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APPENDICES

Appendix A

1. Chromatogram and standard curve of Limonin

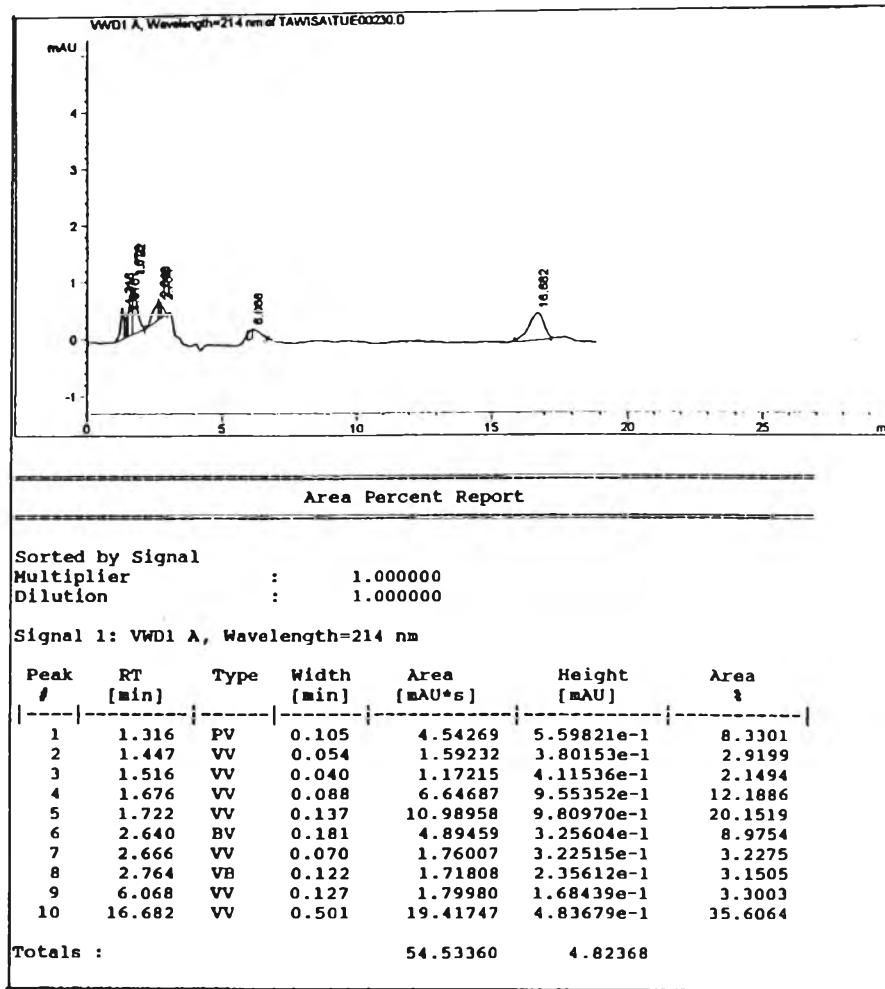


Figure A 1 Chromatogram of standard limonin at 2.5 ppm

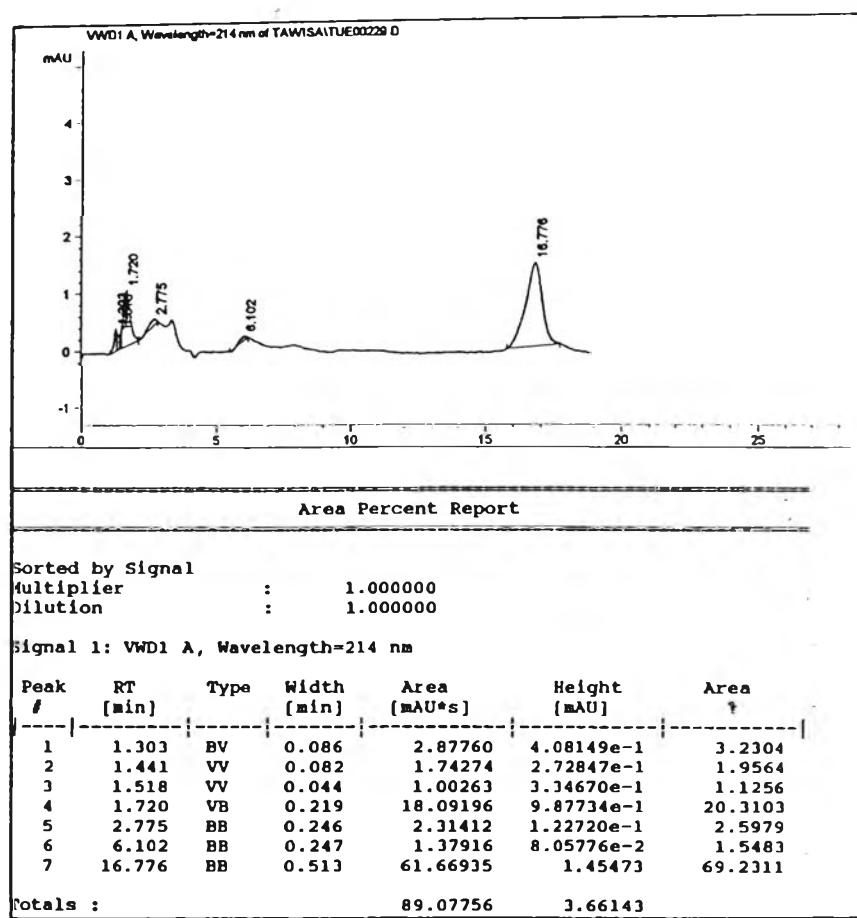


Figure A 2 Chromatogram of standard limonin at 7.5 ppm

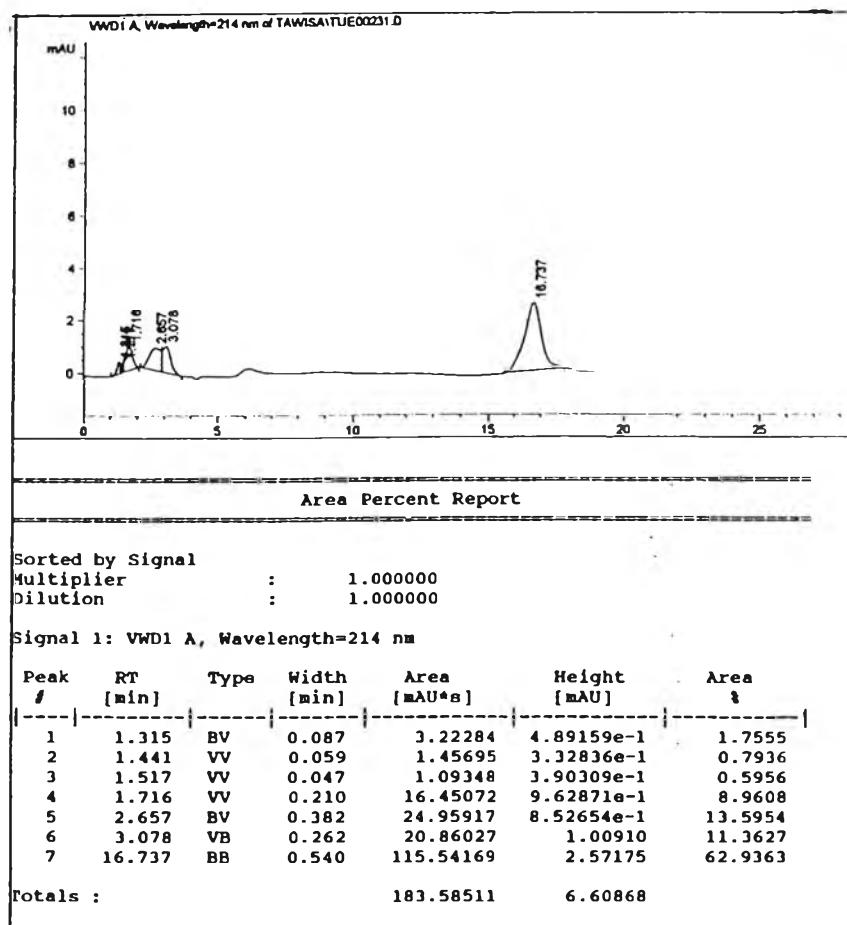


Figure A 3 Chromatogram of standard limonin at 12.5 ppm

