

## References

- Agnihotri, S. A., Malligarjuna, N. N., and Aminabhavi, T. M. Recent advances on chitosan based micro- and nanoparticles in drug delivery. *J. Control. Release.* 100 (2004): 5-28.
- Ahlin, P., Kristl, J., and Smid-Korbar, J. Optimization of procedure parameters and physical stability of solid lipid nanoparticles in dispersions. *Acta Pharm.* 48 (1998): 259-267.
- Aktas, Y., Andrieux, K., Alonso, M. J., Calvo, P., Gursoy, R. N., Couvreur, P., and Capan, Y. Preparation and in vitro evaluation of chitosan nanoparticles containing a caspase inhibitor. *Int. J. Pharm.* 298 (2005) 378-383.
- Alonso, M. J. Nanomedicines for overcoming biological barriers. *Biomed Pharmacother.* 58 (2004):168-172.
- Alvarez-Román, R., Naik, A., Kalia, Y. N., Fessi, H., and Guy, R. H. Visualization of skin penetration using confocal laser scanning microscopy. *Eur. J. Pharm. Biopharm.* 58 (2004): 301-316.
- Alving, C. R., and Glenn, G. M. Transdermal delivery system for antigen. *United States Patents:* 5,910,306. June 8, 1999.
- Ataman-Önal, Y., Munier, S., Ganée, A., Terrat, C., Durand, P., Battail, N., Martinon, F., Grand R. L., Charles, M., Delair, T., and Verrier, B. Surfactant-free anionic PLA nanoparticles coated with HIV-1 p24 protein induced enhanced cellular and humoral immune responses in various animal models. *J. Control. Release.* 112 (2006): 175-185.
- Babiuk, S., Baca-Estrada, M., Babiuk, L., Ewen, C., and Foldvari, M. Cutaneous vaccination: the skin as an immunologically active tissue and the challenge of antigen delivery. *J. Control. Release.* 66 (2000): 199-214.
- Baca-Estrada, M. E., Foldvari, M., Ewen, C., Badea, I., and Babiuk, L. A. Effect of IL-12 on immune responses induced by transcutaneous immunization with antigens formulates in a novel lipid-based biphasic delivery system. *Vaccine* 18 (2000): 1847-1854.

- Baliga, C. S., Maanen, M., Chastain, M., and Sutton, R. E. Vaccination of mice with replication-defective human immunodeficiency virus induces cellular and humoral immunity and protects against vaccinia virus-gag challenge. Mol. Ther. 14 (2006): 432-441.
- Balzarini, J., and Damme, L. V., Microbicide drug candidates to prevent HIV infection. Lancet 369 (2007): 787-797.
- Barry, B. W. Dermatological Formulations: Percutaneous Absorption, NY: Dekker, 1983.
- Barry, B. W. Novel mechanism and devices to enable successful transdermal drug delivery. Eur. J. Pharm Sci. 14 (2001): 101-114.
- Bergmann-Leitner, E. S., and Leitner, W. W. Danger, death and DNA vaccine. Microbes Infect. 6 (2004): 319-327.
- Birchall, J. C., Marichal, C., Campbell, L., Alwan, A., Hadgraft, J., and Glumbleton, M. Gene expression in an intact ex-vivo skin tissue model following percutaneous delivery of cationic liposome-plasmid DNA complexes. Int. J. Pharm. 197 (2000): 233-238.
- Bourgaize, D., Jewell, T. R., and Buiser, R. G. Biotechnology : Demysifying the concept. Addison Wesley Longman: CA, 2000.
- Bråve, A., Ljungberg, K., Boberg, A., Rollman, E., Isagulians, M., Lundgren, B., Blomberg, P., Hinkula, J., and Wahren, B. Multigene/multisubtype HIV-1 vaccine induces potent cellular and humoral immune responses by needle-free intradermal delivery. Mol. Ther. 12 (2005): 1197-1205.
- Calvo, P., Alonso, M. J., Vila-Jato, J. L., and Robinson, J. R. Improved ocular bioavailability of indomethacin by novel ocular drug carriers. J. Pharm. Pharmacol. 48 (1996):1147-1152.
- Cevc, G., Schatzlein, A., and Blume, G. Transdermal drug carrier: basic properties, optimization and transfer efficiency in the case of epicutaneously applied peptides. J. Control. Release 36 (1995): 3-16.
- Chacón, M., Berges, L, Molpeceres, J., Aberturas, M. R., and Guzman, M. Optimized preparation of poly D,L(lactic-glycolic) microspheres and nanoparticles for oral administration. Int. J. Pharm. 141 (1996): 81-91.

