

REFERENCES

- [1] Lim, S. H.; and Hudson, S. M. Synthesis and antimicrobial activity of a water soluble chitosan derivative with a fiber reactive group. Carbohydrate Research 339 (2004): 313-319.
- [2] Kim, T. H.; Jiang, H. L.; Jere, D.; Park, I. K.; Cho, M. H.; Hah, J. W.; Choi, Y. J.; Akaike, T.; and Cho, C. S. Chemical modification of chitosan as gene carrier in vitro and in vivo. Progress in Polymer Science 32 (2007): 726-753.
- [3] Kean, T.; Roth, S.; and Thanou, M. Trimethylated chitosans as non-viral gene delivery vectors: cytotoxicity and transfection efficiency. Journal of Controlled Release 103 (2005): 643-653.
- [4] SayedSweet, Y.; Hedstrand, D. M.; Spider, R.; and Tomalia, D. A. Hydrophobically modified poly (amidoamine) (PAMAM) dendrimers: their properties at the air-water interface and use as nanoscopic container molecules. Journal of Material Chemistry 7 (1997): 1199-1205.
- [5] Majoros, I. J.; Williams, C. R.; and Baker, J. R. Current dendrimer applications in cancer diagnosis and therapy. Current Topics in Medicinal Chemistry 8 (2008): 1165-1179.
- [6] Marty, J. D.; Martinez-Aripe, E.; Mingotaud, A. F.; and Mingotaud, C. Hyperbranched polyamidoamine as stabilizer for catalytically active nanoparticles in water. Journal of Colloid and Interface Science 326 (2008): 51-54.
- [7] Vogtle, F.; Gestermann, S.; Hesse, R.; Schwierz, H.; and Windisch, B. Functional dendrimers. Progress in Polymer Science 25 (2000): 987-1041.
- [8] Chen, C. Z.; Beck-Tan, N. C.; Dhurjati, P.; Dyk, T. K.; LaRossa, R. A.; and Cooper, S. L. Quaternary ammonium functionalized poly(propylene imine) dendrimers as effective antimicrobials : structure-activity studies. Biomacromolecules 1 (2000): 473-480.
- [9] Punyacharoenon, P.; and Srikulkit, K. Preparation of hyperbranched polyamidoamine polymer ultrafine silica hybrid composite. Journal of Applied Polymer Science 109 (2008): 3230-3237.
- [10] Tsubokawa, N.; and Takayama, T. Surface modification of chitosan powder by grafting of 'dendrimer like' hyperbranched polymer onto the surface. Reactive & Functional Polymers 43 (2000): 341-350.

- [11] Purwar, R.; and Joshi, M. Recent developments in antimicrobial finishing of textiles-a review. AATCC Review 4 (2004): 22-26.
- [12] Thiry, M.C.; and Small game hunting: Antimicrobials take the field. AATCC Review, 1 (11) (2001): 11-17.
- [13] Gao, Y.; and Cranston, R. Recent advances in antimicrobial treatments of textiles. Textile Research Journal 78 (2008): 60-72.
- [14] The 2nd European conference on textiles and the skin, Bönningheim, Germany 2004 [Online]. Available from http://www.hohenstein.com.tr/ximages/15676_hohtextile.pdf [2007, June]
- [15] Pavlidou, V., New multifunctional textiles: antimicrobial treatments, in “proceedings of the intelligent textile structures - application, production and testing international workshop” Thessaloniki, Greece 2005 [Online]. Available from <http://centrum.vslib.cz/centrum/itsapt/greece2005.html> [2007, June]
- [16] Antimicrobial fabrics help fight war against germs [Online]. Available from <http://www.textilesintelligence.com/til/press.cfm?priid=325> [2007, June]
- [17] Heywood, D. Textile Finishing. UK: Society of Dyers and Colourists, 2003.
- [18] Siu R. G. H. Microbial Decomposition of Cellulose. New York: Reinhold, 1951.
- [19] Evans E. T.; Wales D. S.; Bratt R. P.; and Sagar B. F. Investigation of an endoglucanase essential for the action of the cellulase system of *Trichoderma reesei* on crystalline cellulose. Journal of General Micro-biology. 138 (1992): 1639–1646.
- [20] Heine E.; Knops H.G.; Schaefer K.; Vangeyte P.; and Moeller M. Multifunctional Barriers for Flexible Structure. Springer-Verlag GmbH, 2007.
- [21] Kim, Y. H.; Choi, H. M.; and Yoon, J.H. Synthesis of a quaternary ammonium derivative of chitosan and its application to a cotton. Textile Research Journal 68 (1998): 428-434.
- [22] Kanazawa, A.; Ikeda, T.; and Endo, T. Polymeric phosphonium salts as a novel class of cationic biocides. VI. Antibacterial activity of fibers surface-treated with phosphonium salts containing trimethoxysilane groups Antimicrobial Finish. Journal of Applied Polymer Science 52 (1994): 641-647.
- [23] Oktem, T. Surface treatment of cotton fabrics with chitosan. Coloration Technology 119 (2003): 241-246.

- [24] Lin, J.; Qiu, S.; Lewis, K.; and Klibanov, A. Mechanism of bactericidal and fungicidal activities of textiles covalently modified with alkylated polyethylenimine. Biotechnology and Bioengineering 83 (2003): 168-172.
- [25] Lin, J.; Murthy, S.; Olsen, B.; Gleason, K.; and Klibanov, A. Making thin polymeric materials, including fabrics, microbicidal and also water-repellent. Biotechnology Letters 25 (2003): 1661-1665.
- [26] Risbud, M.; Karamuk, E.; and Mayer, J. Designing hydrogel coated textile scaffolds for tissue engineering: effect of casting conditions and degradation behavior studied at microstructure level. Journal of Materials Science Letters 21 (2002): 1191-1194.
- [27] Wales, D. S.; Hamlyn, P. F.; Todd, J. E.; and Sagar, B. F. Preservation treatments of textile materials and their assessment, Society for Applied Bacteriology Technical Series No. 22. Oxford, England: Blackwell Scientific Publications, 1987.
- [28] McCarthy, B. J.; Preservatives for use in the wool textile industry, Society for Applied Bacteriology Technical Series No. 22. Oxford, England : Blackwell Scientific Publications, 1987.
- [29] McCarthy, B. J.; Bioluminescent assay of microbial contaminants on textile materials. International Biodeterioration Bulletin (1983): 53-57.
- [30] McDonnell, G.; and Russell, A. D. Antiseptics and disinfectants: activity, action, and resistance. Clinical Microbiology Reviews 12 (1999): 147-179.
- [31] Cupron, Inc. Cupron-News-Antimicrobial [Online]. Available from <http://www.cupron.com/Cupron-News-Antimicrobial/>. Greensboro, NC [2007, June]
- [32] Yadav, A. et al. Functional finishing in cotton fabrics using zinc oxide nanoparticles. Bulletin of Material Science 29 (2006): 641-645.
- [33] Charbonneaux, T.; and Rochat, S. Articles with antibacterial and antifungal activity. United States Patent Application 20060208390 (2006).
- [34] Antelman, M. S.; High performance cobalt (II,III) oxide antimicrobial textile articles. United States Patent no 6228491 (2001).
- [35] Hermans, M. H. Silver-containing dressings and the need for evidence. American Journal of Nursing 106 (2006): 60-68.

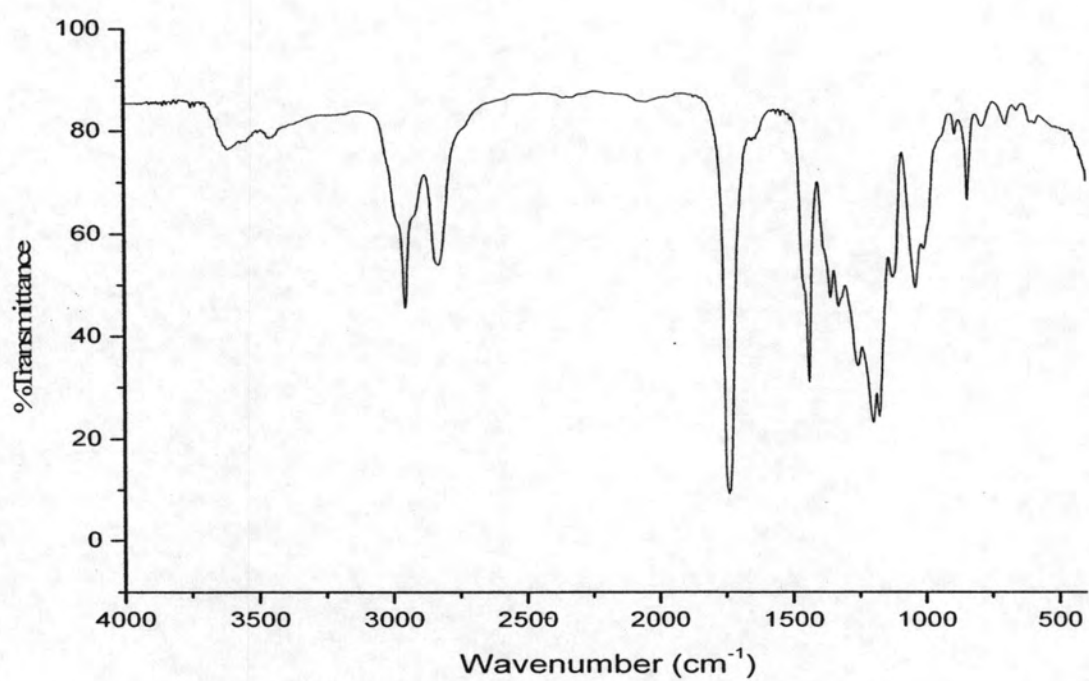
- [36] Butkus, M. A.; Edling, L.; and Labare, M. P. The efficacy of silver as a bactericidal agent: advantages, limitations and considerations for future use. Journal of Water Supply: Research and Technology-AQUA 52 (2003): 407–416.
- [37] Percival, S. L.; Bowler, P. G.; and Russell, D. Bacterial resistance to silver in wound care. Journal of Hospital Infection 60 (2005): 1-7.
- [38] Silver, S.; and Phung le, T.; Silver, G. Silver as biocides in burn and wound dressings and bacterial resistance to silver compounds. Journal of Industrial Microbiology and Biotechnology 33 (2006): 627-634.
- [39] Kim, Y. H.; and Sun, G. Dye molecules as bridges for functional modifications of nylon: antimicrobial functions. Textile Research Journal 70 (2000): 728-733.
- [40] Esfand, R.; and Tomalia, D. A. Poly (amidoamine) (PAMAM) dendrimers: from biomimicry to drug delivery and biomedical applications. Research focus reviews 6 (2001): 427-436.
- [41] Vogtle, F.; Richardt, G.; and Werner, N. Dendrimer Chemistry. Germany: WILEY-VCH Verlag GmbH & Co. KGaA, 2009.
- [42] Esfand, R.; and Tomalia, D. A. Laboratory synthesis of poly (amidoamine) (PAMAM) dendrimers. dendrimers and other dendritic polymers Wiley (2001): 587-604.
- [43] Chen, C. Z.; and Cooper, S.L. Recent advances in antimicrobial dendrimer. Advanced Materials 12 (2000): 843-846.
- [44] Chen, C. Z; and Cooper, S. L. Quaternary ammonium functionalized dendrimers and methods of use therefore. US Patent 64440405 (2002).
- [45] Chen, C. Z; and Cooper, S. L. Interactions between dendrimer biocide and bacterial membranes. Biomaterials 23 (2002): 3359-3368.
- [46] Balogh, L.; Swanson, D. R.; Tomalia, D. A.; Hagnauer, G. L.; and McManus, A. T. Dendrimer-silver complexes and nanocomposites as antimicrobial agents. Nano Letters. 1 (2001): 18-21.
- [47] Khora, E.; Limb, L. Y. Implantable applications of chitin and chitosan Biomaterials 24 (2003): 2339–2349.
- [48] Mourya, V.K.; Inamdar, N. N. Chitosan-modifications and applications: Opportunities galore. Reactive & Functional Polymers 68 (2008): 1013–1051.

