTAGAGATAGGCCGC  
 CTTCGGGACAGAGGAGACAGGTGGTGCATGGTACGTCAGCCC

## 2.2 Strain PBS5 (1493 bp)

ATTAGTTTGTCTGGCTCAGGACGAACGCTGGCGCGTCCTAATACATGCAAGTCAGCGAATCAATAGGAGCTTGCTCTGTGGTAGCGGC  
 GGACGGGTGAGTAACAGTGGGCAACCTGCCGTAAAGACTGGGATAACACCGGAAACCGGTGCTAAATACCGGATAATTCTTCCCTCATGAG  
 GGAAAATTGAAAGTGGTTTCGGCTGACACTAACAGATGGGCCGCGCATTAAAGCTAGTTGGTAGGTAACGGCTACCAAGGCGACOGA  
 TGCGTTAGCGACCTGAGAGGAAATCGGCCACTGGGATTGAGACCCGGCATTCTAACGGAGGCGAGCTAGGAATTCCCATGGGACGAA  
 ATCTGAAGGAGCAAAGCCGCGTGAGCGAGAACGCTTICGGGTGTAACACTCTGTTAGGGAAAGAACAGTACGGAGTAACGCCGTACCTT  
 GACGGTACCTAACAGAAAGCCACGGCTAACACTACGTGCCAGCAGCCGGTAATACGTAGGTGCAAGCGTTGCTCGGAATTATGGGCGTAA  
 GCGCGCGCAGCGCTTAAAGTCTGATGTGAAAGCCCCACGGCTAACCCGTGGAGGGTCACTGGAAACTGGGGACTTGAGTGAGAAGGAGA  
 AACCGGAATTCCACGTGAACCGGTGAAATCGGAGAGATGTGGAGGAACACCAGGGCGAAGGGGGCTTCTGGCTGTAACACTGACGCTGAG  
 GCAGCGAAAGCGTGGGGAGCAACAGGATTAGATACCCCTGGTAGTCCACGCCGTAACCGATGAGTGCTAAGTGTAGGGTTCCGCCCTTCTG  
 TGCTGCAGCTAACGCTTAAGCACTCGCTGGGAGTACGCCGCAAGGCTGAAACTCAAAGGAATTGACGGGGGCCACAAGCGGTGGAG  
 CATGTGGTTAATTGAAAGCAAACCGCAAGAACCTTACCAAGGTCTTGACATCCTCTGACACTCTAGAGATAGGACGTTCCCTCGGGGACAG  
 AGTGACAGGTGGTGCATGGTTGCTCAGCTCGTGTGAGATGGTTAAGTCCCGCAACGAGCGAACCCCTGATCTAGTGGCAGCATT  
 CAGTTGGGACTCTAAGGTGACTGCCGTGACAAACCGGAGGAAGGTGGGATGAGTCAAATCATGCCCTTATGACCTGGGTACACAC  
 TGCTACAATGGATGGTACAAGGCGTCAAGGCTAGCCAATCCCATAAAACCATTCTCAGTCCGATTGAGGCTGCAACTCGCT  
 ACATGAAGCCGAATCGTAGTAATCGGGATCAGCATGCGCGGTGAAATGTTCCCGGCTTGTACACACCGCCGTCACACAGGAGAGT  
 TGTAACACCGAAGTCGGTGGGTAACCGTAAGGAGGCCAGCCGCTAACGGTGGGACAGATGAGGTGAAGT

## 2.3 Strain T3-3 (1204 bp)

TCCGGGGTTGAAAAACCTTGTTTTTAGGGAAAGAACCAAGTCCGGGAGGAAACGCCCTCTAACCCCTTGACGGTCCCCAACCAAG  
 AAAAGCCCCGGGCTAAACTAACCGTGGCCAAGCACCCGCGTTAACCGTAAGGGTGGCAAGCGCTAACCGGGAAATTATGGGCC  
 GTAAAACCGCGCGCCAGGCGTTTAAGTTGATGTTGAAAGCCCCACGGCTAACCGTGGAGGTGCTTGGAAACTGGGGACTGAGTG  
 CAGAAGAGAAAAGCGGAATCCCACTGTAAAGCGGTGAAATCGCTAGAGATGTGGAGGAACACCAGTGCAGGGCTTTGGCTCTGTAAC  
 TGACGCTGAGGCAGCGAAGCGTGGGAGCAAACCGGATTAGATCCCTGGTAGTCCACGCCGTAACCGATGAGTGCTAAGTGTAGGGTT  
 CGCCCTTTAGTGCCTGAGCTAACGCCATTAAAGCCCTCCCCCTGGGGAGAACCGGTTCCCAAAGACCTGAAAACCTCCAAAAGG  
 AATTGAAACGGGGGCCCCGCAAAAGCGGGTGGAGCTTGTGTTAAATTGAAAGCCAACCGCAAGAACCCATTATCCAGGTTCTGGA  
 ACCATCCCTGACAACTCTAAAGATAAGAGCTTITCCCCCTCGGGGACAGAGTGCAGGGTGGTGCTGGTTGTCGTCACCTCGT  
 TCGTGAAGTGGTGGGTTAAAGTCCCGCAAACGAGCGCAACCCCTGATCTAGTGGATTGAGGCTGACATTAAGTGGGACTCTAAGGTGCTGCC  
 GTGACAAACCGGAGGAAGGTGGGATGACGTAAAATCATGCCCTTATGACCTGGCTAACACCGTCTAACATGAGCTGAATCGTAGTAATCG  
 CGGATCAGCATGCCCGGTGAAATGTTCCCGGCTTGTACACACCGCCGTCACACCAACGAGAGGTTGTAACACCCGAAGTCGGTGGAGTAAC  
 CGTAAGGAGCTAGCCGCTAACGGTGGACAGATGATTGGGTGAAGACGTAACAAGGGAACCGC

