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APPENDIX

I. Ascorbic Acid (Vitamin C) Chemistry and Biochemistry

Ascorbic acid ($C_6H_6O_8$) is water soluble, hexonic sugar acid, with a molecular weight of 176.13. Ascorbic acid is synthesized from glucose in most animals.

Vitamin C refers to all compounds that exhibit the biological activity of ascorbic acid, including both ascorbic acid and its oxidized form, dehydroascorbic acid. Removal of one electron from ascorbic acid yields semidehydroascorbic acid (ascorbate radical). This form of the vitamin is a free radical, it contains an unpaired electron. The removal of a second electron yields dehydroascorbic acid. Semidehydroascorbate is an intermediate in this conversion pathway. Dehydroascorbate reductase requires glutathione (GSH) as a source of reducing power. Both ascorbic acid and dehydroascorbic acid have biological activity. The latter compound may break down to form diketogulonic acid. Diketogulonic acid has no biological activity (Levine M, 1986).

II. Plasma ascorbic acid determination

In the present study, a specific enzymatic spectrophotometric method for ascorbic acid in plasma was used. Samples were analyzed indirectly by measuring the absorbance at 593 nm of a reaction product, a complex of ferrous ion and 2, 4, 6-tris (2-pyridyl)-s-triazine (Fe^{2+} -TPTZ). Ascorbic acid is specifically quantified by pretreating one of a pair of replicate samples with ascorbate oxidase, to oxidize the ascorbic acid, then reacting both samples with Fe^{3+} -TPTZ and measuring the difference

between the absorbances at 593 nm of the treated and untreated samples (Liu TZ et al., 1982).

Table 14. Procedure for Enzymatic-assisted plasma ascorbic acid determination.

Material	AA oxidase	AA oxidase
	Treated tube	Free tube
Unknown plasma or (standard and controls)(ml)	0.1	0.1
AA oxidase working solution (ml)	0.05	-----
H ₂ O (ml)	-----	0.05
Vertex-mix gently and incubate at 37°C in a waterbath for 15 minutes		
Acetate buffer , pH 3.6 (ml)	1.25	1.25
TPTZ Solution (ml)	0.15	0.15
FeCl ₃ -6 H ₂ O Solution (ml)	0.1	0.1
Gently mix the contents of every tube after additions. Allow to stand at room temperature for exactly 5 min. Measure absorbance at 593 nm.		

Plasma ascorbic acid concentration can be obtained from a standard curve (LIU TZ et al., 1982).

III. Streptozotocin-Induced Diabetic Rats Model

The animal model of insulin-dependent diabetes mellitus (IDDM) that is used in this study is induced by a single intravenous injection with the dose of 55 mg/kg BW streptozotocin (STZ) (Jariyapongskul A et al., 1996).

In this study, we used STZ-treated rat model as an insulin-dependent diabetes mellitus, because this model closely resemble to

IDDM in human. The dose of 55 mg/kg BW STZ is used by a single intravenous injection. With this dose, the rats become hyperglycemia and hypoinsulinemia within 24-48 hours after induction.

Streptozotocin induce beta-cell damage by initiating biochemical events which cause DNA strand breaks. STZ is able to cause beta –cell specific damage via its ability to interact with the glucose sensing mechanism of the beta-cell specific damage (Like AA, 1976).

IV. Fluorescence Microscopic Technique

Intravital fluorescence microscopy has been used to observe the microvascular of parenchymatous organ, such as brain, heart, lung, liver, pancreas, gut, kidney.

Microscopic viewing may be performed by trans-illumination or by epi-illumination using a modified Leitz Orthoplan microscope (75 W, XBO, xenon lamp or 100 W, HBO, mercury lamp which attached to a ploemo-Pak illuminator with different filter blocks for epi-illumination).

In this study the microscopic pictures are recorded by means of low-light level SIT (Silicon-intensified target) video camera-assisted image analysis system. The availability of different fluorescent markers for in vivo study are fluorescein isothiocyanate dextran (MW 150,000 ; 15 mg/kg BW) (FITC-dextran)-labeled plasma and Rhodamine-6G (0.15 mg/kg BW) to stain mitochondria especially in leukocyte, and their fluorescene emissions are detected separately by using two sets of filters (FITC dextran : λ_{em} =520 nm; R6G : λ_{em} =595 nm). FITC- dextran dissolved in plasma used to measure the diameter of various microvessels.

