CHAPTER V

SUMMARY

The results of the present investigation can be summarized as follows:

- 1. The ¹H-NMR is a suitable method for the determination of glycine betaine in A. halophytica.
- 2. The amount of glycine betaine in A. halophytica grown in non salt-stressed condition (0.5 M NaCl) was 9.7 nmol/10⁶ cells. The glycine betaine accumulation was increased 8-fold in salt-stressed condition (2 M NaCl).
- 3. The A. halophytica BADH was partially purified by ammonium sulfate fractionation and DEAE-cellulose column chromatography. The BADH purification was 18-fold with a specific activity of 290.8 μmol min⁻¹ mg⁻¹ and a recovery of 8.6%. K_m values for betaine aldehyde and NAD⁺ were 91 and 71.4 μM, respectively. NADP⁺ was a less preferred coenzyme (K_m= 100 μM). The V_{max} of BADH was 175.4 μmol min⁻¹ mg⁻¹.
- 4. The optimal condition for A. halophytica BADH was pH 7.5 at 25°C. The enzyme was inhibited by substrate analogs such as ethanolamine and acetaldehyde. Aldehyde analog appeared to be a more potent inhibitor than N-methylated analogs.

- 5. The enzyme activity was initially stimulated by increasing concentration of NaCl and KCl from 0 to 0.1 M above which the BADH activity was decreased. CaCl₂ and MgCl₂ appeared to have a particularly strong inhibition on BADH.
- 6. A.halophytica BADH activity was strongly inhibited by sulfhydryl-reactive compound such as PCMS but incubating the enzyme with reducing agent such as DTT before or after PCMS treatment was able to relieve the inhibition by PCMS.
- 7. The molecular weight of A. halophytica BADH was 120,000 dalton. The holoenzyme was most likely a tetramer of 30,000 dalton subunits.
- 8. The high external salinity resulted in the increased specific activity of A. halophytica BADH.