

## REFERENCES

- Amirtharajah, A., Denett, K.E. and Stustill, A. 1993. Ferric chloride Coagulation for Removal of Dissolved Organic Matter and Trihalomethane Formation Potential. *Wat. Sci. Tech.* 27(11): 113-121.
- Amy, G.L. and Chadlk, A. 1983. Cationic Polyelectrolytes as Primary Coagulants for Removing Trihalomethane Precursors. *J. AWWA.* 75(10):527-536.
- Amy, G. L., Sierka, R.A., Bedessem, J., Price, D., and Tan, L. 1992. Molecular Size Distribution of Dissolved Organic Matter. *J. AWWA.* 84(6): 67.
- Bell-Ajy, K. 2000. Conventional and Optimized Coagulation for NOM Removal. *J. AWWA.* 92(10): 44-58.
- Bottero, J. 1982. Studies of Hydrolyzed Aluminum Chloride Solutions. The Nature of Aluminum Species and Compositions. *J. Phys. Chem.* 84: 2993.
- Bratby, J. 1980. *Coagulation and Flocculation.* Upland Press, London: 54-89.
- Chellam, S. 2000. Effects of Nanofiltration on Trihalomethane and Haloacetic Acid Precursor Removal and Speciation in Waters Containing Low Concentration of Bromide Ion. *Environ. Sci. and Technol.* 34(9): 813-820.
- Childress, Amy, E. 1999. Particulate and THM Precursor Removal with Ferric chloride. *J. Env. Engineering.* 125 (11): 1054-1061.
- Cheng, R.C., Krasner, S.W., Green, J.F. and Wattier, K.L. 1995. Enhanced Coagulation : A preliminary Evaluation. . *J. AWWA.* 87(2):91-103.
- Crozes, G., White, P. and Marshall, M. 1995. Enhanced Coagulation: Its effect on NOM Removal and Chemical Costs. *J.AWWA.* 132 (21): 78-89.
- Dempsey, B.A., Ganho, R. M. and O'Melia, C.R. 1984. The Coagulation of Humic Substances by Means of Aluminum Salts. *J. AWWA.* 76(4):141.
- Diehl, A.C., Symons, G.E., Krasner, S. W., Hwang, C.j. and Barret, S.E. 2000. DBP Formation during Chloramination. *J. AWWA.* 92(6): 78-89.
- Eaton, A. 1995. Measuring UV-Absorbing Organics: A Standard Method. *J. AWWA.* 87(2): 86-90.

- Edzwald, J.K. 1993. Coagulation in Drinking Water Treatment : Particles, Organics and Coagulants. *Wat. Sci. Tech.* 27(11):21.
- Environmental Health Unit Public Health Services. 2000. Environmental Health Guidance Note: Trihalomethanes in Drinking Water. *Environmental Health Unit Public Health Services*. 2<sup>nd</sup> ed., Queensland Health.
- Freese, S.D., Nozaic, D.J. and Smith, R.A. 2001. Enhanced Coagulation: A viable to Advance Treatment Technologies in the South African context. *Wat. Sci. Tech.* 1(1): 33-40.
- Freeman, S. and Shorney ,H. L. 1999 Control of Disinfection By-Product Precursors. *Proceedings of the American Water Works Annual Conference*. Atlanta, GA.
- Gemmell, R.S. 1978. Mixing and Sedimentation. *Water Quality and Treatment*. 3<sup>rd</sup> ed., The AWWA, Inc. McGraw-Hill Publishing: 129-139.
- Hubel, R.E. and Edzwald, J.K. 1987. Removing Trihalomethane Precursors by Coagulation. *J.AWWA*. 79(7):98-106.
- Hoehn, R.C., Dixon, K.L. and Randall, C.W. 1984. Biologically Induced Variations in the Nature and Removability of THM Precursors by Alum Treatment. *J. AWWA*. 76(4):134-140.
- Hundt, T .R. and O'Melia, C.R. 1988. Aluminum-Fulvic Acid Interactions : Mechanisms and Applications. *J. AWWA*. 80(4): 177-186.
- International Agency for Research on Cancer. 1991. IARC Monographs on the Evaluation of Carcinogenic Risks to Humans. World Health Organization, Geneva.
- Joyce, W.S., DiGiano, F.A. and Uden, P.C. 1984. THM Precursors in the Environment. *J.AWWA*. 76(6):102-106.
- Kang, S.F and Yen, H.Y. Peroxide. *Water Supply*. 13 (3): 53-58
- Kavanaugh, M.C. 1978. Modified Coagulation for Improved Removal of Trihalomethane Formation Potential. *J. AWWA*. 70(11): 613-620.
- Knappe, R.U. 2001. Effectiveness of Coagulants and Coagulant Aids for the Removal of Filter-Clogging Synedra. *J. Water SRT-Aqua*.50(3): 135-146.
- Leu, R.J. and Ghosh, M.M. 1988. Polyelectrolyte Characteristics and Flocculation. *J. AWWA*. 80(4): 159-167.