## 2.4 Strain T3-2 (1167 bp)

TTTCCGGGGTCGGAAAAACTCTTTTTAAGGGAAAGACCAGTCCTAACGTTAGAAAAGCCTGGGCCCTTGAACCTTCCCTAAACCCAGAAAA  
AGCCGCGCTAAACTACGGCCAAGCAGCCCGCTTAACCGTAGGTGGCAAGCGTTACCGGAAATTATGGGCGTAAAGCGCGCAGGTG  
GTTCTTAAAGCTGTGAAAGCCCACGGCTAACCGTGGAGGGCTTGGAACTGGGAGACTTGAGTGAGAAGAGGAAAGTGAATTCCA  
TGTGTAGCGGTGAAATCGTAGAGATATGGAGGAACCCAGTGGCGAAGGCAGTTCTGGCTGTAACTGACACTGAGGCGOGAAAGCGTGG  
GAGCAAACAGGATTAGATAACCTGGTAGTCCAOGCGTAAACGATGAGTGCTAACGTTAGAGGTTCCCCCTTAAAGGTGCTGGAAAGGTT  
AACCCGCTTAAGCACTCCCCCTTGGGGAGTTACGCCCTAAAGGCTTGAACCCCTCAAAGGAATTGACCGGGGCCACAAGC  
GGTGGGAGCATGGTGGTTITAATCCGGAAAGCAACCGOGAAAGAACCCCTAACCGAGTCTTGTACATCCCTCTGAAAAACCCAAAGAGATAGG  
GCTTCTCTTCCGGGAGCAGAAGTGAACAGGGTGGTGATTGGTTGTOCGTCCAGCCTCTTGTGAGATGTTGGGGTTAAATCCCC  
GCAAACGAGCGAACCCCTGATTCTAACGGCATCAATTAAATGGGCCCTAACGGGACTGCCCCGTGAAACCGAGGTGGAGCTATTCTCATAAAACCG  
AAATCTCATGCCCTTAACACGGCTAACACGGTCTAACAGCTGCTAACATGGACGTACAAGAGCTGCAAGACCGAGGTGGAGCTATTCTCATAAAACCG  
TTCTCAGTTCGATTGAGGTGCAACTGCCAATGAAAGCGGAATCGAGTAATCCCGGATCACAGCCGCGTGAATACTTCCGGGCTTGT  
ACACACCGCCCGTACACACGAGAGTTGTAACACCGAAGTCGGTGGGTAACCTTTGGAGGCCAGCCTAACGGTGGGACAGATGATTGG  
GGTAAAACGTAACCCCTGGGACCCCCC

## 2.5 Strain N14-2 (803 bp)

ATCCITAGTTGATCCTGGCTCAGATTGAACTGGCGGCAGGCCAACACATGCAAGTCGAGCGGATGAAGAGGCTTGTCTCTGATTAGCTGGC  
GGACGGGGGAGTAATGCCCTAGGGAAATTCTGCCTGGTTAGGTGGGGGATAACCGTCCCCGGAAAACGGGGCGGCCAACCGCATAACG  
CCCTACCCGGGAAAAAGCAGGGGACCTGGGGCTTGGCGCTTACAGATGAAGCCCTAACGGTGGATITAGCCCTAGTTGGTGAGGT  
AAAATGGGCTCACCAAGGCCAGATCCCTAACCTGGCTGAGGGATGATCCAGTCCACCACTGGGAACTTGTAAACACGGTCCAACCTTAGGG  
AGGCAGCGTGGGGAAATATGGCAATGGGAAAGCCTGATCCAGCCATGCCGCGTGTGAAGAAGGTCTCGGATTGAAAAGCACTTAAGTT  
GGGAGGAAGGGCAAGTAAGTTAACCCCTGCTGTTTGACGTTCCGACAGAATAAGCAGGGCTAACCTCGTGCAGCAGCCGCGTAAATACG  
AAGGGTCAAGCGTTAACCGGAAATTACTGGCGTAAAGCGCGTAGGGTGGTAAAGTGGATGTGAAAGCCCGGCTAACCTGGGAAAC  
TGCATCCAAAATGGCAGCTAGAGTACGGTAGAGGGTGGGAAATTCTGTGTTAGCGGTAAATGCGTAGATAGGAAGGAACCCAGTGG  
CGAAGGCGACCACTGGACTGATACTGAATGAGTCAGACTCCC

