



REFERENCES

- Aissam, H., Errachidi, F., Penninckx, M. J., Merzouki, M. and Benlemlih, M. Production of tannase by *Aspergillus niger* HA37 growing on tannic acid and Olive mill waste waters. Microbiology & Biotechnology. 21 (2005): 609-614.
- Akiyama, H., Fujii, K., Yamasaki, O., Takashi, O. and Iwatsuki, K. Anitibacterial action of servaral tannins against *Stephylococcus aureus*. Antimicrobial Chemotherapy. 48 (2001): 487-491.
- Alvarez, O. M., Patel, M., Booker, J. and Markowitz, L. Effectiveness of a biocellulose wound dressing for the treatment of chronic venous leg ulcers: results of a single center randomized study involving 24 patients. Wounds. 16(7) (2004): 224-233.
- Bader, R. A., Herzog, K. T. and Kao, W. J. A study of diffusion in poly(ethyleneglycol)-gelatin based semi-interpenetrating networks for use in wound healing. Polymer Bulletin. 62 (2009): 381-389.
- Baily, A. J. and Light, N. D. Genes, biosynthesis and degradation of collagen in connective tissue in meat and meat products. Elsevier Applied Science. 1 (1989): 39-53.
- Bailey, A. J. and Paul, R. G. Collagen: a not so simple protein. Society of Leather Technologists and Chemists. 82(3) (1998): 104-110.
- Balange, A. K. and Benjakul, S. Effect of oxidised tannic acid on the gel properties of mackerel (*Rastrelliger kanagurta*) mince and surimi prepared by different washing processes. Food Hydrocolloids. 23 (2009): 1693-1701.

- Basavaraju, K. C., Damappa, T. and Rai, S. K. Preparation of chitosan and its miscibility studies with gelatin using viscosity, ultrasonic and refractive index. Carbohydrate Polymers. 66 (2006): 357-362.
- Bigi, A., Cojazzi, G., Panzavolta, S., Roveri, N. and Rubini, K. Stabilization of gelatin films by crosslinking with genipin. Biomaterials. 23 (2002): 4827-4832.
- Brown, Jr. R. M. Advances in Cellulose Biosynthesis Polymers from Biobased Materials. New Jersey: Prentice hall, 1991.
- Buren, J. P. and Robinson, W. B. Formation of Complexes between Protein and Tannic Acid. Agricultural and Food Chemistry. 17(4) (1969): 772-777.
- Cardona, L., Sanzgiri, Y., Benedetti, L., Stella, V. and Topp, E. Application of benzyl hyaluronate membranes as potential wound dressings: evaluation of water vapour and gas permeabilities. Biomaterial. 17 (1996): 1639-1643.
- Cai, Z and Kim, J. Bacterial cellulose/poly (ethylene glycol) composite. Cellulose. 17 (2010): 83-91.
- Cao, N., Fua, Y. and Hea, J. Mechanical properties of gelatin films cross-linked, respectively, by ferulic acid and tannin acid. Food Hydrocolloids. 21 (2007): 575-584.
- Chambi, H. and Grosso, C. Edible films produced with gelatin and casein cross-linked with transglutaminase. Food Research International. 39 (2006): 458-466.
- Chung, T., Lu, Z. and Chou, M. Mechanism of Inhibition of Tannic Acid and Related Compounds on the Growth of Intestinal Bacteria. Food and Chemical toxicology. 36 (1998): 1053-1060.

- Choi, Y. S., Hong, S. R., Lee, Y. M., Song, K. W., Park, M. H. and Nam, Y. S. Study on gelatin-containing artificial skin: I. Preparation and characteristics of navel gelatin-alginate sponge. Biomaterials. 20 (1999a): 409-417.
- Choi, Y. S., Hong, S. R., Lee, Y. M., Song, K. W., Park, M. H. and Nam Y. S. Study on gelatin-containing artificial skin: II. Preparation and characterization of cross-linked gelatin-hyaluronate sponge. Biomedical Materials Research. 48(5) (1999 b): 631-639.
- Choi, Y. S., Lee, S. B., Hong, S. R., Lee, Y. M., Song, K. W., Park, M. H. and Nam, Y. S. Studies on gelatin-based sponges Part III: a comparative study of cross-linked gelatin/alginate, gelatin/hyaluronate and chitosan/hyaluronate sponges and their application as a wound dressing in full-thickness skin defect of rat. Matererials Science Materials in Medicine. 12 (2001): 67-73.
- Ciechańska, D. Multifunctional Bacterial Cellulose/Chitosan Composite Materials for Medical Applications. Fibers & Textiles in Eastern Europe. 12(4) (2004): 69-72.
- Czaja, W., Krystynowicz, A. and Bielecki, S. Microbial cellulose—the natural power to heal wounds. Biomaterials. 27 (2006): 145-151.
- Deng, C. M., He, L. Z., Zhao, M., Yang, D. and Liu, Y. Biological properties of the chitosan-gelatin sponge wound dressing. Carbohydrate Polymers. 69 (2007): 583-589.
- Diepeningen, A., Debets, A., Varga, J., Gaag. M., Swart, K. and Hoekstra, R. Efficient degradation of tannic acid by black *Aspergillus* species. Mycological Research. 108(8) (2004): 919-925.