- [49] Rabea, I. E.; Mohamed, E.; Badawy, T.; Stevens, C. V.; Smagghe, G.; and Steurbaut, W. Chitosan as antimicrobial agent: applications and mode of action. Biomacromolecules 4 (6) (2003): 1457-1465.
- [50] Lim, S. H.; and Hudson, S. M. Review of chitosan and its derivatives as antimicrobial agents and their uses as textile chemicals. Journal of Macromolecular Science-Polymer Reviews 43 (2003): 223–269.
- [51] No, H. K.; Park, N. Y.; Lee, S. H.; and Meyers, S. P. Antibacterial activity of chitosans and chitosan oligomers with different molecular weights. International Journal of Food Microbiology 74 (2002): 65–72.
- [52] Shin, Y.; Yoo, D. I.; and Jang, J. Molecular weight effect on antimicrobial activity of chitosan treated cotton fabrics. Journal of Applied Polymer Science 80 (2001): 2495–2501.
- [53] Lee, S.; Cho, J. S.; and Cho, G. S. Antimicrobial and blood repellent finishes for cotton and nonwoven fabrics based on chitosan and fluoropolymers. Textile Research Journal 69 (1999): 104–112.
- [54] Chung, Y. S.; Lee, K. K.; and Kim, J. W. Durable press and antimicrobial finishing of cotton fabrics with a citric acid and chitosan treatment. Textile Research Journal 68 (1998): 772–775.
- [55] Julia, M. R.; Cot, M.; Erra, P.; Jovic, D.; and Canal, J. M. The use of chitosan on hydrogen peroxide pretreated wool. Textile Chemist and Colorist 30 (1998): 78–83.
- [56] Pascual, E.; and Julia, M. R. The role of chitosan in wool finishing. Journal of Biotechnology 89 (2001): 289–296.
- [57] Roberts, G. A. F.; and Wood, F. A. A study of the influence of structure on the effectiveness of chitosan as an anti-felting treatment for wool. Journal of Biotechnology 89 (2001): 297–304.
- [58] Rybicki, E.; Filipowska, B.; and Walawska, A. Application of natural biopolymers in shrink-proofing of wool. Fibres and Textiles in Eastern Europe 8 (2000): 62–65.
- [69] Onar, N.; and Sariisik, M. Application of enzymes and chitosan biopolymer to the antifelting finishing process. Journal of Applied Polymer Science 93 (2004): 2903–2908.

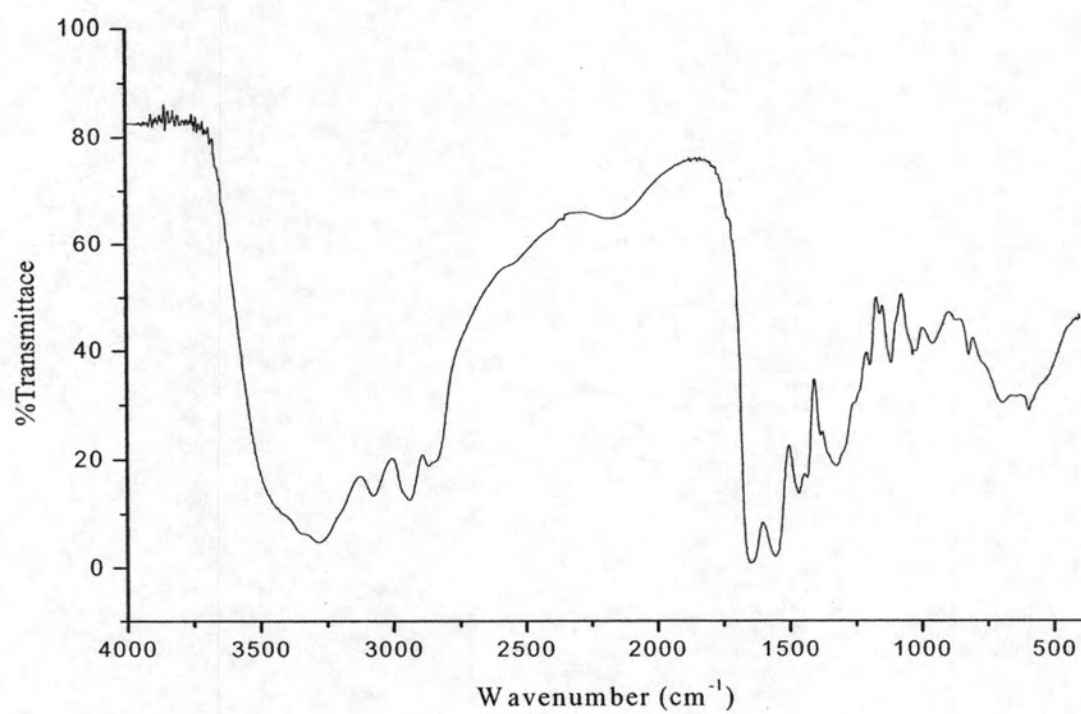
- [60] Erra, P.; Molina, R.; Jovic, D.; Julia, M. R.; Cuesta, A.; and Tascon, J. M. D. Shrinkage properties of wool treated with low temperature plasma and chitosan biopolymer. Textile Research Journal 69 (1999): 811–815.
- [61] Hsieh, S. H.; Huang, Z. K.; Huang, Z. Z.; and Tseng, Z. S. Antimicrobial and physical properties of woolen fabrics cured with citric acid and chitosan. Journal of Applied Polymer Science 94 (2004): 1999–2007.
- [62] Jeong, Y. J. et al. Changes in the Mechanical Properties of Chitosan-treated wool fabric. Textile Research Journal 72 (2002): 70–76.
- [63] Seong, H. S.; Whang, H. S.; and Ko, S. W. Synthesis of a quaternary ammonium derivative of chito-oligosaccharide as antimicrobial agent for cellulosic fibers. Journal of Applied Polymer Science 76 (2000): 2009–2015.
- [64] Kim, J. Y.; Lee, J. K.; Lee, T. S.; and Park, W. H. Synthesis of chitooligosaccharide derivative with quaternary ammonium group and its antimicrobial activity against streptococcus mutans. International Journal of Biological Macromolecules 32 (2003): 23–27.
- [65] Kim, Y. H.; Choi, H. M.; and Yoon, J. H. Synthesis of a quaternary ammonium derivative of chitosan and its application to a cotton antimicrobial finish. Textile Research Journal 68 (1998): 428–434.
- [66] Kim, Y. H.; Nam, C. W.; Choi, J. W.; and Jang, J. H. Durable antimicrobial treatment of cotton fabrics using N-(2-hydroxy)propyl-3-trimethylammonium chitosan chloride and polycarboxylic acids. Journal of Applied Polymer Science 88 (2003): 1567–1572.
- [67] Montazer, M.; and Afjeh, M. G. Simultaneous x-linking and antimicrobial finishing of cotton fabric. Journal of Applied Polymer Science 103 (2007): 178–185.
- [68] Suzuki, K.; Oda, D.; Shinobu, T.; Saimoto, H.; and Shigemasa, Y. New selectively N-substituted quaternary ammonium chitosan derivatives. Polymer Journal 32 (2000): 334–338.
- [69] Kenawy, E. R.; Abdel-Hay, F. I.; Abou El-Magd, A.; and Mahmoud, Y. Biologically active polymers: modification and anti-microbial activity of chitosan derivatives. Journal of Bioactive and Compatible Polymer 20 (2005): 95–111.
- [70] Liang, C. Anti-microbial chitosan composition for textile products. United States Patent Application no 20060008515 (2005).

- [71] Joerger, M. C.; Koniz, R. F.; Sabesan, S.; and Pennias, J. Antimicrobial polyester-containing articles and process for their preparation. United States Patent 7081139 (2006).
- [72] Crabyon, a fibre with crab's pulp.
<http://www.swicofil.com/products/055chitosan.html> (accessed June 2007).
- [73] Illarionova, E. L. et al. Fibre, film, and porous materials based on chitosan. Fibre Chemistry 27 (1995): 392–396.
- [74] Rathke, T. D.; and Hudson, S. M. Review of chitin and chitosan as fiber and film formers. Journal Macromolecular Science-Reviews in Macromolecular Chemistry and Physics C34 (1994): 375-437.
- [75] Srisuk, S.; and Srikulkit, K. Properties evaluation of sodium nitrite treated chitosan-cotton fabric. Journal of Metals, Materials and Minerals 18 (2) (2008): 41-45.