- Cui, Z., Baizer, L., and Mumper, R. J. Intradermal immunization with novel plasmid DNA-coated nanoparticles via a needle-free injection device. J. Biotechnol 102 (2003): 105-115.
- Cui, Z., and Mumper, R. J. Chitosan-based nanoparticles for topical genetic immunization. J. Control. Release. 81 (2001): 409-419.
- Cui, Z., and Mumper, R. J. Topical immunization using nanoengineered genetic vaccine. J. Control. Release. 81 (2002): 173-184.
- Davoren, M., Shúilleabháin, S. N., Hartl, M. G. J., Sheehan, O'Brien, D. N., O'Halloran, M. J., Pelt, F. N. A. M. V., and Mothersill, C. Assessing the potential of fish cell lines as tools for the cytotoxicity testing of estuarine sediment aqueous elutriates. Toxicol In Vitro. 19 (2005): 421-431.
- De Souza, E. F., and Teschke, O. Liposome stability verification by atomic force microscopy. Rev. Adv. Mater. Sci. 5 (2003): 34-40.
- Egan, M. A., Megati, S., Roopand, V., Garcia-Hand, D., Luckay, A., Chong, S., Rosati, M., Sackitey, S., Weiner, D. B., Felber, B. K., Pavlagis, G. N., Israel, Z. R., Eldridge, J. H., and Sidhu, M. K. Rational design of plasmid DNA vaccine capable of eliciting cell-mediated immune responses to multiple HIV antigen in mice. Vaccine 24 (2006): 4510-4523.
- Farhood, H., Serbina, N., and Huang, L. The role of dioleoyl phosphatidylethanolamine in cationic liposome mediated gene transfer. Biochim. Biophys. Acta. 1235 (1995): 289-295.
- Foldvari, M. Non-invasive administration of drugs through the skin: challenges in delivery system design. Pharm. Sci. Technol. Today. 3 (2000): 417-425.
- Freed, E. O. HIV-1 gag protein: Diverse functions in the virus life cycle. Virology. 251 (1998): 1-15.
- Freitas, C., and Müller, R. H. Effect of light and temperature on zeta potential and physical stability in solid lipid nanoparticle (SLN<sup>TM</sup>) dispersions. Int. J. Pharm. 168 (1998): 221-119.
- Gan, Q., Wang, T., Cochrane, C., and McCarron, C. Modulation of surface charged, particle size and morphological properties of chitosan-TPP nanoparticles intended for gene delivery. Colloids surf B: Biointerfaces. 44 (2005): 65-73.