- Dong, Z., Wang, Q. and Du, Y. Alginate/gelatin blend films and their properties for drug controlled release. Membrane Science. 280 (2006): 37-44.
- Fontana, J. D., de Souza, A. M., Fontana, C. K., Torriani, I. L., Moreschi, J. C., Gallotti, B. J., de Souza, S. J., Narcisco, G. P., Bichara, J. A. and Farah, L. F. Acetobacter cellulose pellicle as a temporary skin substitute. Applied Biochemistry and Biotechnology. 24/25 (1990): 253-263.
- Gardner, K. H. and Blackwell, J. The structure of native cellulose. Biopolymer. 13 (1974): 1975-2001.
- Gea, S., Bilotti, E., Reynolds, C. T., Soykeabkeaw, N. and Peijs, T. Bacterial cellulose-poly (vinyl alcohol) composites prepared using an *in situ* process. Materials Letters. 64(8) (2010): 901-904.
- Hanna, J. R. and Giacopelli, J. A. A Review of Wound Healing and Wound Dressing Products. Foot and Ankle Surgery. 36(1) (1997): 2-14.
- Hesse, S and Kondo, T. Behavior of Cellulose Production of *Acetobacter xylinum* in 13C-Enriched Cultivation Media including Movements on Nematic Ordered Cellulose Templates. Carbohydrate Polymer. 60 (2005): 457-465.
- Heijmen, F. H., Pant, J. S., Middelkoop, E., Kreis, R. W. and Hoekstra M. J. Cross-linking of dermal sheep collagen with tannic acid. Biomaterials. 18 (1997): 743-754
- Hirai, A., Tsuji, M. and Horii, F. TEM Study of Band-Like Cellulose Assemblies Produced by *Acetobacter xylinum* at 4 °C. Cellulose. 9 (2002): 105-113.

- Hong, F and Qiu, K. An alternative carbon source from konjac powder for enhancing production of bacterial cellulose in static cultures by a model strain *Acetobacter xylinus* ATCC 23770. Carbohydrate Polymer. 72 (2008): 545-549.
- Hoenich, N. Cellulose for medical applications. BioResource. 1 (2006): 270-280.
- Hong, S. R., Lee, S. J., Shim, J. W., Choi, Y. S., Lee, Y. M., Song, K. W., Park, M. H., Nam, Y. S., and Lee, S. I. Study on gelatin-containing artificial skin IV: a comparative study on the effect of antibiotic and EGF on cell proliferation during epidermal healing. Biomaterials. 22 (2001): 2777-2783.
- Hupkens, I, Boxma, H. and Dokter, J. Tannic acid as a topical agent in burns: historical considerations and implications for new developments. Burns. 21(1) (1995): 57-61.
- Jonas, R. and Farah, L. F. Production and application of microbial cellulose. Polymer degradation and Stability. 59 (1998): 101-106.
- Kanjanamosit, N., Muangnapoh, C. and Phisalaphong, M. Biosynthesis and characterization of bacteria cellulose-alginate film. Applied Polymer Science. 115 (2009): 1581-1588.
- Kasapis, S. The elastic moduli of the microcrystalline cellulose-gelatin blends. Food hydrocolloids. 13 (1999): 543-546.
- Keenan, T. R. Gelatin. Encyclopedia of Polymer Science and Technology. 6 (2003): 311-324.
- Kim, J., Cai, Z. and Chen, Y. Biocompatible Bacterial Cellulose Composites for Biomedical Application. Nanotechnology in Engineering and Medicine. 1 (2010): 1-7.