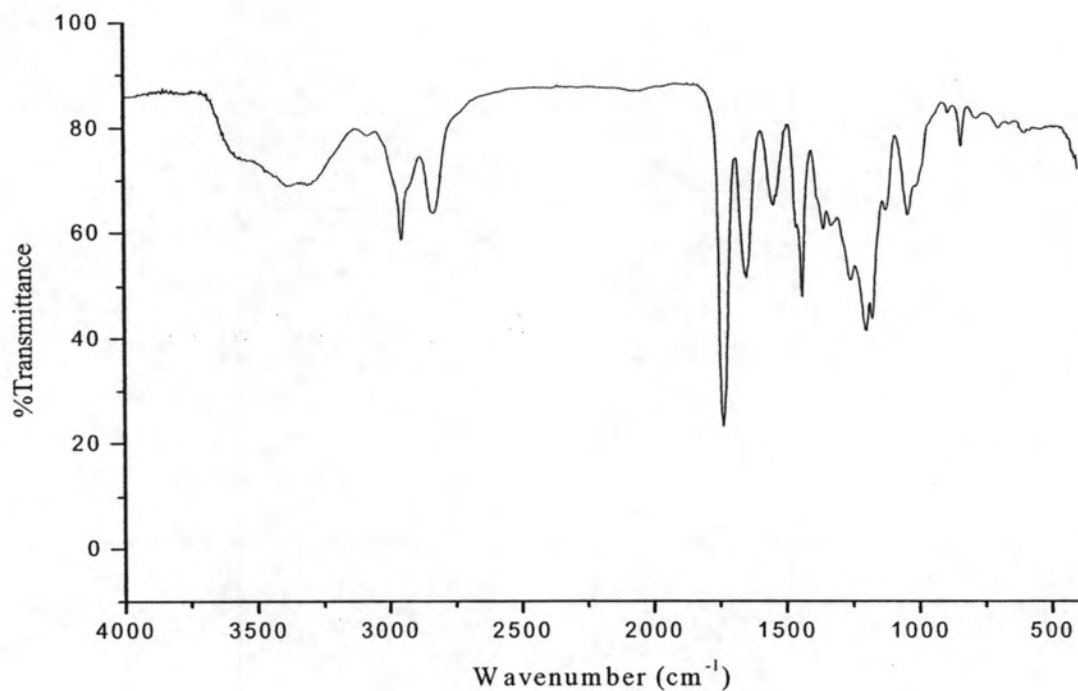
APPENDIX



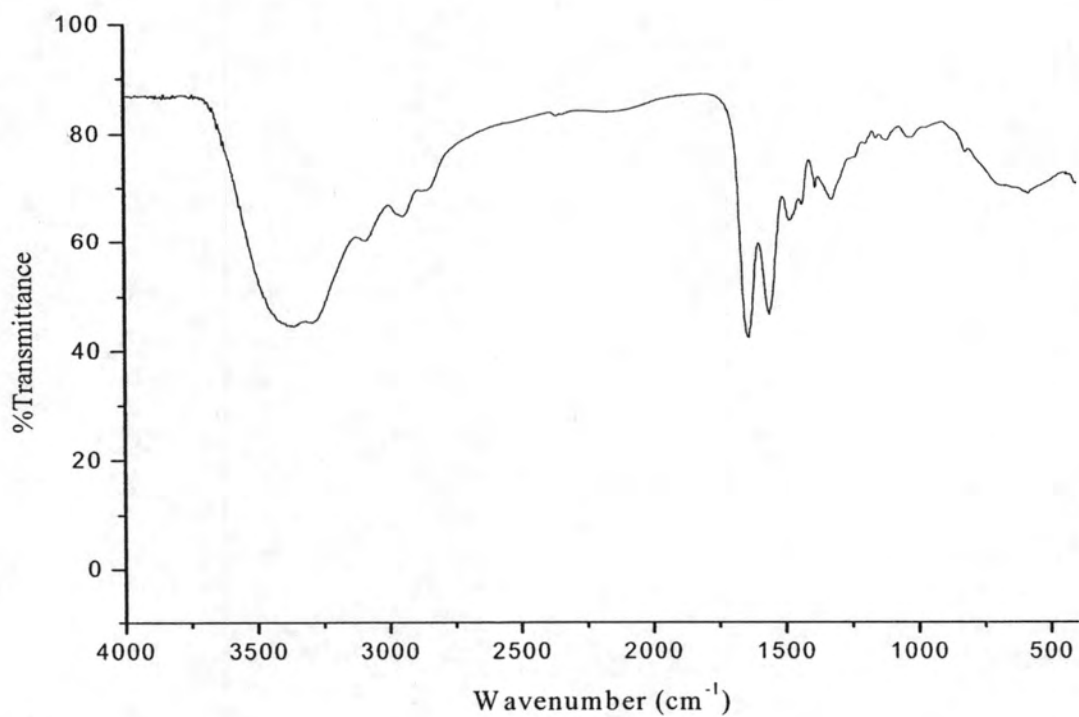
A 1. FTIR spectrum of G-0.5 hyperbranched dendritic PAMAM .



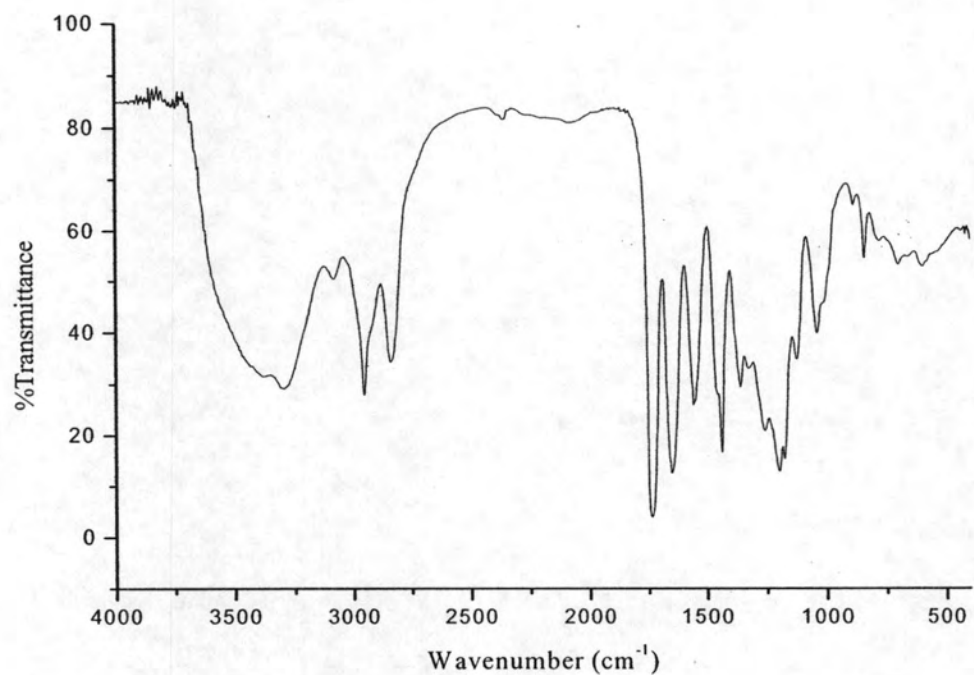
A 2. FTIR spectrum of G0.0 hyperbranched dendritic PAMAM .



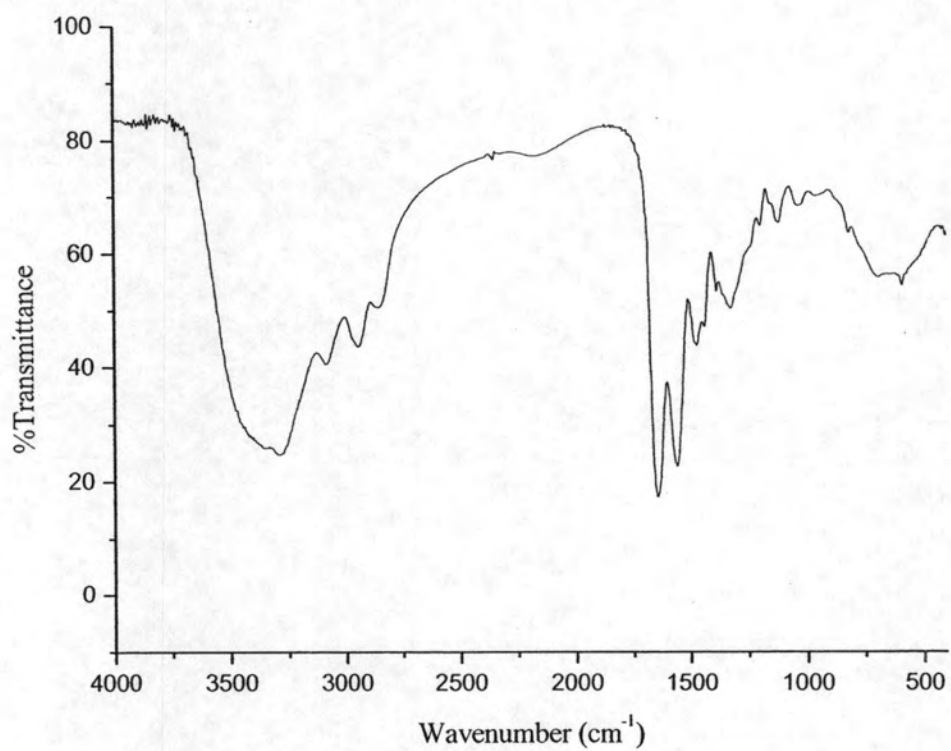
A 3. FTIR spectrum of G0.5 hyperbranched dendritic PAMAM .



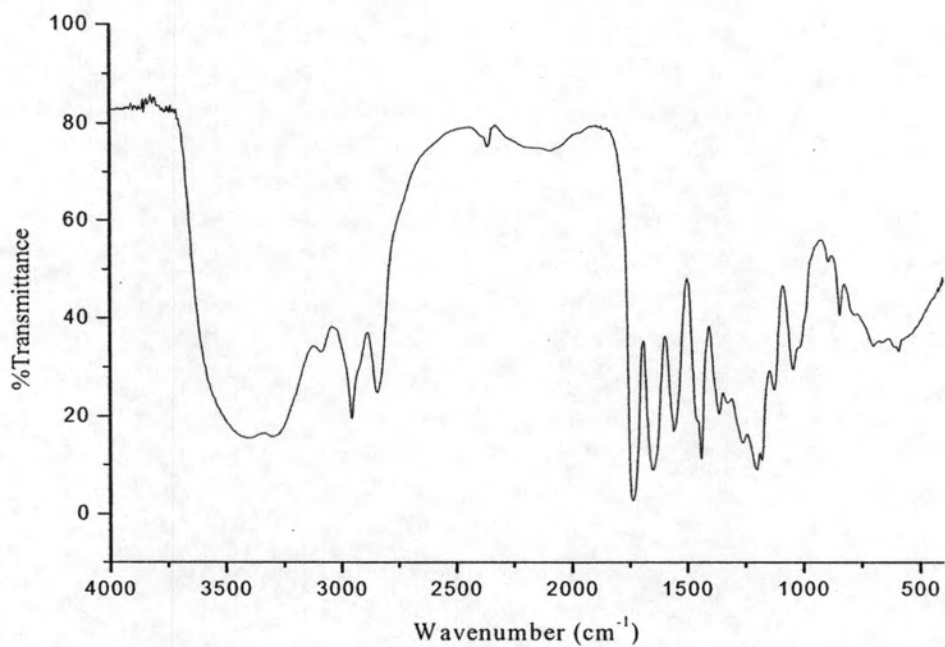
A 4. FTIR spectrum of G1.0 hyperbranched dendritic PAMAM .



