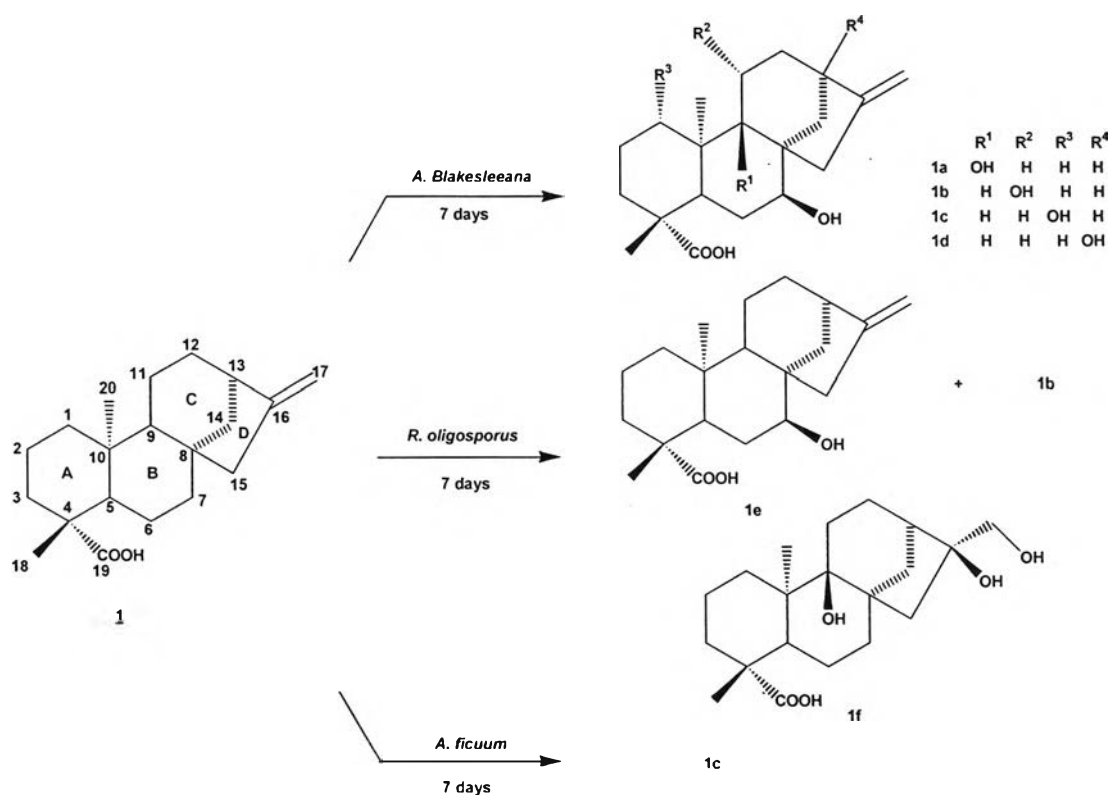


## CHAPTER V

### CONCLUSION

In this research, *ent*-kaur-16-en-19-oic acid (**1**) was isolated from the stem bark of *C. oblongifolius* (Amphur Kuiburi, Prachuab Kirikhan Province) and *ent*-1,2-dehydro-3-oxomanoyl oxide (**2**), *ent*-1 $\beta$ -hydroxy-3-oxomanoyl oxide (**3**) and *ent*-1,2-dehydro-12 $\alpha$ -hydroxy-3-oxomanoyl oxide (**4**) were isolated from the stem bark of *C. oblongifolius* (Loei Province).

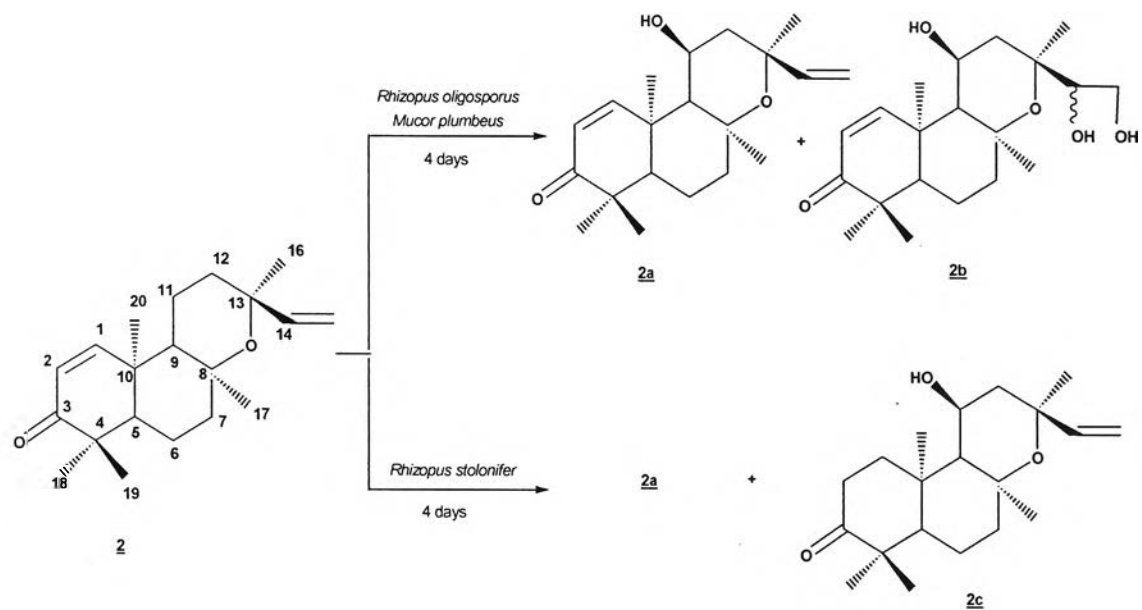
Biotransformation of the kaurane diterpene (*ent*-kaur-16-en-19-oic acid, **1**) with *Absidia blakesleeana*, *Rhizopus oligosporus* and *Aspergillus ficuum* are shown in Table 5.1 and Scheme 5.1. Biotransformation of the labdane diterpene (*ent*-1,2-dehydro-3-oxomanoyl oxide, **2**) by *Rhizopus oligosporus*, *Rhizopus stolonifer* and *Mucor plumbeus* are shown in Table 5.2 and Scheme 5.2.



