การโคลนและลักษณะการแสดงออกที่แตกต่างของยืนในเชื้อ Pythium insidiosum สายพันธุ์ประเทศไทย



นางสาวพัชรี กัมมารเจษฎากุล

วิทยานิพนธ์นี้เป็นส่วนหนึ่งของการศึกษาตามหลักสูตรปริญญาวิทยาศาสตรดุษฎีบัณฑิต สาขาวิชาจุลชีวจิทยาทางการแพทย์ (สหสาขาวิชา) บัณฑิตวิทยาลัย จุฬาลงกรณ์มหาวิทยาลัย ปีการศึกษา 2552 ลิขสิทธิ์ของจุฬาลงกรณ์มหาวิทยาลัย

CLONING AND CHARACTERIZATION OF GENES DIFFERENTIALLY EXPRESSED IN *PYTHIUM INSIDIOSUM* THAI STRAIN

Miss Patcharee Kammarnjassadakul

A Dissertation Submitted in Partial Fulfillment of the Requirements for the Degree of Doctor of Philosophy Program in Medical Microbiology (Interdisciplinary Program)

Graduate School
Chulalongkorn University
Academic Year 2009
Copyright of Chulalongkorn University

Thesis	CLONING AND CHARACTERIZATION OF GENES	
	DIFFERENTIALLY EXPRESSED IN PYTHIUM INSIDIOSUM	
	THAI STRAIN	
Ву	Miss Patcharee Kammarnjassadakul	
Field of Study	Medical Microbiology	
•	. ,	
Thesis Advisor	Associate Professor Ariya Chindamporn, Ph.D.	
Thesis Co-advisor	Assistant Professor Tanapat Palaga, Ph.D.	
Thesis Co-advisor	Kallaya Sritunyalucksana, Ph.D.	
	e Graduate School, Chulalongkorn University in Partial Fulfillment for the Doctoral Degree	
Dean of the Graduate School		
(Associate Professor Pornpote Piumsomboon, Ph.D.)		
THESIS COMMITTEE		
anon Clythalmo Chairman		
(Assistant Professor Anan Chongthaleong, MD.)		
6	Duiya Chindpun Thesis Advisor	
(Asso	ciate Professor Ariya Chindamporn, Ph.D.)	
	Thesis Co-Advisor	
	stant Professor Tanapat Palaga, Ph.D.)	

(Kallaya Sritunyalucksana, Ph.D.)

Arjanny Tanhinaranh External Examiner

Uallaya (b) Thesis Co-Advisor

(Associate Professor Srisurang Tantimavanich, Ph.D.)

พัชรี กัมมารเจษฎากุล : การโคลนและลักษณะการแสดงออกที่แตกต่างของยืนในเชื้อ Pythium insidiosum สายพันธุ์ประเทศไทย (CLONING AND CHARACTERIZATION OF GENES DIFFERENTIALLY EXPRESSED IN PYTHIUM INSIDIOSUM THAI STRAIN) อ.ที่ปรึกษาวิทยานิพนธ์หลัก: รศ.ดร. อริยา จินคามพร, อ.ที่ปรึกษาวิทยานิพนธ์ ร่วม : ผศ.ดร.ธนาภัทร ปาลกะ และ อ.ดร.กัลยาณ์ ศรีธัญญูลักษณา – แดงตั๊บ, 235 หน้า