- Marhaba, T.F.,and Kochar. 2000. Rapid Prediction of Disinfection by-Product Formation Potential by Fluorescence. *Environ. Engg. and Policy*.126(11) :29-36.
- Marhaba, T.F.,and Pipada, N.S. 2000. Coagulation: Effectiveness in Removing Dissolved Organic Matter Fractions. *Environ Eng Sci*. 17(2): 107-118.
- Marhaba, T.F.,and Pu, Y. 2000. Rapid Identification of Humic and Non-humic Organic matter Fraction in Water. *J.of Hazardous Materials*. A73: 221-234.
- Marhaba,T.F., Memberand, P.E.,and Lippnicott 2000. Application of Fluorescence Technique for Rapid Identification of DOM Fractions in Source Waters *Environ Eng Sci*. 17(2): 1039-1044.
- Marhaba, T.F. and Van, D. 1999. Characterizing Dissolved Organic Matter Fractions using Spectral Fluorescent Signatures and processing by Principal Component Analysis. *J.Anal Chem*. 366: 255-268.
- Marhaba, T.F., Van, D.and Lippnicott. 2000. Application of Fluorescence Technique for Rapid Identification of DOM Fractions in Source Waters *Environ Eng Sci*. 17(2): 1039-1044.
- Marhaba, T.F., Washington, M.B. 1998. Drinking Water Disinfection and By-Products : History and Current Practice. *Adv.Environ. Res*. 2(1): 103-115
- Masion, A., Ritter, AV, Rose, J., Stone, W.E., Teppen, D.R., Rybacki, D. and Bottero, J.Y. 2000. Coagulation-Flocculation of Natural Organic Matter with Al Salts; Speciation and Structure of the Aggregates. *Environ. Sci. Technol*. 34 (15): 3242-3246.
- Miller, S. 1993. Disinfection Products in Water Treatment. *Environ. Sci. Technol*. 27(12): 2292-2297.
- Mitsu, Y., Hidetomo, S. and Shigemasa, T. 1975. Separating Liquids from Solids by Pellet Flocculation. *J. AWWA*. 67 (7): 397-402.
- Monteagudo, J.G., Seco, A. and Ferrer, J. 1998. Natural Organic Matter Coagulation in Valencia Water Supply Pilot Plant Studies. *J. Water SRT-Aqua*. 47(4): 199-208.
- Morris, R.D. et al. 1992. Chlorination by products and Cancers: A Meta-analysis. *American Journal of Public Health*. 82(7): 955-963.
- Najm, I., Marcinko, J. 2000. Evaluation TOC Analytical Results. *J.AWWA*. 92(8): 84-92.

- Owem, DM, Amy, G.L., Chowdhury, Z.K. and Viscosil, K. 1995.. NOM Characterization and Treatability. *J. AWWA*,75(10): 46-63.
- Panswad, T. 2000 Turbidity Removal by Pelletization Process for Monsoon Regions. *J. Water SRT-Aqua*. 49 (1): 9-21.
- Panswad, T. and Areeswangkit , K. 2000. Pellet Mass Loading and Pellet Retention time for Turbidity Removal in a Pelletization Process. *J. Water SRT-Aqua*. 49 (1):9-21.
- Panswad T. and Channarong, B. 1998. Turbidity Removal by the upflow Pelletization Process for Low Turbidity Water. *J. Water SRT-Aqua*. 47(1):36-40.
- Panswad T. and Polwanich, B. 1998. Pilot Plant Application of Pelletization Process for Low Turbidity River Water. *J. Water SRT-Aqua*. 47(5):236-244.
- Pontius, F.W. 2000. Regulations in 2000 and Beyond. *J.AWWA*. 12(3): 40-42
- Rand, M.C., Greenberg, A.E. and Taras, M.J. 1995. *Standard Methods for the Examination of Water and Wastewater*.19<sup>th</sup> ed., Baltimore, American Public Health Association American Water Works Association Water Pollution Control Federation New York.
- Randtke, S.R. 1988. Organic Contaminant Removal by Coagulation and Related Process Combination. *J.AWWA*. 85(2): 40-54.
- Reynold, T.D and Richard, P.A. 1996. Unit Operations and Processes in Environmental Engineering. Second Ed., PWS Publishing Company, Boston.
- Rhodes, R. and Mark, D. 1978. The Formation of Trihalomethanes. *J. AWWA*. 70(11): 604-612.
- Shimizu, T., Tambo, N.; Ozawa, G. and Hamaguchi, T. 1994. An Anaerobic Pellet Bed Bioreactor Process for Simultaneous Removal of Organic, Nitrogenous and Phosphorus Substance. *Wat. Res*, 28(9): 1943-1952.
- Singer, P.C. 1993. Trihalomethanes and other By-products Formed by Chlorination of Drinking Water. *Keeping Pace with Science and Engineering*. Washington DC: National Academy Press.141-146.
- Smith, G.C. and Huang, Y.C. 1984. Spectrophotometric Determination of Total Trihalomethanes in Finished Waters. *J. AWWA*. 80(4):168-171.
- Sung, W. 2000. Modeling DBP Formation . *J. AWWA*.92(5): 55-60.