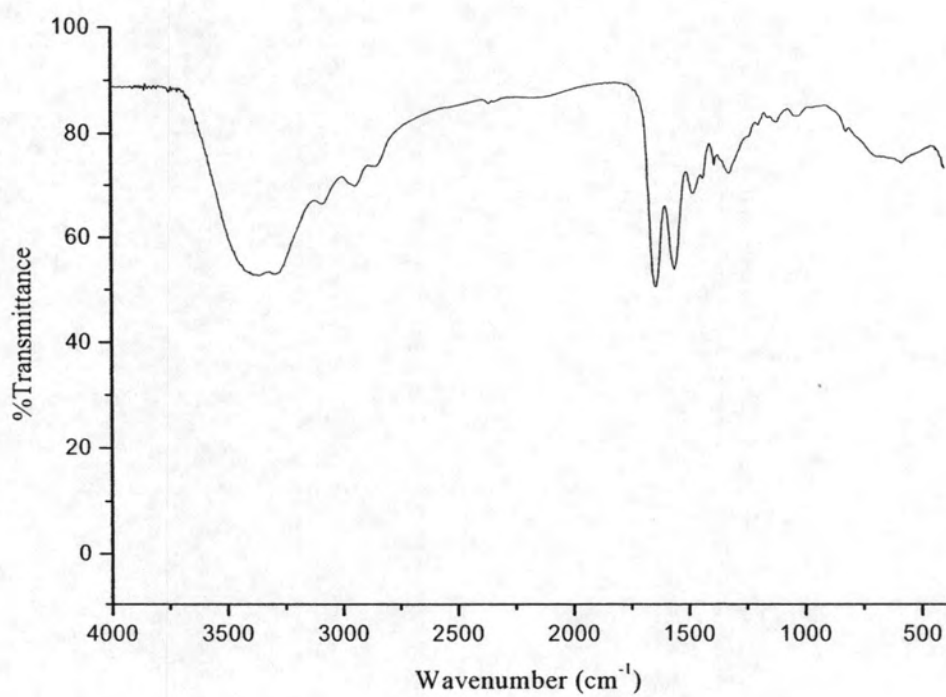
A 5. FTIR spectrum of G1.5 hyperbranched dendritic PAMAM .



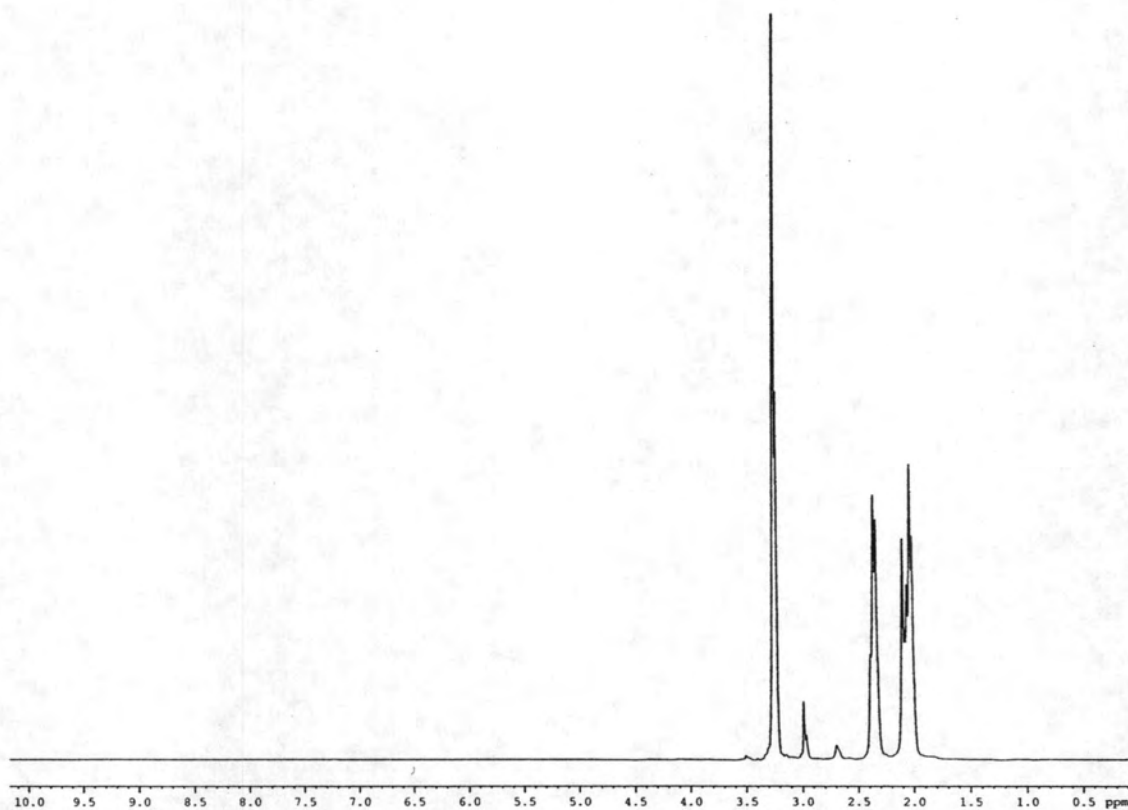
A 6. FTIR spectrum of G2.0 hyperbranched dendritic PAMAM .



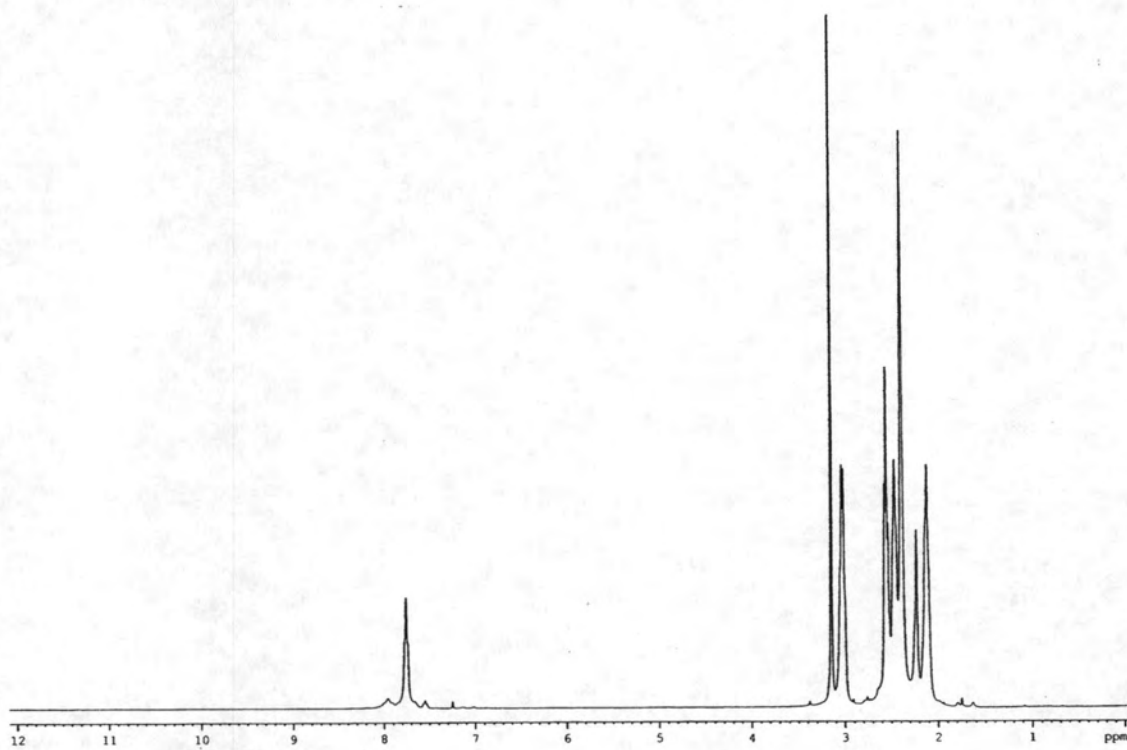
A 7. FTIR spectrum of G2.5 hyperbranched dendritic PAMAM .



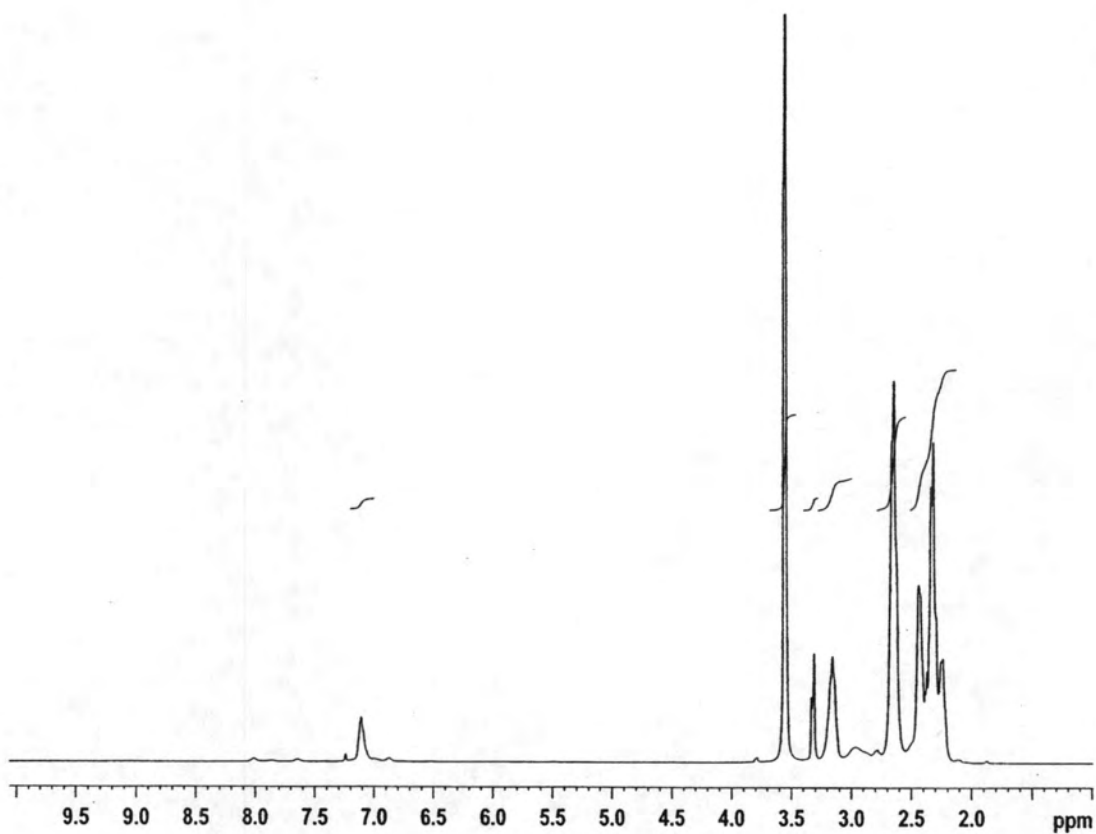
A 8. FTIR spectrum of G3.0 hyperbranched dendritic PAMAM .