Pythium insidiosum เป็นเชื้ออุบัติใหม่อาศัยอยู่ในน้ำมีลักษณะคล้ายเชื้อรา เป็นสาเหตุของโรค pythiosis พบในคนและสัตว์ บริเวณเขตร้อนจนถึงร้อนชื้น มีรายงานพบผู้ป่วย และเสียชีวิตจากโรคนี้สูงที่สุดจาก ประเทศไทย อย่างไรก็ตามข้อมูลเกี่ยวกับตัวเชื้อทางด้านพันธุกรรมและพยาธิสภาพการเกิดโรคมีน้อยมาก งานวิจัยในครั้งนี้จึงมีวัตถุประสงค์ในการหายืนจากเชื้อที่เจริญ ณ อุณหภูมิ 37°C น่าจะมีความสัมพันธ์กับการเกิด พยาธิสภาพของโรค การค้นหายืนดังกล่าวอาศัยวิธี PCR - select cDNA subtraction จากเชื้อที่เพาะ ณ อุณหภูมิ 27°C และ 37°C และจากข้อมูลด้านพันธุกรรมของเชื้อก่อโรคที่คล้ายคลึงจากฐานข้อมูลใน GenBank จากวิธี subtraction พบ 606 โคลนจาก subtracted cDNA libraries ของทั้งสองอุณหภูมิ เมื่อนำไปวิเคราะห์ลำคับเบสค้วย โปรแกรม BLASTN พบโคลนส่วนใหญ่มีลำคับเบสคล้ายกับ mitochondria ของกลุ่มราน้ำ และการวิเคราะห์การ ทำงานของโปรตีนด้วยโปรแกรม Swiss-Prot และ BLASTX พบ candidate genes ที่ควบคุมการทำงานของ โปรตีนหลายชนิด ในที่นี้ผู้วิจัยได้ทำการศึกษาการแสดงออกของยืนที่ควบคุมการทำงานของ cytochrome c oxidase subunit II (COX II) β-tubulin (TUB) และ chitin synthase subunit II (CHI II) โดยอาศัยวิธี semi – quantitative PCR และขึ้นยันผลการศึกษาด้วยวิธี real time RT - PCR ในเชื้อจำนวน 16 สาขพันธุ์ ที่เป็นตัวแทน ของกลุ่มเชื้อที่มาจากแหล่งต่างกันที่ได้จากการจัด phylogenetic distribution ผลการศึกษาพบยืน COX II เท่านั้นที่ มีการแสคงออก ณ อุณหภูมิ 37°C สูงกว่า 27°C ถึง 2.5 เท่าโคยเฉลี่ย จัดว่ามีความแตกต่างอย่างมีนัยสำคัญ (p =0.0347) และ ไม่พบการแสดงออกของยืน TUB และ CHI II ที่แตกต่างกันระหว่างสองอุณหภูมิ จากข้อมูลคั้งกล่าว นี้ชี้ให้เห็นว่าสามารถนำยืน TUB และ CHI II เป็น housekeeping gene ในการศึกษาการแสดงออกของยืนที่ไวค่อ อุณหภูมิที่เปลี่ยนแปลงได้ สำหรับการแสดงออกของยืน *CHI* II ที่37°C พบว่าขึ้นกับแหล่งที่มาของเชื้อ นอกจากนี้ ทางผู้วิจัยยังได้นำ COX II มาประยุกต์ใช้ในการศึกษา Phylogenetic relationship พบว่า COX II สามารถใช้ศึกษา Phylogenetic distribution ของเชื้อ P. insidiosum ได้นอกเหนือ ITS ที่มักนิยมใช้ โดยสรุปผลการศึกษานี้เป็น รายงานแรกที่แสดงว่า การแสดงออกของ COX II ที่ อุณหภูมิ 37°C สูงกว่า 27°C สำหรับความสัมพันธ์ระหว่างยืน COX II กับการก่อให้เกิดพยาธิสภาพ จำเป็นต้องศึกษาเพิ่มเติม

สาขาวิชา	จุลชีววิทยาทางการแพทย์	ลายมือชื่อนิสิต ฟังว์ กัพพา	บฎากุล	
ปีการศึกษา	2552	ลายมือชื่อ อ.ที่ปรึกษาวิทยานิพนธ์หลัก	@30g	Oronsus
		ลายมือชื่อ อ.ที่ปรึกษาวิทยานิพนธ์ร่วม	anni	1/0-
		ลายมือชื่อ อ.ที่ปรึกษาวิทยานิพนธ์ร่วม	moderati	100000

4889665020: MAJOR MEDICAL MICROBIOLOGY
KEYWORDS: *PYTHIUM INSIDIOSUM* / SUPPRESSION SUBTRACTIVE
HYBRIDIZATION/ REAL TIME RT – PCR/ SUBTRACTED CDNA LIBRARIES /
PHYLOGENETIC TREE

PATCHAREE KAMMARNJASSADAKUL: CLONING AND CHARACTERIZATION OF GENES DIFFERENTIALLY EXPRESSED IN *PYTHIUM INSIDIOSUM* THAI STRAIN. THESIS ADVISOR: ASSOC. PROF. ARIYA CHINDAMPORN, Ph.D., THESIS CO-ADVISOR: ASST.PROF.TANAPAT PALAGA, Ph.D., KALLAYA SRITUNYALUCKSANA, Ph.D., 235 pp.