## 2.6 Strain T6-4 (800 bp)

TCCCCCTAGTTGCTGGCTCAGATTGAACTGGCGGCAGGCCAACACATGCAAGTCGAGCGGATGAAGGGAGCTGCTCTGATTCA  
GCCGGACGGGTAAGTAATGCCCTAGGAAATCTGGCCCCGGTAATGGGGGGATAACGTTCCGAAAGGGAACCGCTAAATACCCGGCAT  
ACGTCCCTACCGGGAGAAAAGCGGGGATCTGGACCTCGCTTATOGGATGACGCCAACGGTCCGGATTAGCCAAGTTGGTGGGGTAAAA  
GGCCTACCCAAAGCGACCGATCCGTAACCTGGCTGAGAGGGATGATCAGTCCACACTGGAACTGGAGACCCGGTCCAGACTCCCTACGGG  
GGCAGCGAGGGAAATATTGACATGGTAAGCCTGATCCAGCCATCCCGCTGTGAAGAAGGTCTCGGATTGAAAGCCTTAAGTGG  
GGAGGGAGGGCAGTAAGTTAACCCCTGCTGTTGACGTTACCCAAACAGAATAAGCACCGGCAACTCGTCCAGCAGCCGCGTAAATACGAA  
AGGGTCAAGCGTAATCGGAAATTACCTGGCGTAAAGCGCGTAGGTGGTTGGTAAGATGGATGTGAAATCCCCGGCTAACCTGGGACCGA  
TCCATAACTGCTGACTAGAGTACGGTAGAGGGTGGTGGAAATTCTGTGTTAGCGGTAAATGCGTAGATAGGAAGGAACCCAGTGGCGAA  
GGCGACCACTGGACTGATACTGGAATGAGTCGGACCCCCC

## 2.7 Strain PT4-2 (1510 bp)

AATCCTAGTTGGCTGGCTCAGGACGAACCGGGCGCGTGCTAAACATGCAAGTCGAGCGGATCTTCAAGTAGCTTGACTTGAGAA  
 GTTAGCGCGGACGGGTGAGTAACACGTTAGGCAACCTGCCATAAGACGGGATAACATTGGAAACGAATGCTAAGACGGGATACGCAGAGT  
 GGGGCATCTCACTTGGAAACACGGTCAAGCTGTGGCTATGGATGCCGTACGCCCTAGTAGCTAGTTGGCGGGAAACGGCTACCAAGG  
 CGACGATGCGTAGCCGACCTGAGAGGGTGAACGGCCACACTGGACTGAGACACGGCCAGACTCTACGGGAGGAGCAGTAGGAACTTC  
 CACAATGGCGCAAGCCTGATGGAGCAACCCGCGTGAAGTGGAGGAAGGCTTCGGGTGTAAGCTCTGTTGCCAGGGAAAGAATAAGGGGAGG  
 TCACTACTCGTCCGATGACGGTACCTGAGAAGAAAGCCCCGCTAACACTGTCGAGCAGCCGCGTAATACGTAGGGGGCAAACGTTGTCGG  
 ATTATTGGGCTAAAGCGGCCAGGGCTTAAGTCTGGTTAAGTGCCTGGCTCAACCCGCTGCGCATCGGAAACTGGGAGACT  
 GAGTCAGAAAGAGGAGAGCGGAATTCCACGTGAGCGTGAATGCGTAGAGATGTGGAGGAACACAGTGGCGAAGCGGATCTCTGGACTGT  
 AACTGACGCTGAGGCGCGAAAGCGTGGGAGCAAACAGGATTAGATAACCTGGTAGTCCACGCCGTAAACGATGAGTGCTAGGTGTTGGGGGG  
 TCCACCCCTCGTCCGAAGTTAACATTAAGCACTCTGCCCTGGGAGTACGGTCGAAGACTGAAACTCAAAGGAATTGACGGGACCCG  
 ACAACCAGGGAGATGTGGTTAACCGAAGCAACCGGAAAAAAACCTAACCGAGGCTGGACATCCCTCTGACCGTCCAAGAAAAGGGCTT  
 CCTCGGGCAGAGGAGACAGGTATGCTGGCTAACACGCTGAGGAGGAGCGTGAAGTGGCTGAGATGTGGTTAACGAGCAGCCATGCCC  
 TTAGTTGCCAGCACGTAAGGGGGCACTCTAGATTGACTGCOGGTGACAAACCGGAGGAAGGCGGGATGACGTCAAATCATGCCC  
 GACCTGGGTACACACGTACTAACATGGCCGTACACGGCTGCGAAGGAGCGATCCGGAGCTAACCTATAAAGCCGCTCAGTGGATTG  
 CAGGCTGCAACTCGCTGCATGAAGTGGAAATTGCTAGTAATCGCGGATCAGCATGCCCGGTGAATACGTTCCGGGTCTGTACACACCGCCC  
 GTCACACCACGAGAGTTACAAACACCGAAGCCGGTGGGTAACCGCAAGGAGCCAGCCGTOGAAGGTGGGAAGATGATTCTAGAGAGT