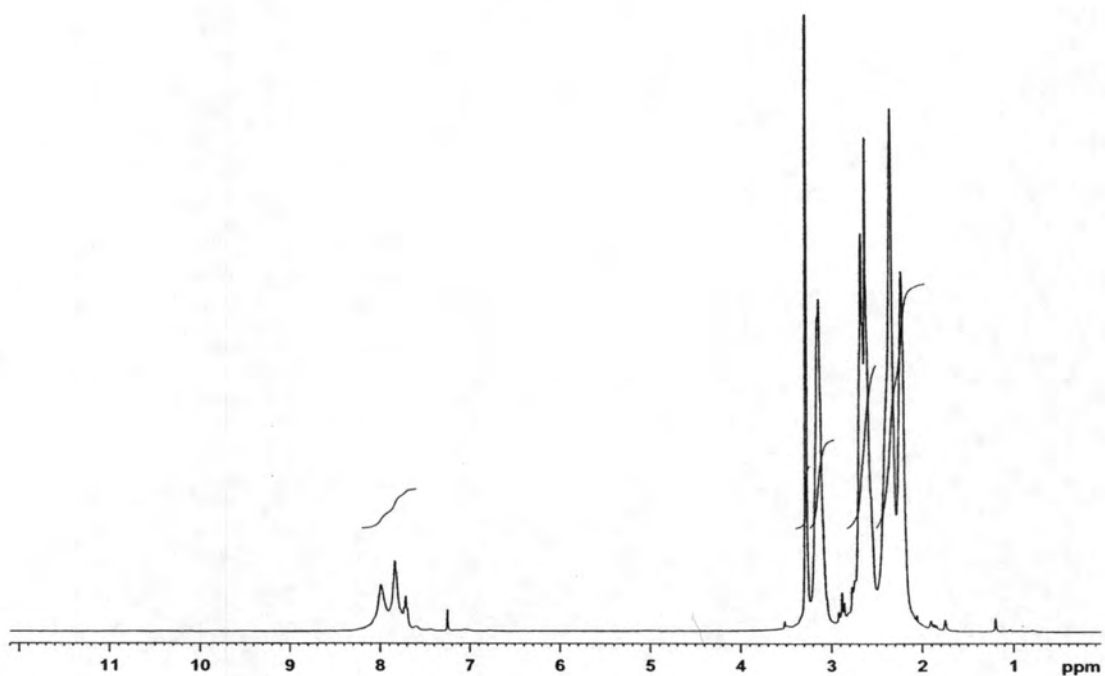
A 9. ^1H NMR spectrum of G-0.5 hyperbranched dendritic PAMAM .



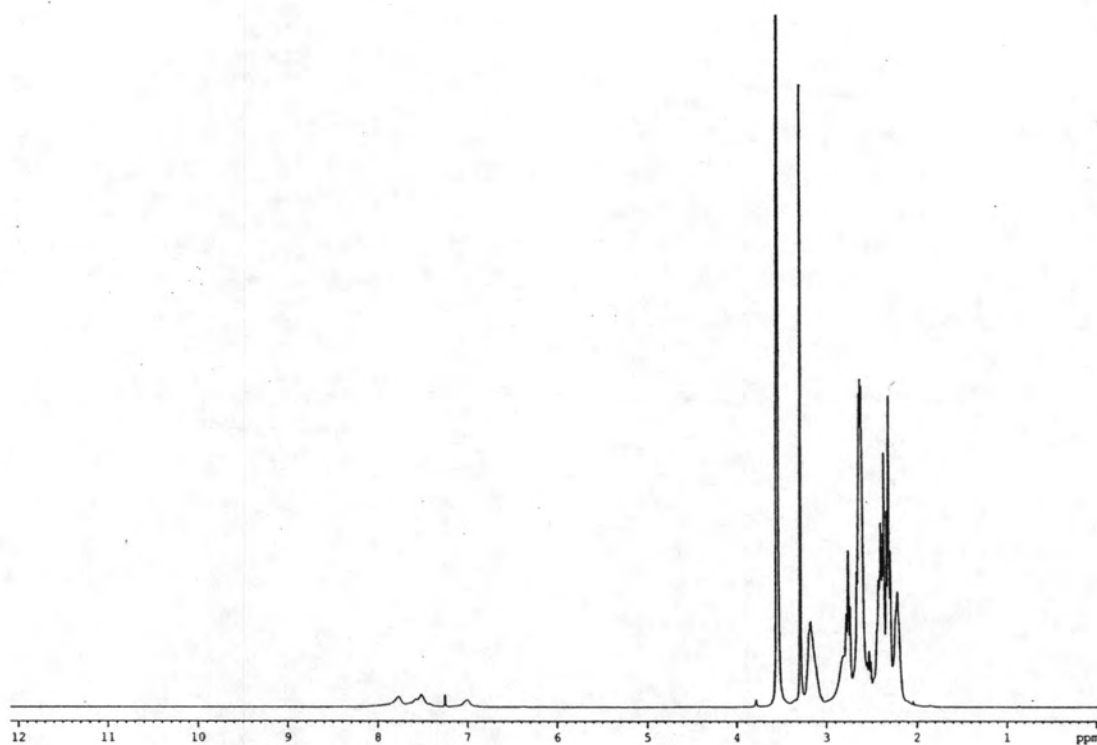
A 10. ^1H NMR spectrum of G0.0 hyperbranched dendritic PAMAM .



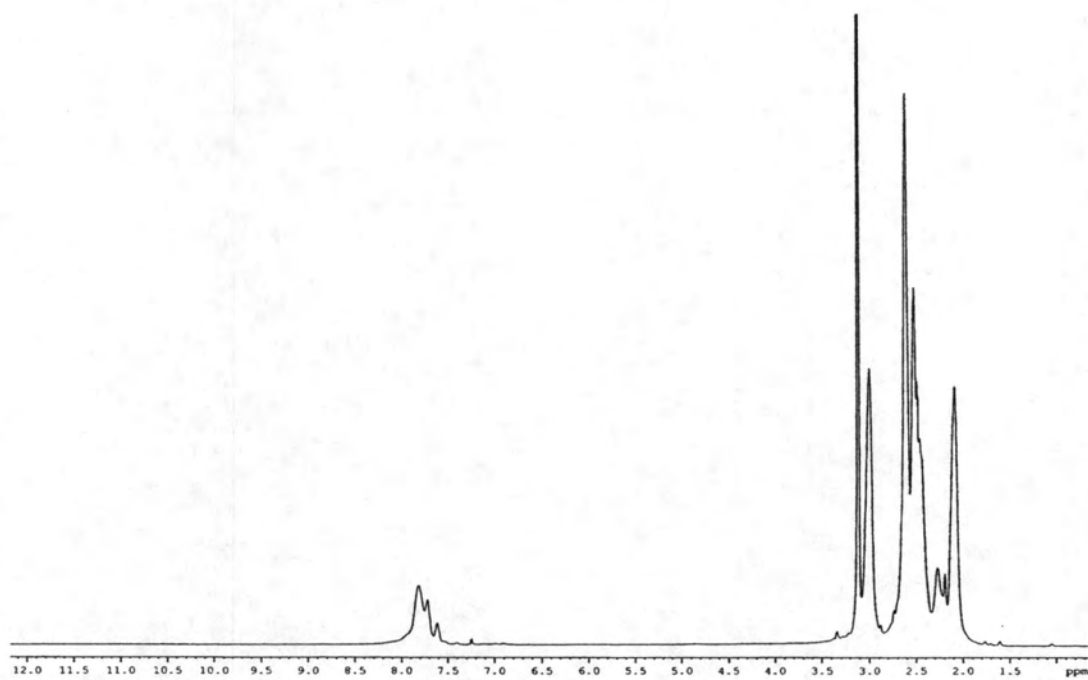
A 11. ^1H NMR spectrum of G0.5 hyperbranched dendritic PAMAM .



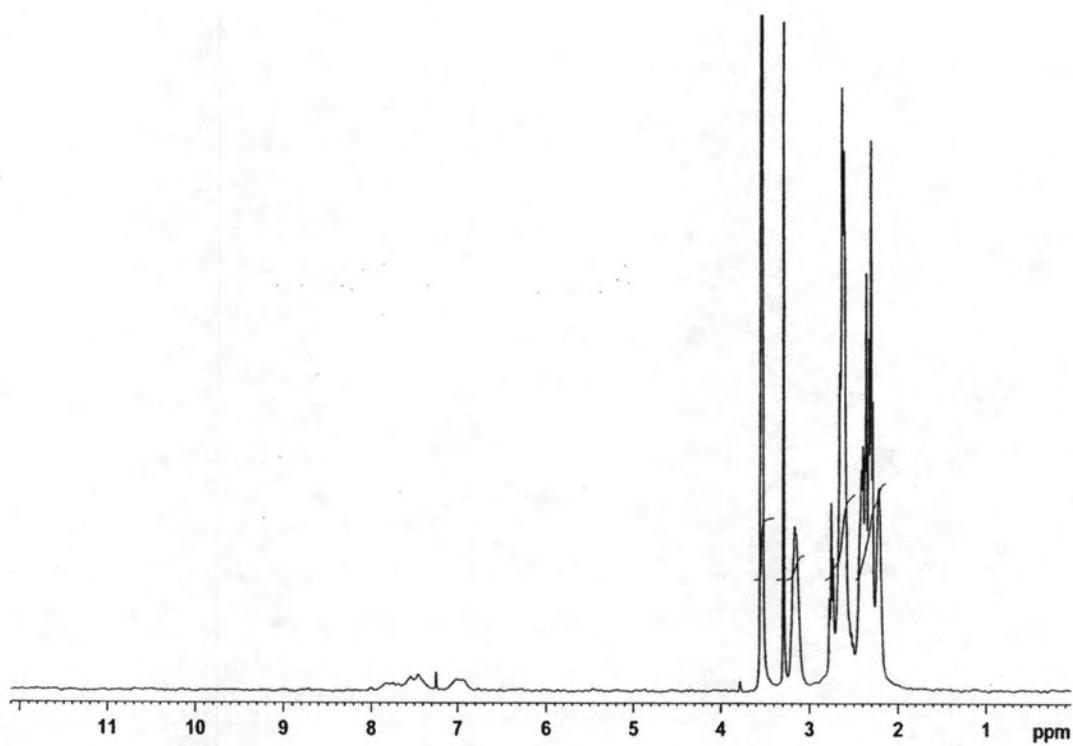
A 12. ^1H NMR spectrum of G1.0 hyperbranched dendritic PAMAM .