- Glenn, G. M., Rao, M., Matyas, G. R., and Alving, C. R. Skin immunization made possible by cholera toxin. Nature 391 (1998): 851.
- Glenn, G. M., Taylor, D. N., Li, X., Frankel, S., Montemarano, A., and Alving, C. R. Transcutaneous immunization: A human vaccine delivery strategy using a patch. Nat. Med. 12 (2000): 1403-1406.
- Goloub, T. P., and Pugh, R. J. The role of the surfactant head group in the emulsification process: binary (nonionic-ionic) surfactant mixtures. J. Colloid Interface Sci. 291 (2005): 256-262.
- Grossman, M. D., and Stawicki, S. P. The impact of human immunodeficiency virus (HIV) on outcome and practice in trauma: past, present and future. Int. J. Care injured. 37 (2006): 1117-1124.
- Gu, Y., and D. Li. The  $\zeta$ -potential of silicone oil droplets dispersed in aqueous solutions. J. Colloid Interface Sci. 206 (1998): 346-349.
- Guebre-Xabier, M., Hammond, S. A., Epperson, D. E., Yu, J., Ellingsworth, L., and Glenn, G. M. Immunostimulant patch containing heat-labile enterotoxin from *Escherichia coli* enhances immune responses to injected influenza virus vaccine through activation of skin dendritic cells. J. Virol. 77 (2003): 5218-5225.
- Hannigan, B. M., and Pallister, C. J. Immunology. Hodder headline: London, 2000.
- Henry, S., Mcallister, D. V., Allen, M. G., and Prausnitz, M. R. Microfabricated microneedles: a novel approach to transdermal drug delivery. J. Pharm. Sci. 87 (1998): 922-925.
- Heurtault, B., Saulnier, P., Pech, B., Proust, J., and Benoit, J. Physico-chemical stability of colloidal lipid particles. Biomaterials. 24 (2003): 4283-4300.
- Hou, D., Xie, C., Huang, K., and Zhu, C. The production and characteristics of solid lipid nanoparticles. Biomaterials. 24 (2003): 1781-1785.
- Huang, L., Farhood, H., Serbina, N., Teepe, A. G., and Barsoum, J. Endosomolytic activity of cationic liposomes enhances the delivery of human immunodeficiency virus-1 trans activator protein (tat) to mammalian cells. Biochim. Biophys. Res. Commun. 217 (1995): 761-768.

- Hung, C., Hwang, T., Chang, C., and Fang, J. Physicochemical characterization and gene transfection efficiency of lipid emulsions with various co-emulsifiers. Int. J. Pharm. 289 (2005):197-208.
- Jain, S. K., Chourasia, M. K., Masuriha, R., Soni, V., Jain, A., Jain, N. K., and Gupta, Y. Solid lipid nanoparticles bearing flurbiprofen for transdermal delivery. Drug Deliv. 12 (2005): 207-215.
- Jenning, V., Thünemann, A. F., and Gohla, S. H. Characterisation of a novel solid lipid nanoparticle carrier system based on binary mixtures of liquid and solid lipids. Int. J. Pharm. 199 (2000): 167-177.
- Katas, H., and Alpar, H. O. Development and characterization of chitosan nanoparticles for siRNA delivery. J. Control. Release. 115 (2006): 215-225.
- Levine, M. M. Can needle-free administration of vaccines become the norm in global immunization? Nat. Med. 9 (2003): 99-103.
- Locher, C. P., Putnam, D., Langer, R., Witt, S. A., Ashlock, B. B., and Levy, J. A.. Enhancement of a human immunodeficiency virus env DNA vaccine using a novel polycationic nanoparticle formulation. Immunol Lett. 90 (2003): 67-70.
- López-León, T., Carvalho, E. L. S., Seijo, B., Ortega-Vinuesa, J. L., and Bastos-Gonzalez, D. Physicochemical characterization of chitosan nanoparticles: electrokinetics and stability behavior. J. Colloid. Interface. Sci. 283 (2005): 344-351.
- Martin, A. Physical pharmacy. 4<sup>th</sup> ed. USA: Lea & Febiger, 1993.
- Mehnert, W., and Mäder, K. Solid lipid nanoparticles production, characterization and applications. Adv Drug Deliv Rev. 47 (2001): 165-196.
- Mikszta, J. A., Alarcon, J. B., Brittingham, J. M., Sutter, D. E., Pettis, R. J., and Harvey, N. G. Improved genetic immunization via micro mechanical disruption of skin-barrier function and targeted epidermal delivery. Nat. Med. 8 (2002): 415-419.
- Mikszta, J. A., Brittingham, J. M., Alarcon, J., Pettis, R. J., and Dekker, J. P. Topical delivery of vaccines. United States Patents: 20030191085. October 9, 2003.
- Montgomery, D. C. Design and analysis of experiments. 6<sup>th</sup> ed. USA: John Wiley & Sons, 2005.