## 2.8 Strain PN8-3 (1491 bp)

AGTTTGAGTTCTGGCTCAGGACGAACCGGGGGTGCTAAACATGCAAGTCGAGCGGTTCAAGGGAGTTGCTCCCGAGAAGGTTAACGGC  
 GGACGGGTGACCTAGTAGGCAACCTGCCCTCAAGCCGGATAACATTTGGAAACGAATTAAAGACCCGATACCAAGGAAGGAGGCTTT  
 TTCTGGAAACCAAGGGGCAACTTGCTGGCTTGAGAAAGGCCCTGACGGCCCTTAAACATTTGGGGGTAAGGCCCCCAGGGCAGT  
 CGTAGCCGACCTGAGAGGGGAACGCCACACTGGACTGAGACACGGCCCGACTCTCACGGGAGGAGCAGTAGGGAACTTCCACAAATGG  
 GCGCAAGCCTGATGGAGCAAACCGCGTGTAGTGGAGGAAGGCCCTGGGTGTAAGGCTCTGTTCCAGGGAAAGAATAAGAGCCAGTTA  
 CTGCTGAGCTGACCTGAGAAGAAAGCCCGCTAACACTACGTGCCAGAGCCGGTAAACGTTAGGGGCAAGCGTTGTCGGAAATTG  
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 ACTGCGCAGAGCGTGGGGAGCAAACAGGATTAGATAACCTGGTAAGTCCACCCGTAACAAATGAGTGCTAGGTGTTGGGGGCTCC  
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 GCTCGGGACAGAGGAGACAGGTAGTGCATGGCTGTGTCAGACTCTGCTGAGATGTGGGTTAAGTCCCGCAACCGAGCGCAAC  
 CCTTGATGAGCTTGGGCACTCTAAGGTGACTGCCGTGACAAACCGGAGGAAGGTGGGGATGACGTCAAATCATGCCC  
 TTAGTTGCCAGCACCTGGGGCACTCTAAGGTGACTGCCGTGACAAACCGGAGGAAGGTGGGGATGACGTCAAATCATGCCC  
 ACCTGGGCTACACCGTACTAACATGGCGGTACAOGGCAGCGAAGGAGCGATCTGGAGCCAATCTATAAAGCCGGTCTAGT  
 CGGAGCTGCAACTCGCTGCATGAAGTGGAAATTGCTAGTAATCGCGGAACAGCATGCCCGGTGAATACGTTCCGGGTCTGT  
 ACACACCACGAGAGTTACAAACACCGAAGCCGGTGGGTAACCGCAAGGACCACCTCCATTGGAAGAT

## 2.9 Strain PN12-3 (1505 bp)

GGGGGGGGATTTGTTTCCCTGCCGACCACCCATTCCCTAAACATAAATTCAAAGATTTTAAAAACCTTCTCTAAAAAGTTAAC  
 CCCCCACCTTAATTAACCTTAGGCAA CGTCCCCATAAACCGGGAAACCTGGAAAGGAATGCTAAAGACCGGATACGCAAAATGGGGCA  
 TCTCATTTGGAAAACACGGGCAAGCTGGCTTATGGATGGCCTGGGCGATAAGCTAGTTGGTGGGTAA CGCCTACCAAGGCGAOGA  
 TGGTAGCGACCTGAGAGGGTAA CGGCCACACTGGGACTGAGACACGCCAGACTCTAOGGAGGCAGCAGTAGGGAATCTTCCACAATG  
 GGCGCAAGCCTGATGGAGCAACCGCGTGAGTGAAGGAAGGCTTCGGGCGTAAGCTCTGTCGGGAAGAATAAGGGCGAGGTAACCTAC  
 TCGTCCGATGACGGTACCTGAGAAGAAGGCCCGCTAACTACGTGCCAGCAGCGOGGTAA TAGTGTAGGGCCAACGTTGTCGGAATTAT  
 TTGGCGTAAAGCGCGCGCAGCGGTTCTTAATTCTGGTGTAAAGTCGGGGCTAACCCGTGTCGATCGGAACTGGGAGACTTGAGTG  
 CAGAAGAGGAGAGCGGATTCACGTGTAGCGGTAAATGCGTAGAGATGTGGAGGAACCCAGTGGCGAAGGGGGCTCTGGACTGTAACG  
 ACGCTGAGGCGCGAAAGCGTGGGAGCAAACAGGATTAGATACCGTGTAGTCCAOGCGTAAACGATGAGTGCTAGGTGTTGGGGGTCCAC  
 CCCTCGGTGCCAGTAAACATTAAGCACCCGCTGGGAGTACGGTOGCAAGACTCAAAGGAATGACGGGACCCCCACAAGC  
 AGTGGAGTAAATGGTTAATTCGAAGCAACCGAAAAACCTTACCAAGGGTCTGGACATCCCTGACCGTCCAAGAGATAGGGCTTCCCTCGG  
 GGCAGAGGAGACAGGTAGTGCATGGTCTGCGTCAGCTGGTGTAGATGGTGGTTAAGTCCCGCAACGAGCGAACCCCTGAATTAGTG  
 CCAGCACTGAAGGGTGGGACTCTAGATTGACTGCCGTGACAAACGGAGGAAGGCGGGATGACGTCAAATCATGCCCCATGACCTGG  
 GCTACACGTACTACAATGGCGGTACAA CGGCTGCGAAGGAGCGATCCGGAGCCAATCTATAAAGCCGGTCTCAGTCGGATTGGAGGCTG  
 CAACTCGCTCCATGAAGTCCAATTGCTAGTAATCGGGATCAGCATGCGGGTGAATACGTTCCGGTCTTGACACACCGCCCGTCACACC  
 ACGAGAGTTACAACACCGAAGCGGTGGGTAACCGCAAGGAGCCAGCGTCGAAGTGGGAAGATGATTCTGGAGAGTA