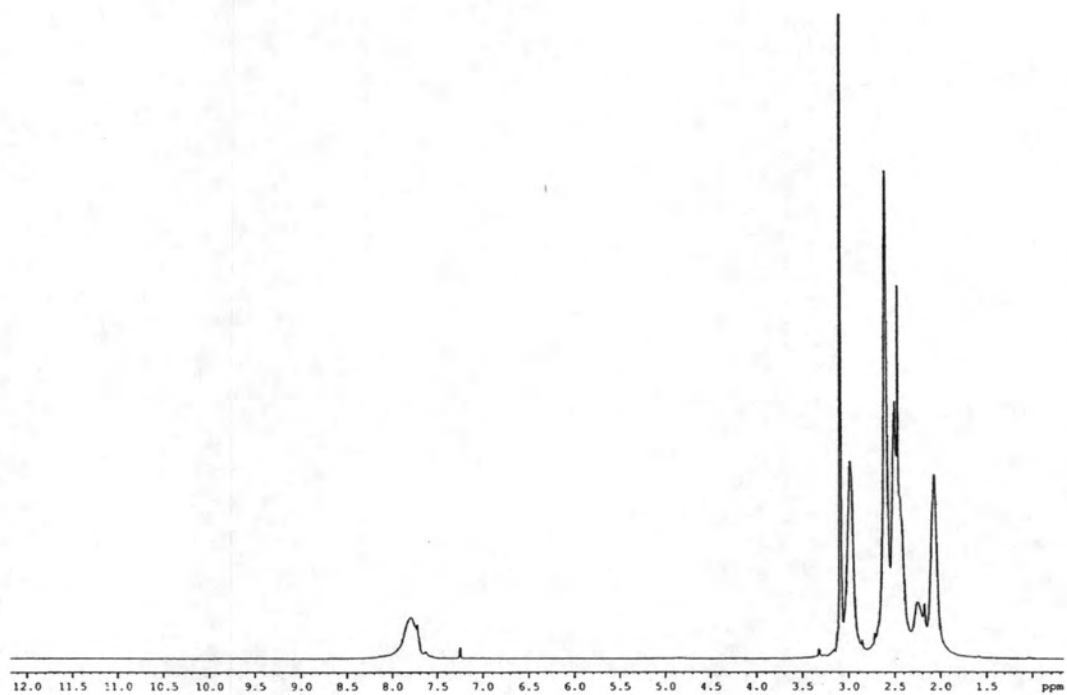
A 13. ^1H NMR spectrum of G1.5 hyperbranched dendritic PAMAM .



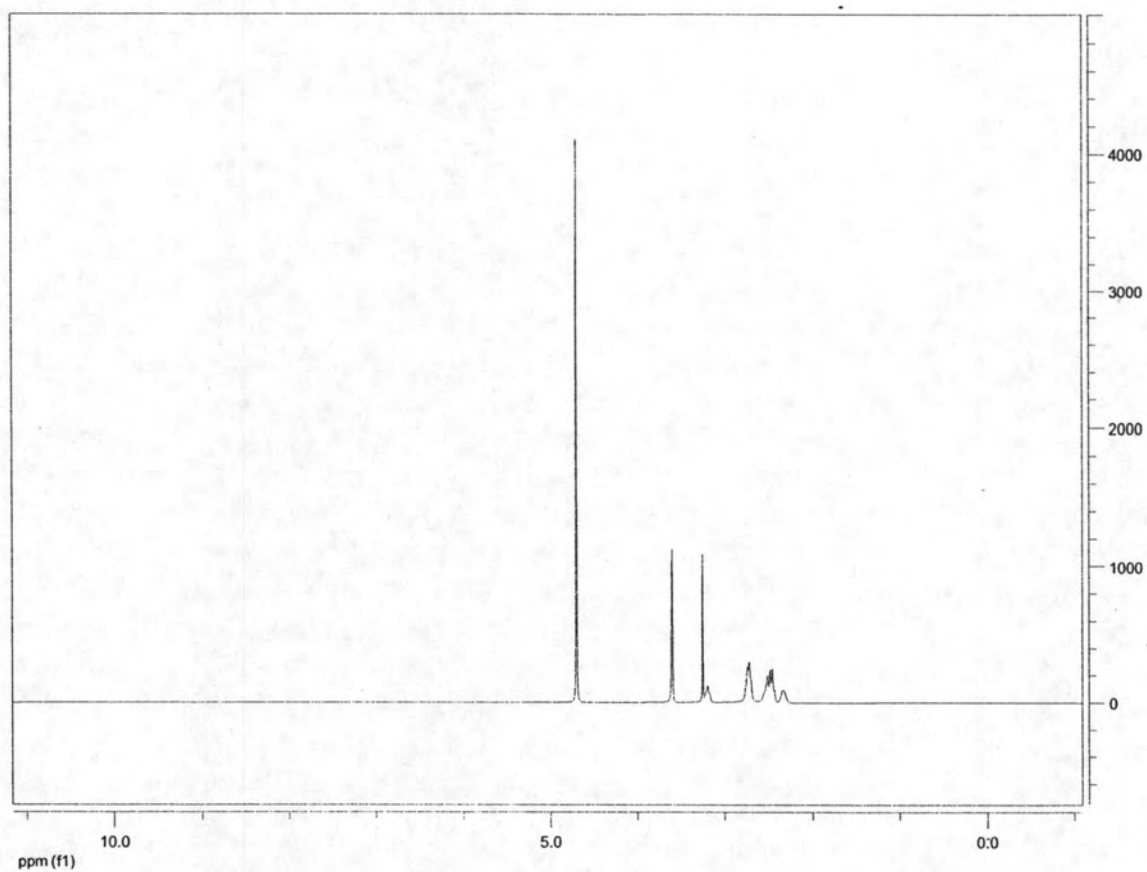
A 14. ^1H NMR spectrum of G2.0 hyperbranched dendritic PAMAM .



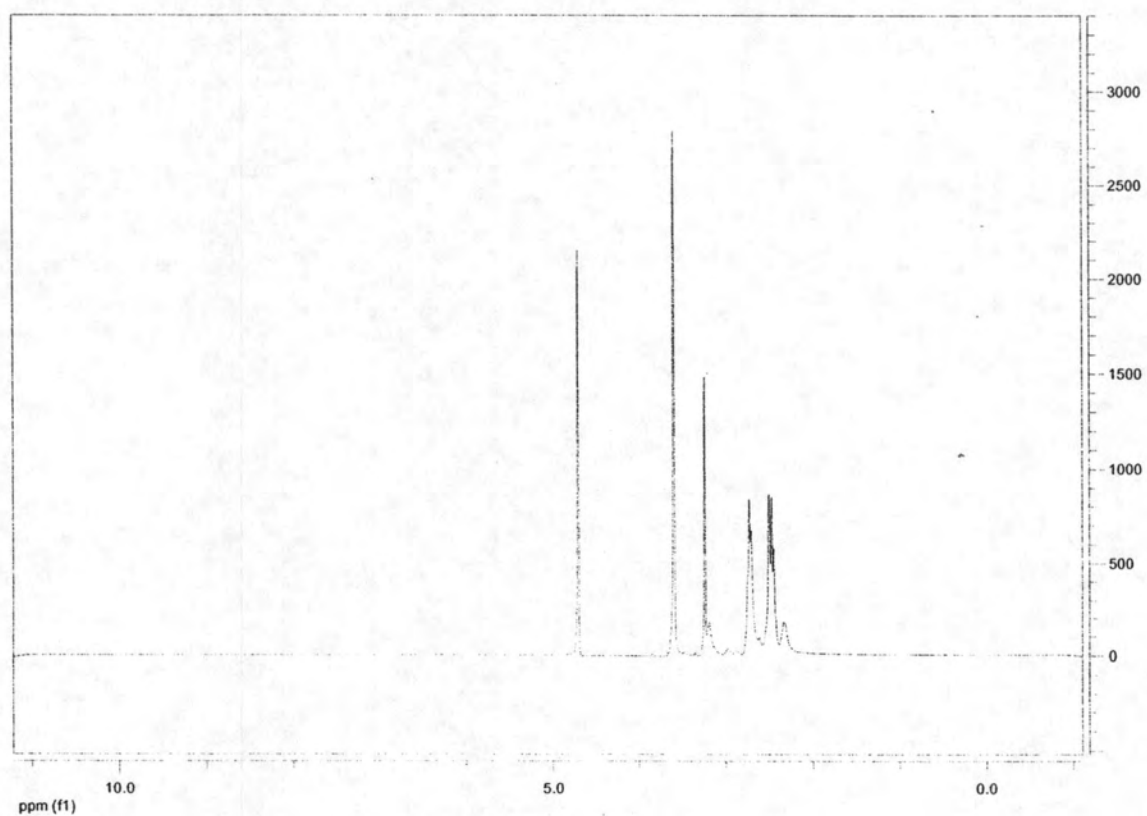
A 15. ^1H NMR spectrum of G2.5 hyperbranched dendritic PAMAM .



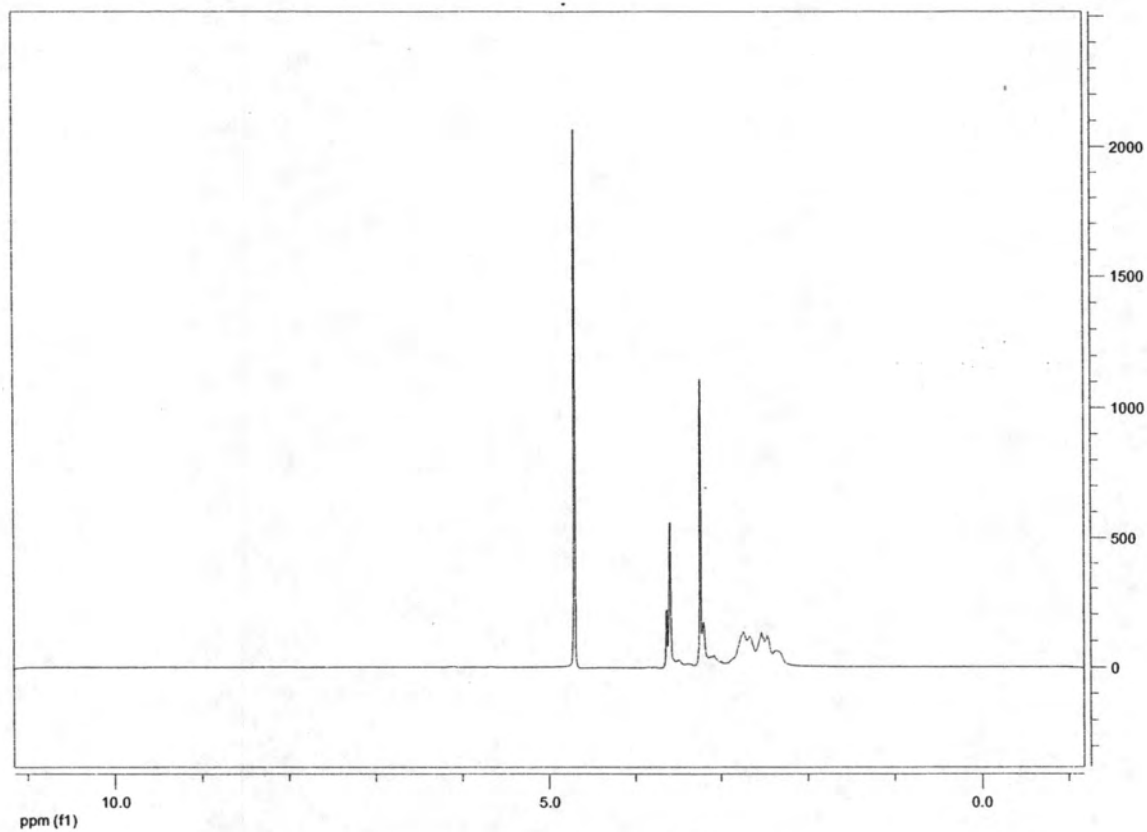
A 16. ^1H NMR spectrum of G3.0 hyperbranched dendritic PAMAM .