- Stevens, A.A., Slocum, C.J., Seeger, D.R. and Robeck, G. 1976. Chlorination of Organics in Drinking Water. *J.AWWA*.68(11): 615-618.
- Tambo, N. and Matsui, Y. 1989. Performance of Fluidized Pellet Bed Separator for High Concentration Suspensions Removal. *J. Water SRT-AQUA*. 38(38): 16-22.
- Tambo, N. and Matsui, Y. 1993. The Mechanism of Pellet Flocculation in a Fluidized Bed Operation. *J. Water SRT-AQUA*. 42(2): 67-76.
- Tambo, N. and Matsui, Y. 1993. Control of Coagulation Condition for Treatment of High-turbidity Water by Fluidized Pellet Bed Separation. *J. Water SRT-AQUA*. 42(4): 212-222.
- Trussell, R.R. and Umphres, M.D. 1978. The Formation of Trihalomethanes. *J.AWWA*. 70(11): 604-611.
- Vrijenhoek, E. M. 1998. Removing Particles and THM Precursors by Enhanced Coagulation. *J. AWWA*. 90(4): 139-150.
- White, M.C., Thompson, J.D., Harrington, G.W. and Singer P.C. 1997. Evaluation Criteria for Enhance Coagulation Compliance. *J. AWW*. 89(5): 64-77.
- Wigle, D.T. 1998. Safe Drinkiing Water: A Public Health Challenge. *Health Canada-CDIC* . 19(3):1-13.

# **APPENDICES**

## APPENDIX A

### UP-FLOW PELLETIZATION PROCESS

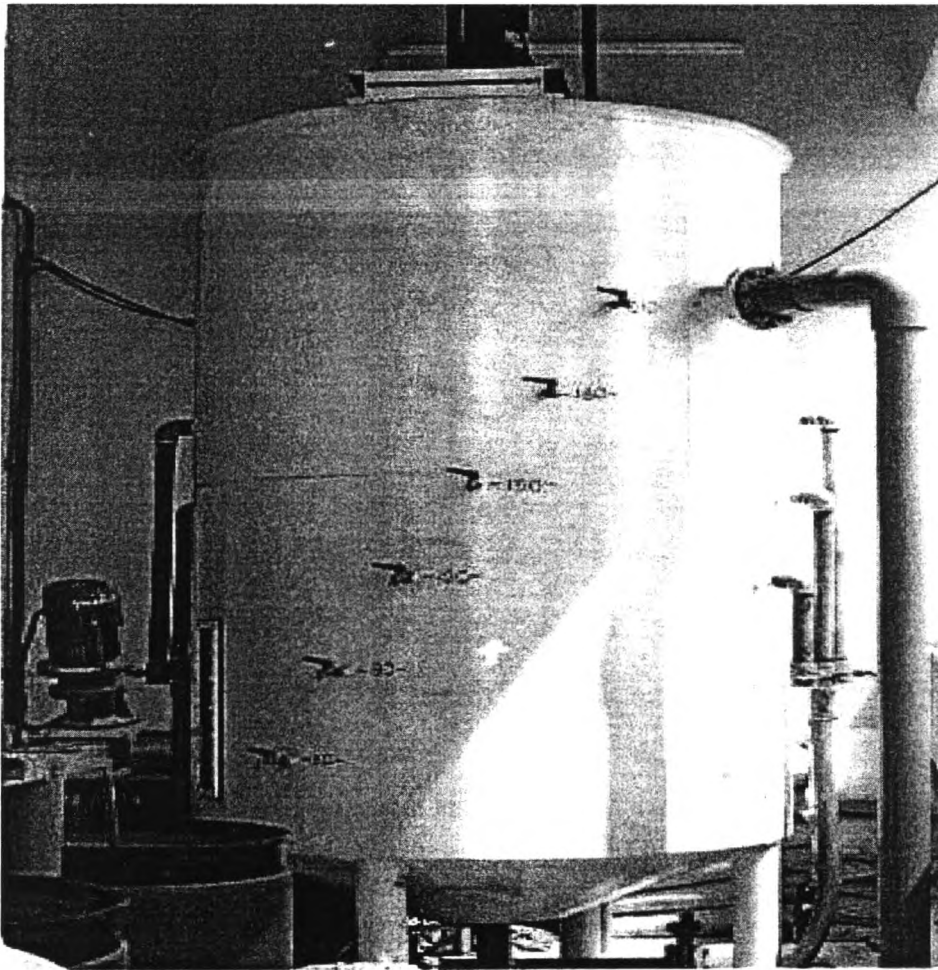
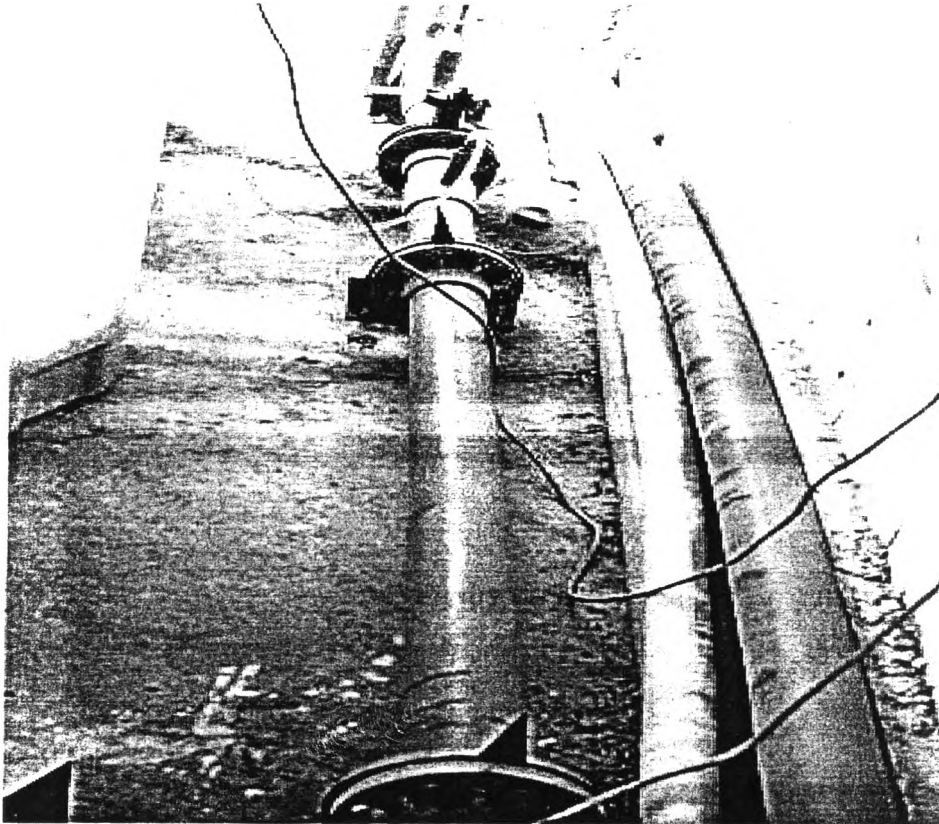
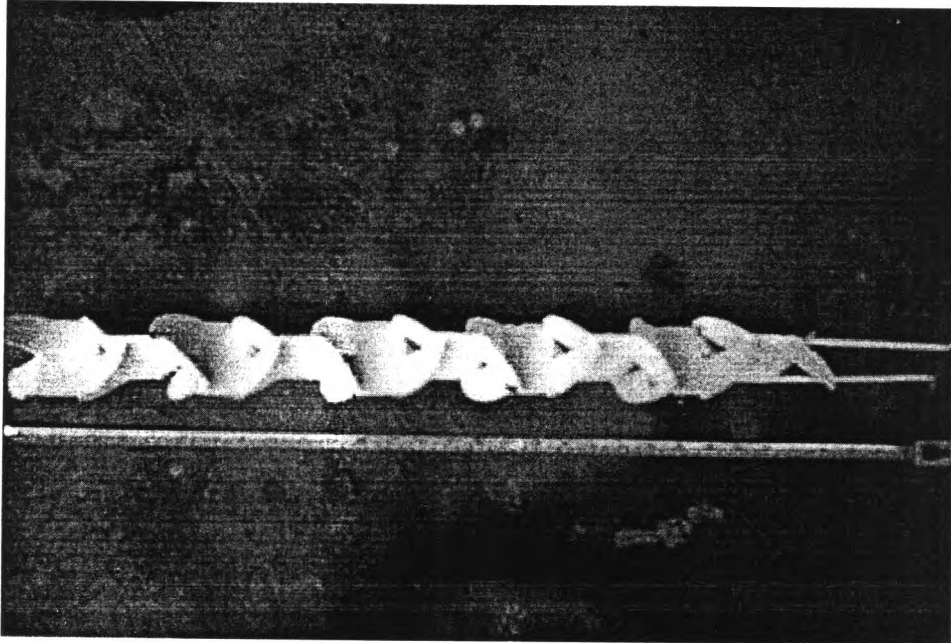