## 2.10 Strain PT6-2 (1508 bp)

GTGAGTTGAACCTGGCTAGGAOGAACGCCGGCGCGTGCCTAATACATTGCAAGTCGAGCGGATCTCAAGGGAGCTTGCTCCGAGAAGGT  
 TAGCGGCCGGACGGGTGAGTAACACGTTAGGCAACCTGCCCTCAAGACCGGATAACATTGGAAA CGAATGCTAAGACGGATA CGCAAGGAGG  
 AGGCATCTCTCTGGAAAACACGGCGCAACTGTGGCTTGAGGATGGGCTGCGGCCCTAATTAGTGGCGGGTAACGGCCACCAAGGCGAC  
 GATGCGTAGCCGACCTGAGAGGGTGAACGCCACACTGGACTGAGACACGCCAGACTCTAOGGGAGCAGCAGTAGGGAATCTTCCACA  
 ATGGCGCAAGCTGATGGAGCAA CGCCGCTGAGTGAGGAAGCCTTGGGCGTAAAGCTCTGTCGAAAGGAGAAGAATAAGAGCCAGTAAAC  
 TGCTGGTTCGATGAOGGTACCTGAGAAGAAGGCCCGCTAACTACGTGCCAGCAGCCGCTAAACGTTAGGGCAAGCGTTGTCGAAATT  
 ATTGGGTAAGCGCGCGAGGGGTCTTAAGTCTGGTGTAAAGTCGGGGCTCAACCCGTGACCGACTGGAAACTGGGAGACTTGAGTGC  
 AGAAGAGGAGAGCGGAATTCCACGTGTAGCGGTGAATGCGTAGAGATGGAGGAACCCAGTGGCGAAGGCGCTCTGGACTGTAACTGA  
 CGCTGAGGCGCGAGAAGCGTGGGAGCAA CAGGATTAGATACCGTGTAGTCCACGCCGTAACGATGAGTGCTAGGTGTTGGGGGTCCACC  
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 TAGTTGCCAGCACTCGGGTGGGCACTCTAAGGTGACTGCCGTGACAAA CGGAGGAAGGAGGGATGACGTCAAATCATGCCCCATG  
 CCTGGGCTACACCGTACTACAATGCCGTACAACGGCAGCGAAGGAGCGATCTGGAGCCAATCTATAAAGCCGGTCTCAGITGGATTGCA  
 GGCTGCAACTCGCCTGATGAAGTGGATTGCTAGTAATCGGGATCAGCATGCCGGTGAATACGTTCCGGGTTGTACACACCGCCCGTC  
 ACACCAAGAGAGTTACAACACCGAAGCCGGTGGGTAACCGCAAGGAGCCAGCGTCGAAGTGGGAAGATGATTCTTAGAGAGTT

