CHAPTER 4

RESULTS

4.1 ISOLATION, SCREENING AND SELECTION OF CHROMIUM-RESISTANT BACTERIA, PHENOL-RESISTANT BACTERIA AND CHROMIUM/PHENOL-RESISTANT BACTERIA

4.1.1 CHROMIUM, PHENOL AND CHROMIUM/PHENOL-RESISTANT BACTERIAL ISOLATES

In four hundreds and ninety-five strains bacterial isolate, one hundred and fifty bacterial strains resisted to 600 µg/ml Cr in ½ strength NA medium were isolated. Resistances to 800, 1200, 1600, 2000 and 2400 µg/ml Cr were found in 58, 24, 11.3, 4.7 and 2 percent; %, respectively. For phenol-resistant bacterial isolates, two hundred and twenty-five bacterial strains resisted to 600 μ g/ml phenol; Ph, in $\frac{1}{2}$ strength NA medium were isolated. Resistances to 800, 1200, 1600 and 2000 µg/ml Ph were found in 48.5, 32.9, 17.3 and 1.3 %, respectively. None of them was resistant to 2400 µg/ml Ph. And chromium/phenolresistant bacterial isolates, one hundred and twenty bacterial strains resisted to 300 µg/ml Cr and 300 µg/ml Ph; CP in ¹/₂ strength NA medium were isolated. Resistances to 400, 600, 800, 1000 and 1200 µg/ml CP were found in 37.5, 30, 19.2, 10.8 and 2.5 %, respectively, all of these resistant bacterial isolates as shown in Table 4.1, on page 65. Three Crresistant isolates, three Ph-resistant isolates and three CP-resistant isolates were selected, namely: CrR-2; CrR-14; CrR-15, PhR-26; PhR-33; PhR-64 and CPR-4; CPR-16; CPR-17. From Table 4.2 on page 66-67, the first

three bacterial strains were rod shape, gram-negative and identified as *Escherichia* sp., *Pseudomonas* sp. and *Enterobacter* sp., respectively, and they were isolated from soil collected from oil refinery and chromium plating. The second three bacterial strains were rod shape, gram-negative and identified as *Klebsiella* sp., *Pseudomonas* sp. and *Escherichia* sp., respectively, and they were isolated from soil collected from soil collected from soil collected from soil collected from wood preserving, painting and chemical production. The last three bacterial strains were also rod shape, gram-negative and identified as *Pseudomonas* sp., *Proteus* sp. and *Escherichia* sp., respectively, and they were isolated from soil collected from the section.

The colonial and cell characteristics of the nine selected bacterial isolates are shown in Figure 4.1-4.9, on page 68-76.

4.1.2 STABILITY OF BACTERIAL RESISTANCE

The results of the stability of Cr resistance, Ph resistance and CP resistance of the nine selected bacterial isolates are briefly summarized in **Table 4.3**, on page 77. Cr-resistant and CP-resistant bacterial isolates were able to maintain the highest Cr and CP resistance, 2400 and 1200 μ g/ml, respectively, after, at least 18 times of subculturing in NA containing small amount of Cr and CP. But Ph-resistant bacterial isolates were able to maintain the highest Ph resistance (2000 μ g/ml) after, at least 15 times of subculturing in NA containing small amount of Ph.

Sets of Experiment	Concentration (µg/ml)	No. of Strains	%				
	800	87	58.0				
	1200	36	24.0				
Chromium	1600	17	11.3				
	2000	7	4.7				
	2400	3	2.0				
Su	b Total	150	100				
	800	109	48.5				
	1200	74	32.9				
Phenol	1600	39	17.3				
	2000	3	1.3				
	2400	0	0				
Su	ıb Total	225	100				
	400	45	37.5				
Chromium/Phenol	600	36	30.0				
	800	23	19.2				
	1000	13	10.8				
	1200	3	2.5				
Su	120	100					
Gra	Grand Total						

Table 4.1Three sets of experiment about resistance in 495 strainsbacterial isolates

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Table 4.2Some characteristics and identification on nine strains of the
chromium-resistant, phenol-resistant and chromium/phenol-
resistant bacterial isolates

Bacterial	Sources	Charac	teristic of	Identified as
Isolate s	(Sampling Site)	Colony	Morphology	
CrR-2	Soil, (Oil	~3 mm in	Rod-shape,	Escherichia sp.
•	refinery, area VI,	diameter,	gram-	
	Samutprakarn)	white	negative,	
CrR-14	Soil, (Chromium	~2 mm in	Rod-shape,	Pseudomonas sp.
	plating, area I,	diameter,	gram-	
	Bangkok)	white	negative,	
CrR-15	Soil, (Chromium	~3 mm in	Rod-shape,	Enterobacter sp.
	plating, area I,	diameter,	gram-	
	Bangkok)	white	negative,	
PhR-26	Soil, (Wood,	~2 mm in	Rod-shape,	Klebsiella sp.
	area I, Bangkok)	diameter,	gram-	
		white	negative,	
PhR-33	Soil, (Painting,	~3 mm in	Rod-shape,	Pseudomonas sp.
	area II, Bangkok)	diameter,	gram-	
~	800.054	white	negative,	
PhR-64	Soil, (Chemical	~3 mm in	Rod-shape,	Escherichia sp.
	Production	diameter,	gram-	
	Industry, area V,	white	negative,	
• 	Bangkok)			

Table 4.2 (Cont.)

Bacterial	Sources	Charac	teristic of	Identified as
Isolates	(Sampling Site)	Colony Morphology		
CPR-4	Soil, (Oil	~3 mm in	Rod-shape,	Pseudomonas sp.
	refinery, area VI,	diameter,	gram-	
	Samutprakarn)	white	negative,	
CPR-16	Soil, (Metal	~2 mm in	Rod-shape,	Proteus sp.
	Plating, area V,	diameter,	gram-	
	Bangkok)	white	negative,	
CPR-17	Soil, (Metal	~2 mm in	Rod-shape,	Escherichia sp.
	Plating, area V,	diameter,	gram-	
	Bangkok)	white	negative,	

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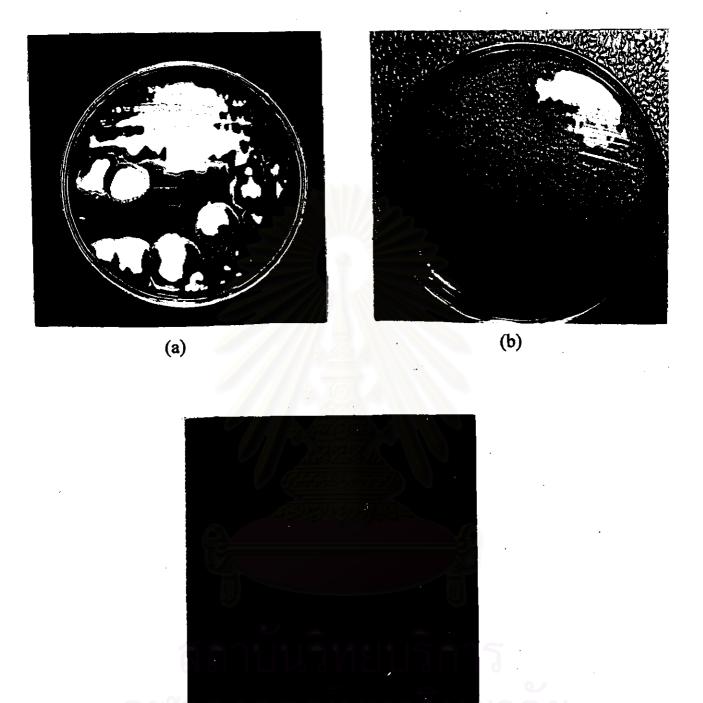