- Kim, T. J., Silva, J. L., Kim M. K. and Jung, Y. S. Enhanced antioxidant capacity and antimicrobial activity of tannic acid by thermal processing. Food Chemistry. 118 (2010): 740-746.
- Klemm, D., Schumann, D., Udhardt, U. and Marsch, S. Bacterial synthesized cellulose artificial blood vessels for microsurgery. Progress in Polymer Science. 26 (2001): 1561-1603.
- Kucharzewski, M., Slezak, A. and Franek, A. Topical treatment of nonhealing venous ulcers by cellulose membrane. Phlebologie. 32 (2003): 147-151.
- Kurosumi, A., Sasaki, C., Yamashita, Y. and Nakamura, Y. Utilization of various fruit juices as carbon source for production of bacterial cellulose by *Acetobacter xylinum* NBRC 13693. Carbohydrate Polymers. 76 (2009): 333-335.
- Lee, K. Y., Shim, J. and Lee, H. G. Mechanical properties of gellan and gelatin composite films. Carbohydrate Polymers. 56 (2004): 251-254.
- Lee, S. B., Jeon, H. W., Lee, Y. W., Lee, Y. M., Song, K.W., Park, M. H., Nam, Y. S. and Ahn, H. C. Bio-artificial skin composed of gelatin and (1-3), (1-6) β -glucan. Biomaterials. 24 (2003): 2503-2511.
- Lee, S. B., Kim, Y. H., Chong, M. S., Hong, S. H. and Lee, Y. M. Study of gelatin-containing artificial skin V: fabrication of gelatin scaffolds using a salt-leaching method. Biomaterials. 26 (2005): 1961-1968.
- Liang X. H., Guo Y. Q., Gu L. Z. and Ding E. Y. Crystalline amorphous phase transition of poly(ethylene glycol)/cellulose blend. Macromolecule. 28(1995): 6551-6555.
- Liang S., Wu J., Zhang L. and Xu J. High-strength cellulose/poly (ethylene glycol) gels. Chem Sus Chem. 1(6) (2008): 558-563.

- Lien, S. M., Ko, L. Y. and Huang, T. J. Effect of pore size on ECM secretion and cell growth in gelatin scaffold for articular cartilage tissue engineering. *Acta Biomaterialia*. 5 (2009): 670-679.
- Lii, C. Y., Tomasik, P., Zalesska, H., Liaw, S. C. and Lai, V. Carboxymethyl cellulose-gelatin complexes. *Carbohydrate Polymers*. 50 (2002): 19-26.
- Lokeswari, N. and Jayaraju, K. Tannase Production By *Aspergillus niger*. *E-Journal of Chemistry*. 4(2) (2007): 192-198.
- Lowe, R.D. Two part wound dressing. U.S. Patent 0051688, Feb. 28, 2008.
- Mayall R. C., Mayall A. C., Mayall L. C., Rocha H. C. and Marques L. C. Tratamento das ulcerações troficas dos membros com um novo substituto da pele. *Revista Brasileira de Cirurgia*. 80(4) (1990): 257-283.
- Nagahama, H., Kashiki, T., Nwe, N., Jayakumar, R., Furuike, T. and Tamura, H. Preparation of biodegradable chitin/gelatin membranes with GlcNAc for tissue engineering applications. *Carbohydrate Polymers*. 73 (2008): 456-463.
- Natarajan, N., Shashirekha, V., Noorjahan, S. E., Rameshkumar, M., Rose, C. and Sastry, T. P. Fibrin-Chitosan-Gelatin Composite Film: Preparation and Characterization. *Macromolecular Science*. 42(7) (2005): 945-953.
- Neumann, P. M., Zur, B. and Ehrenreich, Y. Gelatin-based sprayable foam as skin substitute. *Biomaterial Research*. 15 (1) (1981): 9-18.
- Pal, K., Banthia, A. and Majumdar, D. Preparation and Characterization of Polyvinyl alcohol-Gelatin Hydrogel Membranes for Biomedical Applications. *AAPS Pharmaceutical Science Technology: Article 21*. 8(1) (2007): 1-5.