Figure A-1 Pelletizer

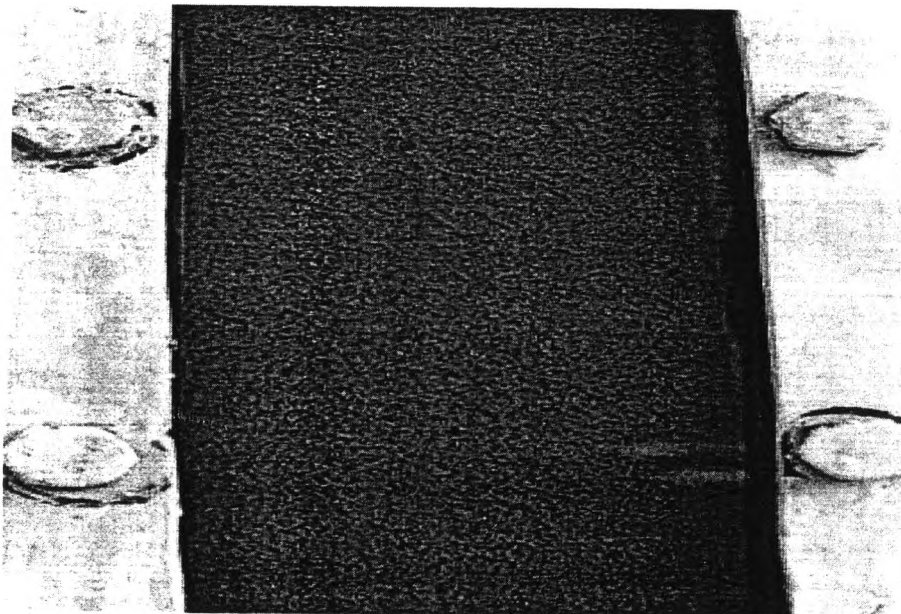


**Figure A-2** Chemicals pipe





**Figure A-3** Static mixer



**Figure A-4** Characteristic of pellets



## APPENDIX B

### DETERMINATION THE MEAN VELOCITY GRADIENT

#### 1. Velocity gradient in Pelletizer

1.1. The root mean square velocity gradient ( $G$ ) can be calculated by equations of Metcalf & Eddy (1991) as follows:

$$G = (P/V\mu)^{0.5}$$

Where:

$G$  = velocity gradient ( $s^{-1}$ )