Figure 4.1 Colonial characteristics of chromium-resistant bacterial strains CrR-2 (*Escherichia* sp.) grown on NA (a) and NA containing 2400 μg/ml chromium (b), incubated at 37°C for 24 hr., and gram staining (c)

(c)

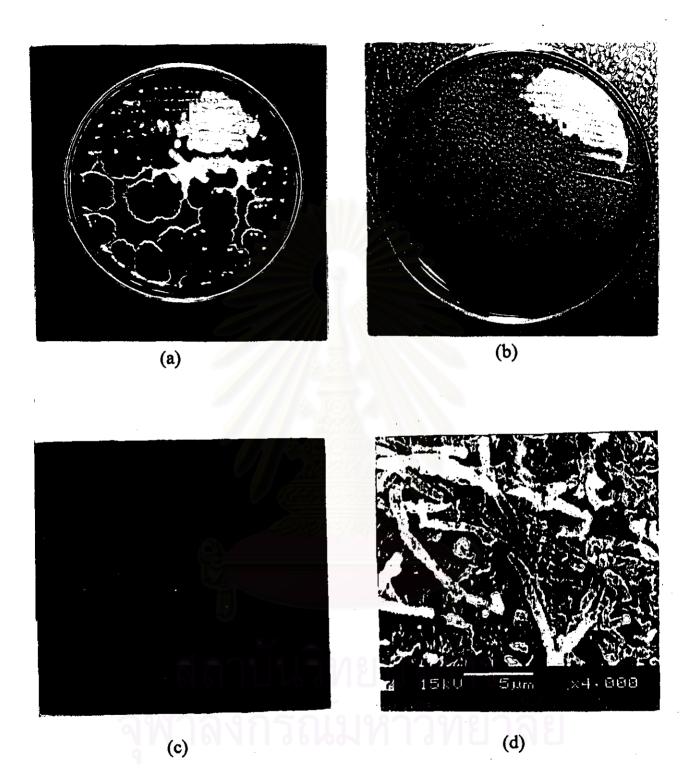
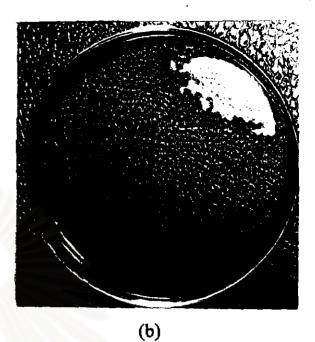


Figure 4.2 Colonial characteristics of chromium-resistant bacterial strains CrR-14 (*Pseudomonas* sp.) grown on NA (a) and NA containing 2400 μg/ml chromium (b), incubated at 37°C for 24 hr., gram staining (c), and Electron Microscope (d)





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Figure 4.3 Colonial characteristics of chromium-resistant bacterial strains CrR-15 (*Enterobacter* sp.) grown on NA (a) and NA containing 2400 μg/ml chromium (b), incubated at 37°C for 24 hr., and gram staining (c)

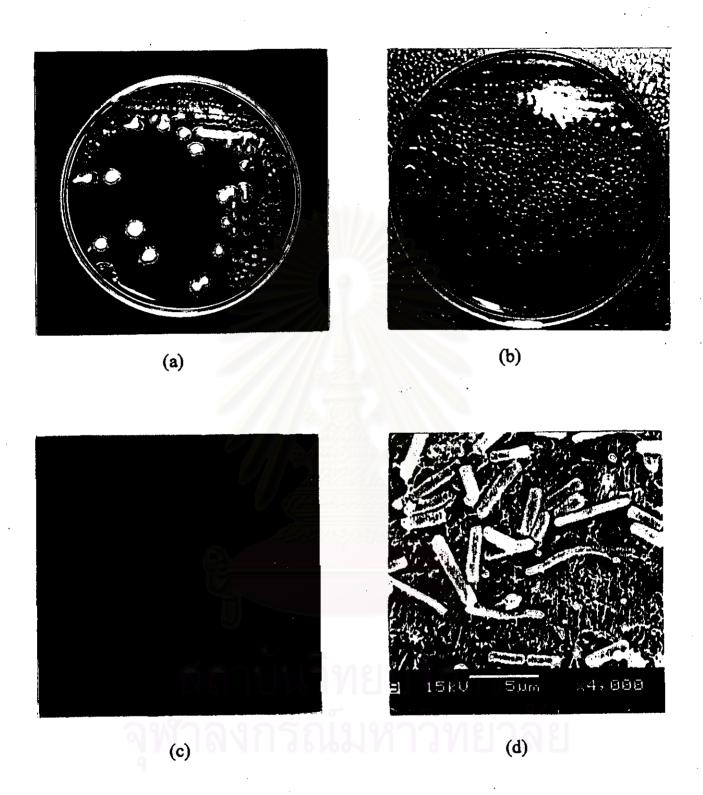
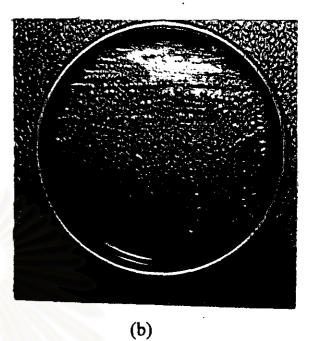


Figure 4.4Colonial characteristics of phenol-resistant bacterial strainsPhR-26 (Klebsiella sp.) grown on NA (a) and NA containing2000 μg/ml phenol (b), incubated at 37°C for 24 hr., gramstaining (c), and Electron Microscope (d)





(a)

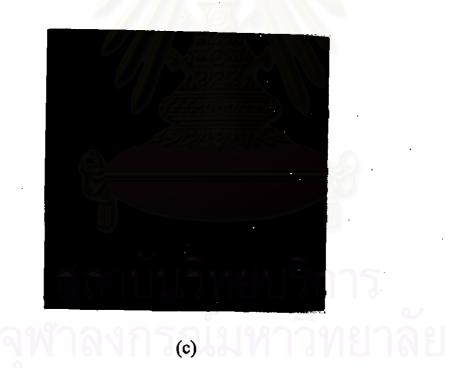
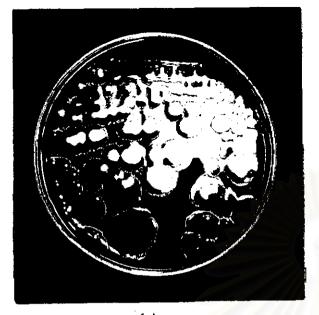
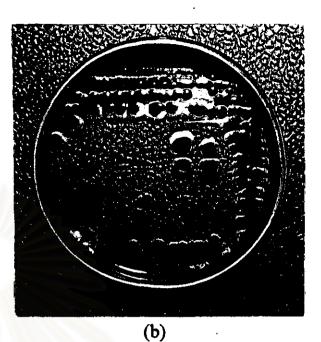