- Petre, G., Zarnea, P., and Adrian, E. Biodegradation and bioconversion of cellulose wastes using bacterial and fungal cells immobilized in radio polymerized hydrogels. Conservation and Recycling. 27 (1999): 309-332.
- Phisalaphong, M. and Jatupaiboon, N. Biosynthesis and characterization of bacteria cellulose-chitosan film. Carbohydrate Polymers. 74 (2008): 482-488.
- Ratanavaraporn, J., Damrongsakkul S., Sanchavanakit N., Banapraser, N., Banaprasert, T. and Kanokpanont, S. Comparison of Gelatin and Collagen Scaffolds for Fibroblast Cell Culture. Metals, Materials and Minerals. 16(1) (2006): 31-36.
- Saied, H. E., Basta, A. H., and Gobran, R. H. Research Progress in Friendly Environmental Technology for the Production of Cellulose Products (Bacterial Cellulose and Its Application). Polymer–Plastics Technology and Engineering. 43(3) (2004): 797–820.
- Sangrungraungroj, W. Development of cellulose membrane from nata-de-coco for material separation. Master's thesis, Department of chemical engineering, Faculty of Engineering, Chulalongkorn University, 2003.
- Sanchavanakit, N., Sangrungraungroj, W., Kaomongkolgit, Banaprasert, T., Pavasant, P. and Phisalaphong, M. Growth of Human Keratinocytes and Fibroblasts on Bacterial Cellulose Film. Biotechnology Progress. 22 (2006): 1194-1199.

- Santos, R., Barbanti, H., Duek, E. and Wada, M. Analysis of the growth pattern of Vero cells cultured on dense and porous poly (L-Lactic Acid) scaffolds. Materials Research. 12(3) (2009): 257-263.
- Segal, L., Creely, J., Martin, A. and Conrad, M. An empirical method for estimating the degree of crystallinity of native cellulose using the X-ray diffractometer. Textile Research. 29 (1959): 786-794.
- Schramm, M., Grommet, Z. and Hestrin, S. Synthesis of Cellulose by *Acetobacter xylinum*. Substrates and Inhibitors Biochemical. 67 (1957): 669-679.
- Shi, S., Chen, S., Zhang, X., Shen, W., Li, X., Hu, W. And Wang, H. Biomimetic mineralization synthesis of calcium-deficient carbonate-containing hydroxyapatite in a three-dimensional network of bacterial cellulose. Chemical Technology & Biotechnology. 84(2) (2009): 285-290.
- Shibasaki, H., Saito, M., Kuga, S. and Okano, T. Native cellulose II production by *Acetobacter xylinum* under physical constraints. Cellulose. 5(1998): 165-173.
- Suwanmajo, T. Development of nanostructure membrane from regeneragerd bacterial cellulose. Master's thesis, Department of chemical engineering, Faculty of Engineering, Chulalongkorn University, 2006.
- Svensson, A., Nicklasson, E., Harrah, T., Panilaitis, B., Kaplan, D. L., Brittberg, M. and Gatenholm, P. Bacterial cellulose as a potential scaffold for tissue engineering of cartilage. Biomaterials. 26 (2005): 419-431.
- Tabata, Y. and Ikada, Y. Protein release from gelatin matrices. Advance Drug Delivery Review. 31(2) (1998): 287-301.

- Tanaka, A., Nagate, T. and Matsuda, H. The biological activity of epidermal growth factor (EGF) include in the gelatin film by cell proliferation assay. Veterinary Medical Science. 67(9) (2005): 909-913.
- Taitzoglou, I. A., Tsantarliotou, M., Zervos, I., Kouretas, D. and Kokolis, N. A. Inhibition of human and ovine acrosomal enzymes by tannic acid *in vitro*. Reproduction. 121 (2001): 131-137.
- Tientanacom, S. Drying characteristics and the effect of drying conditions on the properties of gelatin. Master's thesis, Department of chemical engineering, Faculty of Engineering, Chulalongkorn University, 1979.
- Tomihata, K., Burczak, K., Shiraki, K., Ikada, Y., Shalaby, S. W., Ikada. Y., Langer, R. S. and Williams, J. Polymers of biological and biomedical importance. American Chemical Society Symposium Series. American Chemical Society. 540 (1994): 275–286.
- Vemuri, S. A Screening Technique to Study the Mechanical Strength of Gelatin Formulations. Drug Development and Industrial Pharmacy. 26(10) (2000): 1115–1120.
- Wang, S., Chow, Y., Tan, D. W., Zhang, D. and Zhang, Y. Nanostructured and Transparent Polymer Membranes with Thermosensitivity for Wound Dressing and Cell Grafting. Advanced Materials. 16 (2004): 1790 – 1794.
- Wu, P., Nelson, A., Reid, H., Ruckley, V. and Gaylor, S. Water vapour transmission rates in burns and chronic leg ulcers: influence of wound dressings and comparison with *in vitro* evaluation. Biomaterials. 17 (1996): 1373-1377.

