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APPENDIX

Table A1 List of selective primer combinations used in cDNA-AFLP analysis

E + AA (E1)	E + AT (E2)	E + AG (E3)	E + AC (E4)	E + TA (E5)	E + TT (E6)	E + TC (E7)
M + AA (M1)	M + AA (M1)	M + AA (M1)	M + AA (M1)	M + AA (M1)	M + AA (M1)	M + AA (M1)
M + AT (M2)	M + AT (M2)	M + AT (M2)	M + AT (M2)	M + AT (M2)	M + AT (M2)	M + AT (M2)
M + AC (M3)	M + AC (M3)	M + AC (M3)	M + AC (M3)	M + AC (M3)	M + AC (M3)	M + AC (M3)
M + AG (M4)	M + AG (M4)	M + AG (M4)	M + AG (M4)	M + AG (M4)	M + AG (M4)	M + AG (M4)
M + TA (M5)	M + TA (M5)	M + TA (M5)	M + TA (M5)	M + TA (M5)	M + TA (M5)	M + TA (M5)
M + TT (M6)	M + TT (M6)	M + TT (M6)	M + TT (M6)	M + TT (M6)	M + TT (M6)	M + TT (M6)
M + TC (M7)	M + TC (M7)	M + TC (M7)	M + TC (M7)	M + TC (M7)	M + TC (M7)	M + TC (M7)
M + TG (M8)	M + TG (M8)	M + TG (M8)	M + TG (M8)	M + TG (M8)	M + TG (M8)	M + TG (M8)
M + CA (M9)	M + CA (M9)	M + CA (M9)	M + CA (M9)	M + CA (M9)	M + CA (M9)	M + CA (M9)
M + CT (M10)	M + CT (M10)	M + CT (M10)	M + CT (M10)	M + CT (M10)	M + CT (M10)	M + CT (M10)
M + CC (M11)	M + CC (M11)	M + CC (M11)	M + CC (M11)	M + CC (M11)	M + CC (M11)	M + CC (M11)
M + CG (M12)	M + CG (M12)	M + CG (M12)	M + CG (M12)	M + CG (M12)	M + CG (M12)	M + CG (M12)
M + GA (M13)	M + GA (M13)	M + GA (M13)	M + GA (M13)	M + GA (M13)	M + GA (M13)	M + GA (M13)
M + GT (M14)	M + GT (M14)	M + GT (M14)	M + GT (M14)	M + GT (M14)	M + GT (M14)	M + GT (M14)
M + GC (M15)	M + GC (M15)	M + GC (M15)	M + GC (M15)	M + GC (M15)	M + GC (M15)	M + GC (M15)
M + GG (M16)	M + GG (M16)	M + GG (M16)	M + GG (M16)	M + GG (M16)	M + GG (M16)	M + GG (M16)
E + TG (E8)	E + GT (E10)	E + GC (E11)	E + GG (E12)	E + CT (E14)	E + CC (E15)	E + CG (E16)
M + AA (M1)	M + AA (M1)	M + AA (M1)	M + AA (M1)	M + AA (M1)	M + AA (M1)	M + AA (M1)
M + AT (M2)	M + AT (M2)	M + AT (M2)	M + AT (M2)	M + AT (M2)	M + AT (M2)	M + AT (M2)
M + AC (M3)	M + AC (M3)	M + AC (M3)	M + AC (M3)	M + AC (M3)	M + AC (M3)	M + AC (M3)
M + AG (M4)	M + AG (M4)	M + AG (M4)	M + AG (M4)	M + AG (M4)	M + AG (M4)	M + AG (M4)
M + TA (M5)	M + TA (M5)	M + TA (M5)	M + TA (M5)	M + TA (M5)	M + TA (M5)	M + TA (M5)
M + TT (M6)	M + TT (M6)	M + TT (M6)	M + TT (M6)	M + TT (M6)	M + TT (M6)	M + TT (M6)
M + TC (M7)	M + TC (M7)	M + TC (M7)	M + TC (M7)	M + TC (M7)	M + TC (M7)	
M + TG (M8)	M + TG (M8)	M + TG (M8)	M + TG (M8)	M + TG (M8)	M + TG (M8)	
M + CA (M9)	M + CA (M9)	M + CA (M9)	M + CA (M9)	M + CA (M9)	M + CA (M9)	
M + CT (M10)	M + CT (M10)	M + CT (M10)	M + CT (M10)	M + CT (M10)	M + CT (M10)	
M + CC (M11)	M + CC (M11)	M + CC (M11)	M + CC (M11)	M + CC (M11)	M + CC (M11)	
M + CG (M12)	M + CG (M12)	M + CG (M12)	M + CG (M12)	M + CG (M12)	M + CG (M12)	
M + GA (M13)	M + GA (M13)	M + GA (M13)	M + GA (M13)	M + GA (M13)	M + GA (M13)	
M + GT (M14)	M + GT (M14)	M + GT (M14)	M + GT (M14)	M + GT (M14)	M + GT (M14)	
M + GC (M15)	M + GC (M15)	M + GC (M15)	M + GC (M15)	M + GC (M15)	M + GC (M15)	
M + GG (M16)	M + GG (M16)	M + GG (M16)	M + GG (M16)	M + GG (M16)	M + GG (M16)	

The primers were *Eco*RI (E0) and *Mse*I (M0) pre-selective primers which two selective nucleotides were added to the 3' end.