Figure 4.5 Colonial characteristics of phenol-resistant bacterial strains PhR-33 (*Pseudomonas* sp.) grown on NA (a) and NA containing 2000 μg/ml phenol (b), incubated at 37°C for 24 hr., and gram staining (c)





(a)

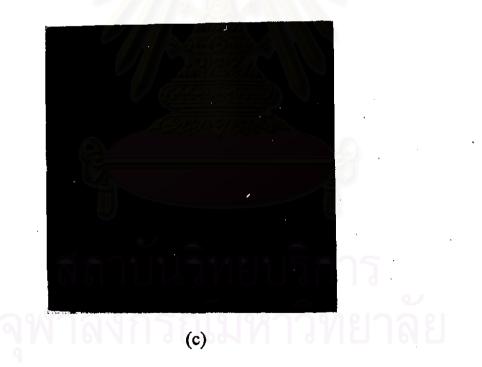
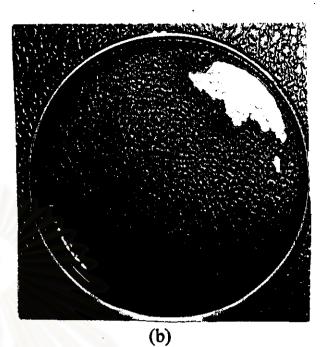


Figure 4.6 Colonial characteristics of phenol-resistant bacterial strains
PhR-64 (*Escherichia* sp.) grown on NA (a) and NA
containing 2000 µg/ml phenol (b), incubated at 37°C for
24 hr., and gram staining (c)





(a)

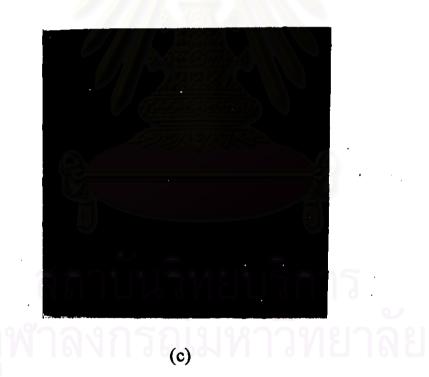


Figure 4.7 Colonial characteristics of chromium/phenol-resistant bacterial strains CPR-4 (*Pseudomonas* sp.) grown on NA (a) and NA containing 1200 μg/ml chromium-phenol (b), incubated at 37°C for 24 hr., and gram staining (c)

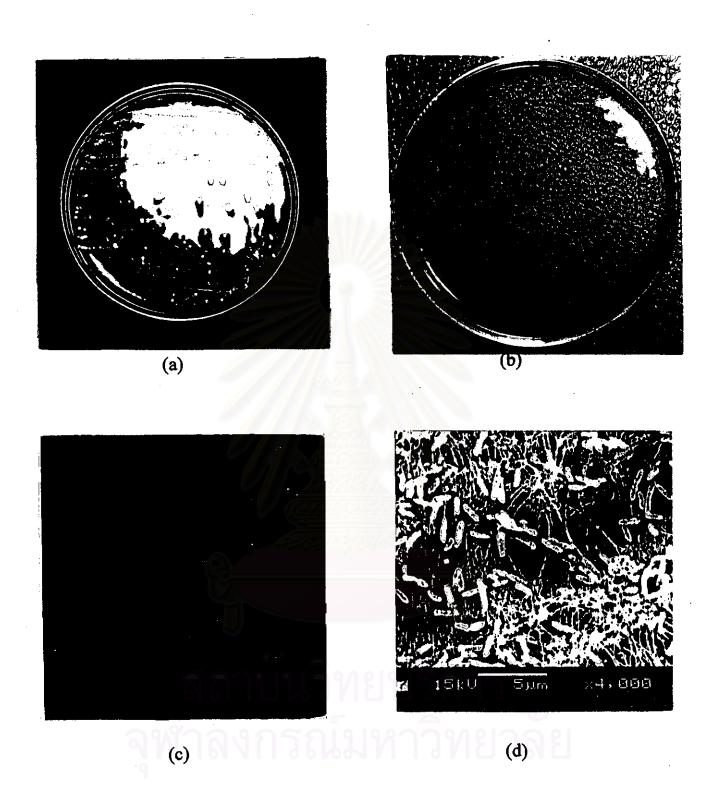


Figure 4.8 Colonial characteristics of chromium/phenol-resistant bacterial strains CPR-16 (*Proteus* sp.) grown on NA (a) and NA containing 1200 μ g/ml chromium-phenol (b), incubated at 37°C for 24 hr., gram staining (c), and Electron Microscope (d)



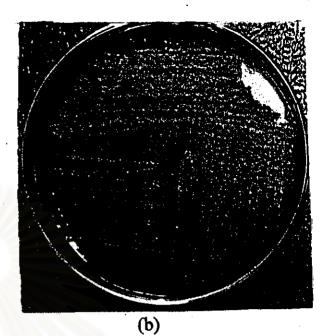


Figure 4.9 Colonial characteristics of chromium/phenol-resistant bacterial strains CPR-17 (*Escherichia* sp.) grown on NA (a) and NA containing 1200 μg/ml chromium-phenol (b), incubated at 37°C for 24 hr., and gram staining (c) Table 4.3Stability of the resistance to chromium or phenol or
chromium/phenol of nine selected bacterial strains after
repeated culturing

Bac	cterial Isolates	Resistant to	Stability of
Strains	Identified as	Concentration (µg/ml)	Resistance
CrR-2	Escherichia sp.	2400	18
CrR-14	Pseudomonas sp.	2400	18
CrR-15	Enterobacter sp.	2400	18
PhR-26	Klebsiella sp.	2000	15
PhR-33	Pseudomonas sp.	2000	15
PhR-64	Escherichia sp.	2000	15
CPR-4	Pseudomonas sp.	1200	18
CPR-16	Proteus sp.	1200	18
CPR-17	Escherichia sp.	1200	18

After at least 20 times of repeated subculturing

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4.1.3 RESISTANCE OF THE SELECTED BACTERIAL STRAINS TO OTHER HEAVY METALS

All of nine strains of the Cr-resistant, Ph-resistant and CPresistant bacterial isolates were found to be resistant to a number of other heavy metals, i.e., As and Zn, but none of them were able to resist to Ag, Cd, Cu, Ni and Mn; detailed results were summarized in **Table 4.4**, on page 79. The bacterial strains CrR-2, CrR-14 and CrR-15 were resistant to the same resistance levels of Cr, Ph, As and Zn, i. e., 2400, 1000, 200 and 100 μ g/ml, respectively; strains, PhR-26, PhR-33 and PhR-64 resisted Ph, Cr, As and Zn at the same resistance levels, i. e., 2000, 500, 200 and 100 μ g/ml, respectively. The bacterial strains, CPR-4, CPR-16 and CPR-17 were resisted Cr, Ph, As and Zn, i. e., 1200, 1200, 200 and 100 μ g/ml.

4.1.4 EFFECTS OF pH AND TEMPERATURE ON VIABLE COUNTS OF THE SELECTED BACTERIAL STRAINS

The optimum pH and temperature of those selected bacterial isolates were found to be 7 and 37°C, respectively (summarized in **Table 4.5**, on page 80). Most pH level 10, the reduction on number of viable cells was higher than at the pH level 4. It may imply that the effect of alkaline condition on growth was stronger than acidic condition, but the effect of temperature at higher (40°C) or lower (30°C) levels was not as strong as pH levels.