- Yamanaka, S. and Watanabe, K. Applications of Bacterial Cellulose, Cellulose Polymers, Blends and Composites. Cincinnati : Hanser / Gardner Publications, 1994: 207-215.
- Yan, Q and Bennick, A. Identification of histatins as tannin-binding proteins in human saliva. Biochemical. 311 (1995): 341-347.
- Yano, H., Sugiyama, J., Nakagaito, A. N., Nogi, M., Matsuura, T., Hikita, M. and Handa, K. Optically transparent composites reinforced with networks of bacterial nanofibers. Advanced Materials. 17(2) 2005: 153-155.
- Yasuda, K., Gong, J. P., Katsuyama, Y., Nakayama, A., Tanabe, Y, Kondo, E., Ueno, M. and Osada, Y. Biomechanical properties of high-toughness double network hydrogels. Biomaterials. 26 (2005): 4468-4475.

APPENDIX

APPENDIX

Table1 Data of Figure 5.7

Gelatin concentration (%w/v) in culture medium	Tensile strength for dried film (MPa)						
	1	2	3	4	5	Average	S.D.
0	52.65	65.86	59.28	64.86	72.45	63.02	7.45
1	57.82	57.67	52.55	57.90	57.72	56.73	2.34
3	62.82	47.81	52.89	57.63	40.12	52.25	8.77
5	32.71	32.65	30.16	30.13	29.31	30.99	1.58
7	33.66	29.23	32.23	28.59	32.13	31.17	2.16
10	27.46	25.06	28.16	34.50	31.49	29.33	3.69

Table2 Data of Figure 5.7

Gelatin concentration (%w/v) in culture medium	Tensile strength for reswollen film (MPa)						
	1	2	3	4	5	Average	S.D.
0	8.82	8.79	8.37	8.45	7.16	8.32	0.68
1	7.99	6.81	6.00	9.02	9.23	7.81	1.40
3	2.17	10.17	5.83	3.17	7.33	5.73	3.22
5	0.01	0.01	0.01	0.04	0.05	0.02	0.02
7	0.01	0.08	0.01	0.03	0.03	0.03	0.03
10	0.08	0.01	0.03	0.01	0.03	0.03	0.03

Table3 Data of Figure 5.8

Gelatin concentration (%w/v) in culture medium	Elongation at break for dried film (%)						
	1	2	3	4	5	Average	S.D.
0	1.27	1.15	1.28	1.59	1.66	1.39	0.22
1	1.04	1.55	1.57	1.11	1.34	1.32	0.24
3	1.49	0.87	0.94	1.75	1.38	1.29	0.37
5	0.60	0.78	0.68	0.79	0.97	0.76	0.14
7	0.93	0.62	1.00	0.65	0.65	0.77	0.18
10	1.59	1.01	0.72	1.02	0.85	1.04	0.33

Table4 Data of Figure 5.8

Gelatin concentration (%w/v) in culture medium	Elongation at break for reswollen film (%)						
	1	2	3	4	5	Average	S.D.
0	20.20	30.21	18.86	17.54	16.69	20.70	5.48
1	20.94	11.61	12.89	21.82	18.99	17.25	4.70
3	17.85	18.72	10.80	15.91	14.29	15.51	3.15
5	7.12	9.26	6.54	8.78	11.91	8.72	2.11
7	11.15	15.27	7.96	7.54	7.20	9.82	3.43
10	6.83	7.91	7.33	7.68	6.94	7.34	0.46

Table5 Data of Figure 5.9

Gelatin concentration (%w/v) in culture medium	Water absorption capacity (%)				
	1	2	3	Average	S.D.
0	618.81	616.49	606.24	613.85	6.69
1	647.92	656.13	655.68	653.24	4.61
3	772.27	745.66	769.66	762.53	14.67
5	708.05	723.27	732.48	721.27	12.34
7	669.36	661.65	697.97	676.33	19.13
10	709.69	700.16	688.94	699.59	10.39

Table6 Data of table 5.1

Gelatin concentration (%w/v) in culture medium	OTR (cc/m ² /day)			
	1	2	Average	S.D.
0	1.62	1.55	1.59	0.05
3	1.62	1.58	1.60	0.03
5	2.65	2.65	2.65	0.00
10	1.69	1.79	1.74	0.07

