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APPENDIX

APPENDIX

I Ascorbic Acid (Vitamin C) Chemistry and Biochemistry

Ascorbic acid ($C_6H_8O_6$) is a water soluble, hexonic sugar acid, with a molecular weight of 176.13. Ascorbic acid is synthesized from glucose in most animals.

Vitamin C refers to all compounds that exhibit the biological activity of ascorbic acid, including both ascorbic acid and its oxidized form, dehydroascorbic acid. Removal of one electron from ascorbic acid yields semidehydroascorbic acid (ascorbate radical). This form of the vitamin is a free radical, it contains an unpaired electron. The removal of a second electron yields dehydroascorbic acid. Semidehydroascorbate is an intermediate in this conversion pathway. Dehydroascorbate reductase requires glutathione (GSH) as a source of reducing power. Both ascorbic acid and dehydroascorbic acid have biological activity. The latter compound may break down to form diketogulonic acid. Diketogulonic acid has no biological activity (Levine M, 1986).

II. Plasma ascorbic acid determination

In the present study, a specific enzymatic spectrophotometric method for ascorbic acid in plasma was used. Samples were analyzed indirectly by measuring the absorbance at 593 nm of a reaction product, a complex of ferrous ion and 2,4,6-tris (2-pyridyl)-s-triazine (Fe^{2+} -TPTZ). Ascorbic acid is specifically quantified by pretreating one of a pair of replicate samples

with ascorbate oxidase, to oxidize the ascorbic acid , then reacting both samples with Fe^{3+} -TPTZ and measuring the difference between the absorbances at 593 nm of the treated and untreated samples (Liu TZ et al., 1982).

Table 14. Procedure for Enzymatic-assisted plasma ascorbic acid determination.

Material	AA oxidase	AA oxidase
	Treated tube	Free tube
Unknown plasma or (standard and controls (ml)	0.1	0.1
AA oxidase working solution (ml)	0.05	-----
H ₂ O (ml)	-----	0.05
Vertex-mix gently and incubate at 37°C in a waterbath for 15 minutes		
Acetate buffer, pH 3.6 (ml)	1.25	1.25
TPTZ Solution (ml)	0.15	0.15
FeCl ₃ -6H ₂ O Solution (ml)	0.1	0.1
Gently mix the contents of every tube after additions. Allow to stand at room temperature for exactly 5 min. Measure absorbance at 593 nm.		

Plasma ascorbic acid concentration can obtained from a standard curve (LIU TZ et al., 1982).

III. Streptozotocin-Induced Diabetic Rats Model

The animal model of insulin-dependent diabetes mellitus (IDDM) that is used in this study is induced by a single intravenous injection with the dose of 55 mg/kg BW streptozotocin (STZ) (Jariyapongskul A et al., 1996).

In this study, we used STZ-treated rat model as an insulin-dependent diabetes mellitus, because this model closely resemble to IDDM in human. The dose of 55 mg/Kg/BW STZ is used by a single intravenous injection. With this dose, the rats become hyperglycemia and hypoinsulinemia within 24-48 hours after induction.

Streptozotocin induce beta-cell damage by initiating biochemical events which cause DNA strand breaks . STZ is able to cause beta-cell specific damage via its ability to interact with the glucose sensing mechanism of the beta-cells and cause beta-cells specific damage (Like AA, 1976)

IV. Fluorescence Microscopic Technique

Intravital fluorescence microscopy has been used to observe the microvascular of parenchymatous organ, such as brain, heart, lung, liver, pancreas, gut, kidney.

Microscopic viewing may be performed by trans-illumination or by epi-illumination using a modified Leitz Orthoplan microscope (75 W, XBO,

xenon lamp or 100 W, HBO, mercury lamp which attached to a Ploemo-Pak illuminator with different filter blocks for epi-illumination).

In this study the microscopic pictures are recorded by means of low-light level SIT (Silicon-intensified target) video camera-assisted image analysis system . The availability of different fluorescent markers for in vivo study are fluorescein isothiocyanate (FITC)-labeled Red blood cells (RBCs) and Rhodamine-B isothiocyanate (RITC)-labeled dextran, and their fluorescence emissions are detected separately by using two sets of filters (FITC: $\lambda_{em}=520$ nm; RITC: $\lambda_{em}=595$ nm). FITC-labeled RBCs flowing in single microveels used to measure the RBC velocity in arterioles, venules and capillaries while fluorescence images of the RITC-labeld dextran dissolved in plasma used to measure the diameter of various microvessels.



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