## 2.11 Strain PN13-1 (1468 bp)

TTAGGTTGAAACCTGGCTAGGAOGAACGCCGGCGCGTGCCCTAATACTGCAAGTCGAGCGGACTTAACTTCGGCAAAGTTAGCG  
 GCGGACGGGTGAGTAACACGTGGTAACCTGCCATAAGACTGGATAACATTGGAAACGAATGCTAATACCGGATACCGAATCGGTG  
 CATGATCGAACCGGGAAAGGGGAGCATTGCCCTTATGGAGGGACCCGOGGCATAACTTAGTTGGGGTAACGGCTCACCAAGGGCGAC  
 ATGCGTAGCCACCTGAGAGGGTGATGTCACACTGGGACTGAGACAOGGCCAGACTCCTACGGGAGGCAGCAGTAGGGATCTCCGCAAT  
 GGACGAAAGTCTGAOGGAGCAACGCCCTGAGTGATAAAAGTTTCTGGATCGTAAAAGCTTGTGAGGGAGAAGAAGCTTGGGCTGAGAGTAAC  
 TGCTCGCAAGGTGAOGGTACCTGAAGAAGAAAGCCCCGGCTAAATTACGTGCCAGCAGCCCCGGTAAACGTTAGGGCCAACGTTGCTCG  
 GAATTATTGGCGTAAAAGGCCCTTGGCTGAGTGAGGGAGCTCAACTTCGAGTCGAGTGGAAACTGCAAAG  
 TTGAGTGCAGAAGAGGAAAGTGGAAATTCAACGTGAGGGTGAATGCGTAGAGATGTGGAGGAACACCAGTGGCGAAGGGCGCTTCTGGGCTG  
 TAATGACGCTGAGGCGCAAGCGTGGGAACAAACAGGATAGATAACCTGGTAGTCCACCCGTAACGATGAATGCTAGGTGTTAGGGGT  
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 CACCAAGCCAGTGGAGTATGGTTAACCGGAAACCTTACCCAGGTCTGACATCCCTTGTGACCGCCCAAGAGAAAGGG  
 CTTCTCGGGACAGAGGAGTCAGGTGGAGCATGGTGGAGCTGGTCTGAGACTCGTGTGAGATGTTGGTTAACGTCACCGAGCGCAACCC  
 TTGATCTTAGTGGCACTTGGTGGGACTCTAGGATGACTGCOGGTGACAAACGGAGGAAGGTGGGGATGACGTCAAATCATCATGCC  
 CCTATGACCTGGGCTACACACGACTACATGGCGATAACACGGAAAGCGAACCGCAGGTGGAGCCTTGAGTTGATCTGGTACAGTTAA  
 TCGACTCAACTCAGTACCAATCGCAATTGAGGATCATAGCACGATGGTAATGTTCCCGCGAGTCAGGCGGCCACAGGTGTTCTGT  
 CCTGGGTACGAAAGCCCCCCCAGGTGGAGATGATCCCTTGGAGAT

## 2.12 Strain T3-2X (1509 bp)

GATTAGTTAATGTCCTGGCTAGGAOGAACGCCGGCGCGTGCCCTAATACTGCAAGTCGAGCGGACTTGCCTTCGGTAAAGTAAGCG  
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 TCACACCAAGAGAGTTTACAACACCGAAGCGGGTGGGTAACCGCAAGGAGCCAGCCGTGAAGGTGGGGTAGATGATGGGTGAAGTC

## 2.13 Strain PT2-3 (1331 bp)

AGGTGGGAAAGGGAGCAATTCTTATGAAAGGCCTTCGCCGCTTAAGTATTGGGAGGAAACGGCTCCCCAAGCGACGATG  
GCGTAACCGCTAGAGGTGATTGCCCTGGGATTGAGACACGGCCCGAGATCTTAOGGAGCAGCAGTTAGGAATTCCCAAAGGAC  
GAAAGTCTGACGGAAGCAACGCCGCTGAGTGATGAAAGTTTGGTGTAAAGTCGTGTCAGGGAGAACGCTAGAGAGAGTAACGCTC  
TTAGGTACGGTACCTGAGAAGAAAGCCCGGCTAACACTGTCAGCAGCOGCCGTAATACTGAGGGGCAAGCGTTGTCGGAAATTATTGGG  
CGTAAAGCCCGCGCAGGGGTTATTAGTCGGTGTAAAGGCTATGGCTCAACCAGATTGCGCACTGGAAACTGGTGTACTGAGTGCAAAA  
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GGTTGCAATTACCTGGTGCAGGTTAACACAATTAGCATCCGCTGGGAGTACGGTGTCAAGACTGAAACTCAAAGGAATTGACGGG  
GACCGCACAAGCCAGTGGAGTATGTTAATTGCAAGCAACGCCGAAAGAACCTTACCAAGGTCTGACATGCCCTGACCGCTAGAGATAGA  
GCTTCTTCGGAGCAGGGACACAGGTGTGATGGTGTAACTGAGGTGAGATGTTAGTCCGCAACGAGCGCAACCC  
CTAAATGTTAGTGCAGCAGGTAAGCTGGGACTCTAACGTCAGGAGATGCGAGATGGAGGAAAGTGGGATGACGTC  
CCCCTTAAGCTGGCTAACACGTACTAACATGGCAGTACAACGGGAAGGAGTCGAGATGGAGGCAATCTCAAAAGCTGGTCTGAGT  
TCGGATTGAGGTGCAACTCGCTGATGAGTCGAAATTGCTAGTAATGGGATCAGCATGCCGGTGAAATGTTCCGGCTTGACAC  
ACGCCCGTCACACCAAGAGAGTTACACCCGAAGCCGGGGTAAACCGCAAGGGAGCCAGCGTCAAGGGTGGGAGTGTGAGGAAAGT  
GAAGT