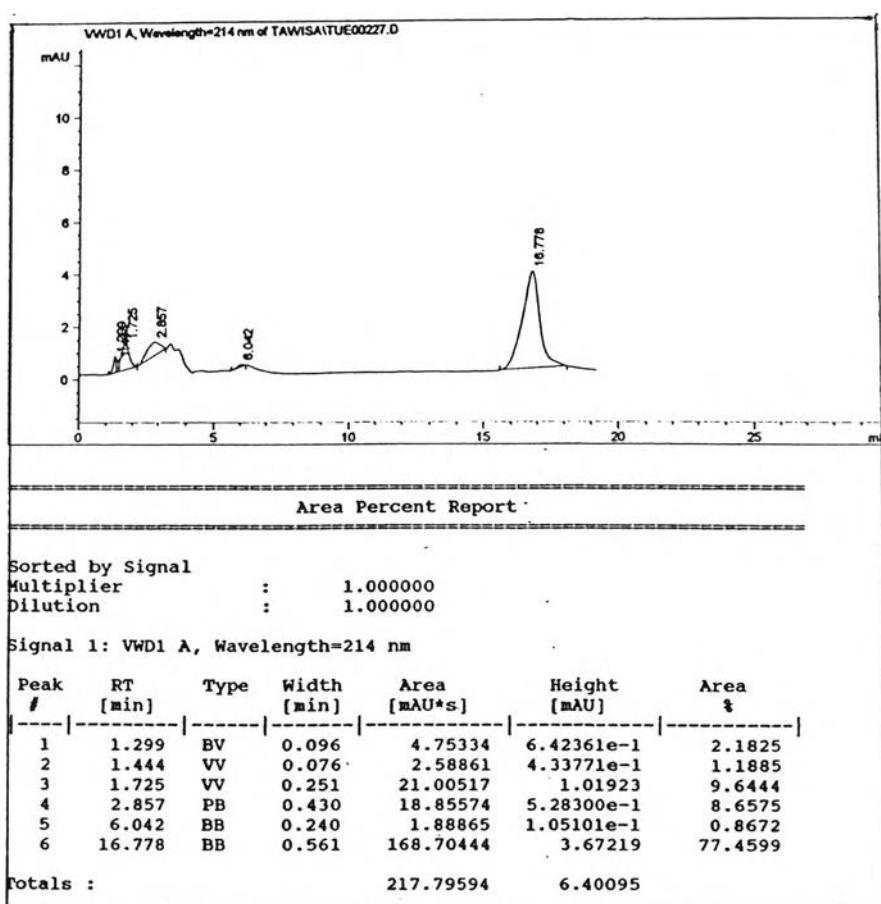


Figure A 4 Chromatogram of standard limonin at 17.5 ppm

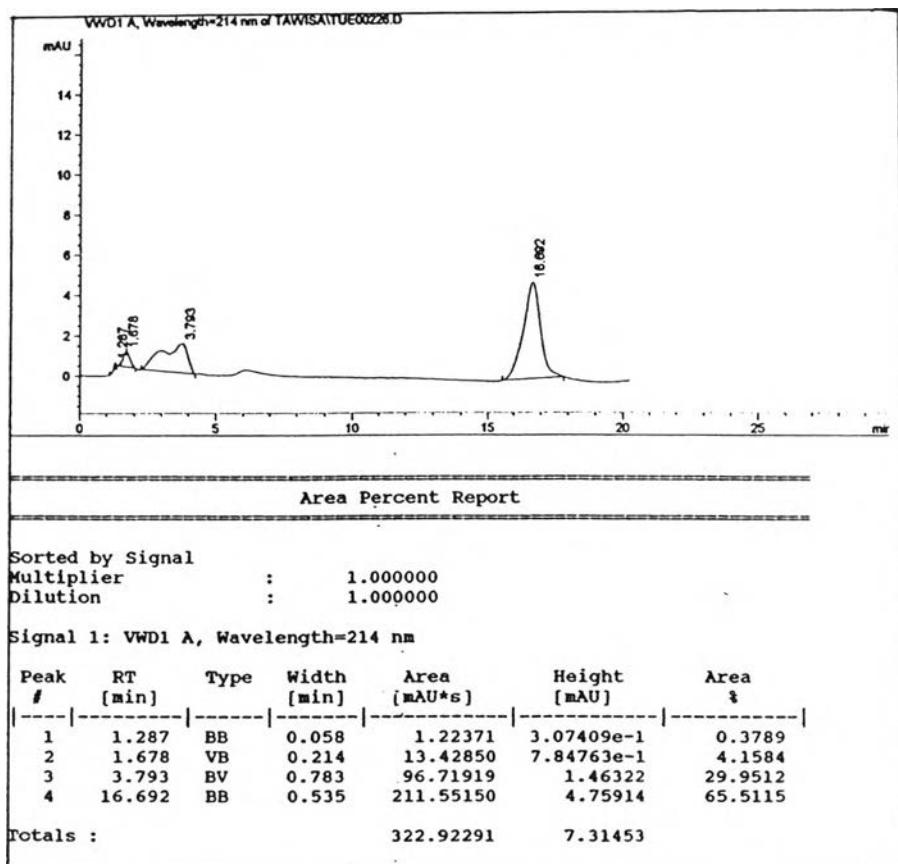


Figure A 5 Chromatogram of standard limonin at 22.5 ppm

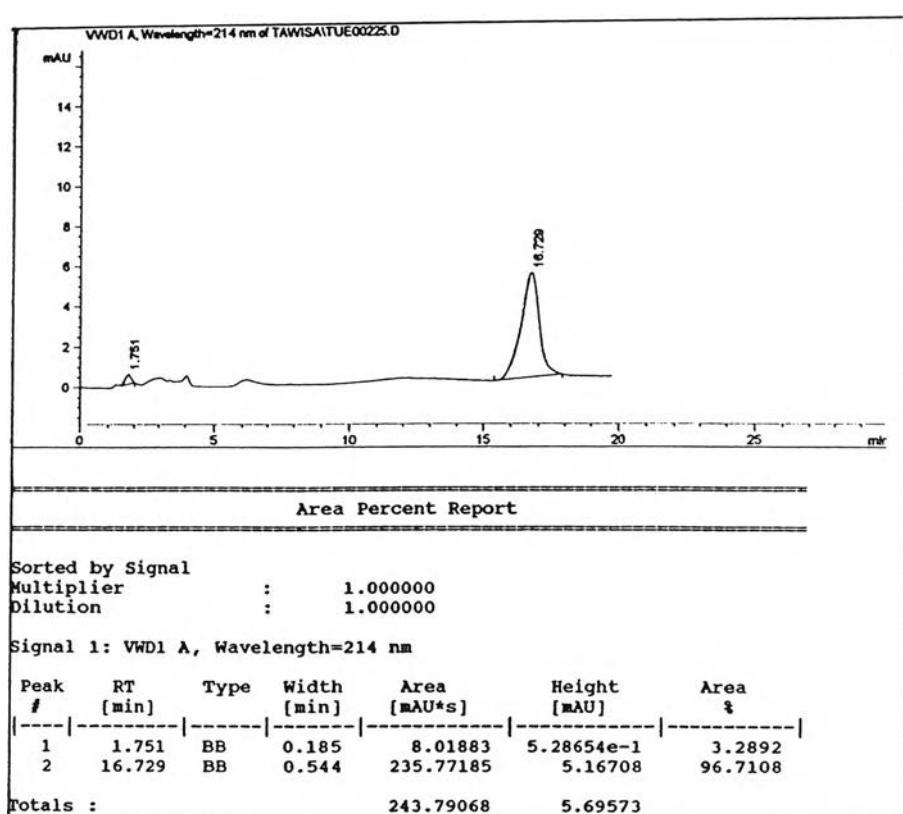


Figure A 6 Chromatogram of standard limonin at 25.0 ppm

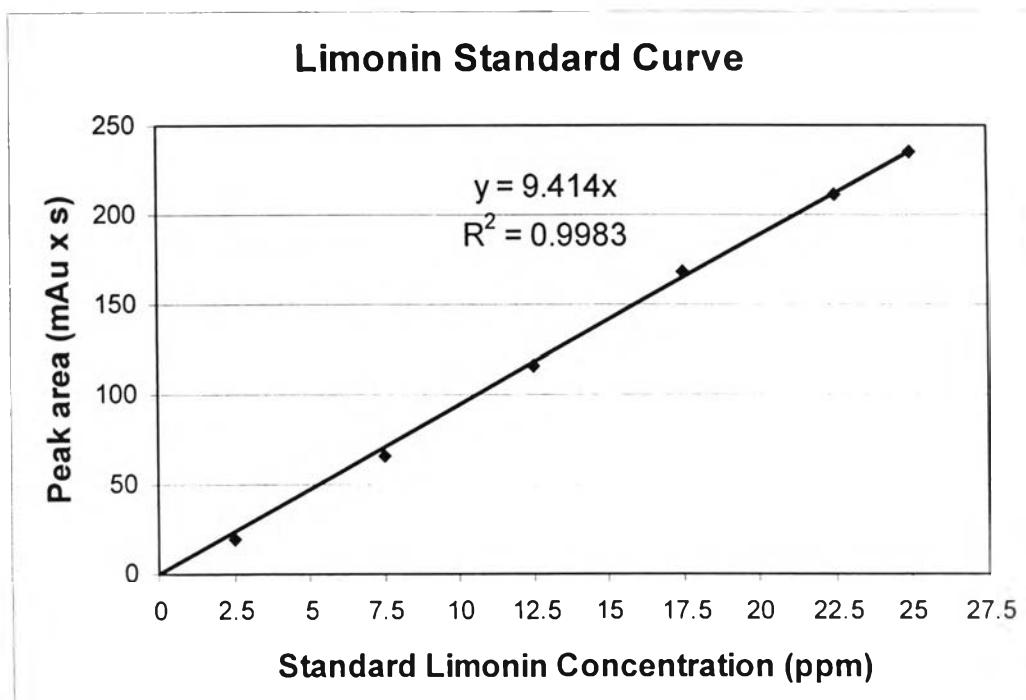


Figure A 7 Standard curve of limonin

Table A 1 % Recoveries of limonin of initial and reused HyperSEP C18 column