Table7 Data of Figure 5.10

Gelatin concentration (%w/v) in culture medium	WVTR (g/m ² /day)			
	1	2	Average	S.D.
0	1128	923	1026	144.96
3	683	500	592	129.40
5	859	911	885	36.77
10	674	668	671	4.24

Table8 Data of Figure 5.16

Gelatin concentration (%w/w) in impregnated solution	Tensile strength for dried film (MPa)						
	1	2	3	4	5	Average	S.D.
0	50.17	50.17	62.67	45.96	62.62	54.32	7.79
15	29.04	40.44	33.43	33.50	32.50	33.78	4.14
30	10.13	10.74	13.95	12.67	10.13	11.53	1.71

Table9 Data of Figure 5.16

Gelatin concentration (%w/w) in impregnated solution	Tensile strength for reswollen film (MPa)						
	1	2	3	4	5	Average	S.D.
0	11.19	9.99	10.77	10.15	9.21	10.26	0.76
15	0.38	0.52	0.31	0.49	0.45	0.43	0.09
30	0.49	0.51	0.13	0.28	0.48	0.38	0.17

Table10 Data of Figure 5.17

Gelatin concentration (%w/w) in impregnated solution	Elongation at break for dried film (%)						
	1	2	3	4	5	Average	S.D.
0	2.12	1.42	1.97	1.33	1.80	1.73	0.34
15	0.32	0.38	0.94	2.63	2.64	1.38	1.17
30	0.46	0.43	0.46	0.50	0.44	0.46	0.03

Table11 Data of Figure 5.17

Gelatin concentration (%w/w) in impregnated solution	Elongation at break for reswollen film (%)						
	1	2	3	4	5	Average	S.D.
0	24.18	22.67	24.56	25.14	24.18	24.15	0.91
15	22.22	21.51	21.16	21.04	21.67	21.52	0.47
30	19.08	15.37	16.94	16.66	18.14	17.24	1.42

Table12 Data of Figure 5.18

Gelatin concentration (%w/w) in impregnated solution	Water absorption capacity (%)				
	1	2	3	Average	S.D.
0	589.11	617.36	611.34	605.94	14.88
15	668.57	670.03	653.93	644.18	8.90
30	664.08	700.46	669.28	677.94	19.68

Table13 Data of table 5.2

Gelatin concentration (%w/w) in impregnated solution	OTR (cc/m ² /day)			
	1	2	Average	S.D.
0	2.44	2.89	2.67	0.32
15	1.68	1.44	1.56	0.17
30	1.66	1.64	1.65	0.01

Table14 Data of Figure 5.19

Gelatin concentration (%w/w) in impregnated solution	WVTR (g/m ² /day)			
	1	2	Average	S.D.
0	1328	1222	1275	75
15	1485	1498	1492	9
30	1540	1513	1527	19

Table15 Data of Figure 5.22

Absorbance					
Type of film		BC	Biosyn-BCG5	Biosyn-BCG10	Impreg-BCG15
0 hr	1	0.314	0.145	0.250	0.218
	2	0.324	0.163	0.251	0.134
	3	0.255	0.169	0.267	0.118
	Average	0.298	0.159	0.256	0.157
	SD	0.037	0.012	0.010	0.054
24 hr	1	0.368	0.248	0.392	0.127
	2	0.351	0.222	0.315	0.148
	3	0.408	0.235	0.318	0.167
	Average	0.376	0.235	0.342	0.147
	SD	0.029	0.013	0.044	0.020
48hr	1	0.523	0.236	0.558	0.128
	2	0.462	0.285	0.561	0.137
	3	0.459	0.34	0.649	0.153
	Average	0.481	0.287	0.589	0.139
	SD	0.036	0.052	0.052	0.013



VITAE

Miss Siriporn Taokaew was born on September 14th, 1985 in Phitsanulok, Thailand. She received the Bachelor Degree of Chemical Engineering from Faculty of Engineering, King Mongkut's University of technology Thonburi in April, 2008. She continued Master degree in chemical engineering at Chulalongkorn University in June, 2008.

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2. BACTERIAL CELLULOSE-GELATIN FILM FROM MICROBIAL SYNTHESIS BY *ACETOBACTER XYLINUM* in the 3rd Technology and Innovation for Sustainable Development International conference 2010 (TISD 2010) on 4-6 March, 2010 at Royal Mekong Nongkhai Hotel, Thailand, pp.1099-1102.