- Müller, R. H., Mäder, K., and Gohla, S. Solid lipid nanoparticles (SLN) for controlled drug delivery-a review of the state of the art. Eur. J. Pharm. Biopharm. 50 (2000): 161-177.
- Müller, R. H., Radtke, M., and Wissing, S. A. Solid lipid nanoparticles (SLN) and nanostructured lipid carriers (NLC) in cosmetic and dermatological preparations. Adv. Drug. Deliv. Rev. 54 suppl. 1 (2002): s131-155.
- Mumper, R. J., and Roland, A. P. Plasmid delivery to muscle: Recent advances in polymer delivery system. Adv. Drug. Del. Rev. 30 (1998): 151-172.
- Myers, R. H., and Montgomery, D. C. Response surface methodology. 2<sup>nd</sup> ed. USA: John Wiley & Sons, 2002.
- Niemiec, S. M., Latta, J. M., Ramachandran, C., Weiner, N. D., and Roessler, B. J. Perifollicular transgenic expression of human interleukin-1 receptor antagonist protein following topical application of novel liposome-plasmid DNA formulations in vivo. J. Pharm. Sci. 86 (1997): 701-708.
- Olbrich, C., Bakowsky, U., Lehr, C., Müller, R. H., and Kneuer, C. Cationic solid-lipid nanoparticles can efficiently bind and transfect plasmid DNA. J. Control. Rel. 77 (2001): 345-355.
- Pang, S., Park, H., Jang, Y., Kim, W., and Kim, J. Effects of charge density and particle size of poly(styrene/(dimethylamino)ethyl methacrylate) nanoparticle for gene delivery in 293 cells. Colloids Surf B: Biointerfaces. 26 (2002):213-222.
- Panyam, J., and Labhasetwar, V. Biodegradable nanoparticles for drug and gene delivery to cells and tissue. Adv. Drug Del. Rev. 55 (2003): 329-347.
- Peachman, K. K., Rao, M., and Alving, C. R. Immunization with DNA through the skin. Method 31(2003): 232-242.
- Pedroso de Lima, M. C., Simões, S., Pires, P., Faneca, H., and Düzgünes, N. Cationic lipid-DNA complexes in gene delivery: from biophysics to biological applications. Adv. Drug. Del. Rev. 47 (2001): 277-294.
- Prather, K. J., Sagar, S., Murphy, J., and Chartrain, M. Industrial scale production of plasmid DNA for vaccine and gene therapy: plasmid design, production, and purification. Enz. Microbiol. Technol. 33 (2003): 865-883.

- Rambali, B., Baert, L., and Massart, D. L. Using experimental design to optimize the process parameters in fluid bed granulation on a semi-full scale. Int. J. Pharm. 220 (2001): 149-160.
- Rekhi, G. S., Nellore, R. V., Hussain, A. S., Tillman, L. G., Malinowski, H. J., and Augsburger, L. L. Identification of critical formulation and processing variables for metoprolol tartrate extended-release (ER) matrix tablets. J. Control. Rel. 59 (1999): 327-342.
- Roland A. Pharmaceutical Particulate Carriers: Therapeutic Application. NY: Marcel Dekker, 1993.
- Roland, A. and Sullivan, S. M. Pharmaceutical gene delivery systems. USA: Marcel Dekker, 2003.
- Schubert, M. A., and Müller-Goymann, C. C. Characterisation of surface-modified solid lipid nanoparticles (SLN): influence of lecithin and nonionic emulsifier. Eur. J. Pharm. Biopharm. 61 (2005): 77-86.
- Senthilkumar, S. R., Ashokkumar, B., Raj, K. C., and Gunasekaran, P. Optimization of medium composition for alkali-stable xylanase production by *Aspergillus fischeri* Fxn 1 in solid-state fermentation using central composite rotary design. Bioresour. Technol. 96 (2005): 1380-1386.
- Shen, W., and Louie, S. G. Immunology for Pharmacy Students. Amsterdam: Harwood Academic Publisher, 1999.
- Shotton, D. M. Confocal scanning optical microscopy and its applications for biological specimens. J. Cell Sci. 94 (1989): 175-206.
- Siekmann, B., and Westensen, K. Thermoanalysis of the reprecipitation process of melt-homogenized glyceride nanoparticles. Colloids surf B: Biointerfaces. 3 (1994): 159-175.
- Sierra, S., Kupfer, B., and Kaiser, R. Basics of the virology of HIV-1 and its replication. J. Clin. Virol. 34 (2005): 233-244.
- Tabatt, K., Kneuer, C., Sameti, M., Olbrich, C., Müller, R. H., Lehr, C., and Bowkosky, U. Transfection with different colloidal systems: comparison of solid lipid nanoparticles and liposomes. J. Control. Release. 97 (2004): 321-332.