Replications	%Recoveries				
	Initial C-18	1 st Reused	2 nd Reused	3 rd Reused	4 th Reused
1	86.55	80.15	77.06	63.56	64.91
2	83.24	80.55	75.66	63.28	63.56
3	82.06	80.18	77.96	63.28	63.50
Average	83.95±2.33 ^a	80.29±0.22 ^b	76.89±1.16 ^c	63.37±0.16 ^d	63.99±0.80 ^d
%C.V.	2.77	0.28	1.51	0.26	1.24

Note: a, b, c and d means with the same letter were significantly different at P>0.05 by Duncan's multiple range test with three replications.

Table A 2 Color values (L, a, b) of each sample juice

Sample	L	a	b	Average / %C.V		
				L	a	b
Fresh juice	102.46	-11.32	+33.50			
	101.17	-10.92	+31.83	101.80 _A ±0.64 /	-11.03 _A ±0.25 /	+32.33 _A ±1.01 /
	101.77	-10.86	+31.66	0.63	2.26	3.14
Clarified juice	92.23	-6.11	+9.90			
	92.34	-6.10	+10.39	92.43 _B ±0.2 /	-6.18 _B ±0.13 /	+10.28 _B ±0.33 /
	92.72	-6.33	+10.54	0.28	2.10	2.60
Debittered juice by β-CD polymer	92.54	-6.54	+10.34			
	92.87	-6.42	+10.23	92.46 _B ±0.44 /	-6.39 _B ±0.17 /	+10.23 _B ±0.12 /
	91.98	-6.21	+10.11	0.49	2.61	4.54
Debittered juice by XAD-16 resin	92.05	-6.86	+10.15			
	92.89	-6.15	+10.17	92.5 _B ±0.42 /	-6.33 _B ±0.47 /	+10.18 _B ±0.04 /
	92.56	-5.97	+10.28	0.46	7.43	0.69
Debittered juice by prepared β-CD polymer	92.19	-6.38	+10.14			
	92.43	-6.19	+9.98	92.39 _B ±0.18 /	-6.26 _B ±0.10 /	+10.16 _B ±0.20 /
	92.54	-6.22	+10.37	0.36	1.63	1.43