In bacterial isolates, the effects of pH and temperature were shown in Figure 4.10 and Figure 4.11, on page 81 and 82.

Strains		Resista	ant Con	centrati	on of H	leavy]	Metal (µg/ml)	
	Cr	Ph	As	Zn	Ag	Cd	Cu	Ni	Mn
CrR-2	2400	1000	200	100	-	-	-		
CrR-14	2400	1000	200	1.00	-	-	-	-	_
CrR-15	2400	1000	200	100	-	-	-	-	-
PhR-26	500	2000	200	100	-	-	-		
PhR-33	500	2000	200	100	-	-	-	-	-
PhR-64	500	2000	200	100	-	-	-	-	_
CPR-4	1200	1200	200	100	-	-			
CPR-16	1200	1200	200	100	4 -	-	-	-	-
CPR-17	1200	1200	200	100	-	-	-	-	-

 Table 4.4
 Resistance of the selected bacterial strains to other heavy metals



	Initial no. of				Viable	Count	(x10 ⁸ ce	lls/ml)			
Strains	Organism			. Temp (°C)							
	(x10 ^s cells/ml)	4	5	6	7	8	9	10	30	37	40
CrR-2	1.12	1.09	1.12	1.14	1.15	1.13	1.09	1.08	1.13	1.17	1.15
CrR-14	1.16	1.02	1.16	1.21	1.24	1.22	1.18	1.17	1.17	1.25	1.20
CrR-15	1.15	1.12	1.16	1.17	1.20	1.17	1.14	1.09	1.15	1.18	1.17
PhR-26	1.14	1.02	1.09	1.21	1.25	1.24	1.15	1.14	1.15	1.17	1.16
PhR-33	1.14	1.01	1.15	1.20	1.25	1.22	1.18	1.17	1.16	1.19	1.18
PhR-64	1.14	1.02	1.13	1.20	1.26	1.25	1.19	1.16	1.17	1.20	1.18
CPR-4	1.15	1.03	1.14	1.21	1.23	1.22	1.20	1.15	1.16	1.21	1.17
CPR-16	1.12	1.02	1.13	1.15	1.22	1.21	1.20	1.18	1.18	1.21	1.20
CPR-17	1.13	1.02	1.07	1.17	1.21	1.19	1.17	1.15	1.15	1.24	1.18

Table 4.5Effect of pH and temperature on growth of the selectedbacterial strains

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→ CrR-2	—■— CrR-14	→ CrR-15
→ PhR-26		PhR-64
-+- CPR-4	CPR-16	CPR-17

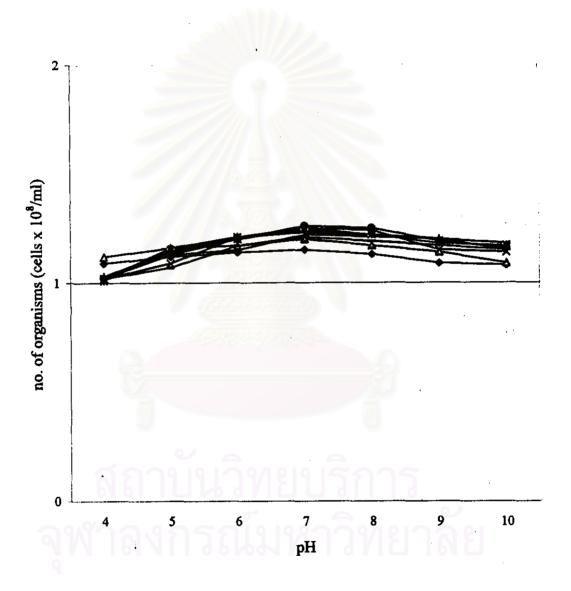


Figure 4.10 Effect of pH on growth of the bacterial strains

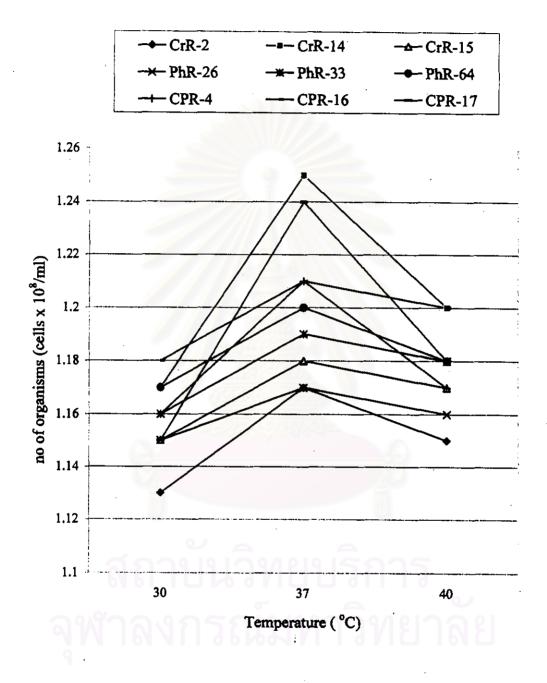


Figure 4.11 Effect of temperature on growth of the bacterial strains

4.2 EFFECTS OF PHENOL ON GROWTH RATE OF THE SELECTED BACTERIAL STRAINS

The results on growth of the Cr-resistant, Ph-resistant and CPresistant bacterial strains; CrR-15, PhR-26, PhR-33, PhR-64 and CPR-16, respectively, were shown in **Table 4.6**, on page 84. Comparing between growth on 0.85% normal saline and phosphate buffer, as control, with 0.85% normal saline and phosphate buffer containing 300 μ g/ml, as testing. It was found that growth of all test bacterial isolates in the medium containing 300 μ g/ml (testing) was better than medium (control), was shown in **Figure 4.12**, on page 85. It is possible to say that, in the condition containing phenol, the selected bacterial strains in this study seems to use phenol as carbon and energy source.

4.3 EFFICIENCY OF CHROMIUM DETOXIFICATION AND PHENOL DEGRADATION BY THE SELECTED BACTERIAL STRAINS

4.3.1 INCUBATION PERIODS AND CONTACT TIME

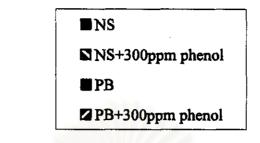
Incubation periods and contact time for growth on chromium detoxification and phenol degradation of the bacterial isolate; CPR-16 were found to be 6-hr. and 15 min.; summarized briefly in **Table 4.7**, on page 86. The results indicated that the equilibrium was taken within 6-hr., 15 min. and prolonged exposure time did not increase Cr(VI) detoxification, Cr(III) production and phenol degradation from the solution. Efficiency of Cr(VI) detoxification, Cr(III) production and phenol degradation were 75.3, 0.3 and 92.7%, respectively., as shown in **Figure 4.13** and **Figure 4.14**, on page 87 and 88.