An emerging aquatic fungal like organism, Pythium insidiosum, causes the disease called 'pythiosis' in both human and animal from tropical, subtropical, and temperate zone. The highest mortality and morbidity in human pythiosis has been reported from Thailand. Very few genetic information and pathogenesis concerning this organism is not known. This study was to investigate genes expressed from culture at 37°C might involved in its pathogenesis. To isolate temperature-induced genes, PCR – select cDNA subtraction from cultures P. insidiosum at 27°C and 37°C and PCR-based searching the genetic information in GenBank Databases. Both subtracted cDNA libraries conditions, total six hundred and six clones with inserted cDNA fragments were detected. Nucleotide analysis by BLASTN showed mostly mitochondria DNA whereas the protein function predicted by Swiss-Prot and BLASTX programs demonstrated candidate genes encoding proteins. Here, gene encoding cytochrome c oxidase subunit II (COX II), β-tubulin (TUB), and chitin synthase subunit II (CHI II) were selected to analyze their expression using semiquantitative PCR and confirmed by Real time RT-PCR. To demonstrate the expression level at 37°C and 27°C conditions, sixteen strains of P. insidiosum, represented three phylogeographic preference were recruited. The results showed that at 37°C growth condition, only COX II gene expressed 2.5 times over 27°C growth condition (p = 0.0347). Even though, TUB and CHI II show indistinguishable expression, these genes can be used as housekeeping genes for temperature susceptible gene expression studies. This is the new findings. However, it is of noted that the expression of CHI II at 37°C depended on the geographic distribution. Not only the expression was analyzed, the phylogenetic relationship was also performed using COX II. The result showed that COX II is the good candidate gene to reveal the genetic association among P. insidiosum. It also is an alternative gene of ITS which is commonly used for study the phylogenetic distribution. In conclusion, these data was the pioneer to demonstrate that COX II expressed at 37°C over 27°C. The insight of this gene and pathogenesis required to perform the further investigation.

Field of Study: Medical Microbiology Student's Signature. Palchan hammanjarsadahrt....

Co-Advisor's Signature Callaya S

ACKNOWLEDGEMENTS

First of all, I would like to express my feeling from the honesty to my great advisor: Assoc. Prof. Ariya Chindamporn, Ph.D. Throughout my candidate Ph.D lifetime, she gave wise advice, good care to support this thesis and other administrative challenges and the many constructive discussions. I am very to special thankful to my co– advisor: Assist. Prof. Tanapat Palaga, Ph.D. With his enthusiasm, his inspiration, and his great efforts to explain things clearly and simply, he provided a lot of suggestions from him improved my thesis and also my scientific thinking. Many thanks are address to my co–advisor to be loved: Kallaya Sritunyalucksana, Ph.D. for supervising this work and asking me to work in her research group at Centex Shrimp, Faculty of Science, Mahidol University, where I carried out a significant part of my thesis. With her talent and expertise for research, she helped me a lot with her great experience in Suppression Subtractive Hybridization System.