- Tabatt, K., Sameti, M., Olbrich, C., Müller, R. H., and Lehr, C. Effect of cationic lipid and matrix lipid composition on solid lipid nanoparticles-mediated gene transfer. Eur. J. Pharm. Biopharm. 57 (2004): 155-162.
- Turner, B. G., and Summers, M. F. Structural biology of HIV. J. Mol. Biol. 285 (1999): 1-32.
- Vandervoort, J., and Ludwig, A. Biocompatible stabilizers in the preparation of PLGA nanoparticles: a factorial design study. Int. J. Pharm. 238 (2002): 77-92.
- Venkateswarlu, V., and Manjunath, K. Preparation, characterization and in vitro release kinetics of clozapine solid lipid nanoparticles. J. Control. Release. 95 (2004): 627-638.
- Watabe, S., Xin, K., Ihata, A., Liu, L., Honsho, A., Aoki, I., Hamajima, K., Wahren, B., and Okuda, K. Protection of influenza virus challenge by topical application of influenza DNA vaccine. Vaccine. 19 (2001): 4434-4444.
- Weiner, N. Targetted follicular delivery of macromolecules via liposomes. Int. J. Pharm. 162 (1998): 29-38.
- Westesen, K. and Bunjes, H. Do nanoparticles prepared from lipids solid at room temperature always possess a solid lipid matrix? Int. J. Pharm 115 (1995): 129-131.
- Wiethoff, C. M., Smith, J. G., Koe, G. S., and Middaugh, C. R. The potential role of proteoglycans in cationic lipid-mediated gene delivery: studies of the interaction of cationic lipid-DNA complexes with model glycosaminoglycans. J. Biol. Chem. 276 (2001): 32806-32813.
- Wissing, S. A., Kayser, O., and Muller, R. H. Solid lipid nanoparticles for parenteral drug delivery. Adv. Drug. Del. Rev. 56 (2004): 1257-1272.
- Wu, H., Ramachandran, C., Bielinska, A. U., Kingzett, K., Sun, R., Weiner, N.D., and Roessler, B. J. Topical transfection using plasmid DNA in a water-in-oil nanoemulsion. Int. J. Pharm 221 (2001): 23-34.



## **APPENDICES**

**Appendix A**

**Details of Statistical Analysis of Model Equations**

**Response: Size**

Source	Sum of Squares	DF	Mean Square	F-value	Prob>F
Mean	4.52E+005	1	4.52E+005		
Linear	1,603.09	3	534.36	4.06	0.0285
2FI	975.78	3	325.26	4.14	0.0343
Quadratic	788.48	3	262.83	27.54	0.0001
Cubic	15.94	4	3.99	0.26	0.8874
Residual	60.40	4	15.10		
Total	4.56E+005	18	25308.46		

***p*-value of Regression Coefficient ( $\alpha=0.05$ )**

Regression Coefficient	<i>p</i> -value
a	<0.0001
b	<0.0001
c	0.0013
a <sup>2</sup>	0.0208
b <sup>2</sup>	0.0002
c <sup>2</sup>	0.0106
ab	0.0001
bc	<0.0001

**Response: PI**

Source	Sum of Squares	DF	Mean Square	F-value	Prob>F
Mean	1.36	1	1.36		
Linear	4.03E-003	3	1.34E-003	0.77	0.5281
2FI	0.011	3	3.54E-003	2.84	0.0867
Quadratic	0.012	3	4.07E-003	21.49	0.0003
Cubic	2.17E-004	4	5.42E-005	0.17	0.9443
Residual	1.30E-003	4	3.24E-004		
Total	1.39	18	0.08		