## 2.14 Strain PN8-2 (1524 bp)

TAGTTTGTCTGGCTCAGGAAGAACGCTGGCGCGTGCTTAATACATGCAAGTCGAGGGACAGATGGGAGCTTGTCTCCCTGATGTTAGCGGC  
GACGGGTGAGTAAACCGTGGTAAACCTGCTGTAAGACTGGGATAACTCCGGAAACCGGGCTAATACCGGATGGTGTGTTGAAACCGCATGGT  
CAAACCAAAAAGGTGGCTTGGCTACCACTTACAGATAGGACCOGCCGCATTAAAGCTTAGTTGGTGGAGGTAACGGCTTACCAAGGCAACGA  
TGCCTAGCCGACCTTGAGAGGGTGTAGCGCCCCACTGGGATTGAGACACGGCCCGAGATCTTACGGAGGCAGCAGTAGGGAACTTCCCAAT  
GGACGAAAGTCTTGACGGAGCAACCCCGCTGAGTGTAGAAGGTTTCGGATGTTAAAGCTCTGGTTAGGGAAAGAACAGTACCCGTTGAA  
TAGGGCGTACCTTGACGGTACCTAACCCAGAAAGCCACGGCTAACTACGTGCCAGCAGCGCGTAATACGTAGGGCAAGCGTTGCTCGGAAT  
TATTGGCGTAAAGGCTCGAGGCGGTCTTAAGTCTGTAGTGAAGGCCCCGGCTAACCGGGGAGGGTCACTGGAAACTTGGGAACCTGGA  
GTGCAAGAGGGAGAGTGGATTCCACCGTGTAGCGTGAAATTCGCTAGGATTGGAGGAACACCACTGGCGAAGGCACCTCTTGGTC  
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CGGGGGCCCGACAAGCGGTGGAGCATGTGGTTAATTGAAAGCAACGCGAAGAACCTTACCAAGGTCTTGACATCCTCTGACAATCTTAGAG  
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TTCGGATCGCAGTGTGCAACTCGACTGCCGTGAGCTGGAATCGCTAGTAATCGCGGATCAGCATGCCCGTGAATACGTTCGGGCTTGTAC  
ACACCGCCCGTCACACCAAGAGAGTTGTAACCCGAAGTCGGTGAGGTAACCTTTAGGAGGCCAGCGCGAACGGTGGGACAGATGAGGAGG  
TGAAGTC

## 2.15 Strain PN1-2 (1083 bp)

GTGCTAGTTAAGAAAAGCACCAACTTGTGCTGGCTCAGATTGACGCTGGGGCAGGCCAACATGCAAGTCAGGGATGAAGGGAGCTTGCTCTGGATTCAGCGGGGACGGGTGAGTAATGCCAGGAATCTGCCTGGTAGTGGGGGATAACGTCGGAAAOGGGCGTAAATACCGCATACGTCTGAGGGAGAAAGTGGGGATCTCGGACCTCACGCTATTAGATGAGCCTAGGTOGGATTAGCTAGTTGGTGGGTAAAGGCCTACCAAGGGACGATCCGTAACCTGGTCCCTGAGAGGAAAATCAGTCCCACCTGGAACCTGAACACGTTCCAATTCTCCGGGCAATTGGGGATTTGACCAAAGGTTAAATGAATCCCAGCCATCCCCGTGTGAGAAAAGGTTTCOGGTTITGAAAGCCTTTAATTGGGAGGAAGGGCAGTAAGTTAAATACCTTGCTGTTTGACGTTACCAACAGAAATAAGCACCGGCTAACCTCGTGCCAGCAGCCGCTAACAGAAGGGTCAAGCCTTAATCGGATTACTGGCGTAAACCGCGCGTAGGTGGTCCAGCAAGTGGATGTGAAATCCCCGGCTCAACCTGGGACTGCATCCAAAACACTGAGCTAGAGTACGGTAGAGGGTGGAAATTCTGTGAGGGTGAATGCGTAGATATAGGAAGGAACACCGAGTGG(GAAGGGCACCACCTGGACTGATACTGACACTGAGGTGCGAAAGCGTGGGAGCAAACAGGATTAGATACCTGGTAGTCCACGCCAACAGTGTGCACTAGCGTTGGGATCTTGAGATCTTAGTGGCGCAGCTAACCGATAAGTCAGCCCTGGGAGTACGCCAACGGTTAAAACCTCAAATGAAATTGACGGGGGCCGCAAAAGCGGTGGAGCATGTGGTTAAATTGAAAGCAAAGGAAGAACCTAACCTGGCCTGACATGCTGAGAACCTTCAGAGATGGATTGGTGCCTCGGGAACTCAGACACAGGTGCTGCATGGGAAACTCAGGCC