$P$  = power applied to stirring, W (ft-lbf/s = HPx 500)

$V$  = reactor volume  $m^3$  ( $ft^3$ )

$\mu$  = dynamic viscosity  $N\cdot s/m^2$  ( $lbf\cdot s/ft^2$ )

1.2. Power requirements are determined from;

$$P = C_D A \rho V^3 / 2$$

where:

$P$  = power applied to stirring, W (ft-lbf/s = HPx 500)

$C_D$  = coefficient of drag (of paddle moving perpendicular to fluid paddles;  $C$  is approximately 1.8)

$A$  = paddle area,  $m^2$  ( $ft^2$ )

$\rho$  = fluid density,  $kg/m^3$  (slugs/ $ft^3$ )(for water at  $25^\circ C$ ;  $\rho = 997 kg/m^3$ )

$V$  = relative velocity of paddles in fluid ( $m/s$ [ $fps$ ]), typically about 0.6 to 0.75 of paddle tip speed ( $v_p$ )

$$v_p = \text{paddle tip speed; } 2\pi rN / 60(\text{m/s}^{-1})$$

Solution

- G values at 6 blades, 2 rpm.

Paddles have 2 size

$$\text{Size A: width 12 cm. length 60 cm.; 4 blades} \quad A = 0.072 \text{ m}^2$$

$$\text{Size B: width 6 cm. length 45 cm.; 2 blades} \quad A = 0.027 \text{ m}^2$$

- a. Calculation  $P$  of paddle size A

$$\begin{aligned} P_A &= 0.5 \times 1.8 \times 0.288 \times 997 \times (0.75 \times 2\pi \times 0.6 \times 2 / 60)^3 \\ &= 0.216 \text{ W} \end{aligned}$$

- b. Calculation  $P$  of paddle size B

$$\begin{aligned} P_B &= 0.5 \times 1.8 \times 0.054 \times 997 \times (0.75 \times 2\pi \times 0.6 \times 2 / 60)^3 \\ &= 0.041 \text{ W} \end{aligned}$$

$$P_A + P_B = 0.257 \text{ W}$$

- d. Calculation  $G$

$$\begin{aligned} G &= (0.257 / (8.9 \times 10^{-4} \times 2.25))^{0.5} \\ G &= 11.33 \text{ s}^{-1} \end{aligned}$$

- G values at 4 paddles, 2 rpm.

$$\text{Paddle size: width 12 cm. length 60 cm.; 4 blades} \quad A = 0.072 \text{ m}^2$$

- a. Calculation  $P$  of paddle

$$\begin{aligned} P_A &= 0.5 \times 1.8 \times 0.288 \times 997 \times (0.75 \times 2\pi \times 0.6 \times 2 / 60)^3 \\ &= 0.216 \text{ W} \end{aligned}$$

- b. Calculation  $G$

$$\begin{aligned} G &= (0.216 / (8.9 \times 10^{-4} \times 2.25))^{0.5} \\ G &= 10.39 \text{ s}^{-1} \end{aligned}$$

➤  $G$  values at 2 paddles, 2 rpm.

Paddles size: width 12 cm. length 60 cm;  $A = 0.072 \text{ m}^2$

a. Calculation  $P$

$$P = 0.5 \times 1.8 \times 0.144 \times 997 \times (0.75 \times 2\pi \times 0.6 \times 2 / 60)^3$$

$$= 0.108 \text{ W}$$

b. Calculation  $G$

$$G = (0.108 / (8.9 \times 10^{-4} \times 2.25))^{0.5}$$

$$G = 7.34 \text{ s}^{-1}$$

## 2. Velocity gradient in static mixer

2.1  $G$  values in static mixer can be calculated by equations as follows:

$$G = (\rho g H_s / \mu t)^{0.5}$$

where:

$G$  = velocity gradient ( $\text{s}^{-1}$ )

$\rho$  = fluid density,  $\text{kg/m}^3$  (slugs/  $\text{ft}^3$ ) (for water at  $25^\circ\text{C}$ ;  $\rho = 997 \text{ kg/m}^3$ )

$g$  =  $9.8 \text{ m}^2/\text{s}^{-1}$

$\rho$  = fluid density,  $\text{kg/m}^3$  (slugs/  $\text{ft}^3$ ) (for water at  $25^\circ\text{C}$ ;  $\rho = 997 \text{ kg/m}^3$ )

$\mu$  = dynamic viscosity  $\text{N}\cdot\text{s}/\text{m}^2$  ( $\text{lbf}\cdot\text{s}/\text{ft}^2$ ) (for water at  $25^\circ\text{C}$ ;  $\mu = 8.95 \times 10^{-4} \text{ kg/m}^3$ )

$t$  = time for water flow through static mixer ( $\text{s}^{-1}$ )

### 2.2 Calculation head loss ( $H_s$ )

$$H_s = h(\text{pipe}) + h(\text{static mixer})$$

where:

$$h(\text{pipe}) = 1.15 \times 10^{-3} L V^{1.95} / D^{1.25}$$

$$h(\text{static mixer}) = 3.24 \times 10^{-3} (1.5 + 0.12 R_C)^{0.5}$$

$L$  = length of static mixer ( $\text{s}^{-1}$ )

$V$  = velocity in pipe ( $\text{m/s}^{-1}$ )

$D$  = diameter of pipe (m)