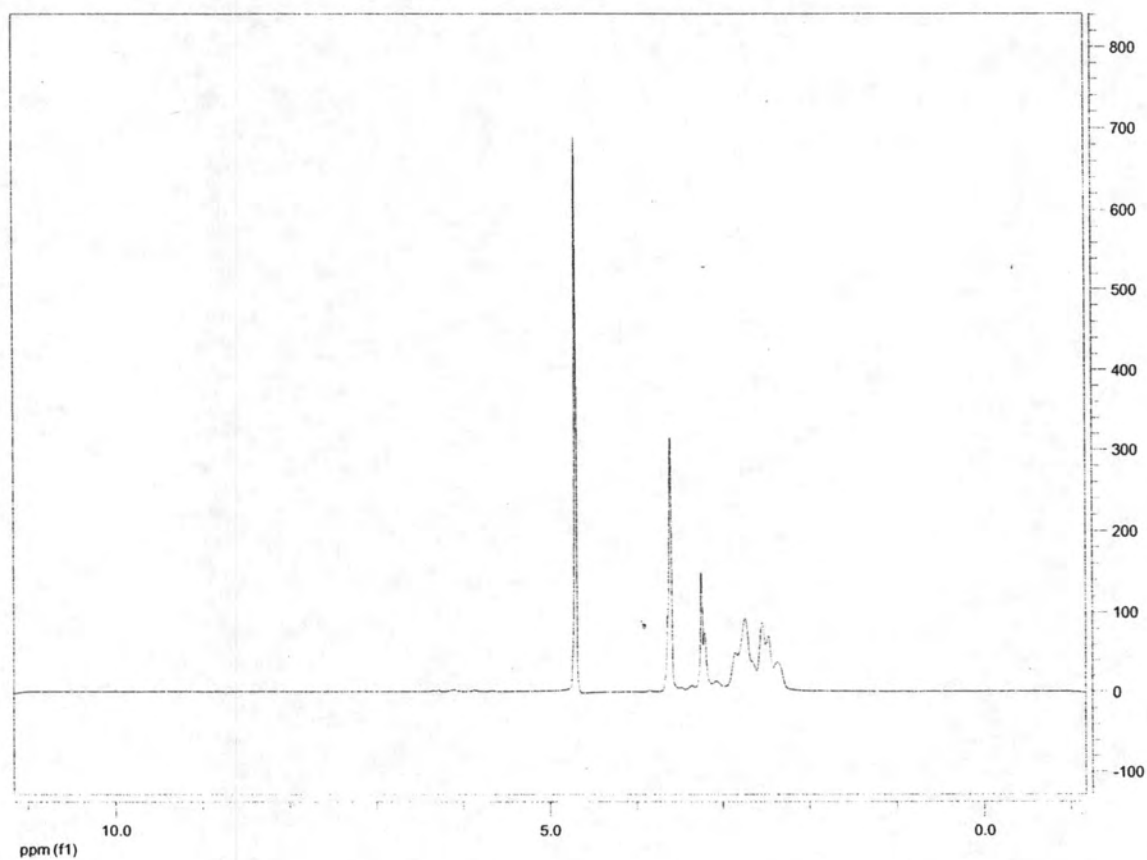
A 17. ^1H NMR spectrum of G3.5 hyperbranched dendritic PAMAM .



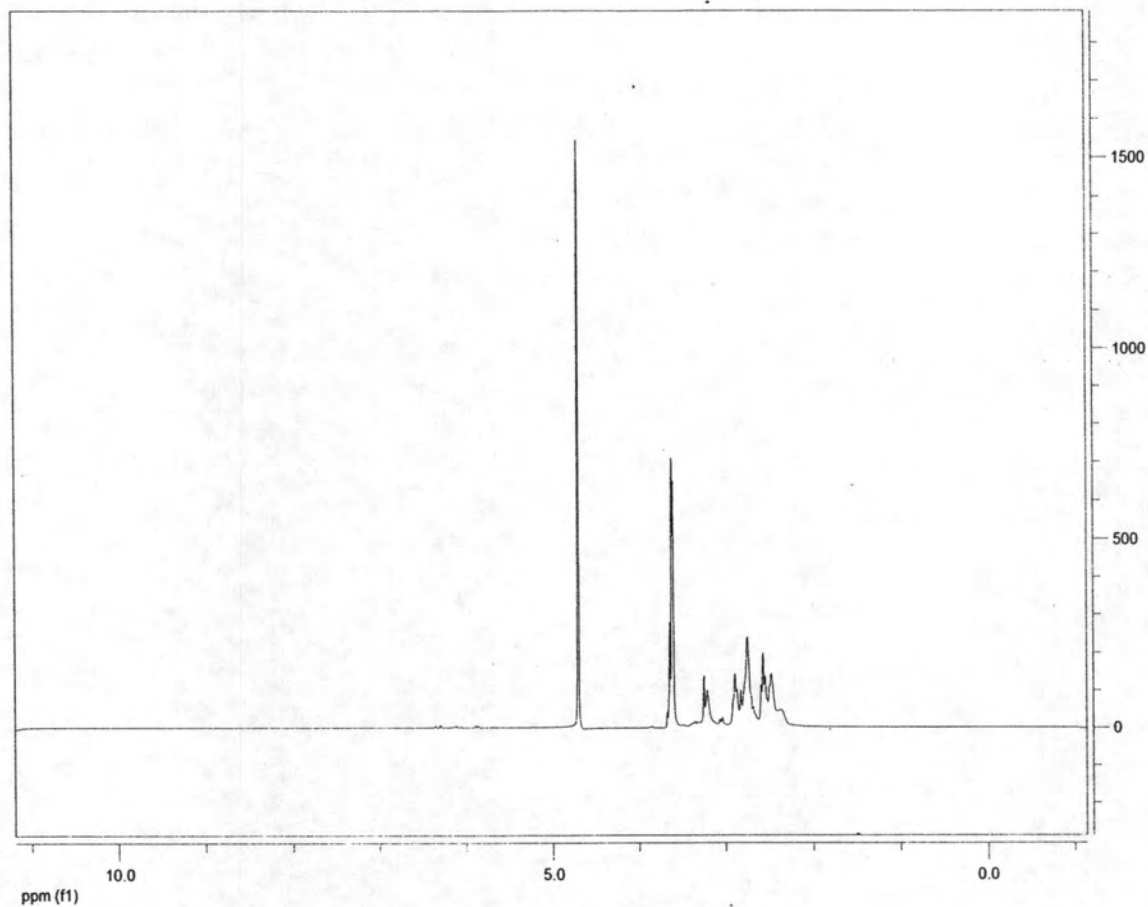
A 18. ^1H NMR spectrum of G4.5 hyperbranched dendritic PAMAM .



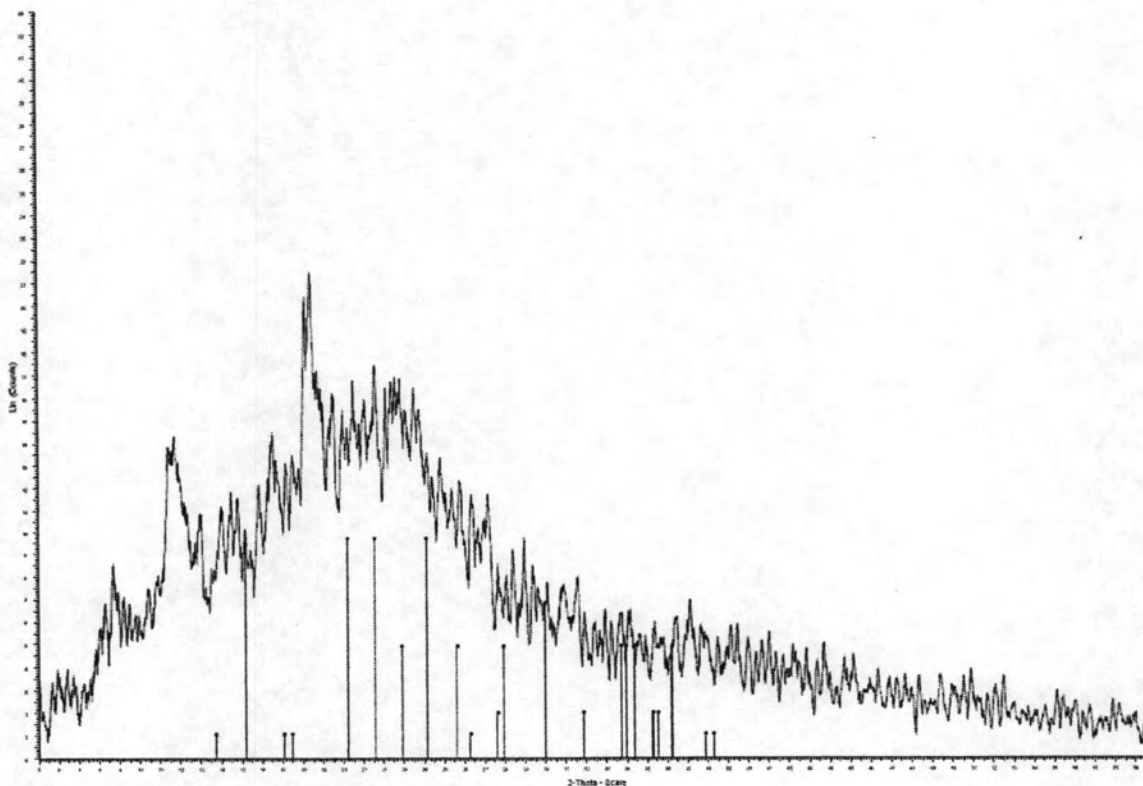
A 19. ^1H NMR spectrum of G2.5 cationic hyperbranched dendritic PAMAM .