Note: L = Lightness, a and b = color coefficient

A and B means with the same letter were significant different at P>0.05 by Duncan's multiple range test with three replications.

Table A 3 Total soluble solids of each sample juice

Juice	Total soluble solids			Average*NS	%C.V
Fresh juice	11.6	11.7	11.6	11.63 \pm 0.06	0.50
Clarified juice	11.6	11.5	11.6	11.55 \pm 0.07	0.50
Debittered juice by β-CD polymer	11.6	11.5	11.5	11.54 \pm 0.05	0.50
Debittered juice by XAD-16 resin	11.7	11.6	11.6	11.63 \pm 0.06	0.50
Debittered juice by prepared β-CD polymer	11.5	11.6	11.7	11.6 \pm 0.10	0.86

Note: *NS= not significantly different at $P\leq 0.05$ by Duncan's multiple range test with three replications.

Table A 4 Vitamin C content of each sample juice

Juice	Vitamin C content (mg/100 ml juice)			Average*NS	%C.V
Clarified juice	5.67	5.80	5.67	5.71±0.08	1.31
Debittered juice by β-CD polymer	5.80	5.80	5.94	5.85±0.08	1.37
Debittered juice by XAD-16 resin	5.80	5.67	5.80	5.76±0.08	1.30
Debittered juice by prepared β-CD polymer	5.94	5.67	5.80	5.80±0.14	2.33

Note: *NS= not significantly different at $P \leq 0.05$ by Duncan's multiple range test with three replications.

Table A 5 Operating cost for β-CD polymer fluidized column (Laboratory scale)

Items	Unit	Quantity	Cost/unit (Bahts/unit)	Cost (Bahts)	(%)
Tangerine fruit	kg	10	7	70	1.56
β-CD polymer	G	11	400	4,400	98.36
NaOH	G	12	0.26	3.12	0.07
Electricity for pump	kw*h	0.004	4	0.016	0.0004
Water	m ³	0.02	10	0.2	0.0045

Total cost (Bahts/column) = 4,473.336

Practical maximum capacity of column (mg limonin/g β-CD polymer) = 0.47

Productivity (ml juice/column/hour) = 6,000

Appendix B

1. Calculation of limonin content in tangerine juice

Standard limonin equation : $Y = 9.414 X$

Slope (S) = 9.414

When Y represents to peak area (mA_U x s) and

X represents to limonin concentration (ppm)

From HPLC analysis

Peak area of the limonin peak at 16 min = P

Thus limonin concentration of the extract = P/S ppm or $\mu\text{g}/\text{ml}$

The 1.5 ml of extract was prepared from 6 ml of sample juice

The efficiency of HyperSEP C18 column = 80%

Limonin content in the 6 ml sample juice = $1.5 \times (P/S) \times 100/80 \quad \mu\text{g}$

= $1.875(P/S) \quad \mu\text{g}$

Finally, the limonin concentration in the sample juice = $1.875(P/S) \quad \mu\text{g}/6 \text{ ml}$

= $0.3215(P/S) \quad \text{ppm}$

2. Calculation of size of fluidized column

The equation of minimum fluidization velocity, for small particles and $Re < 50$:

$$U_{mf} = \frac{688 d_p^{1.82} (\rho_s - \rho_f)^{0.94}}{\mu^{0.88}}$$

where $d_p = 0.015$ in, $\rho_s = 67.8$ lbm/ft³, $\rho_f = 64.74$ lbm/ft³, $\mu = 1.2$ centipoise