$R_C$  = Renold number;  $R_C = \rho VD/\mu$

### Solution

a. Calculation head loss in pipe

$$\begin{aligned} h(\text{pipe}) &= 1.15 \times 10^{-3} \times 0.35 \times 1.47^{1.95} / 0.06^{1.25} \\ &= 0.028 \text{ m} \end{aligned}$$

b. Calculation head loss in static mixer

$$\begin{aligned} R_C &= 997 \times 1.47 \times 0.06 / 0.000895 \\ &= 98804 \end{aligned}$$

$$\begin{aligned} h(\text{static mixer}) &= 3.24 \times 10^{-3} (1.5 + 0.12 \times 98804^{0.5}) \\ &= 0.127 \text{ m} \end{aligned}$$

c. Calculation head loss

$$\begin{aligned} H_s &= 0.028 + 0.127 \\ &= 0.155 \text{ m} \end{aligned}$$

d. Calculation  $Gt$  values

$$\begin{aligned} G &= (997 \times 9.8 \times 0.155) / (0.00089 \times 0.24) \\ &= 2662 \text{ s}^{-1} \end{aligned}$$

$$\begin{aligned} Gt &= 2662 \times 0.24 \\ &= 639 \end{aligned}$$

## APPENDIX C

### COAGULANT AND COAGULANT-AID

**Table C-1** Properties of Polyaluminium Chloride

Code name	PAC
Appearance	Liquid
Al <sub>2</sub> O <sub>3</sub>	10%
SO <sub>4</sub> <sup>-2</sup>	3.5 max
Basicity	45-65%
pH	3.5-5 (at 1%/wt/v)
Specific Gravity	1.19 min (g/cc, at 25°C)

**Table C-2** Properties of Nonionic Polymer

Code name	Kurita C-1330
Type	Nonionic
Main ingredient	Polyacrylamide
Appearance	White granular powder
Molecular weight	Approximately 12 millions
pH	5.5-7.5 (0.1%sol at 25°C)

## APPENDIX D

### REAGENTS FOR TOC ANALYZER

#### Reagents

*Reagent Water:* Distilled or deionized water containing TOC of less than 200 ppbC is recommended.

*Sodium Persulfate ( $\text{Na}_2\text{S}_2\text{O}_8$ ):* The optimum concentration of sodium persulfate solution depends on the range of carbon to be detected. Prepare a 100g/L or 200g/L solution of sodium persulfate by dissolving 100 or 200 g  $\text{Na}_2\text{S}_2\text{O}_8$  into reagent water (1L total volume). Stirring may be necessary, but do not heat. Transfer a portion of this solution to the appropriate reagent bottle provided with the Model 1010. Shelf life is approximately three weeks. Sodium persulfate and reagent are available from OI Analytical.

*Phosphoric Acid ( $\text{H}_3\text{PO}_4$ )(5% vol/vol):* Prepare a 5% by volume solution of phosphoric acid by adding 59mL of ACS Reagent Grade 85%  $\text{H}_3\text{PO}_4$  to reagent water (1L total volume). The acid solution may be purified, if high organic contamination of the solution is suspected, by adding 10 mL of the persulfate solution and immersing the vented container in boiling water for at least two hours at 100 °C. The cooled solution should then be purged for several minutes to remove any  $\text{CO}_2$  from oxidation of organics. The decrease in reagent blank resulting from this procedure is not generally worth the purification effort unless the acid solution is found to be abnormally high in TOC.

*Potassium Biphthalate Stock Solution (KHP) (1000 ppmC)*: Prepare a stock solution by adding 2.128 g of KHP (previously dried to constant mass at 110 C°) into a 1000 ml volumetric flask. Dilute to volume with reagent water. Lower concentration standards can be prepared from this stock. . Self-life is approximately three weeks.

*Sodium carbonate stock solution (Na<sub>2</sub>CO<sub>3</sub>) (100 ppm)*: Prepare a stock solution by adding 8.826 g of Na<sub>2</sub>CO<sub>3</sub> (previously dried to constant mass at 110 C°) to a 1000 ml volumetric flask. Dilute to volume with reagent water. Lower concentration standards can be prepared from this stock. Self-life is approximately three weeks.