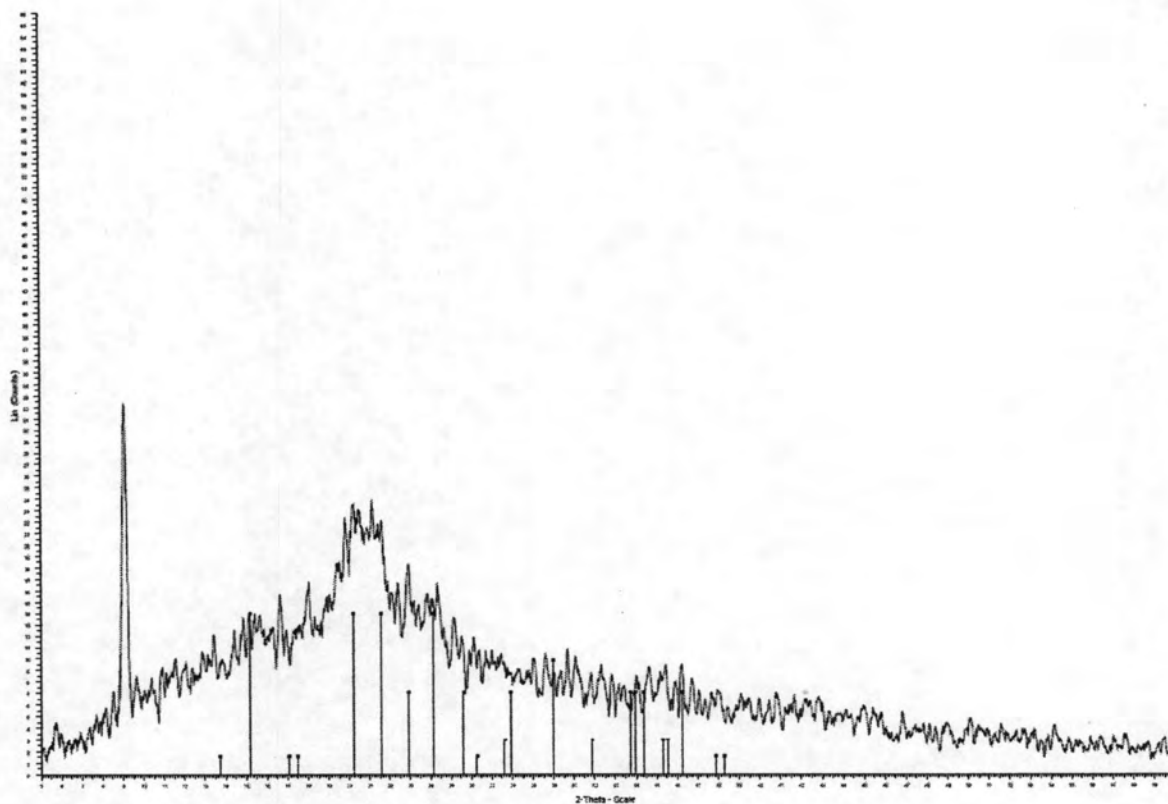
A 20. ^1H NMR spectrum of G3.5 cationic hyperbranched dendritic PAMAM .



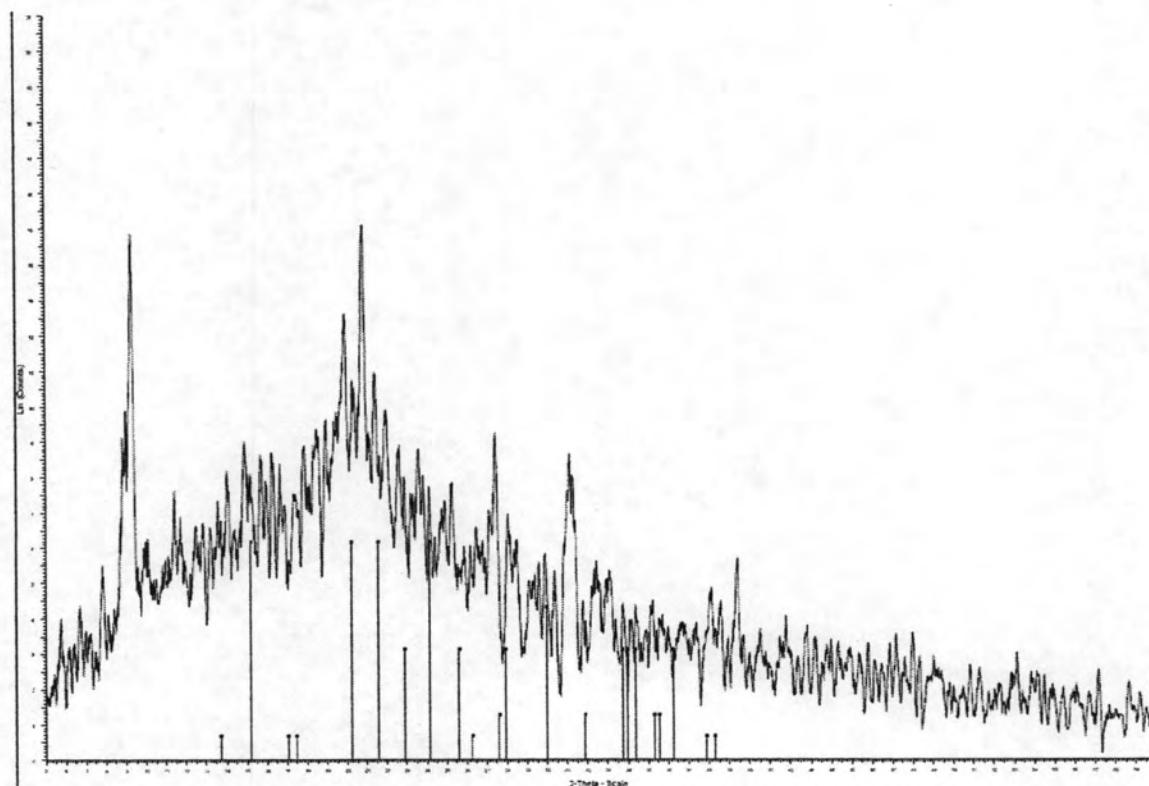
A 21. ^1H NMR spectrum of G4.5 cationic hyperbranched dendritic PAMAM .



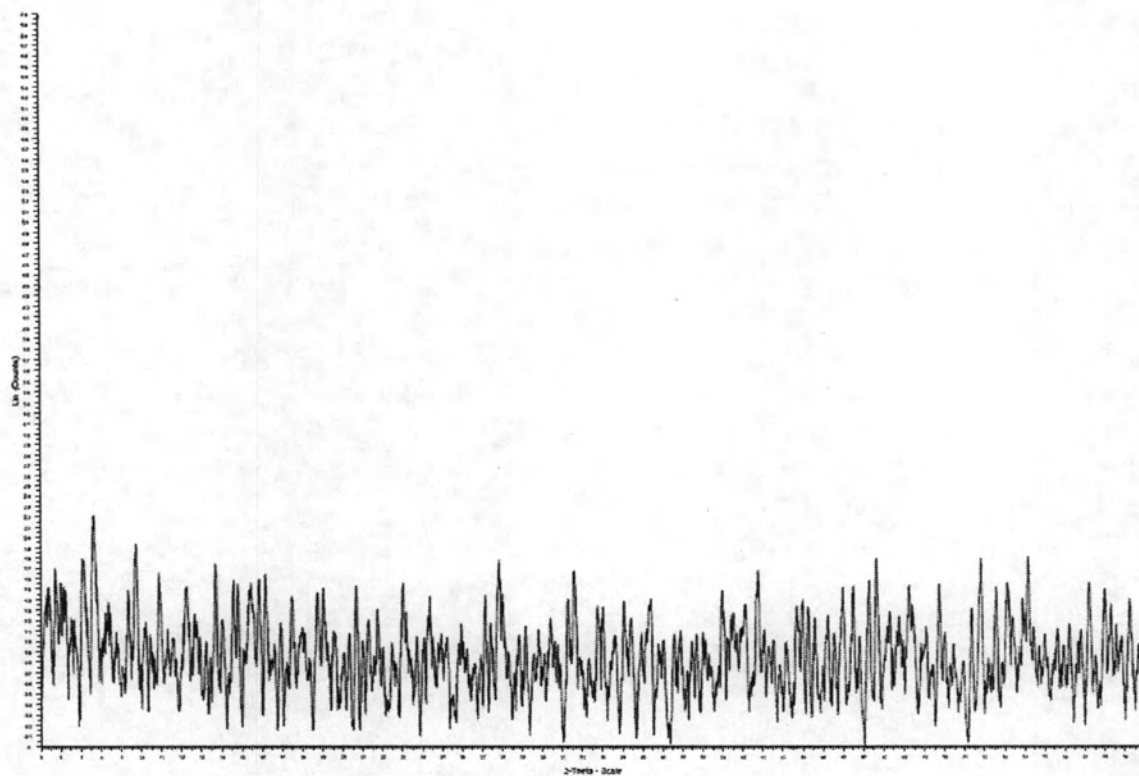
A 22. XRD pattern of Chitosan .



A 23. XRD pattern of 10 wt% PAMAM-CTS.



A 24. XRD pattern of 50 wt% PAMAM-CTS.



A 25. XRD pattern of excessive PAMAM-CTS.

A 26. Antimicrobial activity of cotton fabric treated with cationic hyperbranched dendritic PAMAM

Generation of dendrimer	Dendrimer (% w/v)	The number of bacteria CFU/ml (0 hr.)	The number of bacteria CFU/ml (24 hrs.)	% Reduction
2.5	5	3.8×10^8	8.6×10^7	77.36
	10	4.7×10^8	1.0×10^7	97.87
	15	4.0×10^8	3.5×10^7	91.25
	20	5.8×10^8	5.7×10^7	90.17
3.5	5	5.4×10^8	2.5×10^8	53.70
	10	4.6×10^8	2.6×10^8	43.47
	15	4.2×10^8	1.8×10^8	57.14
	20	5.3×10^8	2.1×10^8	60.37
4.5	5	4.3×10^8	3.4×10^8	20.93
	10	5.0×10^8	3.1×10^8	38.00
	15	3.6×10^8	3.1×10^8	13.88
	20	4.9×10^8	3.1×10^8	36.73

A 27. Antimicrobial activity of cotton fabric treated with chitosan and cationic hyperbranched dendritic polyamidoamine (combined treatment)

Generation of dendrimer	Dendrimer (% w/v)	Chitosan (% w/v)	The number of bacteria CFU/ml (0 hr.)	The number of bacteria CFU/ml (24 hrs.)	% Reduction
2.5	15	0.1	5.3×10^8	1.1×10^8	79.24
		0.5	4.2×10^8	1.0×10^8	76.19
		1.0	6.0×10^8	3.9×10^8	35.0
	20	0.1	1.2×10^8	4.0×10^6	96.66
		0.5	1.2×10^8	3.6×10^6	97.00
		1.0	1.2×10^8	1.5×10^6	98.75
3.5	15	0.1	4.1×10^8	2.1×10^8	48.78
		0.5	4.1×10^8	3.8×10^8	7.31
		1.0	1.9×10^8	1.2×10^8	36.84
	20	0.1	4.1×10^8	1.6×10^8	60.97
		0.5	1.9×10^8	7.1×10^7	62.63
		1.0	3.6×10^8	5.3×10^8	0
4.5	15	0.1	8.5×10^8	3.5×10^8	58.82
		0.5	7.7×10^8	2.1×10^8	72.72
		1.0	7.4×10^8	5.1×10^8	31.08
	20	0.1	8.5×10^8	4.7×10^8	44.70
		0.5	7.1×10^8	4.1×10^8	42.25
		1.0	7.3×10^8	4.8×10^8	34.24

BIOGRAPHY

Miss Benjamas Klaykruayat was born in Phra Nakhon Sri Ayutthaya, Thailand on December 23, 1978 as a daughter of Mr. Nikom Klaykruayat and Mrs. Ubol Klaykruayat. She graduated from Chomsurang Upatham School in March 1997. She received her Bachelor of Science with a major in Materials Science, Chulalongkorn University in 2001 and graduated a Master of Science with a major in Applied Polymer Science and Textile Technology, Chulalongkorn University in 2003. She continued her graduate study in Materials Science Program, Department of Materials Science, Faculty of Science, Chulalongkorn University, and completed her Ph.D. Degree in October 2009.