Thus, $U_{mf} = 0.805$ ft/hr

$= 0.409$ cm/min

The equation of cross-section area of column was

$$Q = U_{mf} \cdot A \quad \text{cm}^3/\text{min}$$

If $Q = 5$ cm³/min

Thus, $A = 12.22$ cm²

The diameter of column was

$$A = (\pi/4) D^2 \quad \text{cm}^2$$

Thus, $D = 3.94$ cm

The bed height was

$$U_{mf} = s/t \quad \text{cm/min}$$

If $t = 30$ min

$$s = 12.27 \quad \text{cm}$$

From this calculation, the fluidized column was approximately designed as the height of column was 50 cm and the diameter of column was between 3 to 5 cm.

Appendix C

Statistical Analysis

Table C.1 ANOVA for the study of regeneration of HyperSEP C18 column for limonin determination

SOURCE	DF	SUM OF SQUARE	MEAN SQUARE	F VALUE	Pr>F
Model	4	1078.9682	269.7420	180.48	0.0001
Error	10	14.9458	1.4946		
Corrected total	14	1093.9140			

Duncan's Multiple Range Test

Duncan Grouping	Mean	N	Between group
A	83.9500	3	1
B	80.2933	3	2
C	76.8933	3	3
D	63.9900	3	5
D			
D	63.3733	3	4

R-square 0.9863 C.V. 1.6588

Root MSE 1.2225 Data Mean 73.7000

Table C 2 ANOVA for the study of color value (L) in sample juice

SOURCE	DF	SUM OF SQUARE	MEAN SQUARE	F VALUE	Pr>F
Model	4	205.4938	51.3734	262.37	0.0001
Error	10	1.9581	0.1958		
Corrected total	14	207.4519			

Duncan's Multiple Range Test

Duncan Grouping	Mean	N	Between group
A	101.8000	3	1
B	92.8200	3	5
B	92.5000	3	4
B	92.4633	3	3
B	92.4300	3	2

R-square 0.9906 C.V. 0.4687

Root MSE 0.4425 Data Mean 94.4027

Table C 3 ANOVA for the study of color value (a) in sample juice

SOURCE	DF	SUM OF SQUARE	MEAN SQUARE	F VALUE	Pr>F
Model	4	54.0706	13.5176	199.26	0.0001
Error	10	0.6784	0.0678		
Corrected total	14	54.7490			

Duncan's Multiple Range Test

Duncan Grouping	Mean	N	Between group
A	-6.1800	3	2
A			
A	-6.2633	3	5
A			
A	-6.3267	3	4
A			
A	-6.3900	3	3
B	-11.0333	3	1

R-square 0.9876 C.V. 3.5982

Root MSE 0.2605 Data Mean 7.2387

Table C 4 ANOVA for the study of color value (b) in sample juice

SOURCE	DF	SUM OF SQUARE	MEAN SQUARE	F VALUE	Pr>F
Model	4	1165.9718	291.4930	1039.68	0.0001
Error	10	2.8037	0.2804		
Corrected total	14	1168.7755			

Duncan's Multiple Range Test

Duncan Grouping	Mean	N	Between group
A	32.3300	3	1
B	10.5600	3	3
B	10.2767	3	2
B	10.2000	3	4
B	10.1300	3	5

R-square 0.9976 C.V. 3.6021

Root MSE 0.5295 Data Mean 14.6993

Table C 5 ANOVA for the study of total soluble solids in sample juice

SOURCE	DF	SUM OF SQUARE	MEAN SQUARE	F VALUE	Pr>F
Model	4	0.0227	0.0057	1.21	0.3639
Error	10	0.0467	0.0047		
Corrected total	14	0.0693			

Duncan's Multiple Range Test

Duncan Grouping	Mean	N	Between group
A	11.6333	3	1
A			
A	11.6333	3	4
A			
A	11.6000	3	5
A			
A	11.5667	3	2
A			
A	11.5333	3	3