## STANDARD CURVE

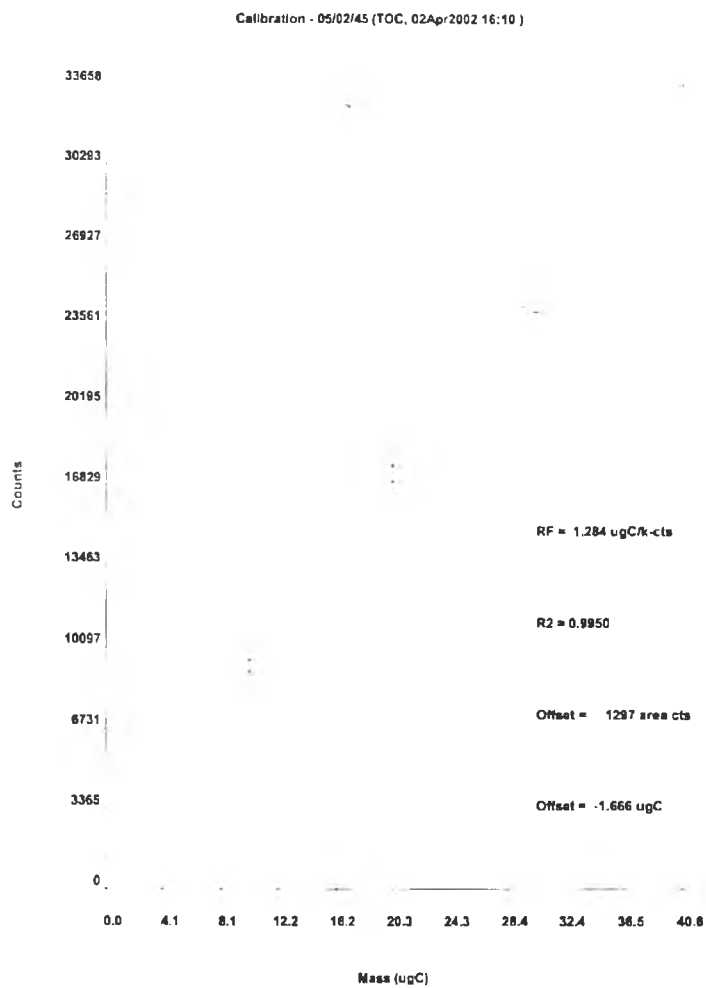


Figure D-1 Calibration curve for TOC

## APPENDIX E

### THM FORMATION POTENTIAL

#### Reagent

##### 1. Chlorine stock solution

Chlorine stock solution was prepared by diluting 10 % sodium hypochlorite (NaOCl) solution in distilled deionized water to the required concentration (1 mg/L). This solution was then stored in a brown, glass-stoppered bottle kept overnight and standardized using iodometric method I, as according in the *Standard Method* (1985), every time before use.

##### 2. Stock THM standard solution

THM mixture standard of 1 mg/mL of chloroform, 20 mg/mL of bromoform, 1 mg/mL of bromodichloromethane, 4 mg/mL of dibromochloromethane and in methanol. The working standard solutions were then prepared by diluting the stock standard mixture to 5, 10, 20 and 40 times with purified methanol. These standards were kept in serum vials, sealed with paraffin paper, septum and aluminum cap and kept in the refrigerator before use.

#### Sample Preparation

Chlorinated two liters of water sample with appropriate amount of hypochlorite solution under a well-mixed condition using a magnetic stirrer. The amount of hypochlorite solution was measured with a pipette and dosed instantaneously. Samples for THM analysis were collected in sterile flask to which 0.5 ml of 5% sodium thiosulphate had already been added ( to neutralize the chlorine in the water)

## Procedure

The Head Space Method as outlined in the Recommended Standard Methods for Water and Wastewater Analysis was adopted for the determination of the THM formation. Exactly 20 mL of the water to be tasted was pipetted into each of the 25 mL serum vials and immediately sealed with silicone septum vial closer. After shaking vigorously the serum vials were allowed to stand upside down for one hour prior to the gas chromatographic analysis. 200  $\mu$ L of headspace vapor were removed with a gas-tight microsyringe and injected into the gas chromatograph. Computed on a Shimadzu C-R4A chromatopac data processor, were obtained by measuring the retention time and peak area on the chromatogram and compared to a standard curve previously prepared using the same method with a known THM previously prepared using the same method with a known THM mixture standard solution.