Table 4.6Effects of phenol on growth rate of the selected bacterialstrains

	Viable Count (x10 ⁸ cells/ml)									
	0.85%	Normal Saline	Phos	phate Buffer						
Strains	Control	NS + 300 µg/ml	Control	$PB + 300 \mu g/ml$						
		Phenol	<u> </u>	Phenol						
CrR-15	0.15	3.05	0.35	9.33						
PhR-26	0.13	3.00	0.06	2.73						
PhR-33	0.08	2.67	0.13	4.83						
PhR-64	0.33	7.00	0.11	4.33						
CPR-16	0.13	3.67	0.11	2.53						

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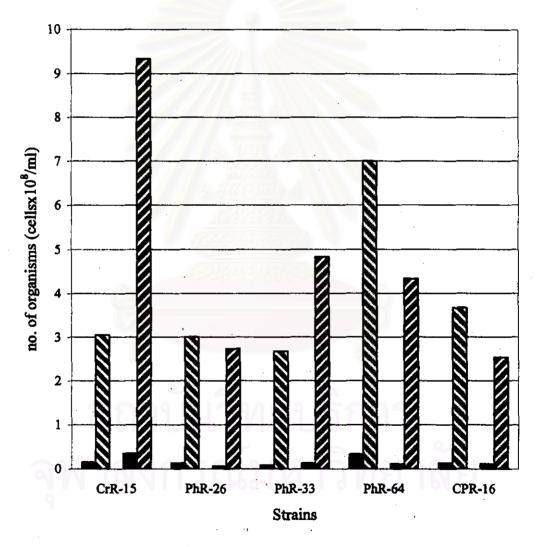


Figure 4.12 Effects of phenol on growth rate of the selected bacterial strains

Incubation	Contact Time (min.)												
Period		15			30			45			60		
(hr.)	Cr(VI) ¹	Cr(III) ²	Ph ³	Cr(VI) ¹	Cr(III) ²	Ph ³	Cr(VI) ¹	Cr(III) ²	Ph ³	Cr(VI) ¹	Cr(III) ²	Ph ³	
	(%)	(%)	(%)	(%)	(%)	(%)	(%)	(%)	(%)	(%)	(%)	(%)	
6	22 6± 0.7	1±0.0	278±0.8	227±0.8	1±0.0	277±0.8	230±0.7	2±0.1	274±0.8	226±0.7	3±0.1	282±0.8	
	(75.3)	(0.3)	(92.7)	(75.7)	(0.3)	(92.3)	(76.7)	(0.7)	(91.3)	(75.3)	(1.0)	(94.0)	
12	231±0.6	5±0.1	252±0.8	230±0.7	3±0.1	258±0.8	228±0.7	3±0.1	277±0.8	227±0.8	5±0.1	276±0.8	
	(77.0)	(1.7)	(84.0)	(76.7)	(1.0)	(86.0)	(76.0)	(1.0)	(92.3)	(75.7)	(1.7)	(92.0)	
24	24 6± 0.8	11±0.3	262±0.8	240±0.7	8±0.2	265±0.8	239±0.7	6±0.2	224±0.7	239±0.7	6±0.2	223±0.7	
	(82.0)	(3.7)	(87.3)	(80.0)	(2.7)	(88.3)	(81.0)	(2.0)	(74.7)	(79.7)	(2.0)	(74.3)	
48	242±0.8	12±0.4	22 6± 0.7	245±0.8	18±0.5	261±0.8	246±0.8	12±0.4	251±0.8	246±0.8	12±0.4	248±0.7	
	(80.7)	(4.0)	(75.3)	(81.7)	(6.0)	(87.0)	(81.3)	(4.0)	(83.7)	(82.0)	(4.0)	(82.7)	

 Table 4.7
 Effect of incubation periods of CPR-16 on contact times

¹ Loss of Cr(VI) Concentration, µg/ml (%)

² Found of Cr(III) Concentration, µg/ml (%)

³ Loss of Phenol Concentration, µg/ml (%)

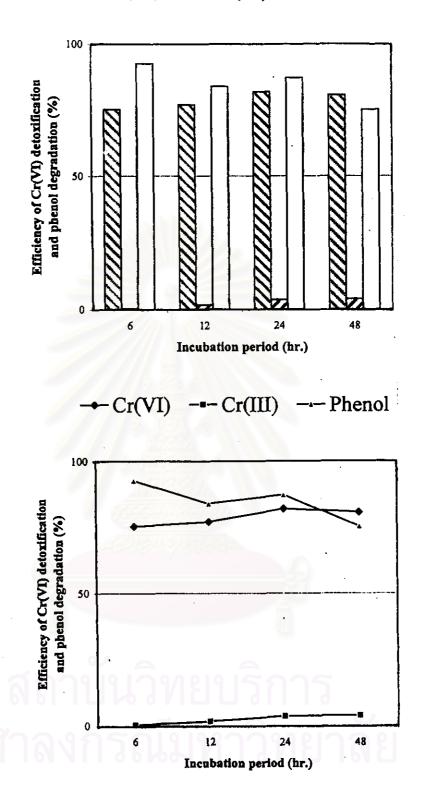


Figure 4.13 Efficiency of Cr(VI) detoxification and phenol degradation at contact time 15 min. varied from incubation period; 6, 12, 24 and 48 hr. by CPR-16

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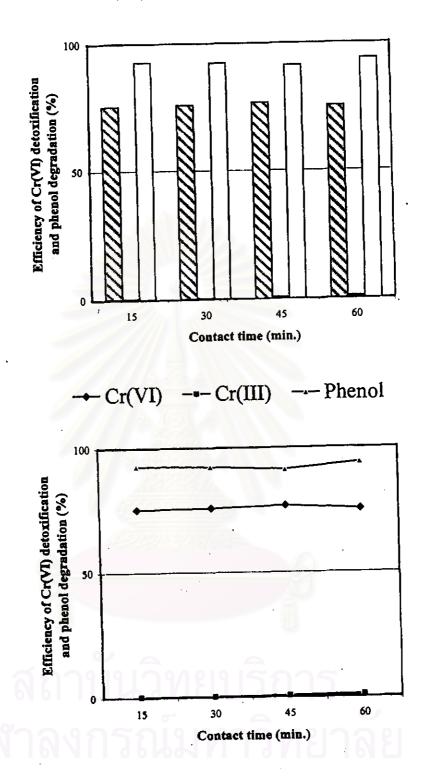


Figure 4.14 Efficiency of Cr(VI) detoxification and phenol degradation at incubation period 6 hr. varied from contact time; 15, 30, 45 and 60 min. by CPR-16

4.3.2 EFFECT OF LOW CONCENTRATIONS OF CHROMIUM DETOXIFICATION AND PHENOL DEGRADATION ON NINE COCULTURES AND THREE SINGLE CULTURES

Efficiency of bacterial isolates, nine cocultures; CrR-2+PhR-26, CrR-2+PhR-33, CrR-2+PhR-64, CrR-14+PhR-26, CrR-14+PhR-33, CrR-14+PhR-64, CrR-15+PhR-26, CrR-15+PhR-33 CrR-15+PhR-64 and three single cultures; CPR-4, CPR-16, CPR-17 at various concentrations; 100, 200, 300 and 400 μ g/ml, summarized in **Table 4.8**, on page 90 were shown in **Figure 4.15**, on page 91.