R-square 0.3269 C.V. 0.5892

Root MSE 0.0683 Data Mean 11.5933

Table C 6 ANOVA for the study of vitamin C content in sample juice

SOURCE	DF	SUM OF SQUARE	MEAN SQUARE	F VALUE	Pr>F
Model	3	0.0535	0.0178	1.98	0.1957
Error	8	0.0721	0.0090		
Corrected total	11	0.1256			

Duncan's Multiple Range Test

Duncan Grouping	Mean	N	Between group
A	5.8933	3	2
A			
A	5.8033	3	4
A			
A	5.7567	3	3
A			
A	5.7133	3	1

R-square 0.4261 C.V. 1.6388

Root MSE 0.0949 Data Mean 5.7917

Appendix D

1. Capacity of β -CD polymer column in limonin adsorption at maximum juice volume

Productivity of the laboratory scale

At maximum load, it produced 900 ml of debittered juice

$$\begin{aligned}
 \text{Flow rate} &= 100 \text{ ml/minute} \\
 \text{Process time for 900 ml juice} &= 9 \text{ minutes or } 0.15 \text{ hours / column} \\
 \text{Productivity of laboratory scale (P)} &= 900 \text{ ml debittered juice / column / } 0.15 \text{ hour} \\
 &= 6000 \text{ ml juice/column/hour}
 \end{aligned}$$

Limonin adsorption capacity of the laboratory scale

When used 15 g β -CD polymer

$$\begin{aligned}
 &= (11.76-6) (\mu\text{g/ml juice}) * 900 \text{ ml juice} / 15 \text{ g } \beta\text{-CD polymer} \\
 &= 345.6 \mu\text{g limonin/g } \beta\text{-CD polymer} \\
 &\approx 0.35 \text{ mg limonin/g } \beta\text{-CD polymer}
 \end{aligned}$$

When used 15 g regenerated β -CD polymer

$$\begin{aligned}
 &= (11.89-6) (\mu\text{g/ml juice}) * 350 \text{ ml juice} / 15 \text{ g } \beta\text{-CD polymer} \\
 &= 137.4 \mu\text{g limonin/g } \beta\text{-CD polymer} \\
 &\approx 0.14 \text{ mg limonin/g } \beta\text{-CD polymer}
 \end{aligned}$$

When used 11 g β -CD polymer

$$\begin{aligned}
 &= (11.76-6) (\mu\text{g/ml juice}) * 900 \text{ ml juice} / 11 \text{ g } \beta\text{-CD polymer} \\
 &= 471.3 \mu\text{g limonin/g } \beta\text{-CD polymer} \\
 &\approx 0.47 \text{ mg limonin/g } \beta\text{-CD polymer}
 \end{aligned}$$

2. Capacity of XAD-16 in limonin adsorption at the optimum condition

Initial limonin content in juice load = 12.07 ppm

Juice load = 1600 ml

Amount of XAD-16 resin used = 11 g

Total limonin in 100 ml juice = 19312 µg

% limonin reduction ~ 90%

Capacity of XAD-16 resin in limonin reduction = $(19312 \times 0.9)/11$

= 1580.07 µg limonin/ g XAD-16 resin

= 1.58 mg limonin/ g XAD-16 resin

3. Capacity of prepared β-CD polymer in limonin adsorption at the maximum juice volume

Initial limonin content in juice load = 12.15 ppm

Maximum juice volume = 25 ml

Amount of prepared β-CD polymer used = 11 g

Total limonin in 100 ml juice = 303.75 µg

Capacity of β-CD polymer in limonin reduction = $303.75/11$

= 27.614 µg limonin/ g β-CD polymer

= 0.028 mg limonin/ g β-CD polymer



BIOGRAPHY

Miss Tawisa Pipatthitikorn was born on May 11, 1982 in Nakhonpathom, Thailand. She graduated her Bachelor degree of Biochemistry, Faculty of Science from Chulalongkorn University in 2001 and then enrolled in the graduate program for Master of Science in Biotechnology at Chulalongkorn University in 2002. She has publication:

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