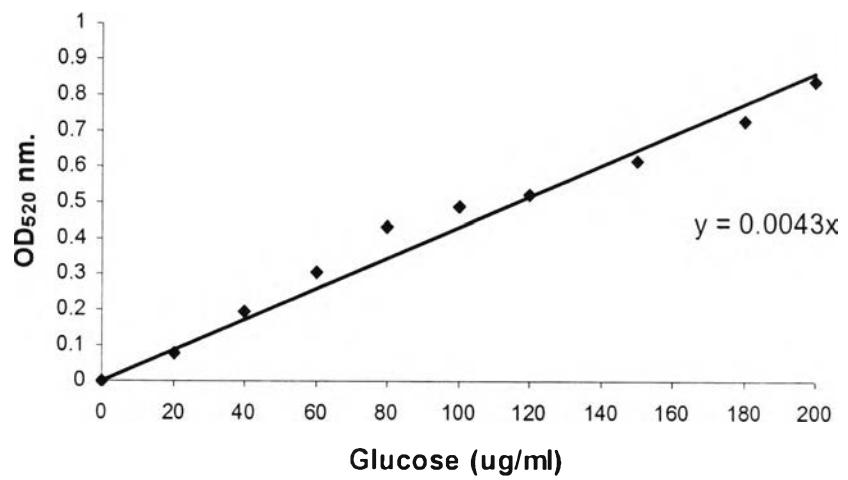
## 2.16 Strain PN9-3 (1563 bp)

## 2.17 Strain N9-2 (691 bp)

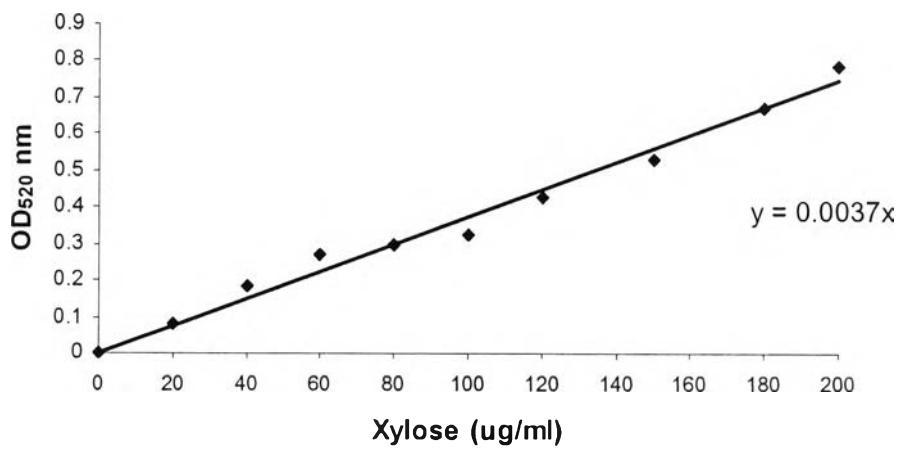
TCCCTCTAACGCTTACCGTAAGTCTTCGTCCAGGGGGCGCCTCGCCACCGGTATCCTCCAGTATCTACGCATTACCGCTACACCTGGAATTCTACCCCCCTCTACGAGACTCAAGCTGCCAGTATCAGATGCAGITCCAGGTTGAGCCCCGGGATTCACTCTGACTTAACAAACGCCCTGCGTGCCTTACGCCAGTAATTCCGATTAACGCTTGACCCCTCGTATTACCGGGCTGCTGCACGGAGTTAGCCGGTGCTCTCTGCGGGTAAOGTCATGAGCAAAGGTATTAACTTACTCCCTCCTCCCGCTGAAAGTACTTACAACCCGAAGGCCTCTCATACACGCGGATGGCTGATCAGGCTTGCGCCATTGTGCAATTCCCCACTGCTGCCTCCCGTAGGAGTCTGGACCGTGTCTCAGTTCCAGTGTGGCTGGTCATCCTCTCAGACCAGCTAGGGATCGCCTAGGTGAGCGTTACCCCACCTACTAGCTAATCCCATCTGGGCACATCCGATGGCAAGAGGCCCGAAGGTCCCCCTTTGGCTTGCGACGTTATGCGGTATTAGCTACCGTTCCAGTATTATCCCCCTCCCATCAGGGAGTTCCCCAGAAATTACTCACCGTCCGCCAATTCGTCAGAAAAAAA

## APPENDIX E

### 1. Standard curve of glucose



### 2. Standard curve of xylose



## BIOGRAPHY

Miss Patcharin Boon-eiam was born in January 8, 1984 in Bangkok, Thailand. She graduated from Department of Microbiology, Faculty of Science, Chulalongkorn University, Thailand with Bachelor Degree of Science since 2006.

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