4.3.3 EFFECT OF HIGH CONCENTRATIONS OF CHROMIUM DETOXIFICATION AND PHENOL DEGRADATION ON THREE COCULTURES AND THREE SINGLE CULTURES

The results of Cr(VI) detoxification, Cr(III) production and phenol degradation by bacterial isolate are briefly summarized in **Table** 4.9, on page 92. The Efficiency of bacterial isolates, three cocultures; CrR-2+PhR-26, CrR-14+PhR-33, CrR-15+PhR-64 and three single cultures; CPR-4, CPR-16, CPR-17 at various concentrations; 500, 1000, 1500 and 2000 μ g/ml were shown in **Figure 4.16**, on page 93.

	Cr(VI) and phenol concentration												
Strains		100 µg/ml (%))		200 µg/ml (%)			300 µg/ml (%))	<u> </u>	400 µg/mi (%)	<u> </u>	
	Cr(VI) ¹	Cr(III) ²	Ph	Cr(VI) ¹	Cr(III) ²	Pb ³	Cr(VI) ¹	Cr(III) ²	Ph ³	Cr(VI) ¹	Ст(Ш) ²	Ph ³	
CrR-2+	86±0.3	3±0.1	92±0.3	166±0.5	1±0.0	188±0.6	241±0.7	17±0.5	265±0.8	342±1.0	9±0.3	378±1.1	
PhR-26	(86.0)	(3.0)	(92.0)	(83.0)	(0.5)	(94.0)	(80.3)	(5.7)	(88.3)	(85.5)	(2.3)	(94.5)	
CrR-2+	85±0.3	2±0.1	92±0.3	170±0.5	2±0.1	188±0.6	225±0.7	1±0.0	267±0.8	345±1.0	7±0.2	379±1.1	
PbR-33	(85.0)	(2.0)	(92.0)	(\$5.0)	(1.0)	(94.0)	(75.0)	(0.3)	(89.0)	(86.3)	(1.8)	(94.8)	
CrR-2+	85±0.3	1±0.0	93±0.3	169±0.5	2±0.1	189±0.6	238±0.7	7±0.2	274±0.8	344±1.0	5±0.1	376±1.	
PhR-64	(85.0)	(1.0)	(93.0)	(84.5)	(1.0)	(94.5)	(79.3)	(2.3)	(91.3)	(86.0)	(1.3)	(94.0)	
CrR-14+	80±0.2	2±0.1	9910.3	155±0.5	3±0.1	191±0.6	239±0.7	8±0.2	284±0.8	319±1.0	6±0.2	399± 1.2	
PhR-26	(80.0)	(2.0)	(99.0)	(77.5)	(1.5)	(95.5)	(79.7)	(2.7)	(94.7)	(79.8)	(1.5)	(99.8)	
CrR-14+	75±0.2	2±0.1	99±0.3	153±0.5	5±0.1	196±0.6	274±0.8	12±0.4	290±0.9	316±0.9	4±0.1	385±1.3	
PhR-33	(75.0)	(2.0)	(99.0)	(76.5)	(2.5)	(98.0)	(91.3)	(4.0)	(96.7)	(79.0)	(1.0)	(96.3)	
CrR-14+	78±0.2	1±0.0	99±0.3	154±0.5	2±0.1	195±0.6	239±0.7	13±0.4	263±0.8	319±1.0	6±0.2	385±1.3	
PhR-64	(78.0)	(1.0)	(99.0)	(77.0)	(1.0)	(97.5)	(79.7)	(4.3)	(87.7)	(79.8)	(1.5)	(96.3)	
CrR-15+	77±0.2	1±0.0	98±0.3	154±0.5	5±0.1	188±0.6	261±0.8	13±0.4	291±0.9	307±0.9	3±0.1	388±1.2	
PhR-26	(77.0)	(1.0)	(98.0)	(77.0)	(2.5)	(94.0)	(87.0)	(4.3)	(97.0)	(76.8)	(0.8)	(97.0)	
CrR-15+	76±0.2	1±0.0	99±0.3	157±0.5	5±0.1.	192±0.6	229±0.7	3±0.1	256±0.8	311±0.9	2±0.1	376±1.	
PhR-33	(76.0)	(1.0)	(99.0)	(78.5)	(2.5)	(96.0)	(76.3)	(1.0)	(85.3)	(77.8)	(0.5)	(94.0)	
CrR-15+	76±0.2	1±0.0	98±0.3	154±0.5	1±0.0	189±0.6	234±0.7	6±0.2	292±0.9	310±0.9	9±0.3	387±1.3	
PhR-64	(76.0)	(1.0)	(98.0)	(77.0)	(0.5)	(94.5)	(78.0)	(2.0)	(97.3)	(77.5)	(2.3)	(96.8)	
CPR-4	85±0.3	3±0.1	92±0.3	165±0.5	4±0.1	196±0.6	241±0.7	13±0.4	271±0.8	333±1.0	4±0.1	390±1.3	
	(85.0)	(3.0)	(92.0)	(82.5)	(2.0)	(98.0)	(80.3)	(4.3)	(90.3)	(83.3)	(1.0)	(97.5)	
CPR-16	49±0.1	3±0.1	53±0.2	1 59±0 .5	4±0.1	174±0.5	241±0.7	18±0.5	274±0.8	352±1.1	10±0.3	341±1.0	
	(49.0)	(3.0)	(53.0)	(79.5)	(2.0)	(87.0)	(80.3)	(6.0)	(91.3)	(88.0)	(2.5)	(85.3)	
CPR-17	84±0.2	2±0.1	95±0.3	166±0.5	2±0.1	197±0.6	24610.8	11±0.3	262±0.8	336±1.0	9±0.3	393±1.	
	(84.0)	(2.0)	(95.0)	(\$3.0)	(1.0)	(98.5)	(82.0)	(3.7)	(87.3)	(84.0)	(2.3)	(98.3)	

Table 4.8Effect of low concentrations; 100, 200, 300 and 400 µg/ml at contact time 15 min., incubation period6 hr. by nine cocultures and three single cultures

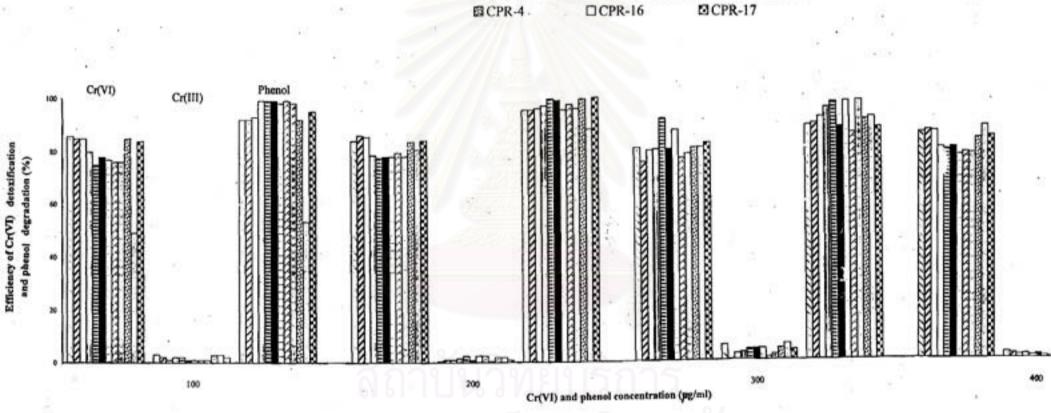


Figure 4.15 Efficiency of Cr(VI) detoxification and phenol degradation at incubation period 6 hr., contact time 15 min. varied from concentration; 100, 200, 300 and 400 µg/ml