My special thanks go to the 90th Anniversary of Chulalongkorn University Fund (Ratchadaphiseksomphot Endowment Fund), Graduate School, Chulalongkorn University, the main financial support and I also thank Research Center, Faculty of Medicine, and Chulalongkorn University. I will forever be indebted to Prof. Dr. Boonsirm Withyachumnarnkul, MD., Ph.D. for the giving me the opportunity to use the excellence facilities at Centex Shrimp for more than three months. At Centex Shrimp, many people have supported me with word and deed. I am very special thankful to Prof. Nongnuch Vanittanakom, Ph.D., Faculty of Medicine, Chiang Mai University, Theerapong Krajaejun, MD., Ph.D. and Assoc. Prof. Srisurang Tantimavanich, Mahidol University for providing P. insidiosum references isolates and DNA samples. I am especially grateful to numerous people were involved and contributed in many ways in this thesis, especially Ms. Supranee Buranapraditkun, Ms. Sirada Koachareon, my sisters and brothers in Mycology research unit and all staffs of Department of Microbiology for providing a stimulating and fun environment in which to learn and grow. Last but not least and most importantly, I am extremely grateful to my parents, younger brother and my husband for all their love! and sharing their life with me. With their understanding, never lost patience with my impatience, they always raised and help me get through the difficult times; without them my work could not succeed.

CONTENTS

	Page
THAI ABSTRACT	iv
ENGLISH ABSTRACT	v
ACKNOWLEDGEMENTS	vi
CONTENTS	vii
LIST OF TABLES	xi
LIST OF FIGURES	xiii
LIST OF ABBREVIATIONS AND SYMBOLS	xvi
CHAPTER	
I INTRODUCTION	1
Hypothesis	7
Objectives	7
Conceptual frame work	8
II REVIEW LIERATURES	9
1. Background History	9
1.1 Mammals pathogen of <i>P. insidiosum</i>	11
2. Biology and Life Cycle	13
3. Hyphae ultrastructure	15
4. Life cycle	16
4.1 Sexual reproduction	16
4.2 Asexual reproduction	17
5. Epidemiology	21
6. Clinical pathogenesis	24
6.1 Clinical manifestation	25
6.1.1 Cutaneous / subcutaneous pythiosis	25
6.1.2 Vascular pythiosis	27
6.1.3 Ocular pythiosis	29
7. Laboratory diagnosis	29
7.1 Specimen collection	29
7.2 Culture	30

CONTENTS (continue)

	Page
CHAPTER	
7.3 Zoospore production	30
7.4 Histopathology	31
7.5 DNA – based diagnostic assays	32
7.6 Serological method	33
8. Treatment	34
9. Genomic data of <i>P. insidiosum</i>	37
10. Differentially Expressed Genes	43
10.1 Suppression Subtractive Hybridization (SSH)	46
10.2 Real – Time Polymerase chain reaction (RT – PCR)	53
10.2.1 SYBR® Green chemistry	57
10.2.2 Melting curve analysis	58
III MATERIALS AND METHODS	62
1. Microorganisms and Identification	62
1.1 Microorganisms	62
1.2 Identification	63
1.2.1 Zoospore production	63
1.2.2 DNA extraction	65
1.2.3 PCR amplification and sequencing for	
P. insidiosum identification	66
2. Temperature sensitive genes selection	67
2.1 Suppression subtractive hybridization	68
2.1.1 RNA extraction	68
2.1.2 Determination of RNA quality	70
2.1.3 mRNA purification.	71
2.1.4 Suppression subtractive hybridization(SSH)	75
2.1.5 The construction of SSH cDNA library	81
2.1.5.1 Competent cells preparation	81
2.1.5.2 pGEM® – T easy vector transformation	82

CONTENTS (continue)

	Page
CHAPTER	
2.1.5.3 Screening cDNA inserted by PCR	
amplification	83
2.1.5.4 Sequencing and data analysis	86
2.2 GenBank databases	87
3. Gene expression Analysis	87
3.1 Semi – Quantitative gene expression determination	
using PCR	87
3.1.1 First strand cDNA synthesis procedure	87
3.1.2 Polymerase Chain Reaction (PCR) Amplification	89
3.2 Quantification of gene expression determination	90
3.2.1 Real time reverse - transcriptase PCR (real time	
RT – PCR)	90
3.2.2 Data analysis	92
4. Genetic diversity	93
4.1 Strains and Genomic DNA	93
4.2 PCR amplification and sequencing	93
4.3 Phylogenetic analysis	94
IV RESULTS	97
1. Isolation and Identification of <i>P. insidiosum</i> Thai isolates	97
1.1 Zoospore production	97
1.2 Verification of P. insidiosum Thai Strain by DNA	
sequencing in ITS region	98
2. Temperature sensitive genes selection	99
2.1 cDNA 37°C library by SSH technique	99
2.1.1 RNA qualification	99
2.1.2 Identification of subtraction efficiency	102
2.1.3 Inserted clones from Subtracted cDNA Libraries	
at 37°C and 27°C conditions	103