Table A2 Genes, primer sequences, annealing temperature (T_m), and number of PCR cycles used in semi-quantitative reverse transcription PCR

Gene	Primers	Sequences (5'⟶3')	Tm (°C)	PCR cycles
Os03g0661900	661900F	TCTGGGAGTCTTGTCGGTTC	62	25
	661900R	CATGGCAGTGAGGCACTCTA		
Os06g0646400	646400F	TACACCACCCAAATGTCGTG	59	25
	646400R	GCATATTGGACGCTGAGGAT		
Os10g0518200	518200F	TTTTATGGATGAGCACCCTGG	59	28
	518200R	GGTGTACTGCCGGATAGCAA		
Os02g0175700	175700F	GAGGAGCACCCAACCCTTTC	67	32
	175700R	TTGAGAGACAGCCATTGCCA		
Os06g0478600	478600F	CTCAGGCTGCTGTCCATGTC	67	25
	478600R	GGACAGAAGAAGGGACGACG		
Os02g0759700	759700F	CAGGTTTCCGGGATGTCAAT	60	28
	759700R	TGCGCTGGCAAAAGTAGAGA		
Os07g0627300	627300F	GGCCAAGGATGACGAGCTAT	59	32
	627300R	CGTCCATGCTTTGGCTTGTA		
OS07g0240300	240300F	GATGGGGATAATGGTGGTGG	66	30
	240300R	AACACGCCCAAGAAGACCTG		
Os07g0645100	645100F	AGGGAACGCGAGGACATCTA	62	25
	645100R	TGAAAAGCAAGCTCGGCATA		
Os02g0560450	560450F	GCTGCCACCGACGACATC	65	28
	560450R	TCTTGTCGGAAACAGCGATCT		
Os08g0179900	179900F	ATCACCACCAAAACACCCCT	65	30
	179900R	AACTCCCAGTCGTCCACCTC		
OS05g0392100	392100F	ACCATGGAAGATGGGTGTGC	59	32
	392100R	CCATCATCAGTTGCGTCACA		
Os03g0569000	569000F	CTTGTCCTCGCGATTCATCA	65	30
	569000R	TGTGCATTGCCTTCTTGACA		
Os12g0624700	624700F	CGAGATACGCCTACACCTGG	67	28
	624700R	CGTTGCTGGTTGAAGACCAA		
Os11g0210500[a]	210500F	ATCAAGGGGAAGCCCATCTT	63	30
	210500R	GTCCACCGTTGGTCATCTCA		
Os05g0536400[a]	536400F	CACACCGTTCAATTTCCCAT	55	25
	536400R	TTTCCAGTCAAAACCAGCATC		
act1 (actin)	actin1F	GAGGCTCCTCTCAACCCCAA	55	25
	actin1R	GTGAGATCACGCCCAGCAAG		

[a]Genes involved in the ethanolic fermentation pathway in rice (Lasanthi-Kudahettige et al., 2007)

VITA

Patipanee Khanthapok received a Bachelor of Science Degree (Biotechnology) from the Faculty of Engineering and Industrial Technology, Silpakorn University in 2003.

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Research publications

1. Submitted manuscripts

1.1 Khanthapok P, Sukrong S, Muangprom A. Identification of ethanol-inducible genes and isolation of the Myb-related protein-like promoter in Oryza sativa L. Journal: Planta, Date of submission: July 8, 2014.

1.2 Khanthapok P, Muangprom A, Sukrong S. Antioxidant activity and DNA protective properties of rice grass juices. Journal: ScienceAsia, Date of submission: June 30, 2014.

2. Conferences

2.1 Khanthapok P, Sukrong S, Muangprom A. Effects of ethanol on growth of rice plants and identification of genes incuced by ethanol, The First National Rice Research Conference "Moving Rice Research Towards Innovation", December 15-17, 2010 Kasetsart University Bangken Campus Bangkok, Thailand.

2.2 Pongsamart, S., Sukrong, S., Khanthapok, P., and Bhusawang, P. Combined molecular and chemical assessments of different cultivars of Tamarindus indica in Thailand, WOCMAP IV: World Conference on Medicinal and Aromatic Plants, November 9-14, 2008. Cape Town, South Africa.

2.3 Pongsamart S, Sukrong S, Khanthapok P, Bhusawang P. DNA sequence and chemical assessment of thai tamarind cultivars with laxative activity, 7th Joint Meeting of AFERP, ASP, GA, PSE & SIF: Natural Products with Pharmaceutical, Nutraceutical, Cosmetic and Agrochemical Interest, August 3-8, 2008. Athens, Greece.

2.4 Chaipornpokin W, Khanthapok P, Pongsamart S. Determination of organic acids in fresh pulps of Thai tamarind cultivars by HPLC/UV, PBP World Meeting: 6th World Meeting on Pharmaceutics, Biopharmaceutics and Pharmaceutical Technology, April 7-10, 2008. Barcelona, Spain.

2.5 Khanthapok P, Bhusawang, P, Pongsamart,S. Analysis of organic acids in certain Tamarindus indica by reversed-phase high-performance liquid chromatography (RP-HPLC), International Workshop on Medicinal and Aromatic Plants, January 15-18, 2007. Chiang Mai, Thailand.

3. Others

3.1 Sukrong S, Khanthapok P, Pongsamart S. 2008. Tamarindus indica chloroplast rbcL gene partial sequences for ribulose 1,5-bisphosphate carboxylase/oxygenase large subunit: GenBank accession numbers AB378725, AB378726, AB378727, AB378728, AB378729, AB378730, AB378731, and AB378732. GenBank, National Center for Biotechnology Information Madison, United States of America.

3.2 Pongsamart S, Sukrong S, Khanthapok P, Bhusawang P. 2008. DNA Sequence and Chemical Assessment of Thai Tamarind Cultivars with Laxative Activity. Planta Med; 74(9): 1207 (Abstracts).