□ CrR-2+PhR-26 □ CrR-2+PhR-33 □ CrR-2+PhR-64 □ CrR-14+PhR-26 □ CrR-14+PhR-33 ■ CrR-14+PhR-64 □ CrR-15+PhR-26 □ CrR-15+PhR-33 □ CrR-15+PhR-64

Table 4.9	Effect of high concentrations; 500, 1000, 1500 and 2000 µg/ml at contact time 15 min., incubation pe	riod
	6 hr. by three cocultures and three single cultures	

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	Cr(VI and phenol concentration												
Strains	50)0 µg/ml (%	6)	1000 µg/ml (%)			1500 µg/ml (%)			2000 μg/ml (%)			
	Cr(VI) ¹	Cr(III) ²	Ph ³	Cr(VI) ¹	Cr(III) ²	Ph ³	Cr(VI) ¹	Cr(III) ²	Ph ³	Cr(VI) ¹	Cr(III) ²	Ph ³	
CrR-2+	390±1.2	100±0.3	461±1.2	920±1.3	80±0.2	783±1.3	1430±1.4	80±0.2	883±1.4	1940±1.2	70±0.2	778±1.3	
PhR-26	(78.0)	(20.0)	(92.2)	(92.0)	(8.0)	(78.3)	(95.3)	(5.3)	(58.9)	(97.0)	(3.5)	(38.9)	
CrR-14+	390±1.2	100±0.3	494±1.2	900±1.3	100±0.3	767±1.3	1410±1.4	90±0.3	867±1.4	1920±1.2	90±0.3	839±1.4	
PhR-33	(78.0)	(20.0)	(98.8)	(90.0)	(10.0)	(76.7)	(94.0)	(6.0)	(57.8)	(96.0)	(4.5)	(41.9)	
CrR-15+	400±1.2	90±0.3	456±1.2	910±1.3	90±0.3	911±1.4	1420±1.4	80±0.2	800±1.4	1950±1.2	100±0.3	956±1.4	
PhR-64	(80.0)	(18.0)	(91.2)	(91.0)	(9.0)	(91.1)	(94.7)	(5.3)	(53.3)	(97.5)	(5.0)	(47.8)	
CPR-4	220±0.7	120±0.4	338±1.0	860±1.2	150±0.4	493±1.2	1380±1.3	70±0.2	662±1.3	1890±1.1	60±0.2	707±1.3	
	(44.0)	(24.0)	(67.6)	(86.0) 🖸	(15.0)	(49.3)	(92.0)	(4.7)	(44.1)	(94.5)	(3.0)	(35.5)	
CPR-16	240±0.7	20±0.1	400±1.2	820±1.2	40±0.2	618±1.3	1350±1.3	50±0.2	707±1.3	1890±1.1	40±0.2	707±1.3	
	(48.0)	(4.0)	(80.0)	(82.0)	(4.0)	(61.8)	(90.0)	(3.3)	(47.1)	(94.5)	(2.0)	(35.3)	
CPR-17	190±0.6	40±0.2	480±1.2	810±1.2	50±0.2	889±1.4	1360±1.3	20±0.1	667±1.3	1900±1.1	30±0.1	756±1.3	
	(38.0)	(8.0)	(96.0)	(81.0)	(5.0)	(88.9)	(90.7)	(1.3)	(44.5)	(95.0)	(1.5)	(37.8)	

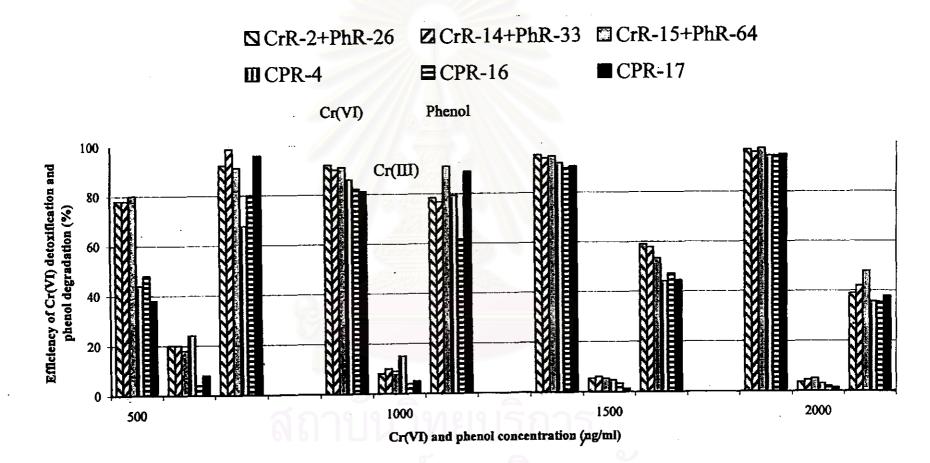


Figure 4.16 Efficiency of Cr((VI) detoxification and phenol degradation at incubation period 6 hr., contact time 15 min. varied from concentration; 500, 1000, 1500 and 2000,ug/ml

4.4 EFFECTS OF SOME ENVIRONMENTAL FACTORS ON CHROMIUM DETOXIFICATION AND PHENOL DEGRADATION

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Optimum pH and optimum temperature for growth of three cocultures and three single cultures were neutral or at 7 and 37°C, summarized in Table 4.10, on page 95. Comparing between bacterial cells of cocultures with single cultures were shown in Figure 4-17 and Figure 4-18, on page 96 and 97.

4.5 EFFICIENCY OF PHENOL DERIVATIVE DEGRADATION BY THE SELECTED BACTERIAL STRAINS

The bacterial strains; PhR-26, PhR-33, PhR-64, CPR-4, CPR-16, CPR-17 were studied the phenol derivative degradation, i. e., p-cresol, p-chlorophenol and p-nitrophenol, the initial concentration 50 μ g/ml,, weekly measured samples for three weeks, summarized in Table 4.11, on page 98. It found that bacterial isolates can degraded p-cresol better than p-chlorophenol and p-nitrophenol, were shown in Figure 4.19, on page 99.