CONTENTS (continue)

	Page
CHAPTER	
2.1.4 Sequence Analysis	104
2.2 GenBank Databases	117
3. Differential expression screening	120
3.1 Validation of reference gene for reference control in	
gene expression, semi – quantitative reverse	
transcriptase (RT) – PCR procedure	120
3.2 Semi – quantitative PCR amplification	121
3.3 Temporal expression of candidate genes by Real time	
reverse transcriptase - PCR (Real time RT – PCR)	124
3.3.1 Identification and validation of endogenous	
control gene	129
3.3.2 Quantitative analysis of COX II, TUB and CHI II	
gene expression using Real time RT – PCR on	
individual samples	132
3.3.3 Analysis of the relationship between COX II,	
TUB and CHI II gene expression and 3	2
phylogeographic regions of P. insidiosum	137
4. Phylogenetic relationship of 33 strains <i>P. insidiosum</i>	139
4.1 Phylogenetic tree based on COX II gene	139
4.2 Phylogenetic tree based on ITS gene	140
V DISCUSSION AND CONCLUSION	146
REFERENCES	156
APPENDICES	189
Appendix A	190
Appendix B	195
Appendix C	206
VITAE	235

LIST OF TABLES

Tab	ole	Page
1.	Important different between the Oomycetes and Fungi	15
2.	Summary of clinical information for human patients with pythiosis	
	diagnosed at 9 tertiary care hospitals across Thailand	23
3.	Oomycete genome projects and their associated web resources	39
4.	Which model system for the oomycetes	42
5.	Comparison of advantage and disadvantage of gene expression profile	46
6.	Dyes available for use in Real - time PCR	59
7.	Hosts/sources and geographical origin where P. insidiosum were	
	isolated	64
8.	The oligonucleotides used for SSH method	79
9.	Oligonucleotides used for checking individual fragments insertion	84
10.	Primers use for RT – PCR and DNA sequencing	91
11.	Real Time RT - PCR condition	92
12.	List of 33 isolates of P. insidiosum and other Pythium species used in	
	phylogenetic study	95
13.	Quality and quantity of total RNA by Hot phenol extraction from 16	
	P. insidiosum isolates	101
14.	Summary sequence analysis of subtracted cDNA libraries at 37°C and	
	27°C condition using BLAST & ExPASy – PROSITE programs	105
15.	List of the nucleotide similarities of SSH cDNA libraries sequences	
	from P. insidiosum strain PC7 at 27°C & 37°C cultivation conditions	107
16.	SSH cDNA libraries (27°C & 37°C conditions) with significant protein	
	matches in The National Center Biotechnology Information (NCBI)	
	protein databases	113
17.	GenBank accession numbers of sixteen isolates <i>P. insidiosum</i> isolates	126
18.	The average melting temperature of each genes with genes specific	
	primer pair	129

LIST OF TABLES (continue)

Tab	Table Table	
19.	Data analysis and validation of endogenous control (rRNA gene) from	
	16 strains of P. insidiosum growth temperature at 27°C and 37°C	
	condition	131
20.	Summary data of cycle threshold (C_T) from 16 strains P . insidiosum	
	growth at 27°C and 37°C conditions	133
21.	The fold change of the candidate temperature sensitive genes (COX II,	
	TUB and CHI II gene) relative to the endogenous control gene (rRNA)	
	at two different temperature	134
22.	The relationship analysis between different of original sources of	
	P. insidiosum isolates	139
23.	GenBank accession numbers of P. insidiosum isolates used in this	
	study	143
24.	Showing the cluster terminology obtained from the phylogenetic	
	analysis of three studies including the present study. Clin: clinical	
	source env. environmental source	154