***p*-value of Regression Coefficient ( $\alpha=0.05$ )**

Regression Coefficient	<i>p</i> -value
a	0.0478
b	0.0019
c	0.0498
a <sup>2</sup>	0.0415
b <sup>2</sup>	<0.0001
c <sup>2</sup>	0.0118
ab	0.0088
ac	0.0008
bc	0.0031

**Response: Zeta potential**

Source	Sum of Squares	DF	Mean Square	F-value	Prob>F
Mean	37753.86	1	37753.86		
Linear	2649.08	3	883.03	10.05	0.0009
2FI	534.33	3	178.11	2.82	0.0884
Quadratic	691.94	3	230.65	561.23	<0.0001
Cubic	0.46	4	0.11	0.16	0.9477
Residual	2.83	4	0.71		
Total	41632.49	18	2312.92		

***p*-value of Regression Coefficient ( $\alpha=0.05$ )**

Regression Coefficient	<i>p</i> -value
a	<0.0001
b	<0.0001
c	0.0040
a <sup>2</sup>	0.0020
b <sup>2</sup>	<0.0001
c <sup>2</sup>	<0.0001
ab	<0.0001
ac	<0.0001
bc	0.0002

**Appendix B**  
**Ingredients of Reagents for ELISA**

**0.05M Carbonate-bicarbonate buffer (pH 9.6)**

Sodium carbonate	0.8 g
Sodium hydrogen carbonate	1.5 g
Purified water added up to make	500 ml

**Phosphate buffer saline pH 7.4 with 0.05% Tween 20 (PBS-T)**

Sodium chloride	8.0 g
Potassium dihydrogen phosphate	0.2 g
Disodium hydrogen phosphate	2.9 g
Potassium chloride	0.2 g
Thimerosol	0.1 g
Tween 20	0.5 ml
Purified water added up to make	1,000 ml

**3% Gelatin in PBS-T**

Gelatin	3.0 g
PBS-T added up to make	100 ml

**Citrate-phosphate buffer pH 5.0**

Citric acid (monohydrate)	10.30 g
Sodium hydrogen phosphate	18.16 g
30% Hydrogen peroxide	1.00 g
Purified water added up to make	1,000 ml

**1% Gelatin in PBS-T**

Gelatin	1.0 g
PBS-T added up to make	100 ml

**4 N Sulfuric acid**

98% Sulfuric acid	54.4 g
Purified water added up to make	500 ml



**Appendix C**

**Physical Stability Study of SLN and CSN**

**Particle size (nm) of SLN and CSN during 60-day storage (mean±SD)**

Formulation	Day 1	Day 7	Day 14	Day 30	Day 60
SLN9	160±2	163±3	163±2	165±1	167±4
SLN10	179±1	179±3	179±3	178±2	180±4
SLN11	142±1	142±4	142±3	148±2	153±3
SLN12	163±1	163±2	163±2	163±3	165±4
SLN13	161±1	162±2	163±4	163±3	164±3
SLN14	154±2	154±2	154±2	154±1	156±5
SLN15	166±1	166±2	166±1	165±2	168±2
CSN2	258±1	259±1	259±1	261±1	269±1

**PI of SLN and CSN during 60-day storage (mean±SD)**

Formulation	Day 1	Day 7	Day 14	Day 30	Day 60
SLN9	0.27±0.01	0.27±0.01	0.27±0.00	0.28±0.01	0.28±0.01
SLN10	0.24±0.01	0.25±0.00	0.25±0.00	0.26±0.01	0.25±0.01
SLN11	0.32±0.01	0.32±0.00	0.33±0.00	0.32±0.00	0.33±0.01
SLN12	0.36±0.01	0.36±0.00	0.37±0.01	0.37±0.01	0.37±0.00
SLN13	0.25±0.01	0.26±0.01	0.26±0.00	0.26±0.00	0.26±0.01
SLN14	0.24±0.00	0.25±0.00	0.25±0.01	0.25±0.00	0.25±0.01
SLN15	0.24±0.00	0.25±0.00	0.25±0.00	0.25±0.00	0.25±0.00
CSN2	0.28±0.00	0.29±0.00	0.29±0.00	0.30±0.01	0.35±0.01