Scheme 5.1 The biotransformation of *ent*-kaur-16-en-19-oic acid.

**Table 5.1** The biotransformation of *ent*-kaur-16-en-19-oic acid.

Fungus	Metabolite	Weight (mg)	% yield from starting material
<i>Absidia blakesleeana</i>	<b>1a</b> , <i>ent</i> -(7 $\alpha$ ,9 $\alpha$ )-dihydroxy-kaur-16-en-19-oic acid	33	6.8
	<b>1b</b> , <i>ent</i> -(7 $\alpha$ ,11 $\beta$ )-dihydroxy-kaur-16-en-19-oic acid	75	15.5
	<b>1c</b> , <i>ent</i> -(1 $\beta$ ,7 $\alpha$ )-dihydroxy-kaur-16-en-19-oic acid	45	9.3
	<b>1d</b> , <i>ent</i> -(7 $\alpha$ ,13)-dihydroxy-kaur-16-en-19-oic acid	65	13.4
<i>Rhizopus oligosporus</i>	<b>1e</b> , <i>ent</i> -7 $\alpha$ -hydroxy-kaur-16-en-19-oic acid	35	8.1
	<b>1a</b> , <i>ent</i> -(7 $\alpha$ ,9 $\alpha$ )-dihydroxy-kaur-16-en-19-oic acid	45	10.4
	<b>1f</b> , <i>ent</i> -(7 $\alpha$ ,16,17)-trihydroxy-kaur-16-en-19-oic acid	23	5.3
<i>Aspergillus ficuum</i>	<b>1b</b> , <i>ent</i> -(7 $\alpha$ ,11 $\beta$ )-dihydroxy-kaur-16-en-19-oic acid	102	28.3



Scheme 5.2 The biotransformation of *ent*-1,2-dehydro-3-oxomanoyl oxide.

**Table 5.2** The Biotransformation of *ent*-1,2-dehydro-3-oxomanoyl oxide.

Fungus	Metabolite	Weight (mg)	% yield from starting material
<i>Rhizopus oligosporus</i>	<b>2a</b> , <i>ent</i> -11 $\alpha$ -hydroxy-1,2-dehydro-3-oxomanoyl oxide	230	46.0
	<b>2b</b> , <i>ent</i> -(11 $\alpha$ ,14 $\xi$ ,15)-trihydroxy-1,2-dehydro-3-oxomanoyl oxide	8	1.6
<i>Rhizopus stolonifer</i>	<b>2a</b> , <i>ent</i> -11 $\alpha$ -hydroxy-1,2-dehydro-3-oxomanoyl oxide	64	12.8
	<b>2c</b> , <i>ent</i> -11 $\alpha$ -hydroxy-3-oxomanoyl oxide	40	8.0
<i>Mucor plumbeus</i>	<b>2a</b> , <i>ent</i> -11 $\alpha$ -hydroxy-1,2-dehydro-3-oxomanoyl oxide	50	10.0
	<b>2b</b> , <i>ent</i> -(11 $\alpha$ ,14 $\xi$ ,15)-trihydroxy-1,2-dehydro-3-oxomanoyl oxide	72	14.4

The systematic use of different microorganisms may greatly facilitate the production of novel hydroxylated derivatives, which can then be tested for possible biological activity.

#### Suggestion for the future work

1. More work is needed to be done in order to understand the relationship between the *ent*-kaurane diterpenoids and the enzymes responsible for their hydroxylation, as well as to perform large-scale biotransformations with improved yields of desirable products.
2. Extension of this strategy to obtain rapid access to other diterpenoid derivatives should be investigated and a study of the potential biological activities of the biotransformed products should be carried out.