LIST OF FIGURES

Fig	ure	Page
1.	Colony of P. insidiosum on SDA	13
2.	TEM micrograph of a transverse section of hyphae of <i>P. insidiosum</i>	16
3.	A developing sexual and asexual reproduction of Oomycota	18
4.	The life cycle of P. insidiosum	20
5.	Distribution of pythiosis in the tropical, subtropical and temperate	
	regions of the world	21
6.	Geographic distribution of patients with pythiosis in Thailand	24
7.	The pathogenesis and histopathology of cutaneous pythiosis	27
8.	The pathogenesis and histopathology of vascular pythiosis	28
9.	Clinical presentation of ocular pythiosis	29
10.	Colony morphology of P. insidiosum	30
11.	Direct examination from tissue biopsy from human vascular pythiosis	32
12.	Diagram representing the main features of the Central Dogma	38
13.	Relationship between incubation temperature and growth rate	41
14.	Overview of the PCR-Select procedure	49
15.	An illustration of the use of melting curve analysis to distinguish	
	between specific and non - specific PCR products	61
16.	Zoosporogenesis of P. insidiosum	65
17.	Scheme of the QuickPrep Micro mRNA Purification procedure	73
18.	Diagram of PCR – select subtraction technique	80
19.	pGEM® - T Vector Map and sequence reference points	83
20.	Drawing of HiYield™ Plasmid Mini Kit protocol	85
21.	The cDNA synthesis procedures	89
22.	Morphology of P. insidiosum clinical isolate	98
23.	Denature agarose gel electrophoresis of total RNA from P. insidiosum	
	strain PC7 which was cultured at 27°C and 37°C	100
24.	Efficiency of cDNA subtraction using specific COX II primers in PCR	
	amplification	103

LIST OF FIGURES (continue)

Fig	ure	Page
25.	PCR products of some positive clones after performing the colony	
	PCR using M13pUC primers	104
26.	Nucleotides analysis of subtracted 150 up-regulated cDNA clones of	
	P. insidiosum strain PC7 at 27°C cultivation condition with global	
	distribution of BLASTN	108
27.	Nucleotides analysis of subtracted 456 up-regulated cDNA clones of	
	P. insidiosum strain PC7 at 37°C cultivation condition with global	
	distribution of BLASTN	109
28.	Translational analysis of subtracted cDNA library of P. insidiosum	
	strain PC7 at 37°C condition	111
29.	Translational analysis of subtracted cDNA library of P. insidiosum	
	strain PC7 at 27°C condition	112
30.	The functional percentage distribution of the genes up-regulated in	
	P. insidiosum strain PC7 was grown at 27°C condition (150 genes in	
	total)	118
31.	The functional distribution percentage of the genes up- regulated in	
	P. insidiosum strain PC7 was grown at 37°C condition (456 genes in	
	total)	119
32.	A: Annealing site for the ITS1 and ITS4 primers and PCR	
	product	120
33.	Comparing PCR product size between cDNA and genomic DNA	
	(gDNA) P. insidiosum template by agarose gel electrophoresis	122
34.	The semi – quantitative PCR of COX II and rRNA at 30 amplification	
	cycles using agarose gel electrophoresis analysis	123
35.	The semi – quantitative PCR of TUB gene and rRNA at 35	
	amplification cycles using agarose gel electrophoresis analysis	124
36.	Real time RT – PCR products of 3 candidate genes were checked	
Ų,	using 1.5% agarose gel electrophoresis	127

LIST OF FIGURES (continue)

Fig	Figure	
37.	Melting peak analysis of each gene amplification using genes specific	
	primer by real time RT – PCR	128
38.	The amplification curve of rRNA	130
39.	Data analysis of the expression of rRNA (endogenous control) under	
	different temperature (27°C & 37°C) conditions	132
4 0.	The relative expression of gene encoded : cytochrome oxidase II, $\boldsymbol{\beta}$ -	
	tubulin, and chitin synthase II (COX II (A), TUB (B) and ,CHI II (C)	
	gene, respectively) at 27°C condition and 37°C condition	136
41.	The relationship analysis gene expression of 3 candidate genes of	
	P. insidiosum between in 3 clades at 27°C &37°C conditions	141
42.	Neighbor Joining (NJ) tree based on cytochrome oxidase II (COXII)	144
43.	Neighbor Joining (NJ) tree based on rDNA ITS region	145

LIST OF ABBREVIATIONS AND SYMBOLS

% = Percent

°C = Degree Celsius

 $\times g$ = g - force

& An ampersand is a logogram representing the

conjunction word "and".