**Zeta potential (mV) of SLN and CSN during 60-day storage (mean±SD)**

Formulation	Day 1	Day 7	Day 14	Day 30	Day 60
SLN9	58.21±1	59.01±0	58.33±1	59.34±0	54.44±1
SLN10	48.56±1	48.77±1	48.66±1	49.12±1	49.89±1
SLN11	21.51±0	22.55±0	21.78±1	19.81±0	19.49±0
SLN12	52.31±1	52.55±1	52.87±1	53.01±1	53.12±0
SLN13	55.72±1	55.82±2	56.12±1	56.39±0	57.31±1
SLN14	53.98±1	53.88±1	52.67±0	53.81±0	53.69±1
SLN15	50.12±1	50.18±1	50.33±1	51.21±0	51.11±0
CSN2	28.25±0	27.70±1	27.31±1	25.12±0	21.67±0

## **Appendix D**

**Powder X-Ray Diffraction Pattern of Bulk Materials, SLN10,  
SLN12, and SLN14**

**Cetylpalmitate**

Angle ( $2\theta^\circ$ )	d-value (Å)	Intensity count	% Intensity
6.04	14.61	57.6	23.9
6.80	12.99	241.0	100.0
10.12	8.73	17.4	7.2
11.40	7.76	91.6	38.0
15.92	5.56	19.5	8.1
21.65	4.10	15.1	6.3
23.84	3.73	14.3	5.9
39.32	2.29	32.7	13.6
41.73	2.16	16.1	6.7

**DDAB**

Angle (2 $\theta$ )	d-value (Å)	Intensity count	% Intensity
7.35	12.01	1308.0	89.3
11.07	7.99	1374.0	93.8
14.83	5.97	1465.0	100.0

<b>Cholesterol</b>			
Angle (2 $\theta$ )	d-value (Å)	Intensity count	% Intensity
5.13	17.23	25549.0	100.0
6.18	14.29	8287.0	32.4
6.96	12.70	3075.0	12.0
7.58	11.65	3756.0	14.7
8.49	10.40	2578.0	10.1
8.95	9.87	2276.0	8.9
9.39	9.41	4374.0	17.1
10.45	8.46	7190.0	28.1
11.16	7.92	4375.0	17.1
11.55	7.66	4389.0	17.2
12.47	7.09	6535.0	25.6
13.00	6.81	7887.0	30.9
14.03	6.31	17946.0	70.2
15.25	5.80	24973.0	97.7
16.05	5.52	6838.0	26.8
16.80	5.27	20444.0	80.0
17.21	5.15	18737.0	73.3
17.99	4.93	17517.0	68.6
18.77	4.72	6142.0	24.0
19.31	4.59	5735.0	22.4
20.07	4.42	3132.0	12.3
20.50	4.33	3792.0	14.8
21.13	4.20	4970.0	19.5
21.48	4.13	4149.0	16.2
22.14	4.01	3294.0	12.9
23.45	3.79	3414.0	13.4
24.75	3.59	2742.0	10.7
25.14	3.54	3093.0	12.1



**Cholesterol (continued)**

Angle (2 $\theta$ )	d-value (Å)	Intensity count	% Intensity
26.20	3.40	3010.0	11.8
28.18	3.16	2032.0	8.0
30.36	2.94	1794.0	7.0
32.56	2.75	1626.0	6.4
33.57	2.67	1504.0	5.9
34.18	2.62	1543.0	6.0
39.79	2.26	1544.0	6.0
42.27	2.14	1707.0	6.7