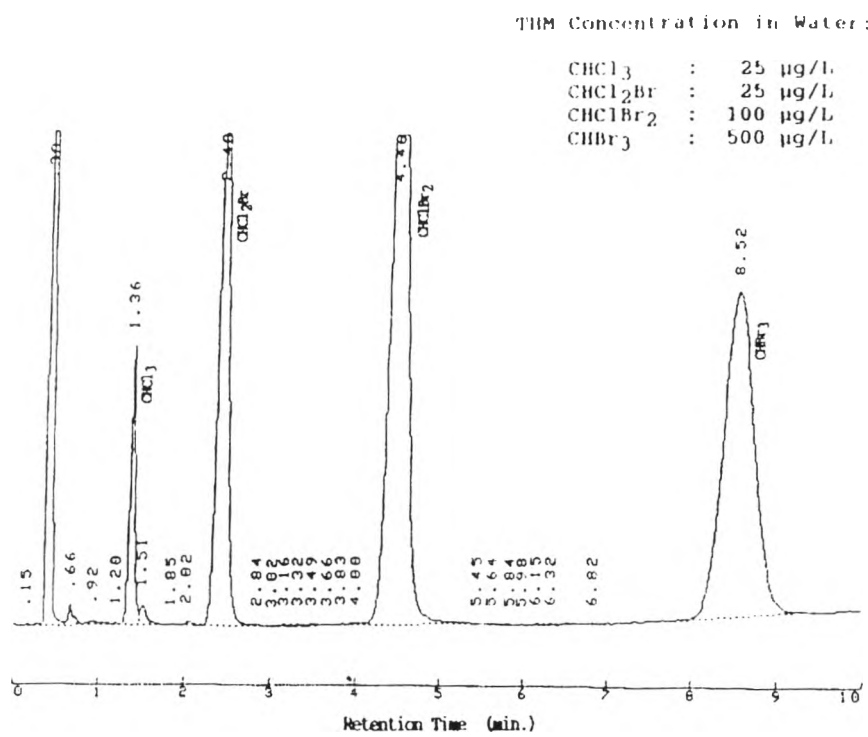


Figure E-1 Typical chromatogram of THM mixture standard in water

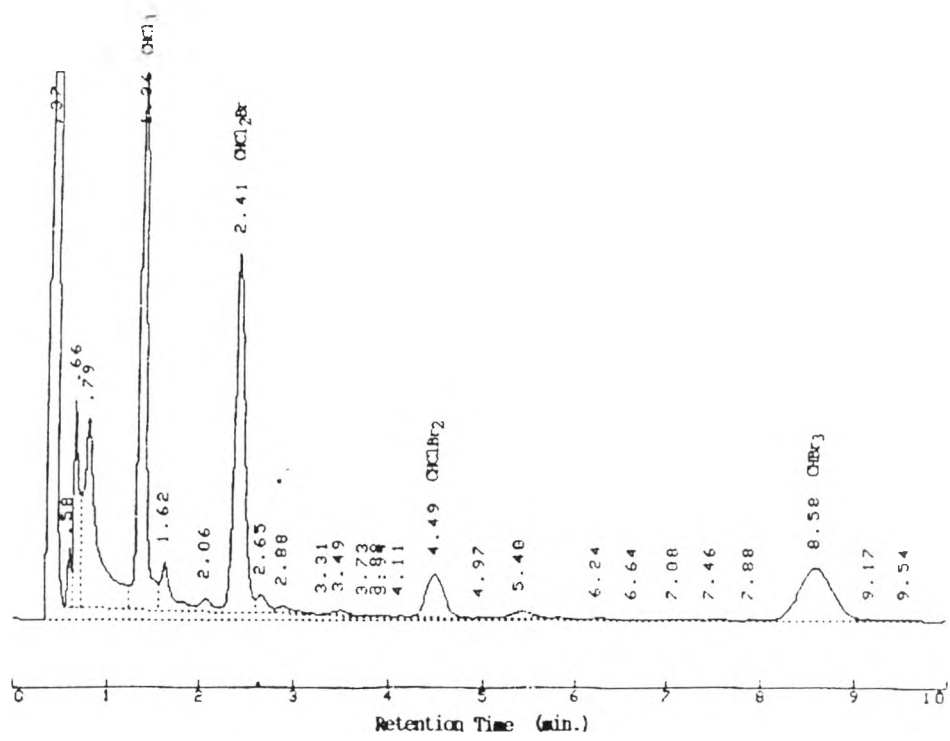


Figure E-2 Typical chromatogram of THMFP occurrence in water sample.

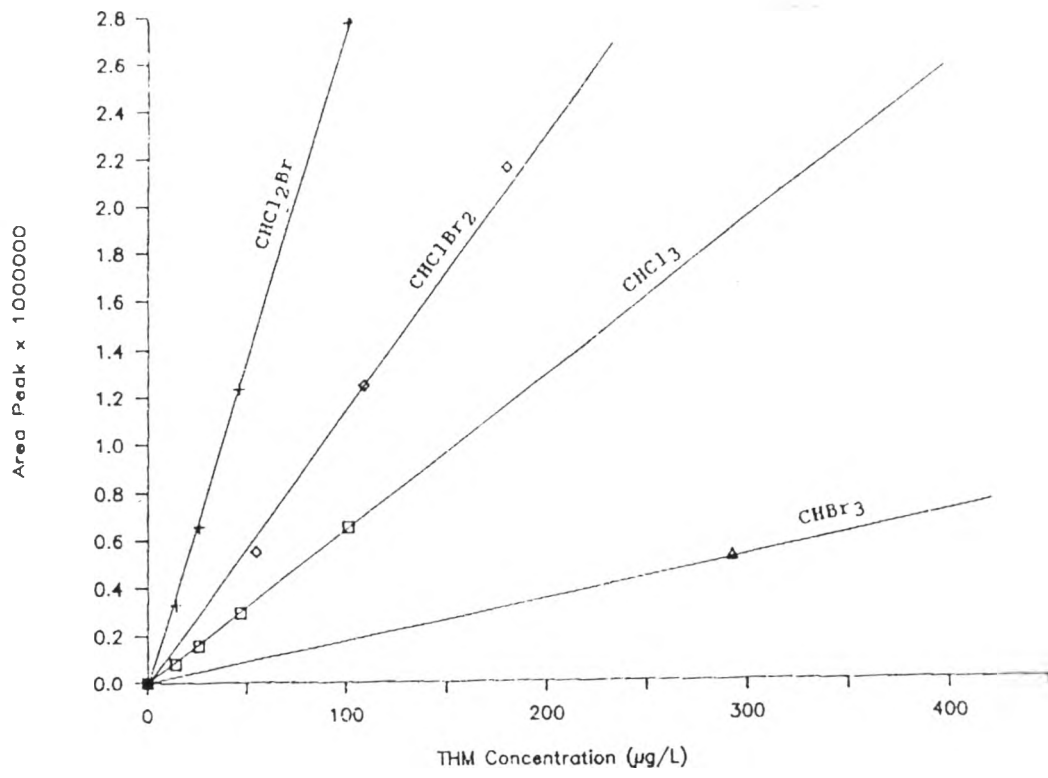


Figure E-3 Calibration curve for THMFP

## BIOGRAPHY



Miss Pornpimon Limtrakul was born on 12 June, 1977 in Bangkok. she studied in the major of General Science in Faculty of Science at Chulalongkorn University and graduated a Bachelor of Science in March,1999. She continued her further study for Master's degree in International Environmental Management Science at Chulalongkorn University and achieved her Master's degree in April, 2002.