 β = Beta

 Δ = Delta

 μg = Microgram (s) = 10^{-6} gram

 μl = Microliter (s) = 10^{-6} liter

 μM = Micromolar

A = Absorbance Spectrum

Ag = Antigen

BLAST = Basic Local Alignment Search Tool

BLASTN = Nucleotide – nucleotide BALST

BLASTX = DNA - protein BLAST

bp = Base pair

cm. = Centimeter

cDNA = Complementary DNA

Cat. = Catalog number

CA = Canada

CHI II = Chitin synthase subunit II gene

COX II = Cytochrome c oxidase subunit II gene

CMA = Cornmeal Agar

 C_T = Cycles threshold

CV = Coefficient of variation

dscDNA = Double strand cDNA

dNTPs = Deoxynucleoside 5'- triphosphates

DEPC = Diethyl pyrocarbonate

DNA = Deoxyribose nucleic acid

DNase = Deoxyribonuclease

DW = Distilled water

et al. = And others

E – value = Expected value

EDTA = Ethylenediaminetetraacetate

EtBr = Ethidium bromide

EtOH = Ethanol

FA gel = Formaldehyde agraose gel

g = Gram(s)

hr = Hour(s)

 H_2O = Water

 H_2O_2 = Hydrogen peroxide

Inc. = Incorporate

ITS region = Internal transcribed spacer region

IGS = Intergenic spacer

kb = Kilobases

KOH = Potassium hydroxide

KCl = Potassium chloride

IPTG = Isopropyl- β -D-thiogalactopyranoside

L = Liter

L. = Lagenidium

LB = Luria-Bertani media

min = Minute (s)

mRNA = Messenger RNA

mg = Milligram(s)

ml = Milliliter (s)

mm = Millimeter

mM = Millimolar

mtDNA = Mitochondria DNA

M = Molar

MOP = 3-[N-Morpholino] propanesulfonic acid]

MEGA = Molecular evolution genetics analysis

MIP = Munich Information Center for Protein

Sequences

MgCl₂ = Magnesium chloride

ng = Nanogram (s) = 10^9 gram

nm = Nanometer (s) = 10^9 meter

no. = Number

NaOAC = Sodium acetate

NaOH = Sodium hydroxide

NCBI = National Center for Biological Information

NJ = Neighbor-joining

nd = The second

OD = Optical density

pg = Pico gram (s)

pmol = Picomole

pH = Potential of hydrogen ion

PCR = Polymerase Chain Reaction

PDA = Potato dextrose agar

P – value = Probability value

P. = Pythium

Ph. = Phytophthora

rpm. = Revolution per minute

rRNA = Ribosomal RNA

rDNA = Ribosomal DNA

RNA = Ribonucleic acid

RT = Room temperature

RNase = Ribonuclease

RT - PCR = Reverse Transcriptase - PCR

s = Second(s)

spp = Specie (s)

st = The first

S = Sevedberg unit

SEM = Standard error

SD = Standard deviation

SDA = Sabouraud dextrose agar

SDB = Sabouraud dextrose broth

SSH = Suppression Subtractive Hybridization

SSKI = Supersaturated potassium iodide

TBE = Tris – borate EDTA

TE = Tris - EDTA (buffer)

Tris = Tris (hydroxymethyl) amino – methane

Taq = Thermus aquaticus

Tm = Melting temperatures

 $TUB = \beta - \text{tubulin gene}$

U = Unit (s) of enzyme

USA = United State of America

UK = United Kingdom

UV = Ultraviolet

V = Volts

V/cm = Volts per centimeter

wt/wt = Weight/weight

w/v = Weight/volume

X -gal = 5-bromo-4-chloro-3-indolyl- β -D-

galactopyranoside