**SLN10**

Angle (2 $\theta$ )	d-value (Å)	Intensity count	% Intensity
6.08	14.52	30.6	6.4
6.80	12.99	476.0	100.0
11.35	7.79	131.0	27.5
15.92	5.56	40.9	8.6
39.32	2.29	32.3	6.8
41.72	2.16	25.7	5.4

**SLN12**

Angle (2 $\theta$ )	d-value (Å)	Intensity count	% Intensity
6.08	14.53	37.3	8.2
6.80	12.99	457.0	100.0
11.32	7.81	118.0	25.9
15.92	5.56	37.0	8.1
39.39	2.29	30.5	6.7
41.72	2.16	19.4	4.3

**SLN14**

Angle (2 $\theta$ )	d-value (Å)	Intensity count	% Intensity
6.05	14.60	55.9	8.8
6.80	12.99	637.0	100.0
11.32	7.81	202.0	31.6
15.91	5.57	56.1	8.8
39.32	2.29	57.4	9.0
41.72	2.16	29.4	4.6

**Appendix E**  
***In Vitro* Cytotoxicity Test**

**%Cell viability of HeLa cells at varying concentrations of SLN (mean±SD)**

Concentration of SLN ( $\mu\text{g/ml}$ )	SLN13	SLN14	SLN15
100	98.03±1.42	98.13±1.88	95.50±3.93
200	74.33±3.87	77.52±6.60	73.54±5.09
300	12.51±2.44	14.03±1.13	13.04±1.00
400	8.38±1.06	9.00±0.90	8.52±0.73
500	1.20±0.14	1.12±0.14	1.31±0.38
1,000	0.86±0.08	0.99±0.11	1.01±0.09
5,000	0.63±0.05	0.86±0.23	0.83±0.10
10,000	0.44±0.07	0.84±0.13	0.84±0.10

**%Cell viability of HeLa cells at varying concentrations of DDAB (mean±SD)**

Concentration of DDAB (concentration of SLN) ( $\mu\text{g/ml}$ )	% Cell viability
0.64 (100)	99.13±2.22
1.28 (200)	80.13±6.27
1.92 (300)	44.54±4.93
2.56 (400)	29.48±7.03
3.2 (500)	25.33±3.63
6.4 (1,000)	20.08±5.31
32 (5,000)	9.50±2.75
64 (10,000)	5.02±1.37

**%Cell viability of HeLa cells at varying concentrations of CSN (mean±SD)**

Concentration of CSN ( $\mu\text{g/ml}$ )	% Cell viability
100	100.62±6.72
500	100.60±5.00
1,000	98.83±4.69
5,000	98.54±4.19
10,000	97.36±6.01



**Appendix F**  
**Mice Immunization Study**

**Mean IgG titers against gag protein in mice serum determined at day 6 after the second and the third pHIS-HIV-hugag immunization (mean±SD)**

Formulation	2 <sup>nd</sup> immunization		3 <sup>rd</sup> immunization	
	Intradermal injection	Topical application	Intradermal injection	Topical application
DOPE-SLN-pHIS-HIV-hugag	700±273.86	82±40.25	900±223.61	340±219.09
Chol-SLN-pHIS-HIV-hugag	520±319.37	64±49.30	700±273.86	180±178.89
CSN-pHIS-HIV-hugag	260±219.09	46±49.30	320±249.00	80±44.7
Naked-pHIS-HIV-hugag	28±40.25	8±4.47	82±40.25	26±41.59

## VITA

Mr. Rathapon Asasutjarit was born on 10<sup>th</sup> December 1973, in Saraburi, Thailand. He got a Bachelor Degree of Science in Pharmacy (1<sup>st</sup> Class Honours) in 1998 and a Master Degree of Science in Pharmacy in 2002 from Faculty of Pharmaceutical Sciences, Chulalongkorn University, Bangkok, Thailand.

He worked as a Pharmacist in Dispensing Unit in 1998 and a Head of Pharmacy Compounding Unit, Department of Pharmacy, Thammasart Chalermprakit Hospital, Thammasart University in 1999-2004. He has been working as a Lecturer in Department of Pharmaceutical Technology, Faculty of Pharmacy, Rangsit University since 2004.