Initial no. of	Viable Count (x10 ^s cells/ml)									
Organism	pH							Temp (°C)		
(x10 ⁸ cells/ml)	4	5	6	7	8	9	10	30	37	40
1.15+1.13	1.32	1.80	1.86	2.45	2.31	1.80	1.48	1.43	1.72	1.68
										ŀ
1.14+1.15	1.56	1.82	1.96	3.44	2.30	1.37	1.15	1.52	1.64	1.54
								4		ļ
1.15+1.14	1.19	1.32	1.72	2.61	1.91	1.25	1.17	1.33	1.65	1.38
								ļ	ļ	
1.14	1.18	1.20	1.31	1.59	1.50	1.17	1.13	1.04	1.40	1.29
1.15 🥖	1.27	1.72	1.99	2.04	1.89	1.16	1.09	1.23	1.53	1.47
1.14	1.34	1.45	1.53	1.71	1.48	1.35	1.26	1.06	1.30	1.10
	Organism (x10 ⁸ cells/ml) 1.15+1.13 1.14+1.15 1.15+1.14 1.15	Organism	Organism Image: style styl	OrganismImage: Second seco	Organism(x108 cells/ml)45671.15+1.131.321.801.862.451.14+1.151.561.821.963.441.15+1.141.191.321.722.611.141.181.201.311.591.151.271.721.992.04	Organism (x10 ⁸ cells/ml)IIII1.15*1.131.321.801.862.452.311.14+1.151.561.821.963.442.301.15*1.141.191.321.722.611.911.151.181.201.311.591.501.151.271.721.992.041.89	Organism (x10 ⁸ cells/ml)Image: Comparison of the tells/ml)Image: Comparison of tells/ml)<	Organism (x108 cells/ml)II </td <td>Organism Image: Sells/ml) Image:</td> <td>Organism (x108 cells/ml)Image: Constraint of the constraint of the cells/ml)Image: Constraint of the cells/ml)</td>	Organism Image: Sells/ml) Image:	Organism (x108 cells/ml)Image: Constraint of the constraint of the cells/ml)Image: Constraint of the cells/ml)

Table 4.10Effect of pH and temperature on growth of the threecocultures and three single cultures



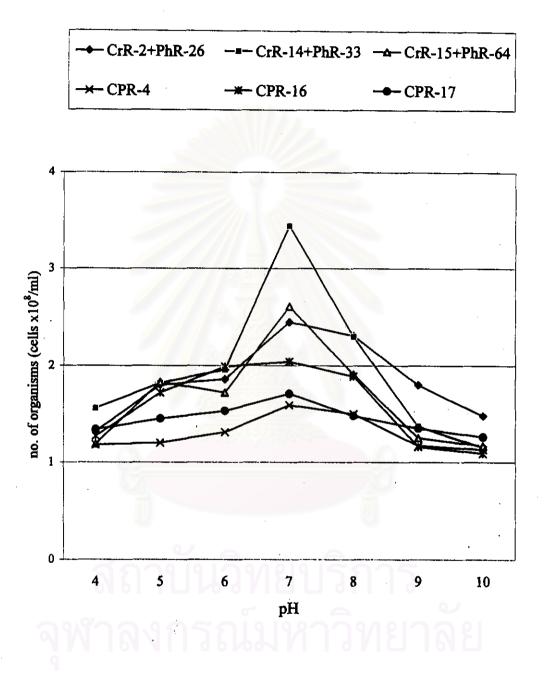


Figure 4.17 Effect of pH on growth of the three cocultures and three single cultures

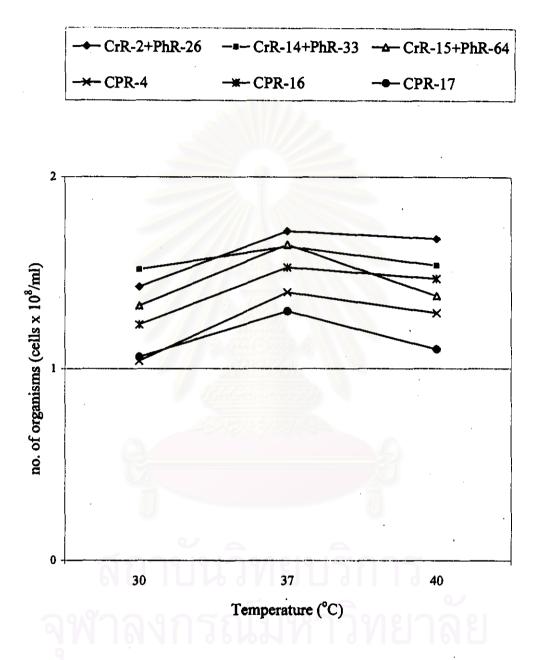


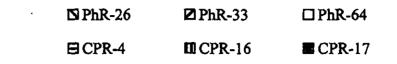
Figure 4.18 Effect of temperature on growth of the three cocultures and three single cultures

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Strains	Week	Loss of Concentration 50 µg/ml (% [*])						
		p-Cresol	p-Chlorophenol	p-Nitrophenol				
	1	49.0±0.1 (98.0)	1±0.0 (2.0)	8±0.2 (16.0)				
PhR-26	2	49.9±0.1 (99.8)	7±0.2 (14.0)	4±0.1 (8.0)				
	3	50.0±0.1 (100.0)	8±0.2 (16.0)	15±0.5 (30.0)				
	1	49.1±0.1 (98.2)	4±0.1 (8.0)	10±0.3 (20.0)				
PhR-33	2	49.6±0.1 (99.2)	3±0.1 (6.0)	11±0.3 (22.0)				
	3	50.0±0.1 (100.0)	11±0.3 (22.0)	12±0.4 (24.0)				
	1	48.3±0.1 (96.6)	2±0.1 (4.0)	8±0.2 (16.0)				
PhR-64	2	49.7±0.1 (99.4)	9± 0.3 (18.0)	8±0.2 (16.0)				
	3	50.0±0.1 (100.0)	13±0.4 (26.0)	13±0.4 (26.0)				
	1	49.4±0.1 (98.8)	3±0.1 (6.0)	13±0.4 (26.0)				
CPR-4	2	50.0±0.1 (100.0)	4±0.1 (8.0)	10±0.3 (20.0)				
	3	50.0±0.1 (100.0)	11±0.3 (22.0)	13±0.4 (26.0)				
	1	49.5±0.1 (99.0)	8±0.2 (16.0)	3±0.1 (6.0)				
CPR-16	2	50.0±0.1 (100.0)	8±0.2 (16.0)	3±0.1 (6.0)				
	3	50.0±0.1 (100.0)	14±0.4 (28.0)	18±0.5 (36.0)				
A N	161	48.6±0.1 (97.2)	8±0.2 (16.0)	11±0.3 (22.0)				
CPR-17	2	50.0±0.1 (100.0)	5±0.2 (10.0)	6±0.1 (12.0)				
	3	50.0±0.1 (100.0)	17±0.5 (34.0)	18±0.5 (36.0)				

Table 4.11Efficiency of phenol derivative degradation on variousweeks; 1, 2 and 3 week.

* Efficiency of Degradation (%)



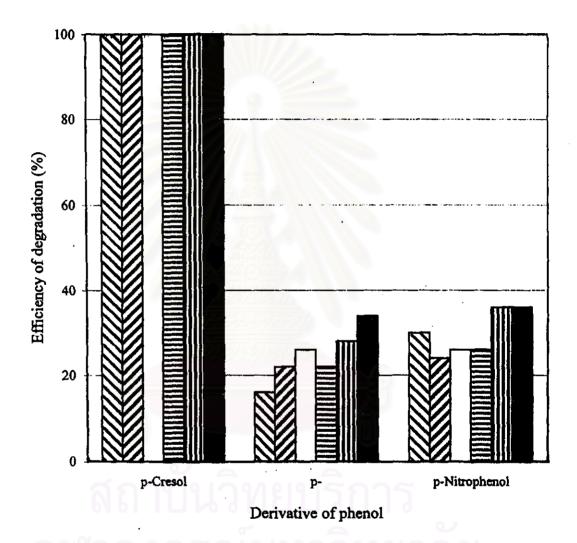


Figure 4.19 Efficiency of degradation of 50 µg/ml p-Cresol, p-Chlorophenol and p-Nitrophenol on the third week