V. Free radical by products: Malondialdehyde (MDA)

Oxidation of polyunsaturated fatty acids leads to numerous peroxidic and aldehydic compound, in particular the volatile low molecular weight aldehyde, malondialdehyde (MDA). The chemical composition of the end products of peroxidation will depend on the fatty acid composition of the lipid substrate used and upon what metal ions are present. Thus copper and iron ions give different end-product distributions as measured by the thiobarbituric acid (TBA) test. This is one of the most commonly used method for detecting and measuring lipid peroxidation. The lipid material is simply heated with TBA at low pH, and the formation of a pink chromogen is measured at or close to 532nm. The chromogen is formed by reaction of one molecule of malondialdehyde (MDA) with two molecules of TBA (Ohgawa H et al., 1979).

Reagents

1. 8.1% Sodium dodecyl sulfate (SDS)

Dissolve SDS 8.1 g in distilled water and allow to stand overnight at room temperature until it is dissolved. Then make up to 100 ml. Do not shake because this solution will product a lot of bubble and store in refrigerator.

2. 20% of acetic acid solution (pH 3.5)

Pipette 26.61 ml of 37% HCl into volumeric flask (1,000 ml) and make up to 1,000ml with distill water. 0.27 M HCl is then obtained. Add 20 ml pure acetic acid in 80 ml 0.27 m HCl. Adjust the solution to pH 3.5 with 1 N NaOH.

3. 0.8% Thiobarbituric acid (TBA)

Weight TBA 0.8 g. Then add distilled water, heat and stir until it is dissolved. Make this solution up to 100 ml and mix.

4. 1,1,3,3- Tetramethoxypropane (TMP) or malondialdehyde bis (dimethyl acetal)

Solution is used as an external standard. The level of lipid peroxide is expressed as nmole of MDA.

Pipette 16.4 μ l stock TMP and make up to 100 ml with distilled water. Then pipette 0.04, 0.08, 0.12, 0.16, 0.20 ml of this stock TMP solution and distilled water to 10 ml in each concentration. These will give the following concentration of standard TMP: 4, 8, 12, 16, 20 nmole/ml. Store the stock solution in refrigerator.

5. 1.15% KCl in 0.1 M KH_2PO_4 to make phosphate buffer pH 7.4 add 1.15 g of KCl to 100 ml of this 0.1 M phosphate buffer (pH7.4), mix thoroughly.

Procedure

1. After washing the isolated muscle in ice-cold 0.9% NaCl, the eye ball homogenate are prepared by homogenizing each gram of wet tissue in 4 ml of 1.15% KCl in 0.1 M phosphate buffer, pH7.4.

2. Pipette the following solutions into a series of glass tubes with screw capped:

Solution (ml)	Blank (ml)	Standard	(ml)
Unknown			
Sample	-	-	0.5
8.1% SDS	0.2	0.2	0.2
20% Acetic acid (pH 3.5)	1.5	1.5	1.5
0.8% TBA	1.5	1.5	1.5
TMP stock standard	-	0.5	-
Distill water	0.8	0.3	0.3

3. Heated the tubes in the water-bath at 95°C for 60 min.

4. After cooling with top water, 1.0 ml of distilled water and 5.0 ml of the mixture of n-butanol and pyridine (15:1 v/v) are added and shaken vigorously (at least 1 min.).

5. After centrifugation at 4,000 rpm for 10 min, the organic layer is taken and its absorbance at 532 nm is measured.

6. The content of lipid peroxide is expressed in terms of nmole MDA/ml.

CALIBRATION CURVE

1. Prepare a series of tube containing TMP stock standard in water in the following concentrations: 2.0 nmole/0.5 ml, 4.0 nmole/0.5 ml, 6.0 nmole/0.5ml, 8.0 nmole/0.5ml and 10.0 nmole/0.5ml.

2. Perform the procedure as in step 2.

3. Determine the absorbance at 532 nm. Then plot the optical density versus nmole of MDA/ml.

BIOGRAPHY

Miss Tippawan Rungjaroen was born on April 26, 1975 in Suratane, Thailand. She graduated Bachelor of Nursing Science from Faculty of Nursing, Mahidol University in 1998.



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